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UNNATURAL β -L-ENANTIOMERS OF NUCLEOSIDE ANALOGUES AS POTENT ANTI-HEPATITIS B VIRUS AGENTS

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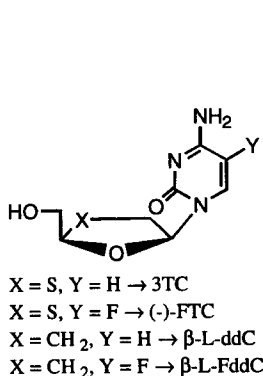
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Abstract: Several 2'- or 3'- substituted 2',3'-dideoxy- β -L-nucleosides bearing adenine as the base were stereospecifically synthesized and their antiviral properties examined. Two of them, namely 2'-azido- and 3'-azido-2',3'-dideoxy- β -L-adenosine (2'-N₃- β -L-ddA and 3'-N₃- β -L-ddA) were found to have some anti-hepatitis B virus (HBV) activity in cell culture.

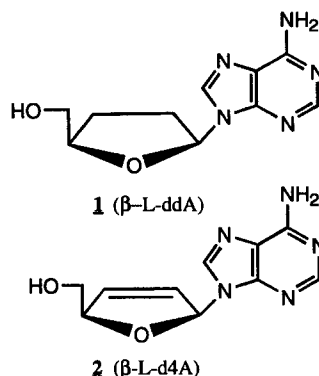
INTRODUCTION

During the last decade there has been some interest in the synthesis and biological evaluation of L-nucleoside analogues, although the activities of most nucleoside analogues had been already associated with the natural D-enantiomers. However, more recently several 2',3'-dideoxy pyrimidine- β -L-nucleoside analogues have been synthesized. Among them, those bearing a cytosine base (3TC, β -L-5FddC, Fig. 1a) have been found to possess potent activity against both human immunodeficiency virus (HIV) and hepatitis B virus (HBV).^{1,2}

As a part of our ongoing research programme on β -L-sugar-modified nucleoside analogues,³⁻¹⁰ we have reported previously the stereospecific



- Figure 1a -



- Figure 1b -

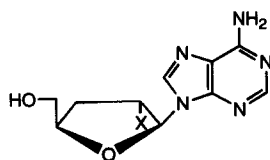
synthesis and the anti-HBV activity of 2',3'-dideoxy- $\beta\text{-L}$ -adenosine ($\beta\text{-L-ddA}$, **1**) and its 2',3'-didehydro derivative ($\beta\text{-L-d4A}$, **2**)¹⁰⁻¹² (Fig. 1b).

Here we described the synthesis of the 2'-fluoro (2'-F- $\beta\text{-L-ddA}$, **3**), 2'-azido (2'-N₃- $\beta\text{-L-ddA}$, **4**), 2'-amino (2'-NH₂- $\beta\text{-L-ddA}$, **5**), 3'-fluoro (3'-F- $\beta\text{-L-ddA}$, **6**), 3'-azido (3'-N₃- $\beta\text{-L-ddA}$, **7**), and 3'-amino (3'-NH₂- $\beta\text{-L-ddA}$, **8**) derivatives (Fig. 2) of $\beta\text{-L-ddA}$ in order to evaluate their antiviral properties in cell cultures.

2'-Substituted $\beta\text{-L-ddA}$ derivatives

From a synthetic view point, coupling of 1,2-di-*O*-acetyl-3-deoxy-5-*O*-benzoyl-L-*erythro*-pentofuranose⁹ with adenine, followed by deacetylation provided 9-(3-deoxy- $\beta\text{-L-erythro}$ -pentofuranosyl)adenine (Scheme 1). Selective bis 5'-*O* and ⁶*N*-tritylation followed by Mitsunobu¹³ inversion of the 2'-hydroxyl function and debenzoylation gave 9-(5-*O*-trityl-3-deoxy- $\beta\text{-L-threo}$ -pentofuranosyl)⁶*N*-trityladenine. Fluoration of this key intermediate was effected using (diethylamino)sulfur trifluoride (DAST) in methylene chloride. Detritylation in acidic conditions afforded the desired 2'-F- $\beta\text{-L-ddA}$ **3** (Scheme 1).

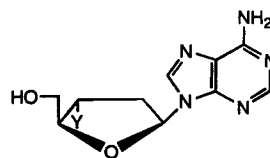
Azidation of 9-(5-*O*-trityl-3-deoxy- $\beta\text{-L-threo}$ -pentofuranosyl)⁶*N*-trityladenine was carried out following a modified Mitsunobu procedure^{13, 14} and



3, X = F (2'-F-β-L-ddA)

4, X = N₃ (2'-N₃-β-L-ddA)

5, X = NH₂ (2'-NH₂-β-L-ddA)

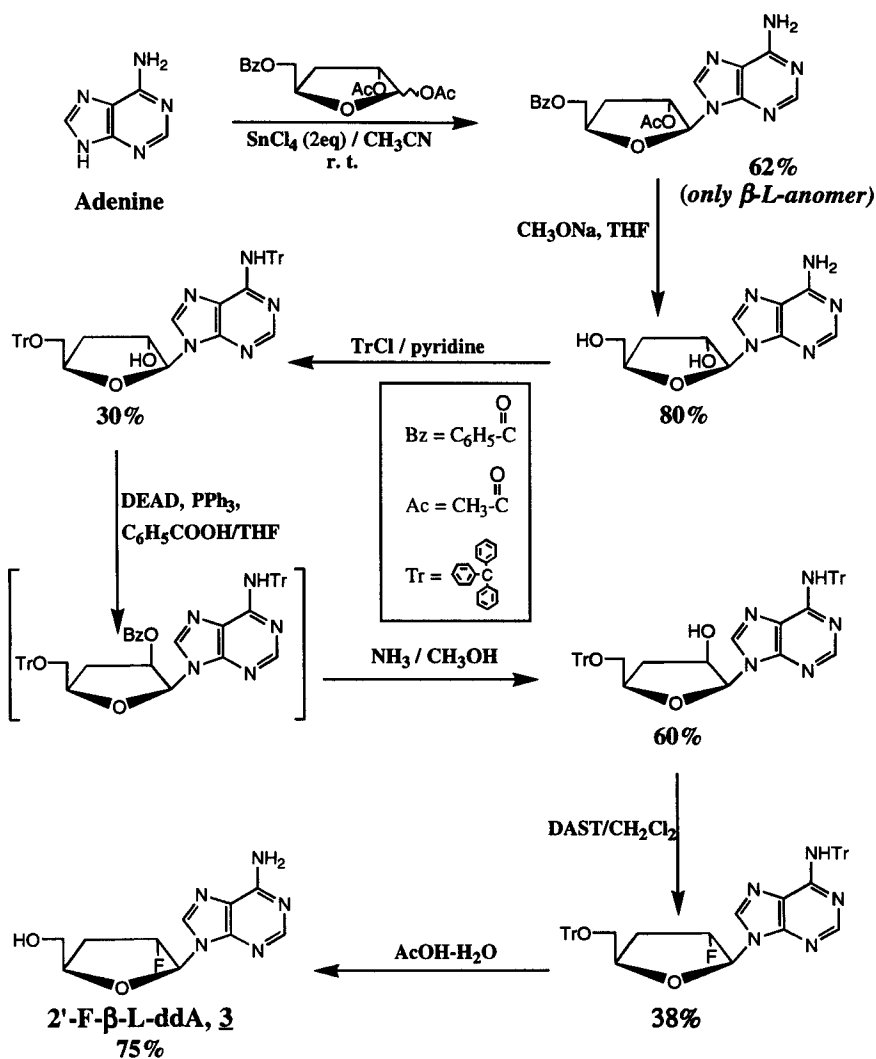


6, Y = F (3'-F-β-L-ddA)

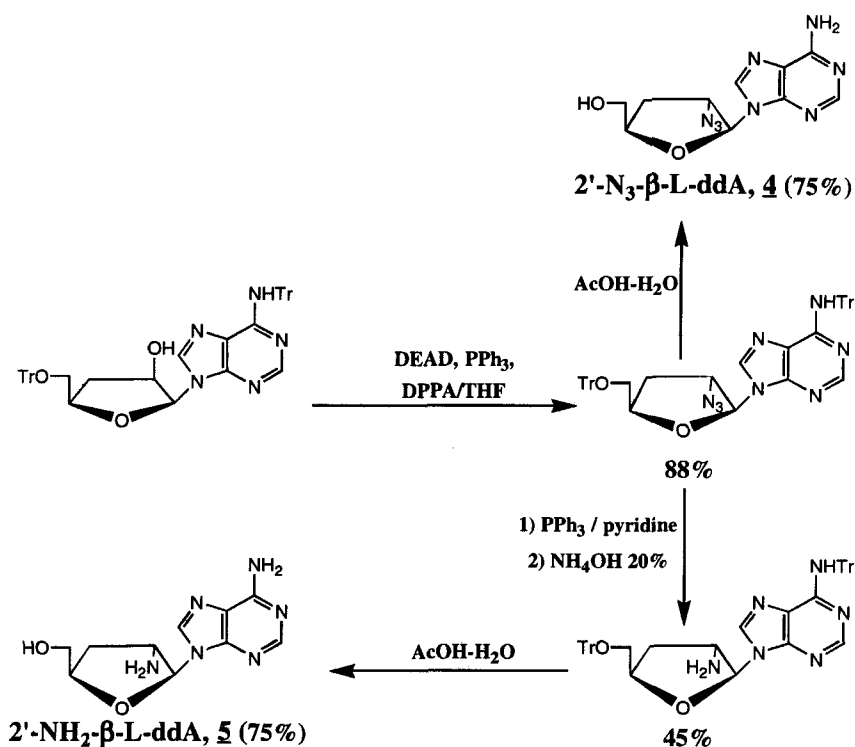
7, Y = N₃ (3'-N₃-β-L-ddA)

8, Y = NH₂ (3'-NH₂-β-L-ddA)

- Figure 2 -



- Scheme 1 -



- Scheme 2 -

afforded, after detritylation, 2'-N₃-β-L-ddA **4** (Scheme 2). The tritylated intermediate azido nucleoside was converted to the corresponding amino derivative 2'-NH₂-β-L-ddA **5** by treatment with triphenylphosphine in pyridine, followed by hydrolysis with concentrated ammonium hydroxide and detritylation, as reported previously in other series.¹⁵

3'-Substituted β-L-ddA derivatives

For the synthesis of these nucleoside analogues, the dissymmetric peracetylated 1,2-di-*O*-acetyl-3,5-di-*O*-benzoyl-L-xylofuranose³ was condensed with adenine to afford exclusively (in accord with Baker's rule¹⁶ owing to 2-*O*-acyl

participation during the condensation) the corresponding fully protected β -L-nucleoside anomer (Scheme 3).

This compound was selectively deacylated at its 2'-position, and then subjected to a deoxygenative hydrogenolysis. Successive 6N -monomethoxytritylation, full debenzoylation and selective 5'-*O*-monomethoxytritylation afforded 9-(2-deoxy- β -L-*threo*-pentofuranosyl) 6N -monomethoxytrityladenine. This intermediate was converted to 3'-fluoro- β -L-ddA **6**, 3'-azido- β -L-ddA **7** and 3'-amino- β -L-ddA **8** following similar procedures as used during the synthesis of the corresponding 2'-substituted derivatives of β -L-ddA.

Antiviral activities

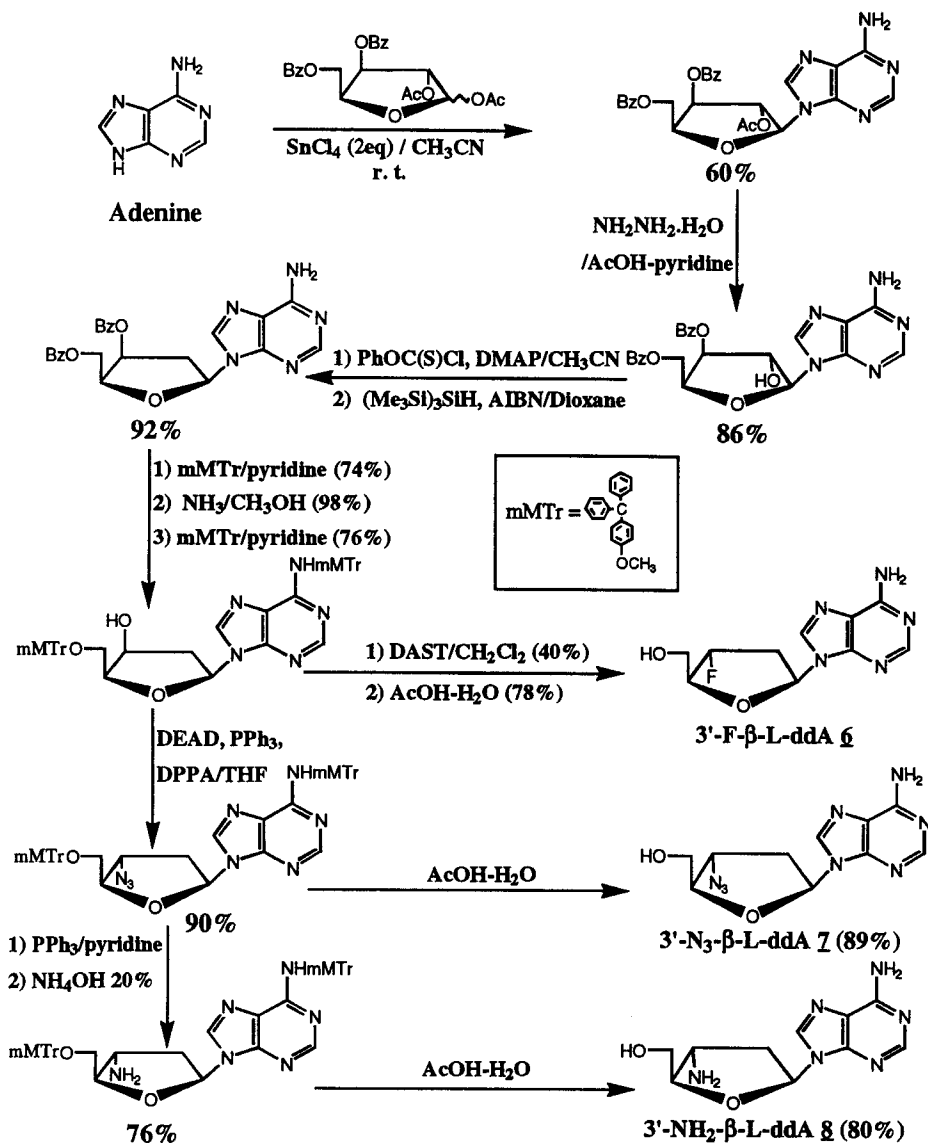
All the 2'- and 3'- substituted derivatives **3-8** of β -L-ddA **1** were evaluated against HIV in MT-4 cells and against HBV in 2.2.15 transfected Hep-G2 cells (Table).

Thus it appears that none of the studied compounds, including the parent nucleoside β -L-ddA **1**, displayed anti-HIV activity at the highest concentration tested (100 μ M). Although we have recently reported that β -L-ddA **1** inhibited HIV replication in PBM cells ($EC_{50} = 8.2 \mu$ M, $CC_{50} > 100 \mu$ M),^{10,11} the lack of anti-HIV activity of this β -L-dideoxynucleoside in MT-4 cells is in accordance with other previously published results ($EC_{50} > 100 \mu$ M in MT-2 cells).¹⁷

On the other hand, both β -L-ddA **1** and its 2'- and 3'-azido derivatives (**4** and **7**, respectively) inhibited HBV replication at concentrations ranging between 2.2 and 5.0 μ M. It is noteworthy that none of these compounds showed cytotoxicity at 200 μ M.

CONCLUSION

Several 2'- and 3'-substituted derivatives (**3-8**) of 2',3'-dideoxy- β -L-adenosine **1** were stereospecifically and conveniently synthesized following a multi-step reaction. They were characterized on the basis of their physical



- Scheme 3 -

Compound	Anti-HIV evaluation ^a MT-4 cells		Anti-HBV evaluation ^a	
	HIV-1 IIIB infected cells	non infected cells	transfected 2.2.15 cells	normal Hep-G2 cells
	EC ₅₀ ^b (μ M)	CC ₅₀ ^c (μ M)	EC ₅₀ ^b (μ M) R.I. ^d	CC ₅₀ ^c (μ M)
β -L-ddA, 1	> 100	> 100	5.0	> 200
2'-F- β -L-ddA, 3	> 100	> 100	> 10	> 200
2'-N ₃ - β -L-ddA, 4	> 100	> 100	2.2	> 200
2'-NH ₂ - β -L-ddA, 5	> 100	> 100	> 10	> 200
3'-F- β -L-ddA, 6	> 100	> 100	> 10	> 200
3'-N ₃ - β -L-ddA, 7	> 100	> 100	5.0	> 200
3'-NH ₂ - β -L-ddA, 8	> 100	> 100	> 10	> 200

^a All data represent average values from at least three separate experiments.

^b EC₅₀ values represent the drug concentration (μ M) requires to inhibit the replication of HIV-1 or HBV by 50%. ^c CC₅₀ represent the drug concentration (μ M) required to reduce the viability of non infected cell growth by 50%. ^d R.I. : Replicative intermediate (intracellular) HBV DNA.

(melting points, optical rotations) and spectroscopic properties (UV, ¹H-NMR, FAB mass spectra) ; their purities were ascertained by combustion analysis and HPLC. When evaluated against HBV in cell cultures, it appeared that the 2'- and 3'-azido derivatives (**4** and **7**, respectively), as well as β -L-ddA **1** showed some anti-HBV activity without cytotoxic effect.

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REFERENCES

1. Nair, V.; Jahnke, T. S. *Antimicrob. Agents Chemother.*, **1995**, 39, 1017.
2. Furman, P. A.; Wilson, J. E.; Reardon, J. E.; Painter, G. R. *Antiviral Chem. Chemother.*, **1995**, 6, 345.
3. Gosselin, G.; Bergogne, M.-C.; Imbach, J.-L. *J. Heterocyclic Chem.*, **1993**, 30, 1229.
4. Gosselin, G.; Mathé, C.; Bergogne, M.-C.; Aubertin, A.-M.; Kim, A.; Schinazi, R. F.; Sommadossi, J.-P.; Imbach, J.-L. *C. R. Acad. Sci., Sciences de la vie*, **1994**, 317, 85.
5. Gosselin, G.; Schinazi, R. F.; Sommadossi, J.-P.; Mathé, C.; Bergogne, M.-C.; Aubertin, A.-M.; Kim, A.; Imbach, J.-L. *Antimicrob. Agents Chemother.*, **1994**, 38, 1292.
6. Schinazi, R. F.; Gosselin, G.; Faraj, A.; Korba, B. E.; Liotta, D. C.; Chu, C. K.; Mathé, C.; Imbach, J.-L.; Sommadossi, J.-P. *Antimicrob. Agents Chemother.*, **1994**, 38, 2172.
7. Faraj, A.; Agrofoglio, L. A.; Wakefield, J. K.; McPherson, S.; Morrow, C. D.; Gosselin, G.; Mathé, C.; Imbach, J.-L.; Schinazi, R. F.; Sommadossi, J.-P. *Antimicrob. Agents Chemother.*, **1994**, 38, 2300.
8. Mathé, C.; Gosselin, G.; Bergogne, M.-C.; Aubertin, A.-M.; Obert, G.; Kim, A.; Imbach, J.-L. *Nucleosides, Nucleotides*, **1995**, 14, 549.
9. Gosselin, G.; Mathé, C.; Bergogne, M.-C.; Aubertin, A.-M.; Kim, A.; Sommadossi, J.-P.; Schinazi, R. F.; Imbach, J.-L. *Nucleosides, Nucleotides*, **1995**, 14, 611.
10. Gosselin, G.; Boudou, V.; Griffon, J.-F.; Pavia, G.; Imbach, J.-L.; Aubertin, A.-M.; Schinazi, R.F.; Faraj, A.; Sommadossi, J.-P. *Nucleosides, Nucleotides*, **1997**, 16, *in press*, and references therein.
11. Bolon, P. J.; Wang, P.; Chu, C. K.; Gosselin, G.; Boudou, V.; Pierra, C.; Mathé, C.; Imbach, J.-L.; Faraj, A.; El Alaoui, M. A.; Sommadossi, J.-P.; Pai, S. B.; Zhu, Y.-L.; Lin, J.-S.; Cheng, Y.-C.; Schinazi, R. F. *Bioorg. Med. Chem. Lett.*, **1996**, 6, 1657.
12. El Alaoui, M. A.; Faraj, A.; Pierra, C.; Boudou, V.; Johnson, R.; Mathé, C.; Gosselin, G.; Korba, B. E.; Imbach, J.-L.; Schinazi, R. F.; Sommadossi, J.-P. *Antiviral Chem. Chemother.*, **1996**, 7, 276.
13. Mitsunobu, O. *Synthesis*, **1981**, 1.
14. Matsuda, A.; Yasuoka, J.; Sasaki, T.; Ueda, T. *J. Med. Chem.*, **1991**, 34, 999.
15. Mungall, W.S.; Greene, G.L.; Heavner, G.A.; Letsinger, R.L. *J. Org. Chem.*, **1975**, 40, 1659.
16. Baker, B. R. The Ciba Foundation Symposium on the Chemistry and Biology of the Purines, G. E. W. Wolstenholme and C. M. O'Connor, eds, Churchill, London, **1957**, 120.
17. Lin T.-S.; Luo, M.-Z.; Zhu, J.-L.; Liu, M.-C.; Zhu, Y.-L.; Dutschman, G. E.; Cheng, Y.-C. *Nucleosides, Nucleotides*, **1995**, 14, 1759.