

Synthesis of [13]-membered macrocyclic stevastelins via a transesterification reaction as the key step: total synthesis of stevastelin C3

Francisco Sarabia,* Miguel García-Castro and Samy Chammaa

*Department of Biochemistry, Molecular Biology and Organic Chemistry, Faculty of Sciences,
University of Malaga, 29071 Malaga, Spain*

Received 25 August 2005; revised 6 September 2005; accepted 7 September 2005

Available online 26 September 2005

Abstract—A new synthesis of stevastelin C3 (**3**), a [13]-membered ring component of the stevastelin family, whose structure was recently revised, is reported. Initially, a macrolactonization approach was attempted to generate the [13]-membered macrolactone but this met with failure, so a transesterification reaction was tried to obtain the targeted stevastelin C3 (**3**) from the corresponding [15]-membered ring counterpart. Unfortunately, this strategy did not prove successful, and, consequently, we opted to undertake a transesterification reaction from **23**, as a means to accommodate the requisite aminoacid moiety at the correct position, to obtain **24**. From **24**, and through intermediates **25–28**, the acyclic precursor of the [13]-membered ring macrolactone, compound **30**, was efficiently prepared. By utilizing the synthetic course developed by Chida, we took **30** forward and completed the total synthesis of stevastelin C3 (**3**).

© 2005 Elsevier Ltd. All rights reserved.

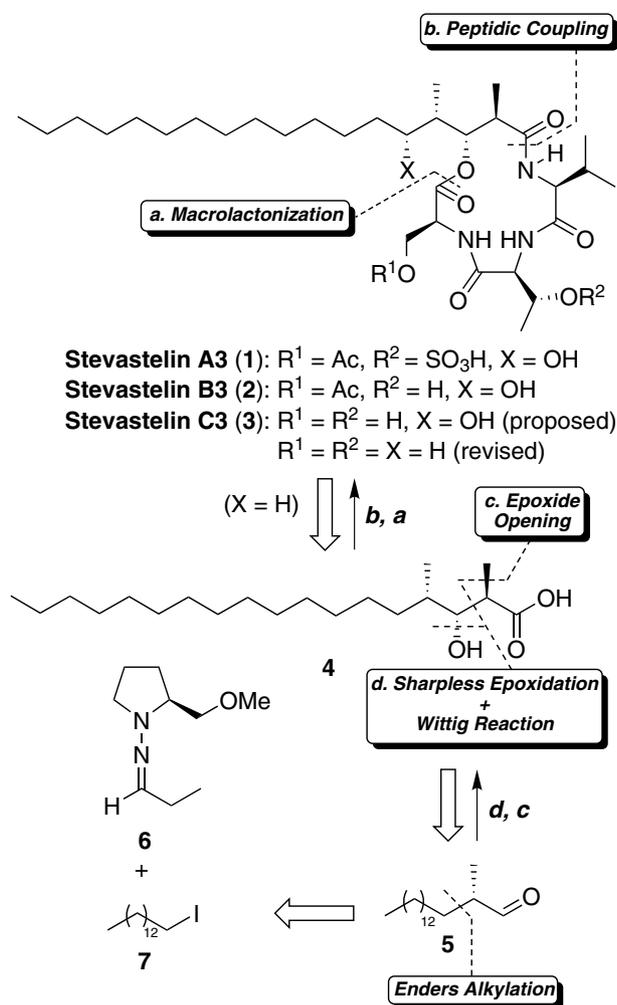
The discovery of novel natural products with unprecedented molecular architectures and intriguing biological modes of action has always represented a stimulating and attractive landmark to initiate a research programme directed towards their chemistry and biology.¹ Presently, the stevastelins, a family of cyclic depsipeptides isolated from the culture broths of *Penicillium* sp. NK374186,² fulfil the features mentioned above. In particular, they exhibit interesting immunosuppressive properties,³ characterized by their inhibition of T-cell proliferation without affecting the phosphatase activity of calcineurin, in contrast to other related immunosuppressive agents.⁴ The use of the stevastelins as potential probes to gain insights into new signal transduction pathways,⁵ as well as their potential as therapeutics in transplantation surgery and their interesting molecular structures, drew our attention and encouraged us to initiate a program directed towards the total synthesis of the [15]- and [13]-membered ring stevastelins. As part of this research, we recently completed the total syn-

theses of stevastelins B, B3 (**2**)⁶ and a set of analogues modified at the lipidic and peptidic regions,⁷ and have published synthetic studies towards the [13]-⁸ and the [15]-membered ring macrolactones.⁹ In a similar direction, other research groups have been engaged in the total syntheses of stevastelin B,^{10,11} stevastelin B3 (**2**),^{12,13} and the recently revised stevastelin C3 (**3**),¹² as an indicator of the increasing interest for this class of products in the scientific community.^{14,15} Inspired by our initial macrolactonization approach,⁸ that gave access to the [13]-membered ring congeners of the stevastelins, we devised a synthesis of stevastelin C3 (**3**) based on this strategy. Accordingly, and after disassembly of the peptidic chain, the hydroxy acid **4** represents the targeted lipidic fragment to be constructed. As is depicted in the retrosynthetic Scheme 1, the synthesis of hydroxy acid **4** would be planned from the chiral aldehyde **5**, involving a sequence composed of a Wittig reaction, ester reduction, Sharpless epoxidation and an oxirane ring opening reaction as a means for the stereoselective construction of the C-2/C-3 system. The chiral aldehyde **5** then would be readily accessible by an Enders alkylation of SAMP-hydrazone **6** with alkyl iodide **7** (Scheme 1).

Thus, using the general strategy outlined in Scheme 1, the SAMP hydrazone **6**¹⁶ was sequentially treated with

Keywords: Cyclic depsipeptide; Immunosuppressant; Stevastelin C3; Total synthesis; Transesterification.

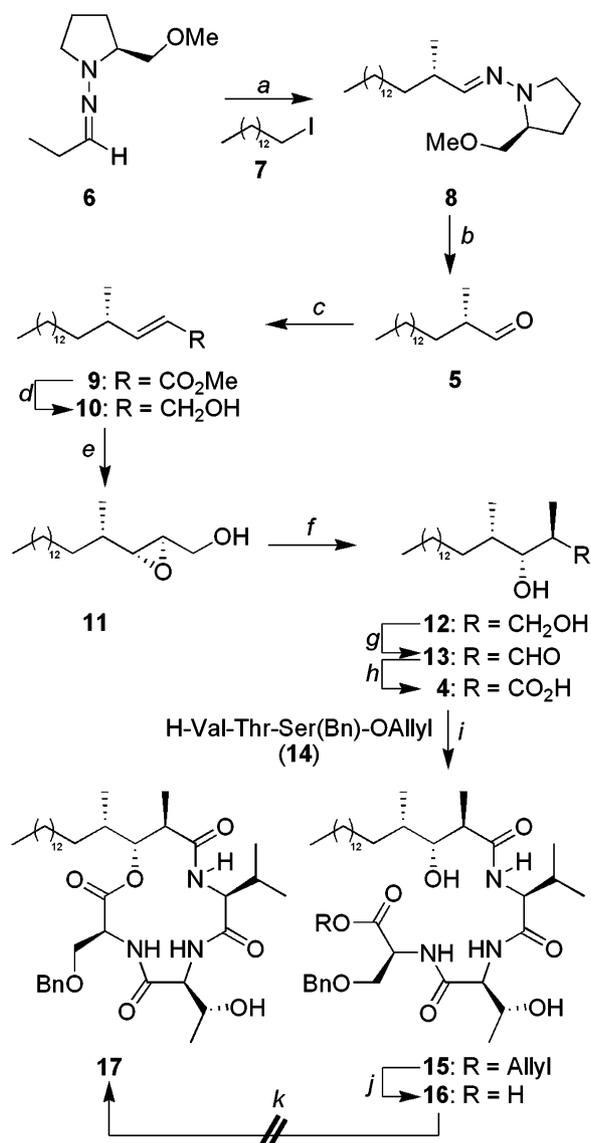
* Corresponding author. Tel.: +34 952 134258; fax: +34 952 131941; e-mail: frsarabia@uma.es



Scheme 1. Structures of [13]-membered stevastelins and retrosynthetic analysis for stevastelin C3 (3).

LDA and 1-iodotetradecane (7) to afford compound 8 in 87% yield and >98% de. Cleavage of the hydrazone moiety by exposure to ozone,¹⁷ followed by Wittig reaction of the resulting aldehyde 5 provided the α,β -unsaturated ester 9 in 93% yield and complete stereoselectivity. After treatment of ester 9 with DIBAL-H, the resulting allylic alcohol 10 was subjected to an asymmetric Sharpless epoxidation (TBHP, Ti(O*i*-Pr)₄, D-(–)-DET)¹⁸ to afford epoxy alcohol 11 in 86% yield and 92% de, according to its ¹H NMR spectra. The reaction of 11 with the Gillman reagent (Me₂CuLi)¹⁹ was followed by a sequential oxidation of the resulting diol 12 that involved oxidation of the primary alcohol to aldehyde 13 by the selective action of TEMPO,²⁰ followed by treatment with NaClO₂²¹ to yield hydroxy acid 4, which was used in the subsequent step without further purification. The coupling of 4 with tripeptide 14 was accomplished under conventional conditions (EDCI, HOBt)²² providing the corresponding coupled product 15 in 47% overall yield from diol 12. The preparation of the key seco acid 16 required the transformation of the allyl ester functional group to the corresponding acid, which was achieved by the action of Pd[PPh₃]₄ and morpholine.²³ The key macrolactonization reaction of 16 was carried out

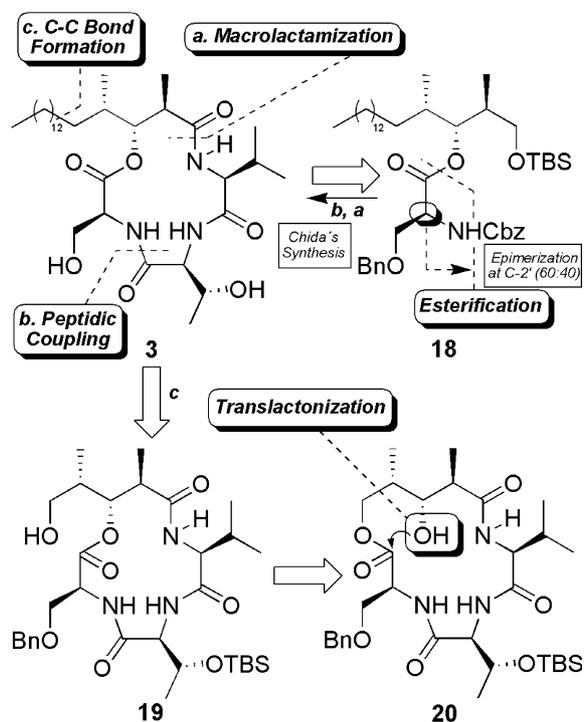
according to the Yamaguchi method²⁴ (2,4,6-trichlorobenzoyl chloride, Et₃N, 4-DMAP) at various temperatures, ranging from 25 up to 80 °C. Unfortunately, in no case, the desired macrocyclic derivative 17 was obtained, yielding a complex mixture of decomposition products (Scheme 2).



Scheme 2. Reagents and conditions: (a) 1.0 equiv 8, 1.0 equiv LDA, THF, –78 °C, 12 h, then 1.0 equiv 7, –78 °C, 6 h, 87%, > 98% ee; (b) O₃, CH₂Cl₂, –78 °C, 1 h, 76%; (c) 2.0 equiv Ph₃P=CHCO₂Me, CH₂Cl₂, 25 °C, 4 h, 93%; (d) 2.4 equiv DIBAL-H, CH₂Cl₂, –78 °C, 0.5 h, 81%; (e) 0.25 equiv D-(–)-DET, 0.23 equiv Ti(O*i*-Pr)₄, 2.1 equiv TBHP, CH₂Cl₂, –20 °C, 24 h, 86% (de 92%); (f) 3.0 equiv Me₂CuLi, THF, 0 °C, 4 h, 77%; (g) 0.01 equiv TEMPO, 0.1 equiv KBr, 15.0 equiv NaHCO₃, 2.4 equiv NaClO, CH₂Cl₂/H₂O, 0 °C, 15 min; (h) 3.0 equiv NaClO₂, 2.4 equiv NaH₂PO₄, 87.0 equiv 2-methyl-2-butene, ¹BuOH/H₂O (4/1), 25 °C, 10 min; (i) 1.2 equiv 15, 1.5 equiv EDCI, 1.1 equiv HOBt, CH₂Cl₂, 25 °C, 2 h, 47% overall yield from 13; (j) 0.15 equiv Pd[PPh₃]₄, 10.0 equiv morpholine, THF, 25 °C, 0.5 h, 60%; (k) 5.0 equiv 2,4,6-trichlorobenzoyl chloride, 6.0 equiv Et₃N, THF, 0 °C, 20 min; then slow addition to a solution of 4-DMAP (10.0 equiv) in toluene (0.002 M based on 4), 25 → 80 °C, 8 h, decomposition products.

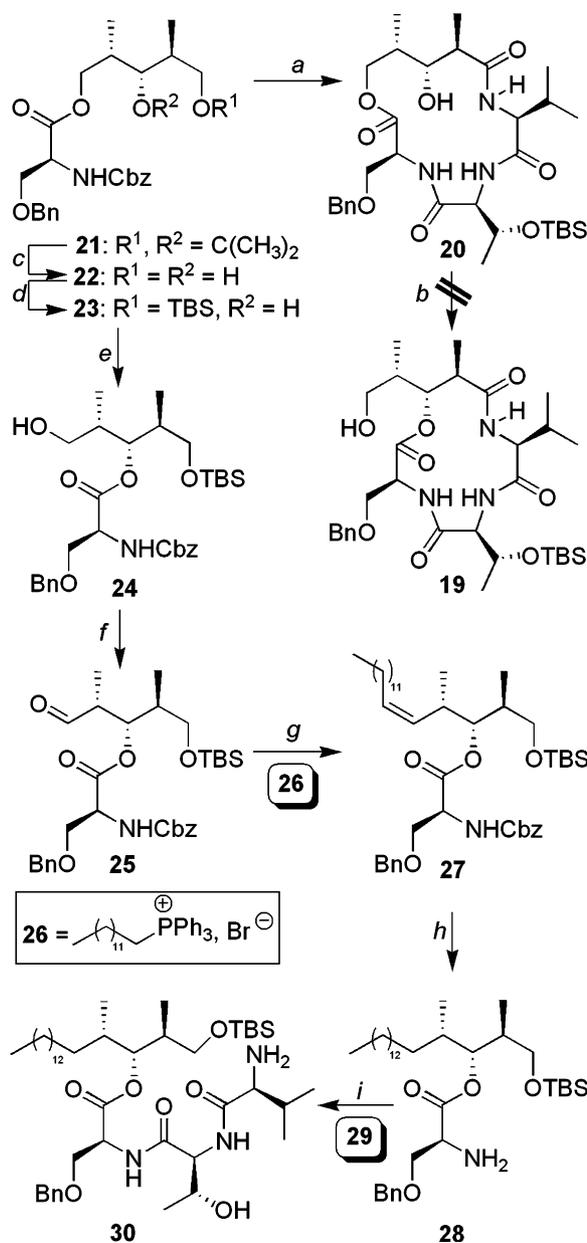
In an effort to find a direct route to the desired [13]-membered ring macrolactone and taking advantage of our previous experience with the synthesis of stevastelin B3 (**2**),⁶ a second protocol involving a [15]- → [13]-membered ring translactonization reaction was explored. According to this new approach, the [15]-membered ring macrodepsipeptide **20** was established as the suitable precursor for the corresponding [13]-membered ring derivative **19**, through a translactonization reaction, which would only require the appending of the appropriate lipidic chain through a suitable C–C-bond formation reaction. This new synthetic strategy would avoid an important drawback found within the reported synthesis by Chida and co-workers,¹² which is the introduction of the serine residue by an esterification process that results in a high degree of epimerization (60:40) at C-2' of compound **18** (Scheme 3).

Having successfully synthesized the cyclic depsipeptide **20** from acyclic precursor **21**,⁷ prepared by esterification of the corresponding alcohol²⁵ with the commercially available amino acid derivative Cbz–Ser(Bn)–OH,²⁶ we studied the translactonization reaction by exposing **20** to different reagents, which proved to be effective for related derivatives. Thus, treatment of **20** with bases (LHMDS or related bases) or with Lewis acids (Ti(Oi-Pr)₄ or Otera catalyst²⁷) did not result in the formation of the desired [13]-membered derivative **19**, and provided only recovered starting material. In pursuit of the targeted compound **20**, we turned our attention to a transesterification reaction as a potential entry into the [13]-membered ring derivatives.²⁸ To this end, ester **21** was transformed into hydroxy ester **23**, through dihydroxy ester **22**. To our delight, treatment of **23** with



Scheme 3. New synthetic strategy for stevastelin C3 based on a translactonization reaction.

2.0 equiv of LHMDS furnished the desired ester **24** in approximately 50% yield, together with the recovered starting material (~50%), which could be incorporated into a recycling process. Other catalysts such as the Otera catalyst proved to be similarly efficient in this reaction, providing **24** in similar yields as the treatments involving basic reagents. Prior to the introduction of the peptidic fragment, we decided to incorporate the lipidic



Scheme 4. Reagents and conditions: (a) See Ref. 10; (b) 2.0 equiv NaHMDS, THF, 0 °C, 15 min, recovery of starting material; (c) AcOH/H₂O, THF, 25 °C, 12 h, 95%; (d) 1.2 equiv TBSCl, 1.5 equiv imidazole, DMF, 25 °C, 15 min, 95%; (e) 2.0 equiv LHMDS, THF, –40 °C, 10 min, 50%, recovering ~50% of starting material; (f) 0.02 equiv TEMPO, 0.1 equiv KBr, 20.0 equiv NaHCO₃, 3.0 equiv NaClO, CH₂Cl₂/H₂O, 0 °C, 45 min; (g) 2.0 equiv **26**, 2.0 equiv NaHMDS, THF, 0 °C, 10 min, then addition to a solution of 1.0 equiv of **25**, 15 min, 42% over two steps from **24**; (h) 0.1 equiv 10% Pd/C-ethylenediamine complex, H₂, MeOH, 25 °C, 0.5 h; (i) 1.2 equiv **29**, 1.0 equiv HOBT, 1.5 equiv EDCI, DMF, 25 °C, 0.5 h, 70% over two steps from **27**.

chain, thus alcohol **24** was transformed into aldehyde **25** by the oxidative action of TEMPO and then reacted with the in situ generated phosphorous ylide derived from phosphonium salt **26** by treatment with NaH-MDS, to afford the *Z* olefin **27** in 42% yield over two steps from **24**. The reduction of alkene **27** to the alkane **28** was followed by the coupling of dipeptide **29**, which was carried out with EDCI and HOBt to provide compound **30** in a 70% overall yield from alkene **27** (Scheme 4).

The completion of the synthesis of stevastelin C3 (**3**) was achieved according to Scheme 5 and following the route reported by Chida. Compound **30** was prepared for the macrocyclization reaction through a synthetic sequence that included silyl ether deprotection (AcOH, H₂O, **31**, 90%), oxidation (TEMPO and NaClO₂, **32**) and Boc-cleavage (TFA) to give amino acid **33**, which was subjected to the action of DEPC under high dilution conditions to afford the macrocyclic derivative **17** in 42% overall yield from alcohol **31**. Finally, the deprotection of the benzyl ether group delivered Stevastelin C3 (**3**)²⁹ in 84% yield, whose physical and spectroscopic properties were identical to those reported for the natural product.³⁰

In conclusion, the chemistry described in this article presents a concise strategy for the construction of the [13]-membered stevastelins based on transesterification and macrolactamization processes as the key reactions, giving efficient access to stevastelin C3 (**3**) and complement-

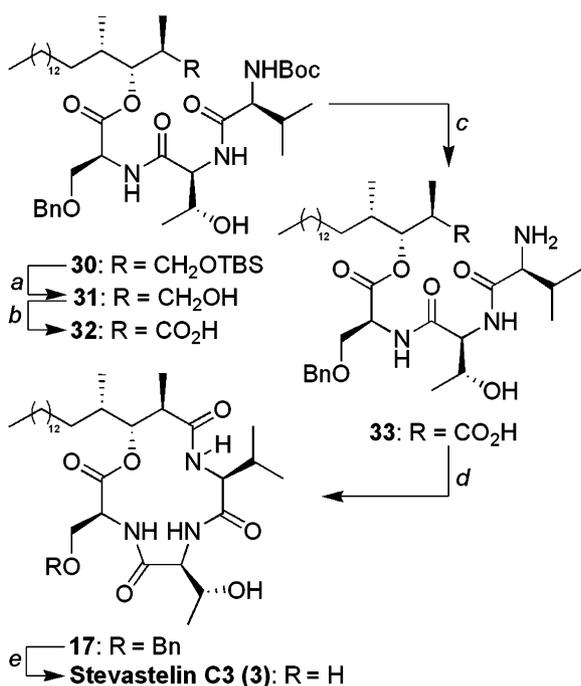
ing our previously described strategy based on a transesterification reaction that was employed for the synthesis of stevastelin B3 (**2**).

Acknowledgements

This work was financially supported by *Fundación Ramón Areces* and the *Dirección General de Investigación y Científica Técnica* (ref. CTQ2004-08141) and fellowship from *Fundación Ramón Areces* (S.C.). We thank Dr. J. I. Trujillo for assistance in the preparation of this manuscript. We thank Unidad de Espectroscopía de Masas de la Universidad de Granada for mass spectroscopic assistance.

References and notes

- Sarabia, F.; Chammaa, S.; Sánchez-Ruiz, A.; Martín-Ortiz, L.; López-Herrera, F. J. *Curr. Med. Chem.* **2004**, *11*, 1309–1332.
- (a) Morino, T.; Masuda, A.; Yamada, M.; Nishimoto, M.; Nishikiori, T.; Saito, S.; Shimada, N. *J. Antibiot.* **1994**, *47*, 1341–1343; (b) Morino, T.; Shimada, K.-I.; Masuda, A.; Nishimoto, M.; Saito, S. *J. Antibiot.* **1996**, *49*, 1049–1051.
- (a) Morino, T.; Shimada, K.-I.; Masuda, A.; Noriyuki, Y.; Nishimoto, M.; Nishikiori, T.; Saito, S. *J. Antibiot.* **1996**, *49*, 564–568; (b) Shimada, K.-I.; Morino, T.; Masuda, A.; Sato, M.; Kitagawa, M.; Saito, S. *J. Antibiot.* **1996**, *49*, 569–574.
- Hamaguchi, T.; Masuda, A.; Morino, T.; Osada, H. *Chem. Biol.* **1997**, *4*, 279–286.
- (a) Fischer, G. In *Drug Discovery from Nature*; Grabley, S., Thiericke, R., Eds.; Springer, 2000; pp 257–280, Chapter 14; (b) Fischer, G. *Angew. Chem., Int. Ed. Engl.* **1994**, *33*, 1415–1436.
- Sarabia, F.; Chammaa, S. *J. Org. Chem.* **2005**, *70*, ASAP (JO0506251).
- Sarabia, F.; Chammaa, S.; García-Castro, M. *J. Org. Chem.* **2005**, *70*, ASAP (JO050628y).
- Sarabia, F.; Chammaa, S.; López-Herrera, F. J. *Tetrahedron Lett.* **2002**, *43*, 2961–2965.
- Sarabia, F.; Chammaa, S.; Sánchez-Ruiz, A.; López-Herrera, F. J. *Tetrahedron Lett.* **2003**, *44*, 7671–7675.
- Kurosawa, K.; Nagase, T.; Chida, N. *Chem. Commun.* **2002**, 1280–1281.
- Kohyama, N.; Yamamoto, Y. *Synlett* **2001**, 694–696.
- Kurosawa, K.; Matsuura, K.; Chida, N. *Tetrahedron Lett.* **2005**, *46*, 389–392.
- Chakraborty, T. K.; Ghosh, S.; Laxman, P.; Dutta, S.; Samanta, R. *Tetrahedron Lett.* **2005**, *46*, 5447–5450.
- Chakraborty, T. K.; Ghosh, S.; Dutta, S. *Tetrahedron Lett.* **2001**, *42*, 5085–5088.
- Manger, M.; Scheck, M.; Prinz, H.; von Kries, J.-P.; Langer, T.; Saxena, K.; Schwalbe, H.; Fürstner, A.; Rademann, J.; Waldmann, H. *ChemBiochem*, in press.
- Enders, D.; Eichenauer, H. *Chem. Ber.* **1979**, *112*, 2933–2960.
- (a) Enders, D.; Tiebes, J.; De Kimpe, N.; Keppens, M.; Stevens, C.; Smagghe, G.; Betz, O. *J. Org. Chem.* **1993**, *58*, 4881–4884; (b) Enders, D.; Plant, A.; Backhaus, D.; Reinhold, U. *Tetrahedron* **1995**, *51*, 10699–10714.
- (a) Katsuki, T.; Sharpless, K. B. *J. Am. Chem. Soc.* **1980**, *102*, 5974–5976; (b) Johnson, R. A.; Sharpless, K. B. In *Catalytic Asymmetric Synthesis*; Ojima, I., Ed.; VCH: Weinheim, New York, 1993; pp 103–158.



Scheme 5. Reagents and conditions: (a) AcOH/H₂O, THF, 25 °C, 18 h, 90%; (b) i. 0.02 equiv TEMPO, 0.1 equiv KBr, 20.0 equiv NaHCO₃, 3.0 equiv NaClO, CH₂Cl₂, 0 °C, 1 h. ii. 2.5 equiv NaClO₂, 2.0 equiv NaH₂PO₄, 100.0 equiv 2-methyl-2-butene, ¹BuOH/THF/H₂O, 25 °C, 1 h; (c) TFA (excess), CH₂Cl₂, 0 °C, 1 h; (d) 5.0 equiv DEPC, 5.5 equiv Et₃N, DMF (1.0 mM based on diol **31**), 0 → 25 °C, 42% for **17** from **31**; (e) H₂, 10% Pd(OH)₂/C, MeOH, 25 °C, 2 h, 84%.

19. (a) Gilman, H.; Jones, R. G.; Woods, L. A. *J. Org. Chem.* **1952**, *17*, 1630–1634; (b) Chakraborty, T. K.; Jayaprakash, S.; Laxman, P. *Tetrahedron* **2001**, *57*, 9461–9467.
20. Leanna, M. R.; Sowin, T. J.; Morton, H. E. *Tetrahedron Lett.* **1992**, *33*, 5029–5032.
21. (a) Lindgren, B. O.; Nilsson, T. *Acta Chem. Scand.* **1973**, *27*, 888–890; (b) Bal, B. S.; Childers, W. E., Jr.; Pinnick, H. W. *Tetrahedron* **1981**, *37*, 2091–2096.
22. (a) Jones, J. *Amino Acid and Peptide Synthesis In Oxford Chemistry Primers Serie, 7*; Oxford University Press: New York, 1992; (b) Hale, K. J.; Bhatia, G. S.; Frigerio, M. In *The Chemical Synthesis of Natural Products*; Hale, K. J., Ed.; Sheffield Academic Press: Sheffield, 2000; pp 349–415.
23. Kogen, H.; Kiho, T.; Nakayama, M.; Furukawa, Y.; Kinoshita, T.; Inukai, M. *J. Am. Chem. Soc.* **2000**, *122*, 10214–10215.
24. (a) Inanaga, J.; Hirata, K.; Saeki, H.; Katsuki, T.; Yamaguchi, M. *Bull. Chem. Soc. Jpn.* **1979**, *52*, 1989–1993; (b) Nicolaou, K. C.; Patron, A. P.; Ajito, K.; Richter, P. K.; Khatuya, H.; Bertinato, P.; Miller, R. A.; Tomaszewski, M. *J. Chem. Eur. J.* **1996**, *2*, 847–868; (c) Nicolaou, K. C.; Ninkovic, S.; Sarabia, F.; Vourloumis, D.; He, Y.; Vallberg, H.; Finlay, M. R. V.; Yang, Z. *J. Am. Chem. Soc.* **1997**, *119*, 7974–7991.
25. (a) Horita, K.; Oikawa, Y.; Yonemitsu, O. *Chem. Pharm. Bull.* **1989**, *37*, 1698–1704; (b) Ziegler, F. E.; Kneisley, A.; Thottathil, J. K.; Wester, R. T. *J. Am. Chem. Soc.* **1988**, *110*, 5434–5442.
26. Aminoacid derivative Cbz–Ser(Bn)–OH was purchased from NovaBiochem.
27. (a) Seebach, D.; Hungerbühler, E.; Naef, R.; Schnurrenberger, P.; Weidmann, B.; Züger, M. *Synthesis* **1982**, 138–141; (b) Otera, J.; Ioka, S.; Nozaki, H. *J. Org. Chem.* **1989**, *54*, 4013–4014; (c) Trost, B. M.; Papillon, J. P. N. *J. Am. Chem. Soc.* **2004**, *126*, 13618–13619.
28. (a) Otera, J.; Dan-oh, N.; Nozaki, H. *J. Org. Chem.* **1991**, *56*, 5307–5311; (b) Orita, A.; Sakamoto, K.; Hamada, Y.; Mitsutome, A.; Otera, J. *Tetrahedron* **1999**, *55*, 2899–2910.
29. All new compounds exhibited satisfactory spectroscopic and analytical and/or accurate mass data.
30. Synthetic Stevastelin C3 (**3**) exhibited identical properties (TLC, $[\alpha]_D$, ^1H and ^{13}C NMR and HRMS) with those reported for the natural compound, matching ^1H and ^{13}C NMR spectra by direct comparison with spectra of natural substance: Morino, T.; Nishimoto, M.; Nishide, M. U.; Masuda, A.; Yamada, M.; Kawano, E.; Nishikiori, T.; Saito, S. EP0525361, 1993.