

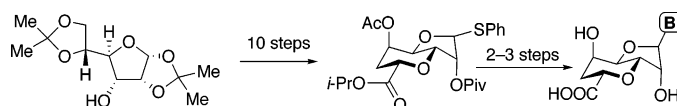
## Short Synthesis of Octosyl Nucleosides

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## ABSTRACT



Commercial 1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-allofuranose was converted to a protected bicyclic octosyl acid thioglycoside donor by a 10-step sequence that features an intramolecular ester enolate alkylation. Glycosylation of *N*-benzoyladenine and methyl uridine-5-carboxylate followed by deprotection gave the respective nucleosides “octosyl adenine” and octosyl acid A.

Complex nucleoside antibiotics are diversely polyfunctional targets.<sup>1</sup> A synthetic strategy can either feature (a) “early glycosylation,” which requires chain, ring, and functional group elaboration of a commercially available or early-route nucleoside, or (b) “late glycosylation,” in which *N*-glycosylation of a purine/pyrimidine acceptor with a higher sugar donor occurs toward the end of the route. The challenges have been successfully met in a variety of instances, but other attractive synthetic approaches have foundered because seemingly basic steps such as glycosylation, C–C bond formation, and protecting group removal are more troublesome in these complex molecular settings than in simpler furanose or nucleoside frameworks. The various approaches<sup>2–11</sup>

to the synthesis of octosyl acid A (**1**, Figure 1)<sup>12</sup> illustrate the difficulties, particularly with respect to the timing of the introduction of the pyrimidine vs formation of the strained trans-fused 1,5-dioxabicyclo[4.3.0]nonane ring system. The three pioneering syntheses<sup>2–4</sup> of **1** made use of the chain extension at C-5' and then intramolecular Williamson ether formation,<sup>2,4</sup> or an oxymercuration,<sup>3</sup> to fuse the tetrahydropyran ring onto an existing nucleoside. In no case was the pyrimidine introduced *after* formation of the bicyclic glycon, even though this might be considered a more convergent and versatile strategy.<sup>13</sup> A promising approach<sup>5</sup> to **1** described the prior assembly of a bicyclic glycon, but standard activation at C-1' for Vorbrüggen coupling<sup>14</sup> to a pyrimidine was unsuccessful (Scheme 1), the failure being “attributed to the susceptibility of the 3,7-anhydrooctose skeleton to

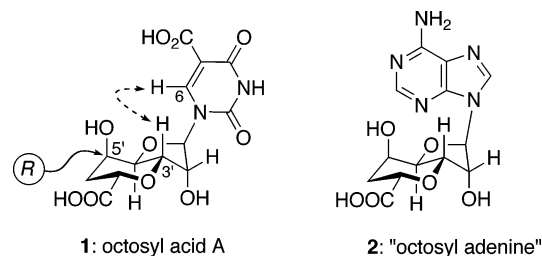
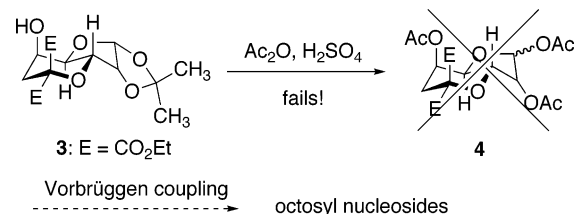
<sup>†</sup> Rutgers - The State University of New Jersey.<sup>‡</sup> Merck & Co.(1) (a) Hanessian, S.; Dixit, D. M.; Liak, T. J. *Pure Appl. Chem.* **1981**, 53, 129–148. (b) Knapp, S. *Chem. Rev.* **1995**, 95, 1859–1876. (c) Ichikawa, S.; Matsuda, A. *Nucleosides Nucleotides Nucleic Acids* **2005**, 24, 319–329.(2) (a) Danishefsky, S.; Hungate, R. *J. Am. Chem. Soc.* **1986**, 108, 2486–2487. (b) Danishefsky, S. J.; Hungate, R.; Schulte, G. *J. Am. Chem. Soc.* **1988**, 110, 7434–7440.(3) Hanessian, S.; Kloss, J.; Sugawara, T. *J. Am. Chem. Soc.* **1986**, 108, 2758–2759.(4) Kozaki, S.; Sakanaka, O.; Yasuda, T.; Shimizu, T.; Ogawa, S.; Suami, T. *J. Org. Chem.* **1988**, 53, 281–286.(5) Anzai, K.; Saita, T. *Bull. Chem. Soc. Jpn.* **1977**, 50, 169–174.(6) Kim, K. S.; Szarek, W. A. *Carbohydr. Res.* **1982**, 100, 169–176.(7) Kim, K. S.; Szarek, W. A. *Can. J. Chem.* **1981**, 59, 878–888.(8) Kim, K. S.; Kang, C. M.; Kim, S. J.; Jung, K. S.; Hahn, C. S. *Bull. Kor. Chem. Soc.* **1985**, 6, 350–354.(9) Araki, Y.; Endo, T.; Arai, Y.; Tanji, M.; Ishido, Y. *Tetrahedron Lett.* **1989**, 30, 2829–2832.

Figure 1. Octosyl nucleoside targets.

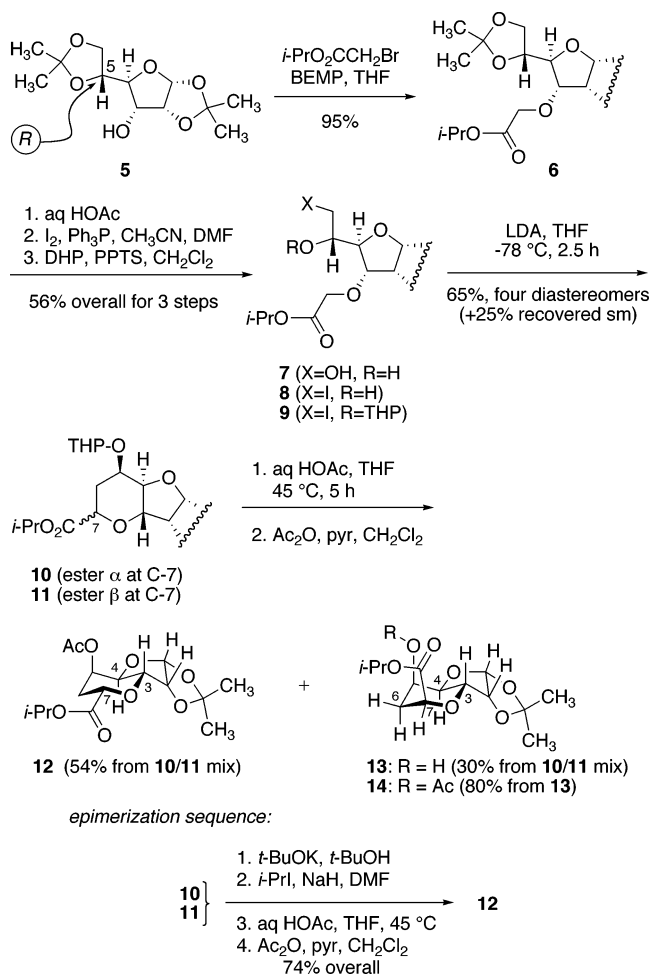
**Scheme 1.** Attempted Preparation of Octosyl Anomeric Acetate



acids”.<sup>5</sup> Given that the adenine analogue of **1**, “octosyl adenine” (**2**) shows more pronounced biological activity than **1** (**2** competes with cAMP for cyclic AMP phosphodiesterases<sup>15</sup>), a late glycosylation approach to this class of complex nucleosides, in which the pyrimidine or purine could be varied, might have value.<sup>13b</sup> Furthermore, synthetic steps would be saved if commercially available 1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-allofuranose (**5**, Scheme 2), which already matches at C-5 the C-5' stereochemistry of **1** and **2**, could be used as the starting material. We are pleased to report the syntheses of **1** and **2** by successful implementation of this late glycosylation strategy.

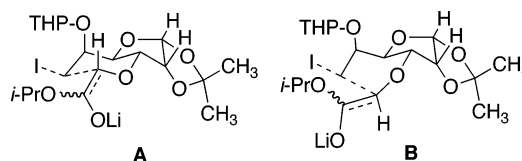
Alkylation of **5** at O-3 with isopropyl bromoacetate occurred smoothly in the presence of the strong soluble base 2-*tert*-butylimino-2-diethylamino-1,3-dimethylperhydro-1,3,2-diazaphosphorine<sup>16</sup> (BEMP, Scheme 2). Selective hydrolysis of the 5,6-*O*-isopropylidene of **6** was followed by conversion of the primary hydroxyl to an iodide (**8**) and then protection of O-5 as the THP ether (**9**). Intramolecular alkylation of the ester lithium enolate<sup>17</sup> of **9** was successful in dilute solution (2.6 mM), whereas at higher concentrations intermolecular Claisen condensation siphoned away the starting material. The product was obtained as a mixture of two stereoisomers at C-7, **10** and **11** (each a mixture of THP diastereomers), along with recovered **9**. The stereochemical picture became clearer after conversion of **10/11** by hydrolysis and acetylation to the desired C-7 isomer **12** (54% yield from **10/11**) and the C-7 epimer **13** (30% from **10/11**), each of which was isolated and characterized. The hindered C-5 hydroxyl of **13** had not acetylated, but the acetyl group could be added in a followup step to provide **14**. First-order vicinal coupling constants, particularly those of the H-6 and H-7 protons, allow assignment of the configuration and confor-

**Scheme 2.** Synthesis of the 3,7-Anhydroocturonate



mation of **12** and **14**, as shown in Scheme 2. For example, H-7 for **12** appears as a dd ( $J = 2.7, 12.3$  Hz), reflecting respective *ax-eq* and *ax-ax* couplings with the H-6's, whereas H-7 for **14** is a d ( $J = 6.8$  Hz; < 1 Hz coupling to the *trans* H-6).

The modest stereoselectivity (**10/11** = 1.8:1) for the intramolecular ester enolate alkylation may be attributed to the availability of reasonably uncongested transition states A (chairlike) and B (boatlike) for the respective modes of cyclization (Figure 2). Fortunately, the undesired epimer **11**



**Figure 2.** Stylized transition states for intramolecular alkylation.

in the **10/11** mixture could be converted to the desired isomer **12** by an epimerization sequence (Scheme 2) that consists

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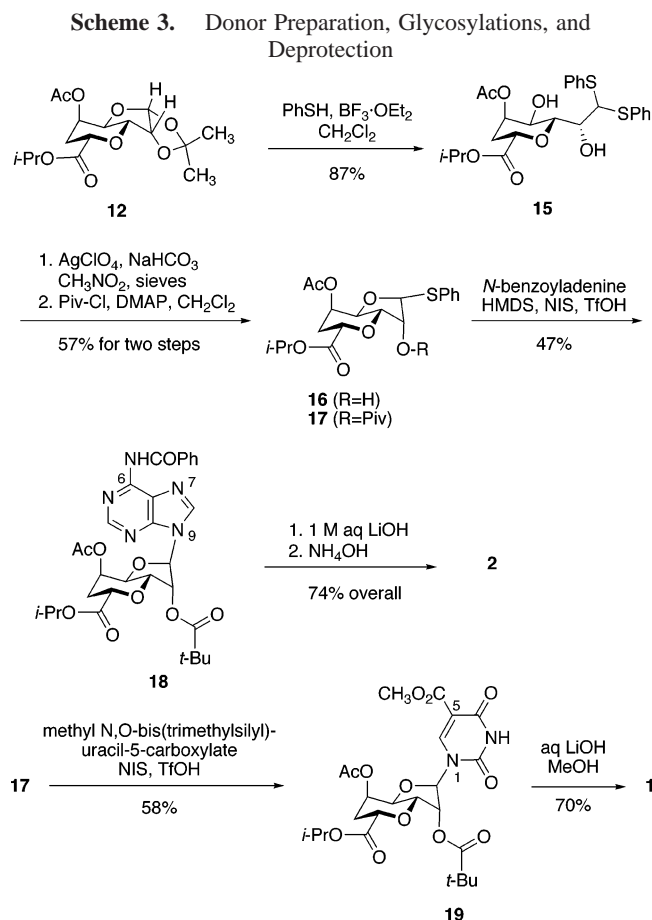
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of treatment of the mixture with *tert*-butoxide, replacement of any isopropyl lost to hydrolysis by carboxylate *O*-alkylation, hydrolysis of the *O*-THP at C-5, and then acetylation. In this way, **12** could be produced from **10/11** in 74% overall yield, 48% from **9** (64% based on recovered **9**). The synthetic route to 3,7-anhydroocturonic ester **12** is relatively short and efficient, but of little use if C-1 cannot be activated for nucleosidation.

In converting **12** to a donor for *N*-glycosylation, we employed the Lewis acid mediated acetal exchange reaction of acetonides with mercaptans.<sup>18</sup> Thus, **12** was converted to the ring-opened bis(phenylthio)acetal **15** (Scheme 3), and



then **15** was closed to the thioglycoside **16** with promotion by Ag(I)<sup>19</sup> and participation of O-4. Although the anomeric stereochemistry was not determined with certainty, **16** was obtained as a single isomer. Subsequent pivaloylation at O-2 to give **17** proceeded very slowly, suggesting that the nearby (*cis*?) phenylthio substituent hinders acylation, as had been observed with a related thioglycoside.<sup>20</sup> As thioglycosides

of either stereochemistry are effective donors for *N*-glycosylation,<sup>21,22</sup> **17** ought to serve as a precursor to a variety of octosyl nucleosides, including **1** and **2**, differing only in the identity of the nucleobase.

Both glycosylations proved successful (Scheme 3). Treatment of a mixture of **17** and silylated *N*<sub>6</sub>-benzoyladenine with *N*-iodosuccinimide and triflic acid<sup>22</sup> led to the protected nucleoside **18** along with a small amount of an isomer, probably *N*-7 glycosylated. Deprotection with aqueous lithium hydroxide removed the acetyl and pivaloyl groups and hydrolyzed the isopropyl ester, but not the *N*-benzoyl, which promotes deprotonation at *N*-6 under these conditions.<sup>23</sup> Subsequent ammonolysis, however, removed the remaining protecting group, and the product **2** was isolated and characterized by its mass spectrum and fully assigned proton and carbon NMR spectra. In particular, the singlet for H-1' of **2** is diagnostic for octosyl nucleosides of the desired stereochemistry, and the respective chemical shifts for C-4 and C-5 (148.5 and 119.1 ppm) match those of adenosine (149.2 and 119.5) but not 7-(β-D-ribofuranosyl)-adenine (160.7 and 110.2).<sup>24</sup> A literature description of **2** (which was prepared by a nucleoside transglycosylation sequence starting with **1**)<sup>25</sup> includes chemical shifts for H-1', H-2, and H-8 that match our values.

The option to activate the anomeric center of **12** for *N*-glycosylation as a thioglycoside, rather than the more usual anomeric acetate, was crucial to the success of this route. The question<sup>26</sup> has been posed: "Why not use the protected 1-*O*-acyl or 1-*O*-alkyl sugars for nucleoside synthesis instead of the corresponding 1-phenylthio sugars, which entail additional reaction steps and bad smelling thiophenols?" The syntheses of **1** and **2** and several additional targets<sup>13b,21,27,28</sup> provide the answer: In a complex synthetic undertaking, the thioglycoside is often a more effective way to prepare the anomeric center for late *N*-glycosylation.

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**Supporting Information Available:** Experimental details and spectral characterization of new compounds, including copies of <sup>13</sup>C and <sup>1</sup>H NMR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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