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Preparation of 2-O-(3-O-Carbamoyl-α-D-mannopyranosyl)-L-gulopyranose: Synthetic Study on the Sugar Moiety of Antitumor Antibiotic Bleomycin

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Abstract: A new practical route to the disaccharide moiety of bleomycin was developed. Both of key building blocks 9 and 16 were prepared from D-mannose in a regioselective manner by applying stannylene acetal methodology. Glycosylation of the allyl alcohol 9 with the trichloroacetimidate 16 proceeded smoothly, and the further incorporation to the disaccharide moiety was successfully accomplished.

Antitumor antibiotic bleomycin² has quite unique disaccharide moiety, 2-O-(3-O-carbamoyl- α -D-mannopyranosyl)-L-gulopyranose. In order to investigate the role of this moiety³, it became necessary to develop a practical method of the preparation of L-gulose equivalent and 3-O-carbamoyl-D-mannose.



Disaccharide Moiety of Bleomycin

Recently Boger and coworkers reported the total synthesis of bleomycin A₂.⁴ During the course of their synthesis, they developed a stereoselective conversion of D-mannose to L-gulose derivative by Rh(I)-catalyzed hydroboration of 6-deoxy-hex-5-enopyranoside derivative.^{4d} This prompts us to disclose our own results on the carbohydrate moiety of bleomycin.

In the previous paper⁵ we reported a novel method for the conversion of D-mannose to L-gulose. The inversion of the stereochemistry at mannose C-5 was achieved through oxidation-reduction method. Described herein are (1) application of our methodology for the preparation of properly protected L-gulose equivalent, (2) regioselective introduction of carbamoyl group onto mannose C-3 hydroxyl, and finally (3) coupling of each fragment and the further incorporation to the disaccharide moiety of bleomycin.

Scheme 1 shows the preparation of L-gulose equivalent. It was necessary to distinguish the gulose C-2 hydroxyl group for introduction of D-mannose portion, and for this purpose CsF-mediated regioselective alkylation of stannylene acetal⁶ was adopted.

Scheme 1



Reagents and Conditions: a (i) Ac₂O, Pyridine; (ii) PhSH, SnCl₄, CH₂Cl₂; (iii) NH₃, MeOH; (iv) PhCH(OMe)₂, HBF₄, DMF (overall 44%), b *n*-Bu₂SnO, PhMe, reflux; BnBr, CsF, DMF (91%), c NaH, MPMCl, DMF (98%), d NBS, aq.acetone (50%, 90% conversion), e Ph₃P⁺CH₃ Br (2.7 equiv.), *n*-BuLi (2.5 equiv.) (66%), f (COCl)₂, DMSO, Et₃N, CH₂Cl₂, g LiB(sec-Bu)₃H, THF, -78 °C (83% from 5), h Ac₂O, Pyridine (94%), i DDQ, aq.CH₂Cl₂ (91%).

Thus, 4,6-O-benzylidene-D-mannothioglycoside 1, prepared from D-mannose (4 steps, 44% overall yields), was converted to 3-O-benzyl-2-O-MPM derivative 3 through selective benzylation of the stannylene acetal of 1 followed by p-methoxybenzylation at the remaining hydroxyl group at C-2. After oxidative hydrolysis of phenylthioglycoside, the resulting hemiacetal 4 was reacted with phosphonium ylide to obtain the hydroxylolefin 5. It was expected that the protection of the C-1⁷ formyl group as vinyl group would result in increasing a nucleophilicity of the hydroxyl group at C-2⁷. The β -dioxanol 5 was oxidized to dioxanone 6 by Swern oxidation. L-Selectride reduction of 6 gave the desired α -dioxanol 7 as a single isomer in 83% yield from the β -dioxanol 5. Protection of C-5 hydroxyl group as an acetate followed by oxidative hydrolysis of MPM ether⁸ provided the allyl alcohol 9⁹, which serves as the glycosyl acceptor for the disaccharide formation.

The synthesis of 3-O-carbamoyl- α -D-mannose portion is shown in Scheme 2. The stannylene acetal method was again employed to distinguish the C-3 hydroxyl of mannose. In this case MPM group was introduced into O-3. Alkylation of the stannylene acetal with MPMBr (generated *in situ* from MPMC1 and *n*-Bu₄NBr) and CsF afforded 3-O-MPM ether 10 in 94% yield. Deprotection of benzylidene acetal followed by benzylation of the triol gave 12. After oxidative cleavage of MPM group, the resulting alcohol 13 was converted to the 3-O-carbamoyl derivative 14 by the conventional method (*p*-NO₂C₆H₄OCOCl, Et₃N, then NH₃). Oxidative hydrolysis of thioglycoside with NBS afforded the hemiacetal 15, which was converted to the trichloroacetimidate 16⁹ by treatment with trichloroacetonitrile and a catalytic amount of DBU.

Scheme 2



Reagents and Conditions: a *n*-Bu₂SnO, PhMe, reflux; MPMCl, *n*-Bu₄NBr, CsF, DMF (94%), b 1%H₂SO₄ in MeOH, 0°C, 45 min (90%), c NaH, BnBr, DMF (99%), d DDQ, aq.CH₂Cl₂, π, 2.5 hr (83%), e *p*-NO₂C₆H₄OCOCl, Et₃N, 4-DMAP, THF; NH₃ (92%), f NBS, aq.acetone (74%), g CCl₃CN, DBU(cat.) (92%).

As expected, glycosylation of the allyl alcohol 9 with the trichloroacetimidate 16 proceeded smoothly at -15°C within 10 minutes to obtain the desired α -mannosyl derivative 17 in 84% yield. After deprotection of the acetate, 18 was subjected to ozonolysis to obtain 19 as an anomeric mixture (α : β =ca 2:1). Hydrogenolysis using Pd(OH)₂ gave free 2-(3-O-carbamoyl- α -D-mannopyranosyl)-L-gulose 20⁹, which was converted to heptaacetate 21 as a single isomer (β -anomer; $J_{1,2}$ =8.4 Hz). Spectral data ([a]_D, MS, ¹H and ¹³C NMR) of 21⁹ were identical to those reported by Bogar *et al.*^{4d}

Scheme 3



Reagents and Conditions: a 16 (1.2 equiv), BF₃-OEt₂ (2.0 equiv), CH₂Cl₂, -15 °C, 10 min (84%), b K₂CO₃, MeOH, 60°C, 10 min (93%), c O₃, MeOH, -78 °C; Me₂S (94%) d 20% Pd(OH)₂ on carbon (cat), H₂, MeOH, 5 hr (97%), e Ac₂O, Pyridine (92%).

In conclusion, we have developed a new practical route to the disaccharide moiety of bleomycin. The key features of the present synthesis are as follows: (1) olefination of C-1 formyl group of 4 not only served as the temporal protection but also increased the nucleophilicity of the hydroxyl group at gulose O-2; and that (2) stannylene acetal methodology was successfully applied for the preparation of both fragments.

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- 9. 9: [a]²⁰ +22.5° (c 1.12, CHCl₃), IR(neat) 3452, 3034, 2862, 1738, 1454 cm⁻¹; CI-MS m/z 399 $(M+H^+)$; ¹H-NMR (400MHz, CDCl₃) δ 2.18 (3H, s), 2.36 (1H, br), 3.93 (1H, dd, J=3.7, 8.4 Hz), 4.04 (1H, dd, J=1.6, 13.1 Hz), 4.09 (1H, m), 4.12 (1H, dd, J=1.6, 8.4 Hz), 4.31 (1H, dd, J=1.6, 13.1 Hz), 4.69 (1H, d, J=11.3 Hz), 4.80 (1H, ddd, J=1.6, 1.6, 1 6 Hz), 4.95 (1H, d, J=11.3 Hz), 5.26 (1H, ddd, J=1.1, 1.2, 10.4 Hz), 5.35 (1H, ddd, J=1.2, 1.2, 17.3 Hz), 5.63 (1H, s), 5.97 (1H, ddd, J=7.2, 10.4, 17.3 Hz), 7.27-7.41 (8H, m), 7.52 (2H, m); Anal calcd. for C₂₃H₂₆O₆ C: 69.33 H: 6.58, found C: 69.09 H: 6.54. 16: IR (neat) 3493, 3381, 3339, 3032, 2918, 2870, 1728, 1672, 1599, 1496, 1454, 1358, 1278, 1101, 1064, 970, 925, 835, 796, 736, 698, 646 cm⁻¹; ¹H-NMR $(400 \text{ MHz}, \text{ CDCl}_3) \delta 3.72 (1\text{H}, \text{ dd}, J=1.9, 11.3\text{Hz}), 3.82 (1\text{H}, \text{ dd}, J=4.0, 11.3\text{Hz}), 4.03 (1\text{H}, \text{ ddd}, \text{J})$ J=1.9, 4.0, 9.7Hz), 4.12 (1H, dd, J=2.4, 3.3 Hz), 4.14 (1H, dd, J=9.0, 9.7Hz), 4.52 (1H, d, J=12.0 Hz and 1H, d, J=11.1 Hz), 4.60-4.71 (5H, m), 4.76 (1H, d, J=12.1 Hz), 5.21 (1H, dd, J=3.2, 9.0 Hz), 6.37 (1H, d, J=2.4 Hz), 7.20-7.38 (15H, m), 8.59 (1H, s). **20**: 13 C-NMR (100MHz, CDCl₃) of the major anomer δ 60.5, 60.8, 63.9, 67.7, 68.5, 69.1, 72.5, 73.1, 73.5, 74.2, 92.4, 96.6, 157.6; ¹³C-NMR (100MHz, CDCl₃) of the minor anomer δ 60.7, 64.1, 65.3, 68.6, 69.4, 69.7, 71.3, 73.1, 73.3, 74.1,93.1, 99.6, 157.5; SIMS (NaCl, 3-NBA) m/z 408 (M⁺+Na). The specific rotation of 21: $[a]_{D}^{20}$ +35.5° (c 0.95, CHCl₃); lit.^{4d} $[a]_{D}^{25}$ +35.4° (c 0.07, CHCl₃).