## MICROSCALE CLEAVAGE REACTION OF (PHENYL)BENZYL ETHERS BY FERRIC CHLORIDE

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SUMMARY : Anhydrous  $\text{FeCl}_3$  in  $\text{CH}_2\text{Cl}_2$  cleaves benzyl and *p*-phenylbenzyl ethers of sugars in 15-30 minutes and 2-3 minutes, respectively, at room temperature in >70% yields; methyl ethers and acyl groups are not affected.

Benzyl ethers are commonly used as protecting groups for polyhydroxy compounds and carbohydrates because of their stability to both acids and bases. Although numerous methods are available for their removal<sup>1</sup>, problems associated with reagent incompatibility, slow debenzylation rate, low yield in microscale reactions and acyl group migration<sup>2</sup> often arise in the presence of multiple functionalities. In connection with our microscale structure determination methods of oligosaccharides by the circular dichroic exciton chirality method<sup>3,4</sup>, it was found that the following method using ferric chloride overcame these difficulties. independent of these studies the cleavage of trityl ethers and benzylidene acetals<sup>5</sup>, and the acetolysis of sugar benzyl ethers (5 min - 12 hr, 60 °C, 55-75% yield)<sup>6</sup> by ferric chloride have been reported recently.

We have found that anhydrous FeCla in CH2Cl2 is highly efficient for the cleavage of benzyl and phenylbenzyl ethers at room temperature. Importantly, methyl ether, acetate and benzoate groups are not affected by the cleavage conditions. A typical procedure involves addition of anhydrous FeCl<sub>3</sub> (2 equiv or more) to a substrate (0.5-50 µmol.) solution in dry CH<sub>2</sub>Cl<sub>2</sub> until the color of the reaction mixture changes to green-blue or brown in the case of p-phenylbenzyl ethers or benzyl ethers, respectively. This color change indicates the reaction end point and occurs in 2-3 minutes. The reaction is guenched by addition of water upon which the color usually disappears. Products are extracted with CH<sub>2</sub>Cl<sub>2</sub>. In the case of water-soluble products the solvents are evaporated, the residual mixture is treated with acetone to remove the FeCl<sub>3</sub>, and the residue is purified by microflash chromatography in an appropriate solvent system.

The results obtained using various carbohydrate benzyl and p-phenybenzyl ethers are shown in the Table. The yields are usually higher than 70 %. The benzylate cleavage reactions presumably involve initial complexation of FeCl<sub>3</sub> to the ethereal oxygen atom and subsequent displacement with a proton to yield the free alcohols and benzyl chloride. Namely, FeCl<sub>3</sub> is reacting stoichiometrically<sup>7</sup>. This reaction does not proceed in protic or lone pair electron rich solvents such as alcohols, diethyl ether and acetone. The acids FeCl<sub>3</sub> 6H<sub>2</sub>O, ZnCl<sub>2</sub> and AlCl<sub>3</sub> are not efficient but SnCl<sub>4</sub> was found to be as effective as FeCl<sub>3</sub>.

Entry	Substrate		Product	Time (min)	Yield (%)			
1		R = OPhBn R <sub>1</sub> = OPhBz		3	82 <sup>a)</sup>			
2	R <sub>1</sub> R OMe	R ≈ OPhBz R <sub>1</sub> = OPhBn	HO HO R OMe	3	83 <sup>a)</sup>			
3	R <sub>1</sub> R OMe	R = OAc R <sub>1</sub> = OPhBn		4	68 <sup>b)</sup>			
4 <sup>Ph<sup>2</sup></sup>		R ≈ OPhBn R <sub>1</sub> = OCH <sub>3</sub>	HO OH HO IO R <sub>1</sub> OMe	4	70 <sup>b)</sup>			
5		R ≈ OPhBn	но со он	Ne 3	73 <sup>b)</sup>			
6	R <sub>1</sub> R <sub>1</sub> OMe	R ≈ OBn R <sub>1</sub> = OPhBz	R <sub>1</sub> OH HO HO OMe	15	85 <sup>a)</sup>			
7		R = OBn	HO HO HO HO HO HO OMe	30	71 <sup>b)</sup>			
a) Yield was determined by UV ( $\epsilon$ 47,800 ) after HPLC ( 0.3-0.5 $\mu\text{mol}$ ).								
b) Isolated yield, SiO <sub>2</sub> column chromatography ( 50 $\mu$ mol ).								

Table. Depr	otection of	(phenyl)benzyl	l ethers of	monosaccharides	by FeCl <sub>3</sub>
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## REFERENCES

- (a) T.W. Greene, Protective Groups in Organic Synthesis, John Wiely & Sons, New York, 1981, p 29; H.G. Fletcher, Jr., Methods Carbohydr. Chem., 2, 166, 386(1963). (b) K. Fuji, T. Kawabata, and E. Fujita, Chem. Pharm. Bull., 28, 3662 (1980). (c) K. Kon, K. Ito and S. Isoe, Tetrahedron Lett., 1984, 3739.
- 2. Under hydrogenolytic conditon: R. Takeda, unpublished.
- 3. R. Takeda, A. Zask, K. Nakanishi and M.H. Park, J. Am. Chem. Soc., 109, 914 (1987).
- 4. K. Nakanishi, M.H. Park, R. Takeda, J.T. Vazquez, and W. Wiesler, *Stereochemistry of Organic and Bioorganic Transformations*, W. Bartmann ed., Verlag Chemie, 1987, pp.303-319.
- 5. K.S. Kim, Y.H. Song, B.H. Lee, C.S. Hahn, J. Org. Chem., 51, 404 (1986).
- 6. K.P.R. Kartha, F. Dasgupta, P.P. Singh and H.C. Srivastava, J. Carbohydr. Chem., 5, 437 (1986).
- 7. T.C. Jempty, K.A.Z. Gogins, Y. Mazur and L.L. Miller, J. Org. Chem., 46, 4545 (1981). (Received in Japan 24 March 1987)