Synthesis of β-D-Galp-(1→4)-β-D-GlcpNAc-(1→2)-α-D-Manp-(1→0)(CH₂)₇CH₃ Mimics to Explore the Substrate Specificity of Sialyltransferases and *trans*-Sialidases

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Eleven trisaccharide octyl glycosides related to the N-glycan sequence $\beta\text{-D-Gal}p\text{-}(1\rightarrow4)\text{-}\beta\text{-D-Glc}p\text{NAc-}(1\rightarrow2)\text{-}\alpha\text{-D-Man}p\text{-}(1\rightarrow0)(\text{CH}_2)_7\text{CH}_3$ (1) designed for detailed exploration of the acceptor specificity of $\alpha\text{-}2\text{,}3\text{-}$ and $\alpha\text{-}2\text{,}6\text{-}\text{sialyltransferases}$ as well as trans-sialidases, have been synthesised: $\beta\text{-D-Gal}p\text{-}(1\rightarrow4)\text{-}\beta\text{-D-Glc}p\text{NPr-}(1\rightarrow2)\text{-}\alpha\text{-D-Man}p\text{-}(1\rightarrow0)(\text{CH}_2)_7\text{CH}_3$ (2), $\beta\text{-D-Fuc}p\text{-}(1\rightarrow4)\text{-}\beta\text{-D-Glc}p\text{NR-}(1\rightarrow2)\text{-}\alpha\text{-D-Man}p\text{-}(1\rightarrow0)\text{-}C\text{H}_2)_7\text{CH}_3$ (3, R = Ac; 4, R = Pr), 6-amino-6-deoxy- $\beta\text{-D-Gal}p\text{-}(1\rightarrow4)\text{-}\beta\text{-D-Glc}p\text{NR-}(1\rightarrow2)\text{-}\alpha\text{-D-Man}p\text{-}(1\rightarrow0)(\text{CH}_2)_7\text{CH}_3$ (5, R = Ac; 6, R = Pr), 2-deoxy- $\beta\text{-D-Gal}p\text{-}(1\rightarrow4)\text{-}\beta\text{-D-Glc}p\text{NR-}(1\rightarrow2)\text{-}\alpha\text{-D-Man}p\text{-}(1\rightarrow0)(\text{CH}_2)_7\text{CH}_3$ (7, R = Ac; 8, R = Pr), $\beta\text{-D-Gal}p\text{NR}^1\text{-}(1\rightarrow4)\text{-}\beta\text{-D-Glc}p\text{NR-}(1\rightarrow2)\text{-}\alpha\text{-D-Man}p\text{-}(1\rightarrow0)$

 $(CH_2)_7CH_3$ (9, $R=R^1=Ac$; 10, $R=R^1=Pr$; 11, R=Ac, $R^1=Pr$; 12, R=Pr, $R^1=Ac$). All trisaccharides were obtained by condensation of suitably modified glycosyl donors based on imidates or thioglycosides with the single disaccharide acceptor octyl (3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl-α-D-mannopyranoside, followed by deprotection. For the trisaccharides containing an N-acylated glucosamine as well as an N-acylated galactosamine unit, use was made of a combination of N-phthaloyl and N-dimethylmaleoyl protection.

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Introduction

Sialic acids occur at the non-reducing termini of many glycoconjugates, and are considered to be key determinants in the regulation of a variety of biological processes.^[1-3] In the biosynthesis of human and animal sialylated glycans at least 18 sialyltransferases are active with different substrate specificities.^[2,4] Furthermore, several protozoal and bacterial sialidases have shown to act as trans-sialidases, transferring sially residues in α -2,3- or α -2,6-linkage from one glycan to the terminal galactose unit of another non-sialylated oligosaccharide or glycoconjugate. [2,5] One of the most abundant elements that is sialylated is the β -D-Galp-(1 \rightarrow 4)β-D-GlcpNAc-(1→2)-α-D-Manp-(1→ sequence in glycoprotein N-glycans. A thorough analysis of the substrate specificity of sialyltransferases and trans-sialidases can be attained by using modified oligosaccharides, probing the contribution of individual hydroxy groups and the N-acetylated amino function in recognition and binding.

Initially, focusing on identifying key polar groups required for transfer of sialic acid from CPM-Neu5Ac by rat liver sialyltransferases, the acceptors β -D-Galp-(1 \rightarrow 3)- β -D-GlcpNAc and β -D-Galp-(1 \rightarrow 4)- β -D-GlcpNAc, using mim-

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ics, mainly deoxygenated forms of the pyranose rings, were studied. [6–8] It turned out that both the HO-6 group of the galactose residue and the N-acetyl group of the N-acetylglucosamine unit are required for the activity of α -2,6-sialyltransferase I (ST6Gal I). α -2,3-Sialyltransferase III (ST3Gal III) required the HO-3, HO-4, and HO-6 groups of the terminal galactose residue, and some influence from the subterminal N-acetylglucosamine was noticed.

Currently, we have studied sialyltransferases involved in the α -2,3- and α -2,6-sialylation of terminal galactose units in N-glycoprotein glycans. At first instance it was shown that rat liver ST6Gal I recognises, in addition to β-D-Galp- $(1\rightarrow 4)$ - β -D-GlcpNAc- $(1\rightarrow OMe)$ and β -D-GalpNAc- $(1\rightarrow 4)$ - β -D-GlcpNAc-(1 \rightarrow OMe), also β -D-Manp-(1 \rightarrow 4)- β -D-GlcpNAc-(1 \rightarrow OMe).^[9] Then, the trisaccharide β -D-Galp- $(1\rightarrow 4)$ - β -D-GlcpNAc- $(1\rightarrow 2)$ - α -D-Manp- $(1\rightarrow O)(CH_2)$ 7CH3 and eleven analogues containing structural variants of Dgalactose were synthesised and employed as substrates for rat liver ST6Gal I, recombinant full length human liver ST6Gal I, and recombinant N-terminal truncated rat liver ST3Gal III.^[10,11,12] Hydroxy groups at either C-3 or C-4 of the D-galactose residue were substituted by hydrogen (3and 4-deoxy-β-D-Galp-R) or fluorine (3- and 4-fluoro-β-D-Galp-R), by amino (3-amino-β-D-Galp-R) or O-methyl groups (3- and 4-O-methyl-β-D-Galp-R), or were inverted (β-D-Gulp-R and β-D-Glcp-R), to determine their involvement in binding to and catalytic activity of the enzyme. In addition, trisaccharides containing α-L-Altp (inverted FULL PAPER

J. P. Kamerling et al.

hydroxymethyl group at C-5) or β-L-Gal*p* (enantiomer) at the non-reducing terminus were constructed as probes. The ST6Gal I tolerated most of the modifications at the D-galactose residue to some extent. The best substrates were the 4-deoxy and 4-fluoro analogues, followed by the 3-deoxy and 3-fluoro analogues. The ST3Gal III displayed a narrower specificity; only the 4-*O*-methyl analogue showed to be a relatively good substrate, and the 4-deoxy and 4-fluoro analogues show a minor activity.^[13] Recently, it was demonstrated that also conformationally constrained oligosaccharides can act as acceptors for rat liver ST6Gal I.^[14]

In the framework of ongoing studies focused on the understanding of the substrate specificity of sialyltransferases and *trans*-sialidases, we have synthesised an additional series of 11 trisaccharide octyl glycosides. The trisaccharides are of the type β -D-Sugp-(1 \rightarrow 4)- β -D-GlcpNR-(1 \rightarrow 2)- α -D-Manp-(1 \rightarrow 0)(CH₂) $_7$ CH₃, with modifications at C-6 (Sug = 6-deoxy-Gal or 6-amino-6-deoxy-Gal) or at C-2 (Sug = 2-deoxy-Gal, GalNAc or GalNPr) of the D-galactose residue, in combination with N-acetylated or N-propionylated D-glucosamine [R = Ac or Pr (note that Pr = propionyl and not propyl throughout the paper)].

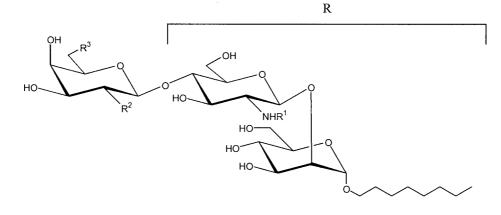
Results and Discussion

In the synthesis of the trisaccharide variants 1-12 (Scheme 1), all containing the element $\rightarrow 4$)- β -D-GlcpNR- $(1\rightarrow 2)$ - α -D-Manp- $(1\rightarrow O)$ (CH₂)₇CH₃ with R = Ac (N-ace-

tyl) for 1, 3, 5, 7, 9, and 11, and R = Pr (*N*-propionyl) for 2, 4, 6, 8, 10, and 12, the key disaccharide acceptor was the earlier synthesised octyl (3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl-α-D-mannopyranoside (13),^[10] having a free HO-4' function for elongation with suitably modified glycosyl residues. The final glycosyl residues incorporated were mimics of the native β-D-galactopyranosyl residue, i.e. a 6-deoxy-β-D-galactopyranosyl (β-D-fucopyranosyl; 3 and 4), a 6-amino-6-deoxy-β-D-galactopyranosyl (2-deoxy-β-D-galactopyranosyl; 7 and 8), a 2-acetamido-2-deoxy-β-D-galactopyranosyl (9 and 12), and a 2-deoxy-2-propionamido-β-D-galactopyranosyl (10 and 11) residue.

Compounds 1 and 2

Trisaccharide 1, octyl β-D-galactopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy-β-D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (Scheme 1), was synthesised as described earlier, starting by coupling 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl trichloroacetimidate $(14)^{[15]}$ with acceptor 13, using trimethylsilyl trifluoromethanesulfonate as a catalyst, to yield trisaccharide derivative $15^{[10]}$ (Scheme 2). Along a similar route octyl β-D-galactopyranosyl- $(1\rightarrow 4)$ -2-deoxy-2-propionamido-β-D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (2) (Scheme 1), the N-propionylated analogue of 1, was prepared. To this end 15 was de-O-acetylated, then de-N-phthaloylated using 1,2-diaminoethane in 1-butanol at 90 °C^[16] (Scheme 2), and N, O-propionylated using propi-



Code	Structure	R^1	R ²	\mathbb{R}^3
1	β-D-Gal-R	Ac	OH	OH
2	β-D-Gal-R	Pr	OH	OH
3	6-deoxy-β-D-Gal-R	Ac	OH	H
4	6-deoxy-β-D-Gal-R	Pr	OH	H
5	6-amino-6-deoxy-β-D-Gal-R	Ac	OH	NH_2
6	6-amino-6-deoxy-β-D-Gal-R	Pr	OH	NH_2
7	2-deoxy-β-D-Gal-R	Ac	H	OH
8	2-deoxy-β-D-Gal-R	Pr	H	OH
9	2-acetamido-2-deoxy-β-D-Gal-R	Ac	NHAc	OH
10	2-deoxy-2-propionamido-β-D-Gal-R	Pr	NHPr	OH
11	2-deoxy-2-propionamido-β-D-Gal-R	Ac	NHPr	OH
12	2-acetamido-2-deoxy-β-D-Gal-R	Pr	NHAc	ОН

Scheme 1. List of structures of synthesized trisaccharide octyl glycosides

Scheme 2. Synthesis of trisaccharide 2: a) NaOMe (pH = 9), CH₂Cl₂, MeOH; b) NH₂CH₂CH₂NH₂, 90 °C, 1-BuOH, molecular sieves (3 Å); c) Pr₂O, pyridine; d) NaOMe (pH = 9), MeOH; e) Ac₂O, pyridine, 72% over five steps; f) 10% Pd/C, H₂C, HOAc, EtOH, EtOAc; g) Ac₂O, pyridine, 65% over two steps; h) NaOMe (pH = 9), CH₂Cl₂, MeOH, 94%

onic anhydride in pyridine. Subsequent de-O-propionylation and O-acetylation (acetic anhydride in pyridine) yielded **16** in an overall yield of 72%. Then, de-O-benzylation of **16** by hydrogenation using 10% Pd/C as a catalyst, followed by O-acetylation (\rightarrow **17**, 65%) and finally de-O-acetylation yielded **2** (94%). The last O-acetylation step was carried out to facilitate chromatographic purification. The ¹H and ¹³C NMR spectroscopic data of **1** and **2** are presented in Tables 1 and 2, respectively.

Compounds 3 and 4

For the syntheses of the 6''-deoxy mimics of 1 and 2, octyl β -D-fucopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy- β -Dglucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (3) and octyl β -D-fucopyranosyl- $(1\rightarrow 4)$ -2-deoxy-2-propionamido- β -Dglucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (4) (Scheme 1), respectively, ethyl 2,3,4-tri-O-acetyl-1-thio-β-D-fucopyranoside (19) was used as a donor (Scheme 3). To this end Dfucose 18 was O-acetylated, after which the thioethyl function was introduced by reaction with ethanethiol in the presence of boron trifluoride-diethyl ether. Separation of the anomeric mixture by column chromatography yielded the β -anomer 19 in a yield of 61%, whereas the α -anomer was isolated in a yield of 36%. Donor 19 was coupled with 13 by in situ activation of the donor with bromine/silver trifluoromethanesulfonate, yielding trisaccharide derivative 20 (74%) (Scheme 3). It should be noted that fucosylation using 2,3,4-tri-O-acetyl-D-fucopyranosyl trichloroacetimidate was not successful (data not shown). De-N-phthaloylation/de-O-acetylation of 20, followed by N,O-acetylation $(\rightarrow 21, 85\%)$, de-O-benzylation, and O-acetylation yielded **22** (72%). Finally, de-*O*-acetylation of **22** afforded **3** (61%). In a similar way, de-N-phthaloylation/de-O-acetylation of **20**, followed by N,O-propionylation (\rightarrow **23**, 87%), de-O-

benzylation, de-*O*-propionylation, and *O*-acetylation (→ **24**, 55%), and de-*O*-acetylation yielded **4** (93%) (Scheme 4). The last *O*-acetylation step was carried out to facilitate chromatographic purification. The ¹H and ¹³C NMR spectroscopic data of **3** and **4** are presented in Tables 1 and 2, respectively.

Compounds 5 and 6

For the syntheses of the 6"-amino-6"-deoxy mimics of 1 and 2, octyl 6-amino-6-deoxy- β -D-galactopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy- β -D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (5) and octyl 6-amino-6-deoxy-β-D-galactopyranosyl- $(1\rightarrow 4)$ -2-deoxy-2-propionamido- β -D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside **(6)** (Scheme 1), respectively, the galactosyl imidate donor 30 with an azide function at C-6 was applied (Scheme 5). This donor was synthesised starting from 1,2:3,4-di-*O*-isopropylidene-α-Dgalactopyranose (25). Treatment of 25 with p-toluenesulfonyl chloride in pyridine (\rightarrow 26, 92%) and subsequent displacement of the tosyl group by an azide group using sodium azide in dimethyl sulfoxide yielded 27 (99%). The introduction of the azide group in 27 was verified by IR spectroscopy [$\tilde{v} = 2110 \text{ cm}^{-1}$]. Then, 27 was de-Oisopropylidenated using aqueous trifluoroacetic acid and Oacetylated (\rightarrow 28, 50%), followed by selective de-O-acetylation at C-1 using hydrazinium acetate in dimethylformamide (\rightarrow 29, 89%), and imidation with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene, to yield 30 (63%). Donor 30 was coupled with 13, using trimethylsilyl trifluoromethanesulfonate as a catalyst, affording trisaccharide derivative **31** (61%) (Scheme 6). De-*N*-phthaloylation/de-O-acetylation and N,O-acetylation of 31 (\rightarrow 32, 99%), followed by de-O-acetylation (\rightarrow 33, 70%) and reduction with hydrogen in the presence of 10% Pd/C of the FULL PAPER

J. P. Kamerling et al.

Table 1. 500 MHz ¹H NMR chemical shifts of the carbohydrate residues in the compounds 1-12

Residue ^[a]	Reporter group	1	2	3	4
Man	H-1	4.858 (1.4)	4.849 (1.5)	4.860 (1.6)	4.808
	H-2 H-3	4.038 (3.2)	4.048 (3.5) 3.79	4.041 (3.3)	4.006
	H-4	3.80 3.53	3.50	3.79 3.51	3.79 3.51
	H-5 H-6a ^[b]	3.60	3.59 3.62	3.58 3.63	3.51 3.68
	H-6b ^[b]	3.63 3.89	3.88	3.88	3.79
GlcNHR ^[c]	H-1	4.583 (7.0)	4.597 (7.6)	4.583 (7.6)	4.609 (7.6)
	H-2 H-3	3.75 3.73	3.75 3.74	3.75 3.72	3.79 3.75
	H-4	3.76	3.76	3.71	3.68
	H-5 H-6a	3.56	3.57	3.58	3.58
	H-6b	3.98 3.84	3.98 3.84	3.97 3.82	3.96 3.84
	CH_2	_	2.323	_	2.324
Gal	CH ₃ H-1	2.052 4.470 (7.7)	1.138 4.470 (7.7)	2.052 4.423 (7.9)	1.142 4.429 (7.8)
Gui	H-2	3.55	3.55	3.51	3.51
	H-3	3.67	3.66	3.66	3.67
	H-4 H-5	3.93 (3.2 ^[d]) 3.74	3.92 (3.3 ^[d]) 3.74	3.75 3.84	3.76 3.84
	H-6a	3.75	3.75	1.252 (6.4 ^[e])	1.257 (6.5 ^[e])
Cnasar	H-6b OC <i>H</i> H	3.75 3.53	3.75 3.52	3.53	3.48
Spacer	OCHH OCH <i>H</i>	3.74	3.79	3.72	3.48
	CH ₃	0.872	0.861	0.861	0.868
Residue	Reporter group	5	6	7	8
Man	H-1 H-2	4.855 4.032	4.845 4.043	4.854 (1.5)	4.820 4.018
	H-3	3.80	3.81	4.033 (3.3) 3.78	3.78
	H-4	3.55 3.56 3.65	3.51	3.51 3.58 3.63	3.57 3.54
	H-5 H-6a ^[b]	3.56 3.65	3.59 3.62	3.58	3.54 3.65
	H-6b ^[b]	3.86	3.87	3.87	3.82
GlcNHR ^[c]	H-1	4.585 (7.8)	4.596 (8.1)	4.569 (8.1) 3.75	4.589 (7.8)
	H-2 H-3	3.75 3.74	3.77 3.73	3.75 3.72	3.75
	H-4	3.78	3.78	3.76	3.72 3.75
	H-5	3.55	3.55	3.51 3.77 ^[b]	3.51
	H-6a H-6b	3.99 3.86	3.99 3.85	3.7/[b] 3.89[b]	3.77 ^[b] 3.88 ^[b]
	CH ₂	_	2.327	_	2.325
Gal	CH ₃ H-1	2.057 4.501 (7.6)	1.139 4.497 (7.7)	2.051 4.715	1.140 4.719
Gai	H-2 _{ax}	3.55	3.53	1.69	1.70
	H-2 _{eq} H-3	_	_	2.08	2.08
	H-3 H-4	3.68 3.93	3.69 3.93	3.89 3.77	3.90 3.77
	H-5	3.84	3.81	3.61	3.61
	H-6a	3.23	3.22	3.77	3.77
Spacer	H-6b OC <i>H</i> H	3.15 3.51	3.15 3.51	3.77 3.51	3.77 3.50
Spacer	OCHH	3.75	3.71	3.73	3.71
	CH ₃	0.865	0.861	0.865	0.867
Residue	Reporter group	9	10	11	12
Man	H-1 H-2	4.852 4.026	4.840 4.027	4.846 4.008	4.837 4.027
				1.000	7.04/
	H-3	3.79		3.79	3.79
	H-3 H-4	3.79 3.50	3.78 3.51	3.79 3.55	3.79 3.50
	H-3 H-4	3.79 3.50 3.58	3.78 3.51	3.79 3.55 3.56 3.64	3.79 3.50 3.57 3.60
	H-3 H-4 H-5 H-6a ^[b] H-6b ^[b]	3.79 3.50 3.58 3.62 3.88	3.78 3.51 3.59 3.61 3.87	3.79 3.55 3.56 3.64 3.85	3.79 3.50 3.57 3.60 3.86
GlcNHR ^{1 [f]}	H-3 H-4 H-5 H-6a ^(b) H-1	3.79 3.50 3.58 3.62 3.88 4.558 (7.9)	3.78 3.51 3.59 3.61 3.87 4.565 (7.9)	3.79 3.55 3.56 3.64 3.85 4.551 (8.0)	3.86 4.571 (7.5)
GlcNHR ^{1 [f]}	H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) H-1 H-2	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71	3.86 4.571 (7.5)
GlcNHR ^{1 [f]}	H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) H-1 H-2 H-3 H-4	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68	3.86 4.571 (7.5) 3.73 3.74 3.64
GlcNHR ^{1 [f]}	H-3 H-6a[b] H-6a[b] H-1 H-2 H-3 H-4	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48
GleNHR ^{1 [f]}	H-3 H-4 H-5 H-6a ^(b) H-0 H-1 H-2 H-3 H-4 H-5 H-6a ^(b)	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84
GlcNHR ^{1 [f]}	H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) H-1 H-2 H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) CH ₂	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32
	H-3 H-4 H-5 H-6b ^(b) H-1 H-2 H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) CH ₂	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84 - 2.047	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133
	H-3 H-6a[b] H-6a[b] H-1 H-2 H-3 H-4 H-5 H-6a[b] H-6b[b] CH ₂ CH ₃ H-1	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84 - 2.045 4.516 (8.4) 3.94	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84 2.047 4.523 (8.4) 3.95	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92
GleNHR ^{1 [f]} GalNHR ^{2[f]}	H-3 H-4 H-5 H-6a ^(b) H-1 H-2 H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) CH ₂ CH ₃ H-1 H-2 H-3	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84 - 2.045 4.516 (8.4) 3.94 3.78	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94 3.75	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.48 - 2.047 4.523 (8.4) 3.95 3.75	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92 3.76
	H-3 H-4 H-5 H-6a ^(b) H-1 H-2 H-3 H-4 H-5 H-6a ^(b) CH ₂ CH ₃ H-1 H-2 H-3 H-4	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84 - 2.045 4.516 (8.4) 3.94 3.78 3.93	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94 3.75 3.93	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84 2.047 4.523 (8.4) 3.95 3.75 3.94	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92 3.76 3.94
	H-3 H-6a ^(b) H-6b ^(b) H-1 H-2 H-3 H-4 H-5 H-6a ^(b) H-6b ^(b) CH ₂ CH ₃ H-1 H-2 H-3 H-4 H-5 H-6a	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84 - 2.045 4.516 (8.4) 3.94 3.78 3.93 3.72 3.78	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94 3.75 3.93 3.73 3.78	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.44 - 2.047 4.523 (8.4) 3.95 3.75 3.94 3.75 3.78	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92 3.76 3.94 3.73 3.81
	H-3 H-4 H-5 H-6a[b] H-6b[b] H-1 H-2 H-3 H-4 H-5 H-6a[b] H-6b[b] CH ₂ CH ₃ H-1 H-2 H-3 H-4 H-5 H-6a	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94 3.75 3.93 3.73 3.78 3.78	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84 2.047 4.523 (8.4) 3.95 3.75 3.94 3.75 3.78	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92 3.76 3.94 3.73 3.81
	H-3 H-6a[b] H-6b[b] H-1 H-2 H-3 H-4 H-5 H-6a[b] H-6b[b] CH ₂ CH ₃ H-1 H-2 H-3 H-4 H-5 H-6a	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84 2.045 4.516 (8.4) 3.94 3.78 3.93 3.72 3.78 3.78	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94 3.75 3.93 3.78 3.78 3.78	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92 3.76 3.94 3.73 3.81 3.81
	H-3 H-4 H-5 H-6a[b] H-6b[b] H-1 H-2 H-3 H-4 H-5 H-6a[b] H-6b[b] CH ₂ CH ₃ H-1 H-2 H-3 H-4 H-5 H-6a	3.79 3.50 3.58 3.62 3.88 4.558 (7.9) 3.73 3.72 3.63 3.47 3.66 3.84 - 2.045 4.516 (8.4) 3.94 3.78 3.93 3.72 3.78	3.78 3.51 3.59 3.61 3.87 4.565 (7.9) 3.73 3.74 3.64 3.48 3.67 3.85 2.32 1.133 4.517 (8.4) 3.94 3.75 3.93 3.73 3.78 3.78	3.79 3.55 3.56 3.64 3.85 4.551 (8.0) 3.71 3.75 3.68 3.46 3.68 3.84 2.047 4.523 (8.4) 3.95 3.75 3.94 3.75 3.78	3.86 4.571 (7.5) 3.73 3.74 3.64 3.48 3.66 3.84 2.32 1.133 4.519 (8.3) 3.92 3.76 3.94 3.73 3.81

[[]a] Data were measured in D_2O at 300 K. Chemical shifts are relative to internal acetone ($\delta=2.225$ ppm). Coupling constants are given in Hertz between parentheses. [b] The assignment of H-6a and H-6b may have to be interchanged within one residue. [c] R=1 acetyl for odd numbers and propionyl for even numbers. [d] $J_{3'',4''}$ in 1 and 2. [e] H-6,6,6 signal and $J_{5'',6''}$ for D-fucose in 3 and 4. [f] I=10 acetyl for 9 and 11, and propionyl for 10 and 12. I=11 acetyl for 9 and 12, and propionyl for 10 and 11.

Table 2. 125 MHz 13 C NMR chemical shifts of the carbohydrate residues in the compounds 1-12

Table 2. (Continued)

Residue ^[a]	Reporter group	1	2	3	4
Man	C-1	97.5	97.5	97.4	98.0
	C-2	77.3	77.4	77.3	78.3
	C-3	70.4	70.4	70.4	70.6
	C-4	67.8	68.0	67.9	67.5
	C-5	73.4	73.5	73.5	73.5
	C-6	62.0	62.1	62.1	61.8
GlcNHR ^[b]	C-1	100.1	100.2	100.1	100.8
	C-2	55.6	55.5	55.5	55.6
	C-3	72.5	72.7	72.6	72.5
	C-4	79.2	79.3	79.8	79.8
	C-5	75.3	75.4	75.3	75.2
	C-6	60.6	60.7	60.6	60.7
	CO	175.2	179.0	175.2	178.5
	CH_2	_	30.0	_	29.9
	CH_3	22.9	9.9	22.9	9.9
Gal	C-1	103.5	103.6	103.6	103.6
	C-2	71.5	71.6	71.3	71.3
	C-3	73.1	73.2	73.3	73.4
	C-4	69.1	69.2	71.8	71.8
	C-5	75.9	76.0	71.7	71.7
_	C-6	61.6	61.6	15.9	16.0
Spacer	OCH_2	68.7	68.7	68.7	68.5
	CH_2	31.7	31.7	31.7	32.1
	CH_2	29.2	29.1	29.1	29.6
	CH_2	29.1	29.0	29.0	29.5
	CH_2	29.0	28.9	28.9	29.4
	CH_2	26.0	26.0	26.0	26.3
	CH_2	22.6	22.6	22.6	22.9
	CH ₃	14.0	14.0	14.0	14.2
Residue	Reporter group	5	6	7	8
) (
Man	C-1	97.5	97.6	97.5	97.8
Man	C-2	97.5 77.5	97.6 77.7	97.5 77.3	97.8 77.9
Man	C-2 C-3	77.5 70.4	77.7 70.4	77.3 70.4	77.9 70.5
Man	C-2 C-3 C-4	77.5 70.4 67.8	77.7 70.4 67.9	77.3 70.4 67.8	77.9 70.5 67.6
Man	C-2 C-3 C-4 C-5	77.5 70.4 67.8 73.5	77.7 70.4 67.9 73.5	77.3 70.4 67.8 73.5	77.9 70.5 67.6 73.5
	C-2 C-3 C-4 C-5 C-6	77.5 70.4 67.8 73.5 62.0	77.7 70.4 67.9 73.5 62.0	77.3 70.4 67.8 73.5 62.1	77.9 70.5 67.6 73.5 62.0
	C-2 C-3 C-4 C-5 C-6 C-1	77.5 70.4 67.8 73.5 62.0 100.2	77.7 70.4 67.9 73.5 62.0 100.4	77.3 70.4 67.8 73.5 62.1 100.2	77.9 70.5 67.6 73.5 62.0 100.6
	C-2 C-3 C-4 C-5 C-6 C-1 C-2	77.5 70.4 67.8 73.5 62.0 100.2 55.8	77.7 70.4 67.9 73.5 62.0 100.4 55.7	77.3 70.4 67.8 73.5 62.1 100.2 55.6	77.9 70.5 67.6 73.5 62.0 100.6 55.7
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6
	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1
	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3
	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1
	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7
	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 - 23.0 101.1	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9
	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 60.4 175.3 - 23.0 103.1 71.5 73.0	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 - 23.0 101.1 34.1 68.3	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 - 23.0 101.1 34.1 68.3 67.3	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 60.4 175.3 - 23.0 103.1 71.5 73.0	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 23.0 101.1 34.1 68.3 67.3 76.2	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 23.0 101.1 34.1 68.3 67.3 76.2 62.0	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3
GlcNHR ^[b] Gal	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7 72.7	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8 72.9	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 23.0 101.1 34.1 68.3 67.3 76.2	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2
GlcNHR ^[b]	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 23.0 103.1 71.5 73.0 69.7 72.7 41.1	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8 72.9 41.1	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 23.0 101.1 34.1 68.3 67.3 76.2 62.0	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2 61.9 68.5 31.9
GlcNHR ^[b] Gal	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 OCH ₂ C-5 C-6 OCH ₂	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7 72.7 41.1 68.7	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8 72.9 41.1 68.7	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 23.0 101.1 34.1 68.3 67.3 76.2 62.0 68.7	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2 61.9 68.5
GlcNHR ^[b] Gal	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 OCH ₂ CH ₂ CH ₂ CH ₂	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7 72.7 41.1 68.7 31.8	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8 72.9 41.1 68.7 31.7	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 23.0 101.1 34.1 68.3 67.3 76.2 62.0 68.7 31.8	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2 61.9 68.5 31.9
GlcNHR ^[b] Gal	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 OCH ₂ CH ₂ CH ₂ CH ₂ CH ₂ CH ₂ CH ₂	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7 72.7 41.1 68.7 31.8 29.3	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8 72.9 41.1 68.7 31.7 29.1	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 - 23.0 101.1 34.1 68.3 67.3 76.2 62.0 68.7 31.8 29.2	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2 61.9 68.5 31.9 29.4
GlcNHR ^[b] Gal	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 OCH ₂ CH ₂	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7 72.7 41.1 68.7 31.8 29.3 29.2	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 9.9 103.1 71.5 73.0 69.8 72.9 41.1 68.7 31.7 29.1 29.1	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 - 23.0 101.1 34.1 68.3 67.3 76.2 62.0 68.7 31.8 29.2 29.2	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 101.1 34.2 68.3 67.3 76.2 61.9 68.5 31.9 29.4 29.4 29.2
GlcNHR ^[b] Gal	C-2 C-3 C-4 C-5 C-6 C-1 C-2 C-3 C-4 C-5 C-6 CO CH ₂ CH ₃ C-1 C-2 C-3 C-4 C-5 C-6 OCH ₂ CH ₂	77.5 70.4 67.8 73.5 62.0 100.2 55.8 72.5 77.5 75.5 60.4 175.3 - 23.0 103.1 71.5 73.0 69.7 72.7 41.1 68.7 31.8 29.3 29.2 29.0	77.7 70.4 67.9 73.5 62.0 100.4 55.7 72.5 77.6 75.5 60.4 179.0 29.9 103.1 71.5 73.0 69.8 72.9 41.1 68.7 31.7 29.1 29.0	77.3 70.4 67.8 73.5 62.1 100.2 55.6 72.6 79.1 75.3 60.7 175.2 - 23.0 101.1 34.1 68.3 67.3 76.2 62.0 68.7 31.8 29.2 29.2 29.0	77.9 70.5 67.6 73.5 62.0 100.6 55.7 72.6 79.1 75.3 60.8 178.7 29.9 9.9 101.1 34.2 68.3 67.3 76.2 61.9 68.5 31.9 29.4 29.4

Residue	Reporter group	9	10	11	12
Man	C-1	97.4	97.4	97.4	97.5
	C-2	77.3	77.3	77.3	77.4
	C-3	70.4	70.4	70.4	70.4
	C-4	67.9	67.9	67.7	67.9
	C-5	73.5	73.5	73.5	73.5
	C-6	62.1	62.1	62.0	62.0
GlcNHR ^{1 [c]}	C-1	100.1	100.1	100.1	100.2
	C-2	55.4	55.2	55.4	55.3
	C-3	72.7	72.8	72.6	72.8
	C-4	79.9	79.6	79.5	79.9
	C-5	75.1	75.2	75.2	75.1
	C-6	60.7	60.8	60.7	60.7
	CO	175.2	179.0	175.2	179.0
	CH_2	_	30.0	_	29.9
	CH ₃	23.0	9.9	23.0	9.9
GalNHR ^{2[c]}	C-1	102.4	102.2	102.3	102.4
	C-2	53.2	53.0	53.0	53.2
	C-3	71.3	71.3	71.3	71.3
	C-4	68.2	68.3	68.3	68.2
	C-5	75.9	75.9	75.9	75.9
	C-6	61.5	61.6	61.6	61.5
	CO	175.2	179.1	178.9	175.3
	CH ₂	_	30.0	29.9	_
	CH ₃	22.8	9.9	9.9	22.8
Spacer	OCH ₂	68.7	68.7	68.6	68.7
•	CH ₂	31.7	31.7	31.9	31.7
	$\overline{\text{CH}_{2}}$	29.1	29.1	29.4	29.1
	$\overline{\text{CH}_2}$	29.1	29.0	29.3	29.1
	CH_2^2	29.0	28.9	29.2	29.0
	CH_2	26.0	26.0	26.2	26.0
	CH_2	22.6	22.6	22.7	22.6
	CH_3^2	14.0	14.0	14.1	14.0

 $^{[a]}$ Data measured in D_2O at 300 K. Chemical shifts are relative to internal acetone ($\delta_{CH3}=30.89$ ppm). $^{[b]}$ R = acetyl for odd numbers and propionyl for even numbers. $^{[c]}$ R 1 = acetyl for 9 and 11, and propionyl for 10 and 12. R^2 = acetyl for 9 and 12, and propionyl for 10 and 11.

azido function (pH = 10, ammonia; conversion into amino) and of the *O*-benzyl groups (pH \approx 5, acetic acid; conversion into hydroxy), yielded 5 (99%). In a similar way, de-*N*-phthaloylation/de-*O*-acetylation and *N*,*O*-propionylation of 31 (\rightarrow 34, 94%), followed by de-*O*-propionylation (\rightarrow 35, 79%), and conversion of the azido function into an amino group combined with the removal of the *O*-benzyl groups, yielded 6 (48%) (Scheme 7). The ¹H and ¹³C NMR spectroscopic data of 5 and 6 are presented in Tables 1 and 2, respectively.

Compounds 7 and 8

For the syntheses of the 2"-deoxy mimics of **1** and **2**, octyl 2-deoxy- β -D-lyxo-hexopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy- β -D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (7) and octyl 2-deoxy- β -D-lyxo-hexopyranosyl- $(1\rightarrow 4)$ -2-deoxy-2-propionamido- β -D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (8) (Scheme 1), respectively, use was made of

Scheme 3. Synthesis of trisaccharide 3: a) Ac_2O , pyridine; b) 4 equiv. $BF_3 \cdot Et_2O$, 1.2 equiv. EtSH, 0 °C, CH_2Cl_2 , molecular sieves (4 Å), 61% over two steps; c) 3.5 equiv. AgOTf, 1.1 equiv. Br_2 , CH_2Cl_2 , toluene, molecular sieves (4 Å), 74%; d) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); e) Ac_2O , pyridine, 85% over two steps; f) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc; g) Ac_2O , pyridine, 72% over two steps; h) NaOMe (pH = 9), CH_2Cl_2 , MeOH, 61%

20 a,b
$$R^{2}O$$
 OR^{2}
 $R^{1}O$
 $R^{1}O$

Scheme 4. Synthesis of trisaccharide 4: a) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); b) Pr_2O , pyridine, 87% over two steps; c) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc; d) NaOMe (PH=9), POH=90, POH=91, POH=92, POH=93, POH=94, PO

Scheme 5. Synthesis of 6-azido galactose donor 30: a) p-TosCl, CH_2Cl_2 , pyridine, 92%; b) NaN_3 , Me_2SO , 160 °C, 99%; c) TFA, H_2O , CH_2Cl_2 ; d) Ac_2O , pyridine, 50% over two steps; e) hydrazinium acetate, DMF, 50 °C, 89%; f) Cl_3CCN , DBU, 0 °C, CH_2Cl_2 , 63%

Scheme 6. Synthesis of trisaccharide 5: a) 17% TMSOTf, CH_2Cl_2 , 0 °C, molecular sieves (4 Å), 61%; b) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); c) Ac_2O , pyridine, 99% over two steps; d) NaOMe (pH = 9), MeOH, 70%; e) 10% Pd/C, H_2 , NH_3 , 2-PrOH, H_2O , HOAc, 99%

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31
$$\xrightarrow{a,b}$$
 RO OR BNO OBN \xrightarrow{BnO} $\xrightarrow{$

Scheme 7. Synthesis of trisaccharide 6: a) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); b) Pr_2O , pyridine, 94% over two steps; c) NaOMe (pH = 9), MeOH, 79%; d) 10% Pd/C, H_2 , NH_3 , 2-PrOH, H_2O , HOAc, 48%

ethyl 2-O-acetyl-3,4,6-tri-O-benzyl-1-thio- β -D-galactopyranoside (36).^[17] Coupling of 36 with 13, using N-iodosuccinimide/trifluoromethanesulfonic acid as the catalytic system, yielded trisaccharide derivative 37 (70%) (Scheme 8). De-N-phthaloylation/de-O-acetylation and N,O-acetylation of 37 (\rightarrow 38, 94%), followed by de-O-acetylation (\rightarrow 39) and treatment with phenyl chlorothionocarbonate in 4-dimethylaminopyridine/acetonitrile,^[18] yielded 40 (61%). 2''-Deoxygenation of 40 with tributylstannane in the presence

of catalytic amounts of α,α' -azoisobutyronitrile (AIBN) at $100 \, ^{\circ}\mathrm{C}^{[18]}$ afforded **41** (70%), which was catalytically de-*O*-benzylated to give **7** (75%). In a similar way, de-*N*-phthaloylation/de-*O*-acetylation and *N*,*O*-propionylation of **37** (\rightarrow **42**, 94%), followed by de-*O*-propionylation (\rightarrow **43**), *O*-phenoxythiocarbonylation at C-2'' (\rightarrow **44**, 43%), 2''-deoxygenation (\rightarrow **45**, 62%), and de-*O*-benzylation yielded **8** (57%) (Scheme 9). The $^{1}\mathrm{H}$ and $^{13}\mathrm{C}$ NMR spectroscopic data of **7** and **8** are presented in Tables 1 and 2, respectively.

Scheme 8. Synthesis of trisaccharide 7: a) 1.4 equiv. NIS, 20% TfOH, CH_2Cl_2 , 0 °C, molecular sieves (4 Å), 70%; b) $NH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); c) Ac_2O , pyridine, 94% over two steps; d) NaOMe (pH = 10), CH_2Cl_2 , MeOH, 40 °C; e) $C_6H_5OC[S]Cl$, DMAP, CH_3CN , 100 °C, 61% over two steps; f) Bu_3SnH , AIBN, 100 °C, toluene, 70%; g) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc, 75%

Scheme 9. Synthesis of trisaccharide 8: a) NH₂CH₂CH₂NH₂, 90 °C, 1-BuOH, molecular sieves (3 Å); b) Pr₂O, pyridine, 94% over two steps; c) NaOMe (pH = 10), CH₂Cl₂, MeOH, 40 °C; d) C₆H₅OC[S]Cl, DMAP, CH₃CN, 100 °C, 43% over two steps; e) Bu₃SnH, AIBN, 100 °C, toluene, 62%; f) 10% Pd/C, H₂, HOAc, EtOH, EtOAc, 57%

FULL PAPER

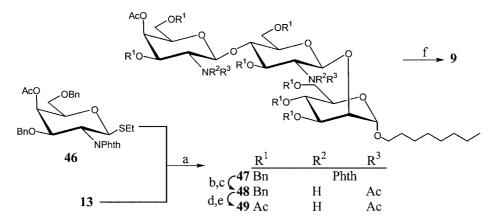
J. P. Kamerling et al.

Compounds 9 and 10

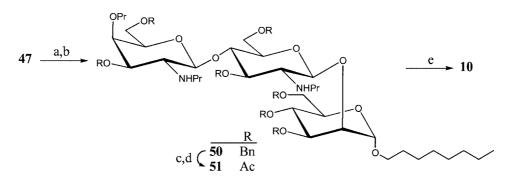
For the syntheses of the 2''-acylamido-2''-deoxy mimics of 1 and 2, octyl 2-acetamido-2-deoxy-β-D-galactopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy- β -D-glucopyranosyl- $(1\rightarrow 2)$ α-D-mannopyranoside (9) and octyl 2-deoxy-2-propionamido- β -D-galactopyranosyl- $(1\rightarrow 4)$ -2-deoxy-2-propionamido- β -D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (10) (Scheme 1), respectively, the earlier reported donor ethyl 4-O-acetyl-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1thio-β-D-galactopyranoside (46)^[19] was used. This fragment was synthesised from a known glucosamine building block, [20] which was applied in the synthesis of disaccharide 13.^[10,11] Compound 46 was coupled to acceptor 13, using bromine/silver trifluoromethanesulfonate as the catalytic system, to yield trisaccharide derivative 47 (51%) (Scheme 10). De-N-phthaloylation/de-O-acetylation and N,O-acetylation of 47 (\rightarrow 48, 61%), followed by de-Obenzylation and O-acetylation (\rightarrow 49, 86%), and final de-O-acetylation yielded 9 (56%). In a similar way, de-Nphthaloylation/de-O-acetylation and N,O-propionylation of 47 (\rightarrow 50, 80%), followed by de-O-benzylation and O-acetylation (\rightarrow 51, 92%), and final de-O-acylation yielded 10 (91%) (Scheme 11). It should been noted that two analogues of 9, β -D-GalpNAc- $(1\rightarrow 4)$ - β -D-GlcpNAc- $(1\rightarrow 2)$ - α -D- $Manp-(1\rightarrow O)(CH_2)_8COOCH_3^{[21]}$ and β-D-GalpNAc- $(1\rightarrow 4)$ - β -D-GlcpNAc- $(1\rightarrow 2)$ - α -D-Manp- $(1\rightarrow O)(CH_2)_2$ - CH₃,^[22] have been synthesised earlier along similar routes. The ¹H and ¹³C NMR spectroscopic data of **9** and **10** are presented in Tables 1 and 2, respectively.

Compounds 11 and 12

The syntheses of the 2"-acylamido-2"-deoxy mimics of 1 and 2, octyl 2-deoxy-2-propionamido-β-D-galactopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy- β -D-glucopyranosyl- $(1\rightarrow 2)$ α-D-mannopyranoside (11) and octyl 2-acetamido-2-deoxy- β -D-galactopyranosyl-(1 \rightarrow 4)-2-deoxy-2-propionamido-β-Dglucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (Scheme 1), respectively, required in addition to the phthalimido function for the glucosamine unit in acceptor 13, the selection of another N-protective group for the galactosamine donor. Introductory experiments with donors containing an N-acetyl or an N-(tetrachlorophthalimido)^[23,24] protective group gave rise to irreproducible results only. Testing of the promising N-(2,2,2-trichloroethoxycarbonyl) protective group^[25-27] in terms of yields, showed problems in the deprotection using activated zinc; [28] here also primary Obenzyl groups were removed, leading to mixtures that could not be fractionated. The N-(dimethylmaleoyl) protective group^[29,30] turned out to be the best choice. According to the earlier reported route for D-glucosamine hydrochloride, [29,30] D-galactosamine hydrochloride (52) was neutralised with sodium methoxide in methanol, then N-acylated



Scheme 10. Synthesis of trisaccharide 9: a) 0.5 equiv. Br_2 , 1.2 equiv. AgOTf, CH_2Cl_2 , toluene, molecular sieves (4 Å), 51%; b) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); c) Ac_2O , pyridine, 61% over two steps; d) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc; e) Ac_2O , pyridine, 86% over two steps; f) NaOMe (pH = 10), MeOH, 56%



Scheme 11. Synthesis of trisaccharide 10: a) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH, molecular sieves (3 Å); b) Pr_2O , pyridine, 80% over two steps; c) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc; d) Ac_2O , pyridine, 92% over two steps; e) NaOMe (pH = 10), MeOH, 91%

OH OH OH
$$a,b$$
 OAC OAC A CO OAC A

Scheme 12. Synthesis of galactose donor 55: a) 1 equiv. NaOMe, DMMA, NEt₃, 60 °C, MeOH; b) Ac₂O, pyridine, 31% over two steps; c) hydrazinium acetate, DMF, 61%; d) CCl₃CN, DBU, CH₂Cl₂, 59%

Scheme 13. Synthesis of trisaccharide 11: a) 10% TMSOTf, 0 °C, CH_2Cl_2 , molecular sieves (4 Å), 84%; b) NaOH, HCl, dioxane, H_2O ; c) Pr_2O , pyridine, 33% over two steps; d) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH; e) Ac_2O , pyridine, 74% over two steps; f) NaOMe (pH = 9), CH_2Cl_2 , MeOH; g) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc; h) Ac_2O , pyridine, 72% over three steps; i) NaOMe (pH = 10), CH_2Cl_2 , MeOH, 77%

with dimethylmaleic anhydride in the presence of triethylamine at 60 °C, and O-acetylated to give 53 (31%) (Scheme 12). Subsequent de-O-acetylation at C-1 using hydrazinium acetate in dimethylformamide (\rightarrow 54, 61%) and imidation with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene, yielded donor 55 (59%). Coupling of 55 [N-(dimethylmaleoyl) function] with 13 (N-phthaloyl group), using trimethylsilyl trifluoromethanesulfonate as a catalyst, afforded trisaccharide derivative 56 (84%) (Scheme 13). De-N-(dimethylmaleoyl)ation and de-O-

acetylation of **56** by sequential incubation with sodium hydroxide in aqueous dioxane (alkaline pH), and, after acidification with HCl at pH = 3,[29,30] followed by N,O-propionylation yielded **57** (33%). De-N-phthaloylation/de-O-propionylation and N,O-acetylation of **57** (\rightarrow **58**, 74%), followed by de-O-acetylation, de-O-benzylation, and O-acetylation (\rightarrow **59**, 72%), and final de-O-acetylation yielded **11** (77%). The last O-acetylation step was carried out to facilitate chromatographic purification. In a similar way, de-N-(dimethylmaleoyl)ation/de-O-acetylation and N,O-acetylation/de-O-acetylation and N,O-acetylation/de-O-acetylation and N,O-acetylation/de-O-acetylation and N,O-acetylation/de-O-acetylation and N,O-acetylation/de-O-acetylation and O-acetylation/de-O-acetylation and O-acetylation/de-O-acetylation and O-acetylation/de-O-acetylation and O-acetylation/de-O-acetylation and O-acetylation/de-O-acetylation and O-acetylation/de-O-acetylation and O-acetylation/de-

Scheme 14. Synthesis of trisaccharide 12: a) NaOH, HCl, AcCl, dioxane, H_2O ; b) Ac_2O , pyridine, 48% over two steps; c) $NH_2CH_2CH_2NH_2$, 90 °C, 1-BuOH; d) Pr_2O , pyridine, 80% over two steps; e) NaOMe (pH = 9), CH_2CI_2 , MeOH; f) 10% Pd/C, H_2 , HOAc, EtOH, EtOAc; g) Ac_2O , pyridine, 67% over three steps; h) NaOMe (pH = 9), CH_2CI_2 , MeOH, 84%

lation of **56** (\rightarrow **60**, 48%) (Scheme 14), followed by de-*N*-phthaloylation/de-*O*-acetylation and *N*,*O*-propionylation (\rightarrow **61**, 80%), and de-*O*-propionylation, de-*O*-benzylation, and *O*-acetylation (\rightarrow **62**, 67%), and final de-*O*-acetylation yielded **12** (84%). The ¹H and ¹³C NMR spectroscopic data of **11** and **12** are presented in Tables 1 and 2, respectively. The NMR spectra of **11** and **12** are almost identical, except for the *N*-acetyl methyl signal in the ¹H NMR spectra. The NAc signal in **11** resonates at δ = 2.047 ppm, and in **12** at δ = 2.068 ppm. Note that in **9** the two NAc signals resonate at δ = 2.045 and 2.067 ppm.

Conclusion

Summarising, to prepare a series of relevant trisaccharide mimics for enzymatic sialyl-transfer studies, the strategy of condensing a series of galactosyl-modified monosaccharide donors with a general glucosaminyl-mannosyl acceptor has proven to be successful. In earlier work, we have shown that such an approach leads to better results than preparing disaccharide donors by introducing modifications at the galactosyl-glucosaminyl level, followed by coupling to a fixed mannosyl acceptor.[12] In the present approach, monosaccharide thioglycosides were used for coupling of synthons containing besides acyl groups also benzyl groups or a deoxy function (19, 36, 46), and trichloroacetimidates for coupling of acylated deactivated synthons (14, 30, 55). For the activation of the thioglycosides, bromine/silver trifluoromethanesulfonate gave the best results for 19 and 46, whereby 19 needed more activation than 46, and N-iodosuccinimide/trifluoromethanesulfonic acid for 36. TMSOTf gave always the best results in trichloroacetimidate couplings.

The trisaccharides 1-12 will be used in α -2,3- and α -2,6-sialyltransferase as well as in *trans*-sialidase kinetic studies in order to investigate the relevance in the substrate/enzyme interaction of the replaced hydroxy functions in the terminal galactose unit of 1 in combination with the effect of replacing the *N*-acetyl function of the middle glucosamine unit by an *N*-propionyl function.

Experimental Section

General: All solvents used were distilled from appropriate drying agents. In the workup procedures of reaction mixtures, organic solutions were washed with appropriate amounts of aqueous solutions as indicated. Solutions were concentrated under reduced pressure at 40 °C (water bath). Reactions were monitored by TLC on Silica Gel 60 F₂₅₄ (Merck), compounds were visualized under UV light, and by charring with either 10% ethanolic H₂SO₄ or 0.2% orcinol in 20% methanolic H₂SO₄. Column chromatography was performed on Kieselgel 60 F₂₅₄ (Merck, 0.063-0.200 mm). Size-exclusion chromatography was carried out on Sephadex LH-20 or Bio-Gel P-2. ¹H (300 MHz) and ¹³C NMR spectra (75.5 MHz) of protected compounds were recorded at 300 K using a Bruker AC 300 spectrometer; only relevant NMR spectroscopic data are included in the experimental procedures. Two-dimensional ¹H-¹H correlated spectra (TOCSY, ROESY) and ¹H-¹³C correlated spectra (HSQC) of compounds 1-12, 59 and 62 were recorded at 300 K using a Bruker

AMX 500 spectrometer. ¹H NMR chemical shifts (δ_H) are given in ppm relative to the signal for internal Me₄Si ($\delta_H = 0$ ppm) for solutions in CDCl₃ or by reference to acetone ($\delta_H = 2.225$ ppm) for solutions in D_2O . ¹³C NMR chemical shifts (δ_C) are relative to the signal for CDCl₃ ($\delta_C = 76.9$ ppm) for solutions in CDCl₃ or by reference to acetone ($\delta_C = 30.89$ ppm) for solutions in D₂O. J values are given in Hz. Optical rotations were determined for solutions in CHCl₃ or H₂O at 20 °C with a Perkin-Elmer 241 polarimeter, using a 10-cm 1-mL cell. Fast-atom-bombardment mass spectrometry (FABMS) was performed with a JEOL JMS SX/SX 102A four-sector mass spectrometer, operated at 10 kV accelerating voltage, equipped with a JEOL MS-FAB 10 D FAB gun operated at 10 mA emission current, producing a beam of 6 keV Xenon atoms. MALDI-TOF mass spectra were recorded with a Voyager-DE (Per-Septive Biosystems) instrument using dihydroxybenzoic acid (DHB) as a matrix. The matrix was dissolved in a 1:1 (v/v) mixture of acetonitrile/H₂O (10 mg DHB/mL), and the sample was dissolved in acetone (5 mg/mL). Subsequently, $0.5 \mu L$ of matrix solution and $0.5 \mu L$ of sample solution were brought on the sample plate of the mass spectrometer. Spectra were generated by summing positive-ion signals of 256 laser shots with constant intensity. Exact masses were measured by nano electrospray time-of-flight mass spectrometry using a Micromass LCToF mass spectrometer at a resolution of 5000 FWHM. Gold-coated capillaries were loaded with 1 µL of sample (conc. 20 µM), dissolved in a 1:1 (v/v) mixture of acetonitrile/H₂O and 0.1% formic acid. Pentafluorophenylalanine was added as internal standard. The capillary voltage was set at 1500 V and the cone voltage was set at 30 V.

Octyl (2,3,4,6-Tetra-O-acetyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-propionamido-β-D-glucopyranosyl)-(1→2)-3,4,6tri-*O*-benzyl-α-D-mannopyranoside (16): To a solution of octyl (2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -(3,6-di-Obenzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl-α-D-mannopyranoside (15; 246 mg, 0.18 mmol)^[10] in $CH_2Cl_2/MeOH$ (20 mL, 3:1) was added NaOMe (pH = 9), and the mixture was stirred for 4 h. After neutralisation with Dowex 50×8 (H⁺), and filtration, the solution was concentrated. A solution of the residue in 1-BuOH (20 mL), containing molecular sieves 3 Å (0.75 g), was stirred for 30 min under Ar, then 1,2-diaminoethane (2.2 mL, 33 mmol) was added. The mixture was stirred at 90 °C overnight, filtered through Celite, and co-concentrated with toluene. The residue was dissolved in pyridine/propionic anhydride (10 mL, 1:1) and stirred overnight, then co-concentrated with toluene. A solution of the residue in MeOH (20 mL) was treated with NaOMe (pH = 9) for 4 h, then neutralised with Dowex 50×8 (H⁺), filtered, and concentrated. The residue was dissolved in pyridine/acetic anhydride (10 mL, 1:1) and stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 3:1) of the residue yielded 16, isolated as a syrup (172 mg, 72%). TLC (toluene/ EtOAc, 1:1): $R_f = 0.69$. $[\alpha]_D^{20} = +3$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.00 (t, 3 H, COCH₂CH₃), 1.26 (m, 10 H, 5 octyl CH₂), 1.50 (m, 2 H, octyl CH₂), 1.97, 1.98, 2.01, and 2.08 (4 s, each 3 H, 4 COCH₃), 2.15 (q, 2 H, COCH₂CH₃), 3.13 and 3.42 (2 m, each 1 H, octyl OCH₂), 4.11 (d, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.12 (dd, 1 H, 2''-H), 5.17 (d, $J_{1',2'} = 7.7$ Hz, 1 H, 1'-H), 5.18 (d, $J_{2',NH} = 7.7$ Hz, 1 H, NH), 5.28 (d, $J_{3'',4''} =$ 2.9, $J_{4'',5''}$ < 1 Hz, 1 H, 4''-H), 7.45-7.10 (m, 25 H, 5 OCH₂C₆H₅) ppm. MS (FAB⁺) of $C_{72}H_{91}NO_{20}$ (1289): found m/z = 1290 [M + H]⁺, 1312 [M + Na]⁺.

Octyl (2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-O-acetyl-2-deoxy-2-propionamido- β -D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-O-acetyl- α -D-mannopyranoside (17): To a solution of 16 (172 mg,

1.13 mmol) in EtOH/EtOAc (20 mL, 1:1) were added 10% Pd/C (140 mg) and HOAc (0.25 mL), and the mixture was stirred for 2.5 h under H₂, then filtered through Celite, and concentrated. A solution of the residue in pyridine/acetic anhydride (22 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 1:3) of the residue yielded 17, isolated as a syrup (88 mg, 65%). TLC (toluene/EtOAc, 1:3): $R_f = 0.14$. $[\alpha]_D^{20} =$ -7 (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.08 (t, 3 H, COCH₂CH₃), 1.28 (m, 10 H, 5 octyl CH₂), 1.59 (m, 2 H, octyl CH₂), 1.96, 1.99, 2.02, 2.04, 2.06, 2.07, 2.11, and 2.14 (8 s, 3, 3, 3, 3, 3, 6, 3, 3 H, 9 COCH₃), 2.15 (m, 2 H, $COCH_2CH_3$), 3.40 (m, 1 H, octyl OCHH), 4.39 (dd, $J_{5',6'a} = 2.5$, $J_{6'a,6'b} = 11.8 \text{ Hz}, 1 \text{ H}, 6'a\text{-H}), 4.48 \text{ (d, } J_{1'',2''} = 7.7 \text{ Hz}, 1 \text{ H}, 1''\text{-}$ H), 4.69 (d, $J_{1,2} = 1.4$ Hz, 1 H, 1-H), 4.73 (d, $J_{1',2'} = 7.7$ Hz, 1 H, 1'-H), 4.96 (dd, $J_{2'',3''} = 10.4$, $J_{3'',4''} = 3.3$ Hz, 1 H, 3''-H), 5.15 (dd, 1 H, 4-H), 5.29 (dd, 1 H, 3'-H), 5.34 (d, $J_{4'',5''}$ < 1 Hz, 1 H, 4"-H), 5.69 (d, $J_{2',NH} = 8.5 \text{ Hz}$, 1 H, NH) ppm. ¹³C NMR (75.5 MHz, CDCl₃): $\delta = 9.4$ (COCH₂CH₃), 13.8 (octyl CH₃), 20.3–20.6 (COCH₃), 22.4, 25.9, 28.9, 29.0, 29.1, 29.4, and 31.6 (6 octyl CH₂, COCH₂CH₃), 54.0 (C-2'), 60.7, 62.6, 62.7, and 68.1 (C-6, C-6', C-6'', octyl OCH₂), 66.0, 66.5, 68.3, 68.9, 70.0, 70.5, 70.6, 71.4, 72.5, 74.6, and 75.9 (C-2, C-3, C-4, C-5, C-3', C-4', C-5', C-2", C-3", C-4", C-5"), 97.2, 99.2, and 100.7 (C-1, C-1', C-1"), 168.9, 169.3, 169.8, 169.9, 170.0 (2 C), 170.1, 170.2, and 170.5 (9 COCH₃), 173.8 (COCH₂CH₃) ppm. MS (FAB⁺) of C₄₇H₇₁NO₂₅ (1049): found $m/z = 1050 \,[M + H]^+$, $1072 \,[M + Na]^+$.

Octyl β-D-Galactopyranosyl-(1 \rightarrow 4)-2-deoxy-2-propionamido-β-D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranoside (2): To a solution of 17 (88 mg, 84 μmol) in MeOH/CH₂Cl₂ (10 mL, 4:1) was added NaOMe (pH = 9), and the mixture was stirred for 2.5 h. After neutralisation with Dowex 50 × 8 (H⁺) and filtration, the solution was concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with H₂O, and subsequent lyophilization yielded **2**, isolated as a white powder (53 mg, 94%). TLC (CH₂Cl₂/MeOH/H₂O, 5:10:3): $R_{\rm f} = 0.88$. [α]²⁰_D = -2 (c = 1, H₂O). High-resolution MS of C₂₉H₅₃NO₁₆ (671.3364): calcd. 694.3262, found 694.3265 [M + Na]. For ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2, respectively.

Ethyl 2,3,4-Tri-O-acetyl-1-thio-\(\beta\)-fucopyranoside (19): A solution of D-fucose (18; 936 mg, 5.7 mmol) in pyridine/acetic anhydride (20 mL, 1:1) was stirred for 7 h, then co-concentrated with toluene. The residue was dissolved in dry CH₂Cl₂ (10 mL), containing molecular sieves 4 Å (200 mg), and ethanethiol (507 µL, 6.84 mmol) was added. The mixture was stirred under Ar for 60 min, then cooled to 0 °C, and BF₃·Et₂O (2.86 mL, 22.8 mmol) was added. After 100 min, during which period the temperature was allowed to reach room temperature, the mixture was neutralised with NEt3, and filtered. The filtrate was diluted with CH₂Cl₂, washed with water and aq. saturated NaHCO₃, dried (MgSO₄), and concentrated. Column chromatography (toluene/EtOAc, 4:1) of the residue yielded 19, isolated as a white foam (1.16 g, 61%), and the α -product (700 mg, 36%). TLC (toluene/EtOAc, 4:1): $R_f = 0.42$. $[\alpha]_D^{20} = -28$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.94$ (d, $J_{5.6} = 6.3$ Hz, 3 H, 6-H), 1.02 (t, 3 H, SCH₂CH₃), 1.70, 1.79, and 1.89 (3 s, each 3 H, 3 COCH₃), 2.37–2.57 (m, 2 H, SCH₂CH₃), 3.63 (m, 1 H, 5-H), 4.26 (d, $J_{1,2} = 9.7$ Hz, 1 H, 1-H), 4.83 (dd, $J_{2,3} = 10.0$, $J_{3,4} = 3.2$ Hz, 1 H, 3-H), 4.95 (t, 1 H, 2-H), 5.01 (dd, $J_{4,5} < 1$ Hz, 1 H, 4-H) ppm. MS (FAB⁺) of $C_{14}H_{22}O_7S$ (334): found $m/z = 335 \text{ [M + H]}^+$, 357 $[M + Na]^+$.

Octyl (2,3,4-Tri-O-acetyl- β -D-fucopyranosyl)-(1 \rightarrow 4)-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranoside (20): A solution of 19 (109 mg, 325

μmol) and 13 (250 mg, 242 μmol) in dry CH₂Cl₂ (10 mL), containing powdered molecular sieves 4 Å (0.3 g), was stirred for 3.5 h under Ar, then a solution of AgOTf (296 mg, 1.15 mmol) in dry toluene (4.3 mL) was added, followed after 20 min by a solution of Br₂ (371 μmol) in dry CH₂Cl₂ (0.69 mL). After additional stirring for 60 min, the mixture was neutralised with NEt₃ and filtered through Celite. The filtrate was diluted with CH₂Cl₂, washed with aq. 10% NaS₂O₃, aq. saturated NaHCO₃, and water, dried (MgSO₄), filtered, and concentrated. Column chromatography (toluene/EtOAc, 3:1) of the residue afforded 20, isolated as a syrup (233 mg, 74%). TLC (toluene/ EtOAc, 3:1): $R_f = 0.56$. $[\alpha]_D^{20} = -6$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.87$ (t, 3 H, octyl CH₃), 1.06 (d, $J_{5'',6''} =$ 6.0 Hz, 3 H, 6"-H), 1.23 (br. s, 10 H, 5 octyl CH₂), 1.42 (m, 2 H, octyl CH₂), 1.96, 2.01, and 2.07 (3 s, each 3 H, 3 COCH₃), 2.97 and 3.19 (2 m, each 1 H, octyl OCH₂), 5.26 (d, $J_{1',2'} = 7.4$ Hz, 1 H, 1'-H), 6.80-7.42 (m, 25 H, 5 OCH₂C₆H₅), 7.45-7.70 (m, 4 H, Phth) ppm. MS (FAB⁺) of $C_{75}H_{87}NO_{19}$ (1305): found m/z = 1306 [M + H]⁺, 1328 [M + Na]⁺.

Octvl (2,3,4-Tri-O-acetvl- β -D-fucopyranosyl)-(1 \rightarrow 4)-(2-acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-Obenzyl-α-D-mannopyranoside (21): A solution of 20 (72 mg, 55 μmol) in 1-BuOH (8 mL), containing molecular sieves 3 Å (0.25 g), was stirred under Ar for 30 min, then 1,2-diaminoethane (0.74 mL, 11 mmol) was added. The mixture was stirred overnight at 90 °C, filtered through Celite, and co-concentrated with toluene and EtOH. A solution of the residue in pyridine/acetic anhydride (6 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, $3:1 \rightarrow 1:1$) of the residue yielded 21, isolated as a syrup (58 mg, 85%). TLC (toluene/EtOAc, 3:1): $R_{\rm f} = 0.29$. $[\alpha]_{\rm D}^{20} = -3$ (c = 0.8, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.07 (d, $J_{5''.6''} = 6.0$ Hz, 3 H, 6"-H), 1.26 (m, 10 H, 5 octyl CH₂), 1.50 (m, 2 H, octyl CH₂), 1.74 (s, 3 H, NHCOCH₃), 1.96, 1.97, and 2.11 (3 s, each 3 H, 3 COCH₃), 4.13 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.73 (d, $J_{2',NH} = 8.7$ Hz, 1 H, NH), 7.10-7.58 (m, 25 H, 5 OCH₂C₆H₅) ppm. MS (FAB⁺) of $C_{69}H_{87}NO_{18}$ (1217): found $m/z = 1218 [M + H]^+$, 1240 $[M + Na]^+$.

(2,3,4-Tri-O-acetyl-β-D-fucopyranosyl)-(1→4)-(2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-Oacetyl-α-D-mannopyranoside (22): To a solution of 21 (106 mg, 86 μmol) in EtOH/EtOAc (15 mL, 1:1) were added 10% Pd/C (48 mg) and HOAc (165 µL), and the mixture was stirred overnight under H₂, then filtered through Celite, and concentrated. A solution of the residue in pyridine/acetic anhydride (43 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 1:3) of the residue yielded 22, isolated as a syrup (61 mg, 72%). TLC (toluene/EtOAc, 1:3): $R_f = 0.17$. $[\alpha]_D^{20} = -21$ $(c = 1, CHCl_3)$. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.19 (d, $J_{5'',6''} = 6.4$ Hz, 3 H, 6''-H), 1.30 (m, 10 H, 5 octyl CH₂), 1.58 (m, 2 H, octyl CH₂), 1.95, 1.98, 2.01, 2.03, 2.08, 2.10, and 2.15 (7 s, 3, 3, 3, 6, 6, 3, 3 H, 9 COCH₃), 3.41 (m, 1 H, octyl OCHH), 4.44 (d, $J_{1'',2''} = 7.8$ Hz, 1 H, 1''-H), 4.63 (d, $J_{1',2'} =$ 7.4 Hz, 1 H, 1'-H), 4.71 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.68 (d, $J_{2',NH} =$ 8.7 Hz, 1 H, NH) ppm. MS (FAB+) of C₄₄H₆₇NO₂₃ (977): found $m/z = 978 [M + H]^+, 1000 [M + Na]^+.$

Octyl β-D-Fucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranoside (3): To a solution of 22 (61 mg, 62 μmol) in MeOH/CH₂Cl₂ (8 mL, 3:1) was added NaOMe (pH = 9), and the mixture was stirred for 5 h. After neutralisation with Dowex 50 \times 8 (H⁺) and filtration, the solution was concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, and subsequent lyophilization yielded 3, isolated as a white powder (24 mg, 61%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1):

 $R_{\rm f}=0.32.~ [\alpha]_{\rm D}^{20}=-7~(c=1,~{\rm H_2O}).$ High-resolution MS of $C_{28}H_{51}NO_{15}$ (641.3259): calcd. 664.3156, found 664.3170 [M + Na]. For $^1{\rm H}$ and $^{13}{\rm C}$ NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (2,3,4-Tri-O-acetyl-β-D-fucopyranosyl)-(1→4)-(3,6-di-O-acetyl-2-deoxy-2-propionamido-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-*O*acetyl-α-D-mannopyranoside (24): A solution of 20 (161 mg, 123 μmol) in 1-BuOH (17 mL), containing molecular sieves 3 Å (0.55 g), was stirred for 30 min under Ar, then 1,2-diaminoethane (1.66 mL, 25 mmol) was added. The mixture was stirred at 90 °C overnight, filtered through Celite, and co-concentrated with toluene and EtOH. A solution of the residue in pyridine/propionic anhydride (12 mL, 1:1) was stirred overnight, then co-concentrated with toluene and EtOH. Column chromatography (toluene/EtOAc, 3:1) of the residue yielded 23, isolated as a syrup (138 mg, 87%). To a solution of 23 in EtOH/EtOAc (16 mL, 1:1) were added 10% Pd/C (60 mg) and HOAc (0.2 mL), and the mixture was stirred overnight under H₂, then filtered through Celite, and concentrated. To a solution of the residue in MeOH (15 mL) was added NaOMe (pH = 9), and the mixture was stirred for 6 h, then neutralised with Dowex 50 \times 8 (H⁺), and concentrated. The residue was dissolved in pyridine/acetic anhydride (16 mL, 1:1) and stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 1:2) of the residue yielded 24, isolated as a syrup (58 mg, 55%). TLC (toluene/ EtOAc, 1:2): $R_f = 0.25$. $[\alpha]_D^{20} = -18$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.89$ (t, 3 H, octyl CH₃), 1.11 (t, 3 H, $COCH_2CH_3$), 1.21 (d, $J_{5'',6''} = 5.8$ Hz, 3 H, 6''-H), 1.33 (m, 10 H, 5 octyl CH₂), 1.59 (m, 2 H, octyl CH₂), 1.89-2.16 (m, 26 H, 8 $COCH_3$, $COCH_2CH_3$), 3.39 (m, 1 H, octyl OCHH), 4.42 (d, $J_{1'',2''} =$ 7.8 Hz, 1 H, 1''-H), 4.62 (d, $J_{1',2'} = 7.3$ Hz, 1 H, 1'-H), 4.67 (s, $J_{1,2}$ $< 1 \text{ Hz}, 1 \text{ H}, 1\text{-H}), 5.60 (d, J_{2',NH} = 7.4 \text{ Hz}, 1 \text{ H}, NH) ppm. MS$ (FAB^+) of $C_{45}H_{69}NO_{23}$ (991): found $m/z = 992 [M + H]^+$, 1014 [M $+ Na]^+$.

Octyl β-D-Fucopyranosyl-(1 \rightarrow 4)-2-deoxy-2-propionamido-β-D-glucopyranosyl-(1 \rightarrow 2)-α-D-mannopyranoside (4): To a solution of 24 (58 mg, 58 μmol) in MeOH (10 mL) was added NaOMe (pH = 9), and the mixture was stirred for 24 h. After neutralisation with Dowex 50 \times 8 (H⁺) and filtration, the solution was concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, and subsequent lyophilization yielded 4, isolated as a white powder (36 mg, 93%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1): $R_{\rm f} = 0.38$. [α] $_{\rm D}^{\rm 20} = -1$ (c = 1, H₂O). High-resolution MS of C₂₉H₅₃NO₁₅ (655.3415): calcd. 678.3313, found 678.3331 [M + Na]. For $^{\rm 1}$ H and $^{\rm 13}$ C NMR spectroscopic data, see Tables 1 and 2, respectively.

6-Azido-6-deoxy-1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (27): To a solution of 1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (25; 1.08 g, 4.15 mmol) in pyridine (10 mL) was added dropwise a solution of p-toluenesulfonyl chloride (1.3 g, 6.3 mmol) in CH₂Cl₂ (10 mL), and the mixture was stirred for 5 h. After the addition of water (1 mL), the mixture was stirred for 10 min, then coconcentrated with toluene. A solution of the residue in CH₂Cl₂ was washed with water and aq. NaHCO₃, dried (MgSO₄), and concentrated. Column chromatography (toluene/EtOAc, 5:1) of the residue yielded 26, isolated as a syrup (1.59 g, 92%). To a solution of 26 (1.31 g, 3.17 mmol) in Me₂SO (30 mL) was added NaN₃ (1 g, 15.7 mmol), and the mixture was stirred for 2 h at 160 °C, then cooled to room temperature, poured into iced water, extracted with EtOAc, and the organic phase was dried (MgSO₄), and concentrated to yield 27, isolated as a syrup (900 mg, 99%). TLC (toluene/EtOAc, 3:1): $R_f = 0.62$. IR (KBr): $\tilde{v} = 2110 \text{ cm}^{-1} \text{ (N_3)}$. $[\alpha]_D^{20} = +43 \text{ (}c =$ 1, CHCl₃). ¹H NMR (300 MHz, [D₆]Me₂SO): $\delta = 1.31, 1.32, 1.43,$ and 1.52 (4 s, each 3 H, 4 COCH₃), 3.33 (dd, $J_{5,6a} = 5.4$, $J_{6a,6b} = 12.7$ Hz, 1 H, 6a-H), 3.47 (dd, $J_{5,6b} = 7.8$ Hz, 1 H, 6b-H), 3.89 (ddd, 1 H, 5-H), 4.17 (dd, $J_{3,4} = 7.9$, $J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.30 (dd, $J_{1,2} = 5.0$, $J_{2,3} = 2.5$ Hz, 1 H, 2-H), 4.60 (dd, 1 H, 3-H), 5.52 (d, 1 H, 1-H) ppm. MS (FAB⁺) of $C_{12}H_{19}N_3O_5$ (285): found m/z = 286 [M + H]⁺, 308 [M + Na]⁺.

2,3,4-Tri-O-acetyl-6-azido-6-deoxy-α-D-galactopyranosyl roacetimidate (30): To a solution of 27 (900 mg, 3.15 mmol) in CH₂Cl₂ (30 mL) were added trifluoroacetic acid (5.6 mL, 73 mmol) and water (0.7 mL), and the mixture was stirred for 2 h, then coconcentrated with toluene. A solution of the residue in pyridine/ acetic anhydride (20 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 3:1) of the residue yielded 28, isolated as a syrup (583 mg, 50%). To a solution of 28 in DMF (3 mL) was added hydrazinium acetate (158 mg, 1.72 mmol), and the solution was stirred for 70 min at 50 °C, then diluted with EtOAc, washed with water and aq. saturated NaCl, dried (MgSO₄), and concentrated to give 29, isolated as a foam (461 mg, 89%). To a solution of 29 (461 mg, 1.4 mmol) in CH₂Cl₂ (8 mL) was added, at 0 °C, trichloroacetonitrile (1.4 mL, 14 mmol) and 1,8-diazabicyclo[5.4.0]undec-7-ene (104 μL, 0.7 mmol), and the mixture was stirred for 1 h, then concentrated. Column chromatography (toluene/EtOAc, 4:1) of the residue yielded 30, isolated as a syrup (380 mg, 63%). TLC (toluene/EtOAc, 4:1): $R_{\rm f} = 0.33$. $[\alpha]_{\rm D}^{20} = +68$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.94$, 1.97, and 2.12 (3 s, each 3 H, 3 COCH₃), 3.16 (dd, $J_{5,6a} = 4.9$, $J_{6a,6b} = 12.9$ Hz, 1 H, 6a-H), 3.39 (dd, $J_{5,6b} =$ 7.7 Hz, 1 H, 6b-H), 4.30 (m, 1 H, 5-H), 5.28 (dd, $J_{1,2} = 3.3$, $J_{2,3} =$ 10.8 Hz, 1 H, 2-H), 5.35 (dd, $J_{3,4} = 2.9$ Hz, 1 H, 3-H), 5.47 (dd, $J_{4.5} = 1.1 \text{ Hz}, 1 \text{ H}, 4\text{-H}, 6.55 (d, 1 \text{ H}, 1\text{-H}), 8.60 (s, 1 \text{ H}, NH) ppm.$ MS (FAB⁺) of $C_{14}H_{17}Cl_3N_4O_8$ (474): found $m/z = 475 \text{ [M + H]}^+$, $497 [M + Na]^+$

(2,3,4-Tri-O-acetyl-6-azido-6-deoxy-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl- α -D-mannopyranoside (31): A solution of 13 (260 mg, 251 μmol) and **30** (195 mg, 450 μmol) in CH₂Cl₂ (12.5 mL), containing molecular sieves 4 Å (50 mg), was stirred under Ar at 0 °C for 2.5 h, then TMSOTf (17 µL, 80 µmol) was added. After stirring for 35 min, the mixture was neutralised with NEt₃, filtered, diluted with CH₂Cl₂, washed with water, dried (MgSO₄), and concentrated. Column chromatography (toluene/EtOAc, 3:1) of the residue yielded 31, isolated as a syrup (206 mg, 61%). TLC (toluene/EtOAc, 3:1): $R_f = 0.59$. $[\alpha]_D^{20} = +1$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.90$ (t, 3 H, octyl CH₃), 1.25 (m, 10 H, 5 octyl CH₂), 1.46 (m, 2 H, octyl CH₂), 2.00, 2.05, and 2.10 (3 s, each 3 H, 3 COCH₃), 4.67 (d, $J_{1'',2''}$ = 8.0 Hz, 1 H, 1''-H), 5.17 (dd, $J_{2'',3''}$ = 10.4 Hz, 1 H, 2''-H), 5.28 (d, $J_{1',2'} = 7.6$ Hz, 1 H, 1'-H), 5.28 (d, $J_{3'',4''} = 3.0, J_{4'',5''} < 1 \text{ Hz}, 1 \text{ H}, 4''-\text{H}), 6.90-7.48 \text{ (m, 25 H, 5)}$ $OCH_2C_6H_5$), 7.51-7.68 (2 m, each 2 H, Phth) ppm. MS (FAB⁺) of $C_{75}H_{86}N_4O_{19}$ (1346): found $m/z = 1347 [M + H]^+$, 1369 $[M + Na]^+$.

Octyl (2,3,4-Tri-*O*-acetyl-6-azido-6-deoxy-β-D-galactopyranosyl)-(1 \rightarrow 4)-(2-acetamido-3,6-di-*O*-benzyl-2-deoxy-β-D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl- α -D-mannopyranoside (32): A solution of 31 (103 mg, 76 μmol) in 1-BuOH (10 mL), containing molecular sieves 3 Å (0.3 g), was stirred under Ar for 30 min, then 1,2-diaminoethane (1.05 mL, 15.7 mmol) was added. The mixture was stirred overnight at 90 °C, filtered through Celite, and co-concentrated with toluene and EtOH. A solution of the residue in pyridine/acetic anhydride (30 mL, 1:1) was stirred overnight, then co-concentrated with toluene and EtOH. Column chromatography (toluene/EtOAc, 2:1) of the residue yielded 32, isolated as a syrup (96 mg, 99%). TLC (toluene/EtOAc, 2:1): $R_f = 0.50$. [α] $_D^{20} = +5$ (c = 1, CHCl₃). ¹H NMR

(300 MHz, CDCl₃): δ = 0.88 (t, 3 H, octyl CH₃), 1.27 (m, 10 H, 5 octyl CH₂), 1.51 (m, 2 H, octyl CH₂), 1.72 (s, 3 H, NHCOC*H*₃), 1.96, 2.01, and 2.10 (3 s, each 3 H, 3 COCH₃), 4.11 (d, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.23 (d, $J_{3'',4''}$ = 3.3, $J_{4'',5''} < 1$ Hz, 1 H, 4''-H), 5.69 (d, $J_{2',NH}$ = 7.1 Hz, 1 H, NH), 7.15–7.45 (m, 25 H, 5 OCH₂C₆*H*₅) ppm. MS (FAB+) of C₆₉H₈₆N₄O₁₈ (1258): found m/z = 1259 [M + H]⁺, 1281 [M + Na]⁺.

Octyl (6-Azido-6-deoxy-β-D-galactopyranosyl)-(1→4)-(2-acetamido-3,6-di-O-benzyl-2-deoxy-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl- α -D-mannopyranoside (33): To a solution of 32 (96 mg, 76 μmol) in MeOH (15 mL) was added NaOMe (pH = 9), and the mixture was stirred for 2 h. After neutralisation with Dowex 50 × 8 (H⁺), and filtration, the solution was concentrated. Column chromatography (CH₂Cl₂/MeOH, 95:5) of the residue yielded 33, isolated as a syrup (61 mg, 70%). TLC (CH₂Cl₂/MeOH, 95:5): R_f = 0.77. [α] $_D^{20}$ = -8 (c = 1, MeOH). $_1^1$ H NMR (300 MHz, CDCl₃): δ = 0.90 (t, 3 H, octyl CH₃), 1.27 (m, 10 H, 5 octyl CH₂), 1.52 (m, 2 H, octyl CH₂), 1.66 (s, 3 H, NHCOC H_3), 3.04 (m, 1 H, octyl OCHH), 4.13 (d, $J_{1,2}$ < 1 Hz, 1 H, 1-H), 5.13 (d, $J_{1',2'}$ = 7.3 Hz, 1 H, 1'-H), 5.87 (d, $J_{2',NH}$ = 7.1 Hz, 1 H, NH), 7.25 - 7.40 (m, 25 H, 5 OCH₂C₆ H_5) ppm. MS (FAB⁺) of C₆₃H₈₀N₄O₁₅ (1132): found m/z = 1133 [M + H] $^+$, 1155 [M + Na] $^+$.

Octyl 6-Amino-6-deoxy-β-D-galactopyranosyl-(1→4)-2-acetamido-2deoxy-β-D-glucopyranosyl-(1 \rightarrow 2)-α-D-mannopyranoside (5): To a solution of 33 (61 mg, 53 µmol) in 2-propanol (6 mL) and water (4 mL) were added 10% Pd/C (160 mg) and aq. 25% NH₃ (pH = 10), and the mixture was stirred under H_2 for 3 h. After the removal of NH₃ by bubbling N₂ through the mixture, the solution was acidified with HOAc, stirred under H₂ for 23 h, filtered through Celite, and concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, yielded crude 5. The residue was dissolved in water (1 mL), and applied on a SepPak C₁₈ cartridge that was eluted first with water, then with MeOH. The MeOH fraction was concentrated and lyophilized to yield 5, isolated as a white powder (34.5 mg, 99%). TLC $(1-\text{BuOH/EtOH/HOAc/H}_2\text{O}, 4:2:2:1)$: $R_f =$ 0.16. $[\alpha]_D^{20} = -15$ ($c = 1, H_2O$). High-resolution MS of $C_{28}H_{52}N_2O_{15}$ (656.3368): calcd. 657.3446, found 657.3441 [M + H]. For ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (6-Azido-6-deoxy-2,3,4-tri-*O*-propionyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-propionamido-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl-α-D-mannopyranoside (34): A solution of 31 (103 mg, 76 µmol) in 1-BuOH (10 mL), containing molecular sieves 3 Å (0.3 g), was stirred under Ar for 30 min, then 1,2diaminoethane (1.05 mL, 15.7 mmol) was added. The mixture was stirred at 90 °C overnight, filtered through Celite, and co-concentrated with toluene and EtOH. A solution of the residue in pyridine/ propionic anhydride (30 mL, 1:1) was stirred overnight, then co-concentrated with toluene and EtOH. Column chromatography (toluene/EtOAc, 4:1) of the residue yielded 34, isolated as a syrup (91 mg, 94%). TLC (toluene/EtOAc, 3:1): $R_f = 0.56$. $[\alpha]_D^{20} = +7$ (c = 1, CHCl₃). ${}^{1}H$ NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 0.94-1.23 (m, 12 H, 3 COCH₂CH₃, NHCOCH₂CH₃), 1.29 (m, 10 H, 5 octyl CH₂), 1.51 (m, 2 H, octyl CH₂), 1.95 (m, 2 H, NHCOC H_2 CH₃), 2.18–2.53 (m, 6 H, 3 COC H_2 CH₃), 4.12 (d, $J_{1,2}$ $< 1 \text{ Hz}, 1 \text{ H}, 1\text{-H}), 4.87 \text{ (dd, } J_{2'',3''} = 10.3, J_{3'',4''} = 3.1 \text{ Hz}, 1 \text{ H},$ 3''-H), 5.14 (dd, $J_{1'',2''}$ = 8.0 Hz, 1 H, 2''-H), 5.18 (d, $J_{1',2'}$ = 7.7 Hz, 1 H, 1'-H), 5.24 (d, $J_{4'',5''}$ < 1 Hz, 1 H, 4''-H), 5.74 (d, $J_{2',NH}$ = 6.8 Hz, 1 H, NH), 7.13-7.42 (m, 25 H, 5 OCH₂C₆H₅) ppm. MS (FAB^+) of $C_{73}H_{94}N_4O_{18}$ (1314): found $m/z = 1315 [M + H]^+$, 1337 $[M + Na]^+$.

Octyl 6-Amino-6-deoxy- β -D-galactopyranosyl- $(1\rightarrow 4)$ -2-deoxy-2-propionamido- β -D-glucopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranoside (6): To a

solution of 34 (91 mg, 71 µmol) in MeOH (15 mL) was added Na-OMe (pH = 9), and the mixture was stirred for 2.5 h. After neutralisation with Dowex 50×8 (H⁺) and filtration, the solution was concentrated. Column chromatography (CH2Cl2/MeOH, 9:1) of the residue yielded 35, isolated as a syrup (65 mg, 79%). To a solution of 35 (65 mg, 53 µmol) in 2-propanol/water (10 mL, 3:2) were added 10% Pd/C (160 mg) and aq. 25% NH₃ (pH = 9), and the mixture was stirred under H₂ for 3.5 h. After the removal of NH₃ by bubbling N₂ through the mixture, the solution was acidified with HOAc, stirred under H₂ for 23 h, filtered through Celite, and concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, yielded crude 6. The residue was dissolved in water (1 mL), and applied on a SepPak C18 cartridge that was eluted first with water, then with MeOH. The MeOH fraction was concentrated and lyophilized to yield 6, isolated as a white powder (18 mg, 48%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1): $R_f = 0.15$. $[\alpha]_D^{20} = -1$ (c = 1, H_2O). High-resolution MS of $C_{29}H_{54}N_2O_{15}$ (670.3524): calcd. 671.3602, found 671.3597 [M + H]. For ${}^{1}H$ and ${}^{13}C$ NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (2-O-Acetyl-3,4,6-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl-α-D-mannopyranoside (37): A solution of ethyl 2-O-acetyl-3,4,6-tri-O-benzyl-1-thio-β-D-galactopyranoside (486 mg, 906 μmol) and 13 (541 mg, 522 μmol) in CH₂Cl₂ (25 mL), containing powdered molecular sieves 4 Å (0.5 g), was stirred under Ar for 3 h. The mixture was cooled to 0 °C, then N-iodosuccinimide (289 mg, 1.28 mmol) and triflic acid (16 μ L, 181 μ mol) were added. After 40 min, the solution was neutralised with NEt₃, filtered, diluted with CH₂Cl₂, washed with aq. 10% NaHSO₃, aq. saturated NaHCO₃, and water, dried (MgSO₄), and concentrated. Column chromatography (toluene/EtOAc, 8:1) of the residue yielded 37, isolated as a syrup (548 mg, 70%). TLC (toluene/EtOAc, 6:1): $R_{\rm f}$ = 0.61. $[\alpha]_D^{20} = +4$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta =$ 0.88 (t, 3 H, octyl CH₃), 1.22 (m, 10 H, 5 octyl CH₂), 1.41 (m, 2 H, octyl CH₂), 1.98 (s, 3 H, COCH₃), 2.75 and 3.19 (2 m, each 1 H, octyl OCH₂), 5.23 (d, $J_{1',2'}$ = 8.0 Hz, 1 H, 1'-H), 5.38 (dd, $J_{1'',2''}$ = 7.7, $J_{2'',3''} = 9.9 \,\text{Hz}$, 1 H, 2''-H), 6.75-7.38 (m, 40 H, 8 $OCH_2C_6H_5$), 7.45-7.70 (m, 4 H, Phth) ppm. MS (FAB⁺) of $C_{92}H_{101}NO_{18}$ (1507): found $m/z = 1508 [M + H]^+$, 1530 [M + Na]⁺.

Octyl (2-O-Acetyl-3,4,6-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-(2acetamido-3,6-di-O-benzyl-2-deoxy-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl-\alpha-D-mannopyranoside (38): A solution of 37 (238 mg, 158 µmol) in 1-BuOH (22 mL), containing molecular sieves 3 Å (0.2 g), was stirred under Ar for 1 h, then 1,2-diaminoethane (2.0 mL, 30 mmol) was added. The mixture was stirred overnight at 90 °C, filtered through Celite, and co-concentrated with toluene. A solution of the residue in pyridine/acetic anhydride (60 mL, 1:1) was stirred overnight, then co-concentrated with toluene to give 38, isolated as a syrup (211 mg, 94%). TLC (toluene/ EtOAc, 3:1): $R_f = 0.29$. $[\alpha]_D^{20} = -1$ (c = 1, CHCl₃). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.87 \text{ (t, 3 H, octyl CH}_3), 1.25 \text{ (m, 10 H, 5)}$ octyl CH₂), 1.52 (m, 2 H, octyl CH₂), 1.76 (s, 3 H, NHCOCH₃), 1.96 (s, 3 H, COCH₃), 4.11 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.33 (dd, $J_{1'',2''} = 8.1$, $J_{2'',3''} = 10.1$ Hz, 1 H, 2''-H), 5.82 (d, $J_{2',NH} =$ 7.8 Hz, 1 H, NH), 7.10-7.45 (m, 40 H, 8 OCH₂C₆H₅) ppm. MS (FAB^{+}) of $C_{86}H_{101}NO_{17}$ (1419): found $m/z = 1420 [M + H]^{+}$, 1442 $[M + Na]^+$.

Octyl (3,4,6-Tri-O-benzyl-2-O-phenoxythiocarbonyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(2-acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranoside (40): To a solution of 38 (211 mg, 148 μ mol) in MeOH/CH₂Cl₂ (100 mL, 3:1) was added NaOMe (pH = 10), and the mixture was stirred at 40

°C for 17 h, then neutralised with Dowex 50×8 (H⁺), filtered, and concentrated to give 39. To a solution of the residue in acetonitrile added 4-(dimethylamino)pyridine (25 mL)were (825 mg, 6.75 mmol) and phenyl chlorothionocarbonate (540 μL , 3.90 mmol), and the mixture was stirred at 100 °C for 4.5 h, then at room temperature overnight. After dilution with CH2Cl2, the solution was washed with aq. 0.5 m HCl, aq. saturated NaHCO₃, and water, dried (MgSO₄), and concentrated. Column chromatography (toluene/EtOAc, 4:1) of the residue yielded 40, isolated as a syrup (139 mg, 61%). TLC (toluene/EtOAc, 4:1): $R_f = 0.50$. $[\alpha]_D^{20} =$ $-7 (c = 1, CHCl_3)$. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.91 (t, 3)$ H, octyl CH₃), 1.31 (m, 10 H, 5 octyl CH₂), 1.58 (m, 2 H, octyl CH₂), 1.73 (s, 3 H, NHCOCH₃), 3.22 (m, 1 H, octyl OCHH), 4.13 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.18 (d, $J_{1',2'} = 7.1$ Hz, 1 H, 1'-H), 5.66 (d, $J_{2',NH} = 7.1 \text{ Hz}$, 1 H, NH), 5.95 (dd, $J_{1'',2''} = 7.8$, $J_{2'',3''} =$ 10.0 Hz, 1 H, 2''-H), 7.00-7.50 (m, 45 H, 9 OCH₂C₆ H_5) ppm. MS (FAB^+) of $C_{91}H_{103}NO_{17}S$ (1513): found $m/z = 1514 [M + H]^+$, $1536 [M + Na]^{+}$.

Octyl (2-Deoxy-3,4,6-tri-*O*-benzyl- β -D-*lyxo*-hexopyranosyl)-(1 \rightarrow 4)-(2-acetamido-3.6-di-O-benzyl-2-deoxy-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl-α-D-mannopyranoside (41): To a solution of 40 (139 mg, 91 µmol) in dry toluene (3 mL) was added tributylstannane (295 µL, 1.1 mmol), and the mixture was stirred at 100 °C. A catalytic amount of α,α' -azoisobutyronitrile was added, and the mixture was stirred at 100 °C for 2 h, then concentrated. A solution of the residue in acetonitrile was extracted with hexane $(3 \times)$, and concentrated. Column chromatography (toluene/EtOAc, 3:1) of the residue yielded 41, isolated as a syrup (87 mg, 70%). TLC (toluene/ EtOAc, 3:1): $R_f = 0.53$. $[\alpha]_D^{20} = -5$ (c = 1, CHCl₃). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.88 \text{ (t, 3 H, octyl CH}_3)$, 1.26 (m, 10 H, 5 octyl CH₂), 1.50 (m, 2 H, octyl CH₂), 1.67 (s, 3 H, NHCOCH₃), 1.90-2.11 (m, 2 H, 2_{ax} "-H, 2_{eq} "-H), 3.12 (m, 1 H, octyl OC*H*H), 4.13 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.11 (d, $J_{1',2'} = 7.7$ Hz, 1 H, 1'-H), 5.61 (d, $J_{2',NH} = 6.8 \text{ Hz}$, 1 H, NH), 7.10-7.45 (m, 40 H, 8 $OCH_2C_6H_5$) ppm. MS (FAB⁺) of $C_{84}H_{99}NO_{15}$ (1361): found $m/z = 1362 [M + H]^+, 1384 [M + Na]^+.$

Octyl 2-Deoxy-β-D-*lyxo*-hexopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2)-α-D-mannopyranoside (7): To a solution of 41 (107 mg, 79 μmol) in EtOH/EtOAc (14 mL, 1:1) were added 10% Pd/C (45 mg) and 6 drops of HOAc, and the mixture was stirred under H₂ for 21 h, then filtered through Celite, and concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, and subsequent lyophilization yielded 7, isolated as a white powder (38 mg, 75%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1): $R_{\rm f} = 0.29$. [α] $_{\rm D}^{20} = -9$ (c = 1, H₂O). Highresolution MS of C₂₈H₅₁NO₁₅ (643.3259): calcd. 664.3156, found 664.3175 [M + H]. For $_{\rm I}^{\rm I}$ H and $_{\rm I}^{\rm I}$ C NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (3,4,6-Tri-*O*-benzyl-2-*O*-phenoxythiocarbonyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-*O*-benzyl-2-deoxy-2-propionamido-β-D-glucopyranosyl)-(1→2)-3,4,6-tri-*O*-benzyl-α-D-mannopyranoside (44): A solution of 37 (310 mg, 205 μmol) in 1-BuOH (28 mL), containing molecular sieves 3 Å (1 g), was stirred under Ar for 1 h, then 1,2-diaminoethane (2.6 mL, 39 mmol) was added. The mixture was stirred at 90 °C overnight, filtered through Celite, and co-concentrated with toluene. A solution of the residue in pyridine/propionic anhydride (60 mL, 1:1) was stirred overnight, then co-concentrated with toluene and EtOH. Column chromatography (toluene/EtOAc, 2:1) of the residue yielded 42 (280 mg, 94%), which was immediately employed in the next reaction. To a solution of 42 (280 mg, 193 μmol) in MeOH/CH₂Cl₂ (75 mL, 2:1) was added NaOMe (pH = 10), and the solution was stirred at 40 °C overnight, then

neutralised with Dowex 50×8 (H⁺), filtered, and concentrated. To a solution of the residue (43) in dry acetonitrile (22 mL) were added 4-(dimethylamino)pyridine (825 mg, 6.75 mmol) and phenyl chlorothionocarbonate (534 $\mu L,\,3.86\,mmol),$ and the mixture was stirred at 100 °C for 2 h, then overnight at room temperature. After dilution with CH₂Cl₂, the solution was washed with aq. 0.5 M HCl, aq. saturated NaHCO3, and water, dried (MgSO4), and concentrated. Column chromatography (toluene/EtOAc, 5:1) of the residue yielded 44, isolated as a syrup (128 mg, 43%). TLC (toluene/ EtOAc, 5:1): $R_f = 0.61$. $[\alpha]_D^{20} = -2$ (c = 1, CHCl₃). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.88 \text{ (t, 3 H, octyl CH}_3), 0.97 \text{ (t, 3 H, octyl CH}_3)$ NHCOCH₂CH₃), 1.26 (m, 10 H, 5 octyl CH₂), 1.52 (m, 2 H, octyl CH₂), 1.82-2.05 (m, 2 H, NHCOCH₂CH₃), 3.17 (m, 1 H, octyl OC*H*H), 4.13 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.18 (d, $J_{1',2'} = 7.9$ Hz, 1 H, 1'-H), 5.62 (d, $J_{2',NH} = 6.8$ Hz, 1 H, NH), 5.90 (dd, $J_{1'',2''} =$ 7.8, $J_{2'',3''} = 9.9 \,\text{Hz}$, 1 H, 2''-H), 6.92-7.47 (m, 45 H, 9 $OCH_2C_6H_5$) ppm. MS (FAB⁺) of $C_{92}H_{105}NO_{17}S$ (1527): found $m/z = 1528 [M + H]^+, 1550 [M + Na]^+.$

Octvl (2-Deoxy-3,4,6-tri-*O*-benzyl- β -D-*lyxo*-hexopyranosyl)-(1 \rightarrow 4)-(3.6-di-O-benzyl-2-deoxy-2-propionamido-β-D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl- α -D-mannopyranoside (45): To a solution of 44 (127 mg, 83 µmol) in dry toluene (3 mL) was added tributylstannane (270 µL, 1.0 mmol), and the mixture was stirred at 100 °C. A catalytic amount of α,α' -azoisobutyronitrile was added and the mixture was stirred at 100 °C for 2 h, then concentrated. A solution of the residue in acetonitrile was extracted with hexane (3 ×), and concentrated. Column chromatography (toluene/EtOAc, 4:1) of the residue yielded 45, isolated as a syrup (71 mg, 62%). TLC (toluene/EtOAc, 4:1): $R_f = 0.45$. $[\alpha]_D^{20} = -1$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 0.98 (t, 3 H, NHCOCH₂CH₃), 1.25 (m, 10 H, 5 octyl CH₂), 1.50 (m, 2 H, octyl CH₂), 1.82–2.12 (m, 4 H, NHCOC H_2 CH₃, 2_{ax} ''-H, 2_{eq} ''-H), 3.10 (m, 1 H, octyl OC*H*H), 4.12 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.15 (d, $J_{1',2'}$ = 7.6 Hz, 1 H, 1'-H), 5.63 (d, $J_{2',NH}$ = 6.8 Hz, 1 H, NH), 7.10-7.43 (m, 40 H, 8 OCH₂C₆H₅) ppm. MS (FAB⁺) of $C_{85}H_{101}NO_{15}$ (1375): found $m/z = 1376 [M + H]^+$, 1398 [M +

Octyl 2-Deoxy-β-D-*lyxo*-hexopyranosyl-(1 \rightarrow 4)-2-deoxy-2-propionamido-β-D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranoside (8): To a solution of 45 (70 mg, 51 μmol) in EtOH/EtOAc (9 mL, 1:1) were added 10% Pd/C (30 mg) and 6 drops of HOAc, and the mixture was stirred under H₂ overnight, then filtered through Celite, and concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, and subsequent lyophilization yielded 8, isolated as a white powder (19 mg, 57%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1): $R_{\rm f}=0.44$. [α] $_{\rm D}^{20}=-14$ (c=1, H₂O). Highresolution MS of C₂₉H₅₃NO₁₅ (655.3415): calcd. 678.3313, found 678.3326 [M + Na]. For $^{\rm 1}$ H and $^{\rm 13}$ C NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (4-*O*-Acetyl-3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-β-D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl- α -D-mannopyranoside (47): A solution of ethyl 4-*O*-acetyl-3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-galactopyranoside^[19] (46; 228 mg, 0.40 mmol) and 13 (169 mg, 0.16 mmol) in dry CH₂Cl₂ (5 mL), containing powdered molecular sieves 4 Å (0.46 g), was stirred under Ar for 4 h. Then a solution of AgOTf (83 mg, 0.32 mmol) in dry toluene (1 mL) was added and the stirring was continued for 20 min. After the addition of a solution of Br₂ (0.21 mmol) in dry CH₂Cl₂ (0.5 mL) and stirring for 1 h, a second solution of AgOTf (43 mg, 0.17 mmol) in dry toluene (0.5 mL) was added. When TLC (toluene/EtOAc, 5:1) showed the reaction to be complete, the mix-

ture was neutralised with NEt₃, filtered through Celite, diluted with CH₂Cl₂, washed with aq. 10% NaHSO₃, aq. saturated NaHCO₃, and water, dried (MgSO₄), and concentrated. Column chromatography (toluene/EtOAc, 5:1) of the residue yielded **47**, isolated as a white foam (135 mg, 51%). TLC (toluene/EtOAc, 5:1): $R_{\rm f} = 0.58$. [α] $_{\rm f}^{20} = -3$ (c = 1, CHCl₃). 1 H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.27 (m, 10 H, 5 octyl CH₂), 1.60 (m, 2 H, octyl CH₂), 2.05 (s, 3 H, COCH₃), 2.98 and 3.10 (2 m, each 1 H, octyl OCH₂), 3.97 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 5.13 (d, $J_{1',2'} = 7.5$ Hz, 1 H, 1'-H), 5.37 (d, $J_{1'',2''} = 8.2$ Hz, 1 H, 1''-H), 5.64 (d, $J_{3'',4''} = 2.9$, $J_{4'',5''} < 1$ Hz, 1 H, 4''-H), 6.85–7.91 (m, 43 H, 2 Phth, 7 OCH₂C₆ H_5) ppm. MS (FAB⁺) of C₉₃H₉₈N₂O₁₉ (1546): found m/z = 1547 [M + H] $^+$, 1569 [M + Na] $^+$.

Octyl (2-Acetamido-4-O-acetyl-3,6-di-O-benzyl-2-deoxy-β-D-galactopyranosyl)-(1→4)-(2-acetamido-3,6-di-O-benzyl-2-deoxy-β-Dglucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-O-benzyl- α -D-mannopyranoside (48): A solution of 47 (186 mg, 120 μmol) in 1-BuOH (17 mL), containing molecular sieves 3 Å (0.5 g), was stirred under Ar for 30 min, then 1,2-diaminoethane (1.7 mL, 25.3 mmol) was added. The mixture was stirred overnight at 90 °C, filtered through Celite, and co-concentrated with toluene, EtOH and CH2Cl2. A solution of the residue in pyridine/acetic anhydride (10 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 2:3) of the residue yielded 48, isolated as a white foam (102 mg, 61%). TLC (toluene/EtOAc, 2:3): $R_{\rm f} = 0.59$. $[\alpha]_{D}^{20} = -2 (c = 1, \text{CHCl}_3).$ ¹H NMR (300 MHz, CDCl₃): $\delta = 0.91$ (t, 3 H, octyl CH₃), 1.29 (m, 10 H, 5 octyl CH₂), 1.53 (m, 2 H, octyl CH₂), 1.77 and 1.82 (2 s, each 3 H, 2 NHCOCH₃), 2.02 (s, 3 H, COCH₃), 4.13 (s, $J_{1.2} < 1$ Hz, 1 H, 1-H), 5.06 (d, $J_{1'.2'} = 7.5$ Hz, 1 H, 1'-H), 5.12 (d, $J_{1'',2''}$ = 8.0 Hz, 1 H, 1''-H), 5.59 (dd, $J_{3'',4''}$ = 2.9, $J_{4'',5''}$ < 1 Hz, 1 H, 4"-H), 5.87 (d, 2 H, 2 NH), 7.18-7.39 (m, 35 H, 7 OCH₂C₆ H_5) ppm. MS (FAB⁺) of C₈₁H₉₈N₂O₁₇ (1370): found $m/z = 1371 [M + H]^+, 1393 [M + Na]^+.$

Octyl (2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy-\beta-D-galactopyranosyl)-(1→4)-(2-acetamido-3,6-di-O-acetyl-2-deoxy-β-D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-O-acetyl- α -D-mannopyranoside (49): To a solution of 48 (102 mg, 74 µmol) in EtOH/EtOAc (7 mL, 1:1) were added 10% Pd/C (62 mg) and 3 drops of HOAc. The mixture was stirred under H₂ overnight, then filtered through Celite, and concentrated. A solution of the residue in pyridine/acetic anhydride (10 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (CH₂Cl₂/MeOH, 9:1) of the residue yielded 49, isolated as a syrup (67 mg, 86%). TLC (CH₂Cl₂/MeOH, 9:1): $R_f = 0.59$. $[\alpha]_D^{20} = -2$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.85$ (t, 3 H, octyl CH₃), 1.25 (m, 10 H, 5 octyl CH₂), 1.54 (m, 2 H, octyl CH₂), 1.90, 1.91, 1.94, 1.96, 1.99, 2.02, 2.05, 2.06, 2.07, and 2.11 (10 s, each 3 H, 8 COCH₃, 2 NHCOCH₃), 3.37 (m, 1 H, octyl OCHH), 5.05 (dd, $J_{2'',3''} = 9.9$, $J_{3'',4''} = 3.3$ Hz, 1 H, $3^{\prime\prime}\text{-H}),~6.10$ and 6.27 (2 d, each 1 H, 2 NH) ppm. ^{13}C NMR $(75.5 \text{ MHz}, \text{CDCl}_3)$: $\delta = 13.9 \text{ (octyl CH}_3), 20.5 - 20.8 \text{ (CO}_{3})$ 23.0 and 23.1 (NHCOCH₃), 22.5, 26.0, 29.0, 29.2, 29.3, and 31.6 (6 octyl CH₂), 51.6 and 54.1 (C-2', C-2"), 61.0, 62.6, 62.7, and 68.2 (C-6, C-6', C-6", octyl OCH2), 97.4, 99.0, and 100.5 (C-1, C-1', C-1"), 169.3, 170.0, 170.1, 170.2 (3 C), 170.3, 170.5 (2 C), and 170.7 (COCH₃, NHCOCH₃) ppm. MS (FAB⁺) of C₄₆H₇₀N₂O₂₄ (1034): found $m/z = 1035 [M + H]^+$, 1057 $[M + Na]^+$.

Octyl 2-Acetamido-2-deoxy-β-D-galactopyranosyl- $(1\rightarrow 4)$ -2-acetamido-2-deoxy-β-D-glucopyranosyl- $(1\rightarrow 2)$ -α-D-mannopyranoside (9): To a solution of 49 (65 mg, 63 μmol) in MeOH (4 mL) was added NaOMe (pH = 10), and the mixture was stirred overnight. After neutralisation with Dowex 50 × 8 (H⁺) and filtration, the solution was concentrated. Gel-filtration through a Bio-Gel P-2

column, eluted with water, and subsequent lyophilization yielded 9, isolated as a white foam (24.9 mg, 56%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1): $R_{\rm f}=0.39$. [α] $_{\rm D}^{\rm 20}=-6$ (c=0.5, H₂O). Highresolution MS of C₃₀H₅₄N₂O₁₆ (698.3473): calcd. 721.3371, found 721.3351 [M + Na]. For $^{\rm 1}$ H and $^{\rm 13}$ C NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (3,6-Di-O-acetyl-2-deoxy-2-propionamido-4-O-propionyl-β-Dgalactopyranosyl)-(1→4)-(3,6-di-O-acetyl-2-deoxy-2-propionamidoβ-D-glucopyranosyl)-(1→2)-3,4,6-tri-*O*-acetyl-α-D-mannopyranoside (51): A solution of 47 (138 mg, 89 μmol) in 1-BuOH (13 mL), containing molecular sieves 3 Å (0.4 g), was stirred under Ar for 30 min, then 1,2-diaminoethane (1.3 mL, 18.6 mmol) was added. The mixture was stirred at 90 °C overnight, filtered through Celite, and co-concentrated with toluene and EtOH. A solution of the residue in pyridine/propionic anhydride (10 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 5:1) of the residue yielded 50, isolated as a white foam (100 mg, 80%). TLC (CH₂Cl₂/MeOH, 9:1): $R_f = 0.90$. To a solution of 50 (100 mg, 71 μmol) in EtOH/EtOAc (7 mL, 1:1) were added 10% Pd/C (102 mg) and 5 drops of HOAc, and the mixture was stirred under H2 overnight, then filtered through Celite and concentrated. A solution of the residue in pyridine/acetic anhydride (10 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (CH₂Cl₂/MeOH, 9:1) of the residue yielded 51, isolated as a syrup (71 mg, 92%). TLC (CH₂Cl₂/ MeOH, 9:1): $R_{\rm f} = 0.70$. $[\alpha]_{\rm D}^{20} = -7$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.85$ (t, 3 H, octyl CH₃), 1.02–1.16 (m, 9 H, COCH₂CH₃, 2 NHCOCH₂CH₃), 1.26 (m, 10 H, 5 octyl CH₂), 1.55 (m, 2 H, octyl CH₂), 1.92-2.12 (m, 27 H, 7 COCH₃, COCH₂CH₃, 2 NHCOCH₂CH₃), 3.39 (m, 1 H, octyl OCHH), 5.07 (dd, $J_{2'',3''} = 9.9$, $J_{3'',4''} = 3.3$ Hz, 1 H, 3''-H), 5.93-6.05 (m, 2 H, 2 NH) ppm. MS (FAB⁺) of $C_{49}H_{76}N_2O_{24}$ (1076): found m/z = $1077 [M + H]^+, 1099 [M + Na]^+.$

Octyl 2-Deoxy-2-propionamido-β-D-galactopyranosyl-(1 \rightarrow 4)-2-deoxy-2-propionamido-β-D-glucopyranosyl-(1 \rightarrow 2)-α-D-mannopyranoside (10): To a solution of 51 (71 mg, 66 μmol) in MeOH (5 mL) was added NaOMe (pH = 10), and the mixture was stirred overnight. After neutralisation with Dowex 50 × 8 (H⁺) and filtration, the solution was concentrated. Gel-filtration of the residue on a Bio-Gel P-2 column, eluted with water, and subsequent lyophilization yielded 10, isolated as a white foam (44 mg, 91%). TLC (1-BuOH/EtOH/HOAc/H₂O, 4:2:2:1): $R_{\rm f}=0.32$. [α] $_{\rm D}^{\rm 20}=-12$ (c=1, H₂O). High-resolution MS of C₃₂H₅₈N₂O₁₆ (726.3786): calcd. 749.3684, found 749.3681 [M + Na]. For $^{\rm 1}$ H and $^{\rm 13}$ C NMR spectroscopic data, see Tables 1 and 2, respectively.

1,3,4,6-Tetra-O-acetyl-2-deoxy-2-(dimethylmaleimido)-β-D-galactopyranose (53): To a solution of D-galactosamine HCl (52; 2.0 g, 9.3 mmol) in MeOH (69 mL) was added NaOMe (0.5 g, 9.3 mmol) and dimethylmaleic anhydride (0.6 g, 4.8 mmol), and the mixture was stirred at 60 °C for 30 min. Then, NEt₃ (0.93 mL) and dimethylmaleic anhydride (0.6 g, 4.8 mmol) were added, and the stirring was continued at 60 °C for 1.5 h. After concentration of the mixture, a solution of the residue in pyridine/acetic anhydride (28.5 mL, 2:1) was stirred overnight, then co-concentrated with toluene. A solution of the residue in CH₂Cl₂ (50 mL) was washed with aq. 3% HCl, aq. saturated NaHCO₃, and water, dried (MgSO₄), filtered, and concentrated. Column chromatography (hexane/EtOAc, 1:1) of the residue vielded 53, isolated as a white foam (1.3 g, 31%). TLC (hexane/EtOAc, 1:1): $R_f = 0.30$. $[\alpha]_D^{20} =$ $+48 (c = 1, CHCl_3)$. ¹H NMR (300 MHz, CDCl₃): $\delta = 1.91, 1.98$, 2.04, and 2.17 (4 s, 3, 6, 6, 3 H, 2 CH₃, 4 COCH₃), 4.41 (dd, $J_{1,2}$ = 8.9, $J_{2,3} = 11.4 \text{ Hz}$, 1 H, 2-H), 5.47 (d, $J_{3,4} = 3.4$, $J_{4,5} < 1 \text{ Hz}$, 1

H, 4-H), 5.77 (dd, 1 H, 3-H), 6.28 (d, 1 H, 1-H) ppm. MS (MALDI-TOF) of $C_{20}H_{25}NO_{11}$ (455): found m/z=478 [M + Na]⁺, 494 [M + K]⁺.

3,4,6-Tri-*O*-acetyl-2-deoxy-2-(dimethylmaleimido)-D-galactopyranose (54): To a solution of **53** (0.78 g, 1.7 mmol) in DMF (6 mL) was added hydrazinium acetate (0.19 g, 2.1 mmol). The mixture was stirred at room temperature for 1 h, diluted with EtOAc, washed with cold aq. saturated NaHCO₃ and water, dried (MgSO₄), filtered, and concentrated. Column chromatography (toluene/EtOAc, 1:1) of the residue yielded **54**, isolated as a white foam (0.43 g, 61%). TLC (toluene/EtOAc, 1:1): $R_f = 0.37$. ¹H NMR β-anomer (300 MHz, CDCl₃): $\delta = 1.89$, 1.96, 2.05, and 2.17 (4 s, 3, 6, 3, 3 H, 2 CH₃, 3 COCH₃), 4.83 (dd, $J_{1,2} = 7.0$, $J_{2,3} = 11.5$ Hz, 1 H, 2-H), 5.33 (d, 1 H, 1-H), 5.43 (d, $J_{3,4} = 3.3$, $J_{4,5} < 1$ Hz, 1 H, 4-H), 5.65 (dd, 1 H, 3-H) ppm. MS (MALDI-TOF) of $C_{18}H_{23}NO_{10}$ (413): found m/z = 436 [M + Na]⁺, 452 [M + K]⁺.

3,4,6-Tri-O-acetyl-2-deoxy-2-(dimethylmaleimido)-β-D-galactopyranosyl trichloroacetimidate (55): To a solution of 54 (0.30 g, 0.73 mmol) in CH₂Cl₂ (6 mL) were added trichloroacetonitrile (1.05 g, 7.3 mmol) and 1,8-diazabicyclo[5.4.0]undec-7-ene (29 mg, 0.19 mmol), and the mixture was stirred for 30 min at room temperature, then concentrated. Column chromatography (toluene/ EtOAc, 1:1) of the residue yielded 55, isolated as a white foam (0.24 g, 59%). TLC (toluene/EtOAc, 1:1): $R_f = 0.56$. $[\alpha]_D^{20} = +36$ $(c = 1, \text{CHCl}_3)$. ¹H NMR (300 MHz, CDCl₃): $\delta = 1.93, 2.05, \text{ and}$ 2.20 (3 s, 9, 3, 3 H, 2 CH₃, 3 COCH₃), 4.58 (dd, $J_{1,2} = 8.6$, $J_{2,3} =$ 11.5 Hz, 1 H, 2-H), 5.50 (d, $J_{3,4} = 3.4$, $J_{4,5} < 1$ Hz, 1 H, 4-H), 5.78 (dd, 1 H, 3-H), 6.40 (d, 1 H, 1-H), 8.67 (s, 1 H, NH) ppm. ¹³C NMR (75.5 MHz, CDCl₃): $\delta = 8.6$ (CH₃), 20.3–20.5 (CO*C*H₃), 49.9 (C-2), 60.8 (C-6), 66.2, 67.6, and 71.6 (C-3, C-4, C-5), 93.9 (C-1), 137.3 (C=C NDMM), 160.4 (CONHCCl₃), 169.5-170.2 ($COCH_3$, CO NDMM).

Octyl (3,4,6-Tri-O-acetyl-2-deoxy-2-dimethylmaleimido-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-O-benzyl- α -D-mannopyranoside (56): A solution of 55 (147 mg, 264 μmol) and 13 (182 mg, 176 μmol) in dry CH₂Cl₂ (2.2 mL), containing molecular sieves 4 Å (30 mg), was stirred under Ar at room temperature for 45 min. After cooling to 0 °C, a solution of TMSOTf (2.64 μmol) in dry CH₂Cl₂ (26.4 μL) was added. When TLC (hexane/EtOAc, 1:1) showed the reaction to be complete, the mixture was neutralised with NEt₃ and concentrated. Column chromatography (hexane/EtOAc, 1:1) of the residue yielded 56, isolated as a colourless syrup (212 mg, 84%). TLC (hexane/EtOAc, 1:1): $R_f = 0.59$. $[\alpha]_D^{20} = +6$ (c = 1, CHCl₃). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.86 \text{ (t, 3 H, octyl CH}_3), 1.23 \text{ (m, 10 H, 5)}$ octyl CH₂), 1.36 (m, 2 H, octyl CH₂), 1.88, 1.93, 2.02, and 2.04 (4 s, 3, 3, 6, 3 H, 2 CH₃, 3 COCH₃), 2.91 and 3.13 (2 m, each 1 H, octyl OCH₂), 5.32 (d, $J_{3'',4''} = 3.4$, $J_{4'',5''} < 1$ Hz, 1 H, 4''-H), 5.32 $(d, J_{1'2'} = 8.2 \text{ Hz}, 1 \text{ H}, 1'\text{-H}), 5.61 (dd, J_{2''3''} = 11.5 \text{ Hz}, 1 \text{ H}, 3''\text{-}$ H), 6.90-7.34 (m, 25 H, 5 OCH₂C₆H₅), 7.55 and 7.68 (2 m, each 2 H, Phth) ppm. ¹³C NMR (75.5 MHz, CDCl₃): $\delta = 8.9$ (CH₃), 14.0 (octyl CH₃), 20.4–20.5 (COCH₃), 22.5, 25.9, 29.1, 29.2, 29.6, and 31.7 (octyl CH₂), 51.8 and 55.3 (C-2', C-2''), 60.9, 67.6, 68.7, 69.8, 70.5, 72.7, 73.1, 73.7, and 74.7 (C-6, C-6', C-6'', 5 OCH₂C₆H₅, octyl OCH₂), 66.6, 67.8, 70.5, 72.5, 73.3, 74.6, 74.7, 75.9, 76.3, and 77.6 (C-2, C-3, C-4, C-5, C-3', C-4', C-5', C-3'', C-4", C-5"), 96.6 (2 C) and 97.1 (C-1, C-1", C-1"), 169.6-170.2 (COCH₃, CO NDMM) ppm. MS (MALDI-TOF) of C₈₁H₉₂N₂O₂₁ (1429): found $m/z = 1452 [M + Na]^+$, 1468 $[M + K]^+$.

Octyl (2-Deoxy-2-propionamido-3,4,6-tri-O-propionyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-Dglucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-O-benzyl- α -D-mannopyranoside (57): To a solution of 56 (364 mg, 255 µmol) in dioxane/water (14.4 mL, 4:1) was added NaOH (287 mg, 7.18 mmol), and the mixture was stirred at room temperature overnight. Then, the pH was adjusted to 3 using aq. 4 m HCl, and the stirring was continued overnight. The mixture was neutralised with solid K₂CO₃, and coconcentrated with toluene. A solution of the residue in pyridine/ propionic anhydride (20 mL, 1:1) was stirred overnight, then coconcentrated with toluene. The residue was dissolved in CH₂Cl₂, and the solution was washed with aq. 1 m HCl, aq. saturated NaHCO₃, and water, dried (MgSO₄), filtered, and concentrated. Column chromatography (hexane/EtOAc, 1:2) of the residue yielded 57, isolated as a colourless syrup (116 mg, 33%). TLC (hexane/EtOAc, 1:1): $R_f = 0.45$. $[\alpha]_D^{20} = +9 (c = 1, CHCl_3)$. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.87$ (t, 3 H, octyl CH₃), 1.02-1.14 (m, 12 H, 3 COCH₂CH₃, NHCOCH₂CH₃), 1.24 (m, 10 H, 5 octyl CH₂), 1.44 (m, 2 H, octyl CH₂), 1.90 (m, 2 H, NHCOCH₂CH₃), 2.20-2.36 (m, 6 H, 3 COC H_2 CH₃), 3.20 (m, 1 H, octyl OCHH), 5.29 (d, $J_{4'',5''}$ < 1 Hz, 1 H, 4''-H), 6.99-7.41 (m, 29 H, Phth, 5 $OCH_2C_6H_5$) ppm. ¹³C NMR (75.5 MHz, CDCl₃): $\delta = 8.7, 8.8, 9.0,$ and 9.6 (3 COCH₂CH₃, NHCOCH₂CH₃), 13.9 (octyl CH₃), 22.5, 25.9, 27.2 (2 C), 29.0, 29.2 (2 C), 29.5 (2 C), and 31.6 (6 octyl CH₂, 3 COCH₂CH₃, NHCOCH₂CH₃), 50.8 (2 C) (C-2', C-2''), 60.8, 67.4, 69.0, 69.6, 70.5, 72.7 (2 C), 73.8, and 74.0 (C-6, C-6', C-6'', 5 OCH₂C₆H₅, octyl OCH₂), 63.9, 66.1, 70.5 (2 C), 71.4, 74.7, 74.8, 77.0, 78.1, and 82.1 (C-2, C-3, C-4, C-5, C-3', C-4', C-5', C-3'', C-4", C-5"), 97.5, 100.8, and 101.7 (C-1, C-1", C-1"), 173.4 and 173.5 (COCH₂CH₃, NHCOCH₂CH₃) ppm. MS (MALDI-TOF) of $C_{81}H_{98}N_2O_{20}$ (1419): found $m/z = 1442 [M + Na]^+$, 1458 [M +

Octyl (3,4,6-Tri-O-acetyl-2-deoxy-2-propionamido-β-D-galactopyranosyl)-(1→4)-(2-acetamido-3,6-di-O-benzyl-2-deoxy-β-D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-O-benzyl- α -D-mannopyranoside (58): A solution of 57 (116 mg, 84.2 µmol) in 1-BuOH (22 mL) was stirred at room temperature for 30 min, then 1,2-diaminoethane (1.2 mL, 18 mmol) was added. The solution was stirred at 90 °C overnight, then co-concentrated with toluene. A solution of the residue in pyridine/acetic anhydride (9.6 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/ EtOAc, 2:3) of the residue yielded 58, isolated as a colourless syrup (78 mg, 74%). TLC (toluene/EtOAc, 2:3): $R_f = 0.41$. $[\alpha]_D^{20} = +3$ $(c = 1, CHCl_3)$. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.06 (t, 3 H, NHCOCH₂CH₃), 1.26 (m, 10 H, 5 octyl CH₂), 1.57 (m, 2 H, octyl CH₂), 1.78 (s, 3 H, NHCOCH₃), 1.96, 2.01, and 2.07 (3 s, each 3 H, 3 COCH₃), 2.05 (m, 2 H, NHCOC H_2 CH₃), 3.34 (m, 1 H, octyl OCHH), 4.09 (s, $J_{1,2} < 1$ Hz, 1 H, 1-H), 4.91 (dd, $J_{2'',3''} = 11.3$, $J_{3'',4''} = 3.2$ Hz, 1 H, 3''-H), 5.25 (d, $J_{4'',5''}$ < 1 Hz, 1 H, 4"-H), 5.91 (d, 1 H, NH), 7.20-7.39 (m, 25 H, 5 OCH₂C₆ H_5) ppm. ¹³C NMR (75.5 MHz, CDCl₃): δ = 9.6 (NHCOCH₂CH₃), 13.9 (octyl CH₃), 20.4–20.5 (COCH₃), 23.1 (NHCOCH₃), 22.5, 26.0, 29.0-29.8, and 31.6 (octyl CH₂, NHCOCH₂CH₃), 50.9 and 55.7 (C-2', C-2''), 61.0, 67.7, 69.2, 71.1, 73.1 (2 C), 73.3, 73.5, and 74.9 (C-6, C-6', C-6'', 5 $OCH_2C_6H_5$, octyl OCH₂), 66.3, 70.1, 70.4, 71.5, 73.6, 74.1, 74.5, 76.2, 77.0, and 78.4 (C-2, C-3, C-4, C-5, C-3', C-4', C-5', C-3'', C-4'', C-5''), 97.5, 97.9, and 100.2 (C-1, C-1', C-1"), 170.1-171.0 (COCH₃, NHCOCH₃), 173.8 (NHCOCH₂CH₃) ppm.

Octyl (3,4,6-Tri-O-acetyl-2-deoxy-2-propionamido- β -D-galactopyr-anosyl)-(1 \rightarrow 4)-(2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyr-anosyl)-(1 \rightarrow 2)-3,4,6-tri-O-acetyl- α -D-mannopyranoside (59): To a

solution of 58 (78 mg, 63 μmol) in MeOH/CH₂Cl₂ (4.0 mL, 1:1) was added NaOMe (pH = 9), and the mixture was stirred overnight. After neutralisation with Dowex 50×8 (H⁺) and filtration, the solution was concentrated. To a solution of the residue in EtOH/EtOAc (6.0 mL, 1:1) were added 10% Pd/C (52 mg) and 3 drops of HOAc, and the mixture was stirred under H2 overnight, then filtered, and concentrated. A solution of the residue in pyridine/acetic anhydride (9.0 mL, 1:1) was stirred overnight, then coconcentrated with toluene. Column chromatography (CH₂Cl₂/acetone, 2:1) of the residue yielded **59**, isolated as a white solid (48 mg, 72%). TLC (CH₂Cl₂/acetone, 2:1): $R_f = 0.32$. $[\alpha]_D^{20} = -18$ (c = 1, CHCl₃). ¹H NMR (500 MHz, CDCl₃; 2D TOCSY, ROESY, HSQC): $\delta = 0.89$ (t, 3 H, octyl CH₃), 1.09 (t, 3 H, NHCOCH₂CH₃), 1.29 (m, 10 H, 5 octyl CH₂), 1.59 (m, 2 H, octyl CH₂), 1.96, 1.97, 2.00, 2.03, 2.06, 2.09, 2.11, and 2.15 (8 s, 3, 3, 3, 3, 3, 6, 3, 3 H, 8 COCH₃, NHCOCH₃), 2.16 (m, 2 H, NHCOC H_2 CH₃), 3.41 and 3.63 (2 ddd, each 1 H, octyl OCH₂), 3.63-3.79 (m, 3 H, 2'-H, 4'-H, 5'-H), 3.84-3.90 (m, 2 H, 5-H, 5"-H), 3.92 (dd, $J_{1",2"} = 8.3$, $J_{2",3"} = 11.2$ Hz, 1 H, 2"-H), 4.05-4.14 (m, 4 H, 2-H, 6a-H, 6''a-H, 6''b-H), 4.18 (dd, $J_{5.6b}$ = 5.8, $J_{6a,6b} = 12.1 \text{ Hz}$, 1 H, 6b-H), 4.29 (dd, $J_{5',6'b} = 5.0$, $J_{6'a,6'b} =$ 11.8 Hz, 1 H, 6'b-H), 4.34 (dd, $J_{5',6'a} = 3.0$ Hz, 1 H, 6'a-H), 4.66 (d, 1 H, 1''-H), 4.72 (d, $J_{1,2} < 1$ Hz, 2 H, 1-H, 1'-H), 5.10 (dd, $J_{2,3} = 3.4$, $J_{3,4} = 10.1$ Hz, 1 H, 3-H), 5.20 (dd, $J_{3'',4''} = 3.4$ Hz, 1 H, 3''-H), 5.22 (t, 1 H, 4-H), 5.30 (t, 1 H, 3'-H), 5.32 (d, $J_{4'',5''}$ 1 Hz, 1 H, 4"-H), 5.87 (d, $J_{2",NH"} = 8.7$ Hz, 1 H, NH"), 5.94 (d, $J_{2',NH'} = 8.6 \text{ Hz}, 1 \text{ H}, NH') \text{ ppm.}^{13}\text{C NMR} (75.5 \text{ MHz}, \text{CDCl}_3)$: $\delta = 9.5 \text{ (NHCOCH}_2\text{CH}_3), 13.9 \text{ (octyl CH}_3), 20.4-20.8 \text{ (CO}_2\text{CH}_3),$ 23.0 (NHCOCH₃), 22.5, 26.0, 29.1-29.6, and 31.7 (octyl CH₂, NHCOCH₂CH₃), 51.5 (C-2"), 53.9 (C-2"), 61.0, 62.7 (2 C), and 68.3 (C-6, C-6', C-6", octyl OCH₂), 66.1, 66.3, 68.4, 69.6, 70.2, 70.5, 71.2, 72.7, 74.5, and 75.2 (C-2, C-3, C-4, C-5, C-3', C-4', C-5', C-3'', C-4'', C-5''), 97.4 and 99.1 (C-1, C-1'), 100.5 (C-1''), 169.3-170.7 (COCH₃, NHCOCH₃), 174.2 (NHCOCH₂CH₃) ppm. MS (MALDI-TOF) of $C_{47}H_{72}N_2O_{24}$ (1048): found m/z = 1049 [M $+ H]^+, 1071 [M + Na]^+, 1087 [M + K]^+.$

Octyl 2-Deoxy-2-propionamido-β-D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranoside (11): To a solution of 59 (33 mg, 31 μmol) in MeOH/CH₂Cl₂ (2.0 mL, 1:1) was added NaOMe (pH = 10), and the mixture was stirred overnight. After neutralisation with Dowex 50 × 8 (H⁺) and filtration, the solution was concentrated. The residue was applied on a C18-Bakerbond spetm column (500 mg), eluted with water and MeOH, and the MeOH fractions were concentrated. Gel-filtration of the residue on a HW-40S Toyopearl column, eluted with aq. 5 mm NH₄OAc, and subsequent lyophilization yielded 11, isolated as a white foam (17 mg, 77%). TLC (MeOH/CH₂Cl₂, 3:1): $R_f = 0.65$. [α] $_D^{20} = -9$ (c = 1, H₂O). High-resolution MS of C₃₁H₅₆N₂O₁₆ (712.3630): calcd. 735.3528, found 735.3507 [M + Na]. For ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2, respectively.

Octyl (2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranoside (60): To a solution of 56 (50 mg, 35 μmol) in dioxane/water (1.75 mL, 4:1) was added NaOH (35 mg, 0.88 mmol), and the mixture was stirred at room temperature overnight. Then the pH was adjusted to 3 using aq. 4 m HCl, and the stirring was continued overnight. Subsequently, the solution was made alkaline with aq. 1 m NaOH followed by the addition of AcCl (43.8 μL). After stirring at pH = 10 for 45 min, the pH was lowered to 3 using aq. 4 m HCl, and the mixture was stirred overnight, neutralised with solid K_2 CO₃, and

co-concentrated with toluene. A solution of the residue in pyridine/ acetic anhydride (5.3 mL, 2:1) was stirred overnight, then co-concentrated with toluene. The residue was dissolved in CH₂Cl₂, and the solution was washed with aq. 1 M HCl, aq. saturated NaHCO₃, and water, dried (MgSO₄), filtered, and concentrated. Column chromatography (hexane/EtOAc, 1:2) of the residue yielded 60, isolated as a colourless syrup (23 mg, 48%). TLC (hexane/EtOAc, 1:2): $R_{\rm f} = 0.36$. $[\alpha]_{\rm D}^{20} = +4$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.87$ (t, 3 H, octyl CH₃), 1.22 (m, 10 H, 5 octyl CH₂), 1.43 (m, 2 H, octyl CH₂), 1.76 (s, 3 H, NHCOCH₃), 1.97 and 2.05 (2 s, 3, 6 H, 3 COCH₃), 2.99 and 3.17 (2 m, each 1 H, octyl OCH₂), 5.24 (d, $J_{4'',5''}$ < 1 Hz, 1 H, 4''-H), 6.85-7.62 (m, 29 H, Phth, 5 $OCH_2C_6H_5$) ppm. ¹³C NMR (75.5 MHz, CDCl₃): $\delta = 14.0$ (octyl CH₃), 20.5-20.6 (COCH₃), 23.1 (NHCOCH₃), 22.5, 25.9, 29.0, 29.2, 29.5, and 31.7 (octyl CH₂), 50.7 (C-2"), 55.5 (C-2"), 96.9 (2 C) and 100.8 (C-1, C-1', C-1''), 169.6-170.2 (COCH₃, NHCOCH₃) ppm.

Octyl (2-Acetamido-2-deoxy-3,4,6-tri-O-propionyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-O-benzyl-2-deoxy-2-propionamido-β-Dglucopyranosyl)-(1→2)-3,4,6-tri-O-benzyl-α-D-mannopyranoside (61): A solution of 60 (54 mg, 40 μmol) in 1-BuOH (5.6 mL) was stirred for 30 min at room temperature, then 1,2-diaminoethane (0.56 mL, 8.4 mmol) was added. The mixture was stirred overnight at 90 °C, then co-concentrated with toluene. A solution of the residue in pyridine/propionic anhydride (4.4 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (toluene/EtOAc, 2:3) of the residue yielded 61, isolated as a colourless syrup (42 mg, 80%). TLC (toluene/EtOAc, 2:3): $R_{\rm f} = 0.39$. $[\alpha]_{D}^{20} = +1$ (c = 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (t, 3 H, octyl CH₃), 1.01 (m, 3 H, NHCOCH₂CH₃), 1.04-1.16 (m, 9 H, 3 COCH₂CH₃), 1.26 (m, 10 H, 5 octyl CH₂), 1.51 (m, 2 H, octyl CH₂), 1.75 (s, 3 H, NHCOCH₃), 1.96 (m, 2 H, NHCOCH₂CH₃), 2.25 (m, 6 H, 3 COCH₂CH₃), 3.33 (m, 1 H, octyl OCHH), 4.09 (s, $J_{1.2} < 1$ Hz, 1 H, 1-H), 5.24 (d, $J_{4'',5''} < 1$ Hz, 1 H, 4"-H), 7.18-7.41 (m, 25 H, 5 OCH₂C₆H₅) ppm. ¹³C NMR $(75.5 \text{ MHz}, \text{ CDCl}_3)$: $\delta = 8.7, 8.8, 9.0, \text{ and } 9.4 (\text{COCH}_2\text{CH}_3,$ NHCOCH₂CH₃), 14.0 (octyl CH₃), 23.1 (NHCOCH₃), 22.5, 26.0, 29.1-29.5, and 31.7 (octyl CH_2 , $COCH_2CH_3$, NHCOCH₂CH₃), 51.2 (C-2"), 56.5 (C-2"), 97.6, 98.0, and 100.2 (C-1, C-1', C-1''), 169.8 (NHCOCH₃), 173.5-174.5 (COCH₂CH₃, NHCOCH₂CH₃) ppm.

Octyl (2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-galactopyranosyl)-(1→4)-(3,6-di-*O*-acetyl-2-deoxy-2-propionamido-β-D-glucopyranosyl)- $(1\rightarrow 2)$ -3,4,6-tri-O-acetyl- α -D-mannopyranoside (62): To a solution of **61** (42 mg, 32 μmol) in MeOH/CH₂Cl₂ (2.0 mL, 1:1) was added NaOMe (pH = 9), and the mixture was stirred overnight. After neutralisation with Dowex 50×8 (H⁺) and filtration, the solution was concentrated. To a solution of the residue in EtOH/EtOAc (3.0 mL, 1:1) were added 10% Pd/C (27 mg) and 2 drops of HOAc, and the mixture was stirred under H₂ overnight, then filtered through Celite, and concentrated. A solution of the residue in pyridine/acetic anhydride (5.0 mL, 1:1) was stirred overnight, then co-concentrated with toluene. Column chromatography (CH₂Cl₂/acetone, 1:1) of the residue yielded **62**, isolated as a white solid (22 mg, 67%). TLC (CH₂Cl₂/acetone, 1:1): $R_f = 0.53$. $[\alpha]_D^{20} =$ -21 (c = 1, CHCl₃). ¹H NMR (500 MHz, CDCl₃; 2D TOCSY, ROESY, HSQC): $\delta = 0.89$ (t, 3 H, octyl CH₃), 1.11 (t, 3 H, NHCOCH₂CH₃), 1.28 (m, 10 H, 5 octyl CH₂), 1.59 (m, 2 H, octyl CH₂), 1.94, 1.98, 2.01, 2.03, 2.06, 2.08, 2.12, and 2.14 (8 s, 3, 3, 3, 3, 3, 6, 3, 3 H, 8 COCH₃, NHCOCH₃), 2.17 (m, 2 H, NHCOCH₂CH₃), 3.40 and 3.63 (2 ddd, each 1 H, octyl OCH₂), 3.65-3.73 (m, 3 H, 2'-H, 4'-H, 5'-H), 3.83-3.91 (m, 3 H, 5-H, FULL PAPER _______ J. P. Kamerling et al.

2"-H, 5"-H), 4.05 (dd, $J_{5,6a} = 2.8$, $J_{6a,6b} = 11.9$ Hz, 1 H, 6a-H), 4.07-4.13 (m, 3 H, 2-H, 6''a-H, 6''b-H), 4.17 (dd, $J_{5,6b} = 5.7$ Hz, 1 H, 6b-H), 4.27 (dd, $J_{5',6'b} = 5.2$, $J_{6'a,6'b} = 12.0$ Hz, 1 H, 6'b-H), 4.33 (dd, $J_{5',6'a} = 2.6$ Hz, 1 H, 6'a-H), 4.66 (d, $J_{1'',2''} = 8.1$ Hz, 1 H, 1''-H), 4.67 (s, $J_{1,2}$ < 1 Hz, 1 H, 1-H), 4.77 (d, $J_{1',2'}$ = 7.6 Hz, 1 H, 1'-H), 5.08 (dd, $J_{2,3}=3.5, J_{3,4}=10.1$ Hz, 1 H, 3-H), 5.19 (dd, $J_{2'',3''} = 10.8$, $J_{3'',4''} = 3.3$ Hz, 1 H, 3''-H), 5.21 (t, 1 H, 4-H), 5.32 (d, $J_{4'',5''}$ < 1 Hz, 1 H, 4''-H), 5.38 (dd, $J_{2',3'}$ = 8.2, $J_{3',4'}$ = 9.4 Hz, 1 H, 3'-H), 5.68 (d, $J_{2',NH'} = 8.3$ Hz, 1 H, NH'), 5.76 (d, $J_{2'',NH''} = 8.8 \text{ Hz}, 1 \text{ H}, NH'') \text{ ppm.}^{13}\text{C NMR} (75.5 \text{ MHz}, CDCl_3)$: $\delta = 9.5 \text{ (NHCOCH}_2\text{CH}_3), 13.9 \text{ (octyl CH}_3), 20.4-20.7 \text{ (COCH}_3),}$ 23.1 (NHCOCH₃), 22.5, 25.9, 29.1 (2 C), 29.3, 29.5, and 31.6 (octyl CH₂, NHCOCH₂CH₃), 51.6 (C-2"), 54.4 (C-2"), 61.0, 62.6, 62.7, and 68.2 (C-6, C-6', C-6'', octyl OCH₂), 66.1, 66.3, 68.5, 69.6, 70.2, 70.5, 71.3, 72.6, 74.5, and 75.2 (C-2, C-3, C-4, C-5, C-3', C-4', C-5', C-3'', C-4'', C-5''), 97.3 and 99.1 (C-1, C-1'), 100.3 (C-1''), 169.3-170.8 (COCH₃, NHCOCH₃), 173.9 (NHCOCH₂CH₃) ppm.

Octyl 2-Acetamido-2-deoxy-β-D-galactopyranosyl-(1 \rightarrow 4)-2-deoxy-2-propionamido-β-D-glucopyranosyl)-(1 \rightarrow 2)- α -D-mannopyranoside (12): To a solution of 62 (22 mg, 21 μmol) in MeOH/CH₂Cl₂ (2.0 mL, 1:1) was added NaOMe (pH = 9), and the mixture was stirred overnight. After neutralisation with Dowex 50 × 8 (H⁺) and filtration, the solution was concentrated. The residue was applied on a C18-Bakerbond spetm column (500 mg), eluted with water and MeOH, and the MeOH fractions were concentrated. Gel-filtration of the residue on a HW-40S Toyopearl column, eluted with aq. 5 mm NH₄OAc, and subsequent lyophilization yielded 12, isolated as a white foam (13 mg, 84%). TLC (MeOH/CH₂Cl₂, 3:1): $R_f = 0.65$. [α] $_D^{20} = -8$ (c = 0.5; H₂O). High-resolution MS of C₃₁H₅₆N₂O₁₆ (712.3630): calcd. 735.3528, found 735.3515 [M + Na]. For ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2, respectively.

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