

imately a 60:40 mixture of the desired 1-(cyclopenten-3-yl)ethanol and the coupling product 3-(cyclopenten-3-yl)cyclopentene. About 3 g of the product mixture was chromatographed on a 2 × 15 cm activity grade 3 alumina column using pentane and ether. The alcohol-containing fraction was dried and distilled; bp 81–82 °C (32 mm). GLC analysis on a 46-m Carbowax 20M glass capillary column showed the material was a 65:35 mixture of diastereomers: NMR (CCl₄) δ 1.1 (d, *J* = 6 Hz, 3 H, CH₃), 1.3–2.9 (m, 5 H), 3.2 (s, 1 H, OH), 3.5 (quintet, *J* = 6 Hz, 1 H, CHOH), 5.4–5.8 (m, 2 H, vinylic).

Anal. Calcd. for C₇H₁₂O: C, 75.00; H, 10.71 Found: C, 74.78; H, 10.69.

Samples of the 1-(cyclopenten-3-yl)ethanol diastereomers were collected from a 20% Carbowax 20M column. NMR analysis showed that they have identical absorptions except in the vinylic region. For the major product, NMR δ (CCl₄) 5.5 and 5.7 (a pair of symmetrical multiplets); for the minor product, NMR δ (CCl₄) 5.7 ppm (a singlet). The major product was identical with the material obtained from hydrolysis of the *anti*-6-methyl-2-bicyclo[3.1.0]hexyl 3,5-dinitrobenzoates. Thus, it was assigned as being the (*RR*)(*SS*) isomer and the minor product was assigned as being the (*RS*)(*SR*) isomer.

Hydrolysis Products from the *anti*-9-Methyl-2-bicyclo[6.1.0]nonyl 3,5-Dinitrobenzoates. The hydrolysis products in 80% aqueous acetone at 80 °C were obtained by following a similar procedure to that described above for the 2-bicyclo[3.1.0]hexyl system, and the product compositions were determined by using a combination of 47-m Carbowax 20M glass capillary column GLC and ¹H NMR techniques. A control experiment with the *anti*-endo alcohol, the most reactive of the various product alcohols, showed that it was stable under the reaction conditions.

Of the three minor hydrolysis products obtained from the *anti*-endo 3,5-dinitrobenzoate, the one that was formed in about 3% yield has an identical GLC retention time as one of the diastereomeric 1-(cycloocten-3-yl)ethanols prepared independently as described below. The other two, present in about equal amounts, have similar GLC retention times as those of the other C₁₀ alcohols encountered in this study. However, they could not be isolated in sufficient quantities for identification.¹¹

The sole minor product (2% yield) from hydrolysis of the *anti*-exo 3,5-dinitrobenzoate had the same GLC retention time as the major product (95% yield) obtained from perchloric acid in acetic acid catalyzed rearrangement of the *anti*-exo alcohol followed by lithium aluminum hydride in ether reduction of the

acetates formed. On the basis of its spectra and from mechanistic expectations, this has been tentatively identified as *trans*-8-hydroxy-*cis*-9-methyl-*trans*-bicyclo[5.2.0]nonane: NMR (CCl₄) δ 1.1 (d, *J* = 7 Hz, 3 H, CH₃), 1.1–2.1 (m, 13 H), 1.7 (s, 1 H, OH), 3.1 (t, *J* = 7 Hz, 1 H, CHOH). A precise molecular weight was determined by mass spectrometry. Calcd for C₁₀H₁₈O: 154.1358. Found: 154.1375.

1-(Cycloocten-3-yl)ethanol. Following an analogous procedure to that described above for preparing 1-(cyclopenten-3-yl)ethanol but starting with 3-chlorocyclooctene,¹² a 60:40 mixture of two diastereomeric 1-(cycloocten-3-yl)ethanols together with some 3-(cycloocten-3-yl)cyclooctene coupling product was obtained. The major isomer had the same GLC retention time as that of one of the minor hydrolysis products of *anti*-9-methyl-*endo*-2-bicyclo[6.1.0]nonyl 3,5-dinitrobenzoate. A sample of the 1-(cycloocten-3-yl)ethanol mixture was collected by GLC and a precise molecular weight determined by mass spectrometry. Calcd for C₁₀H₁₈O: 154.1358. Found: 154.1354. For the major product, NMR (CCl₄) δ 1.1 (d, *J* = 6 Hz, 3 H, CH₃), 1.0–2.6 (m, 11 H), 1.6 (s, 1 H, OH), 3.5 (quintet, *J* = 6 Hz, 1 H, CHOH), 5.0–5.7 (m, 2 H, vinylic); for the minor product, NMR (CCl₄) 1.1 (d, *J* = 6 Hz, 3 H, CH₃), 1.0–2.6 (m, 11 H), 1.5 (s, 1 H, OH), 3.6 (quintet, *J* = 6 Hz, 1 H, CHOH), 5.2–5.9 ppm (m, 2 H, vinylic).

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Registry No. (±)-5, 85249-33-8; (±)-5 3,5-DNB, 85203-30-1; (±)-6, 85249-34-9; (±)-6 3,5-DNB, 85249-35-0; (±)-7, 85203-31-2; (±)-7 3,5-DNB, 85203-32-3; (±)-8, 85203-33-4; (±)-8 3,5-DNB, 85203-34-5; (±)-12, 85203-35-6; (±)-*anti*-6-methyl-2-bicyclo[3.1.0]hexanone, 85249-36-1; bicyclo[6.1.0]nonan-2-one, 29800-55-3; (±)-cycloocten-3-ol, 62249-35-8; 1,1-diiodoethane, 594-02-5; (±)-*syn*-9-methyl-2-bicyclo[6.1.0]nonanone, 85249-37-2; (±)-*anti*-9-methyl-2-bicyclo[6.1.0]nonanone, 85249-38-3; (±)-*syn*-9-methyl-*endo*-2-bicyclo[6.1.0]nonanol, 85249-39-4; (±)-*syn*-6-methyl-*endo*-2-bicyclo[3.1.0]hexanol, 85203-36-7; (±)-cyclopenten-3-ol, 62894-08-0; (±)-*syn*-6-methyl-*exo*-2-bicyclo[3.1.0]hexanol, 85203-37-8; (±)-3-chlorocyclopentene, 62894-09-1; acetaldehyde, 75-07-0; 3-(cyclopenten-3-yl)cyclopentene, 2690-18-8; (±)-(*R**,*S**)-1-(cyclopenten-3-yl)ethanol, 85203-38-9; (±)-*trans*-8-hydroxy-*cis*-9-methyl-*trans*-bicyclo[5.2.0]nonane, 85203-39-0; (±)-(*R**,*R**)-1-(cycloocten-3-yl)ethanol, 85203-40-3; (±)-(*R**,*S**)-1-(cycloocten-3-yl)ethanol, 85203-41-4; (±)-3-chlorocyclooctene, 85249-40-7; (±)-*syn*-9-methyl-*exo*-2-bicyclo[6.1.0]nonanol, 85249-41-8.

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Synthesis of the Carbapenam System from Glutamic Acid and Acetoacetic Acid Derivatives¹

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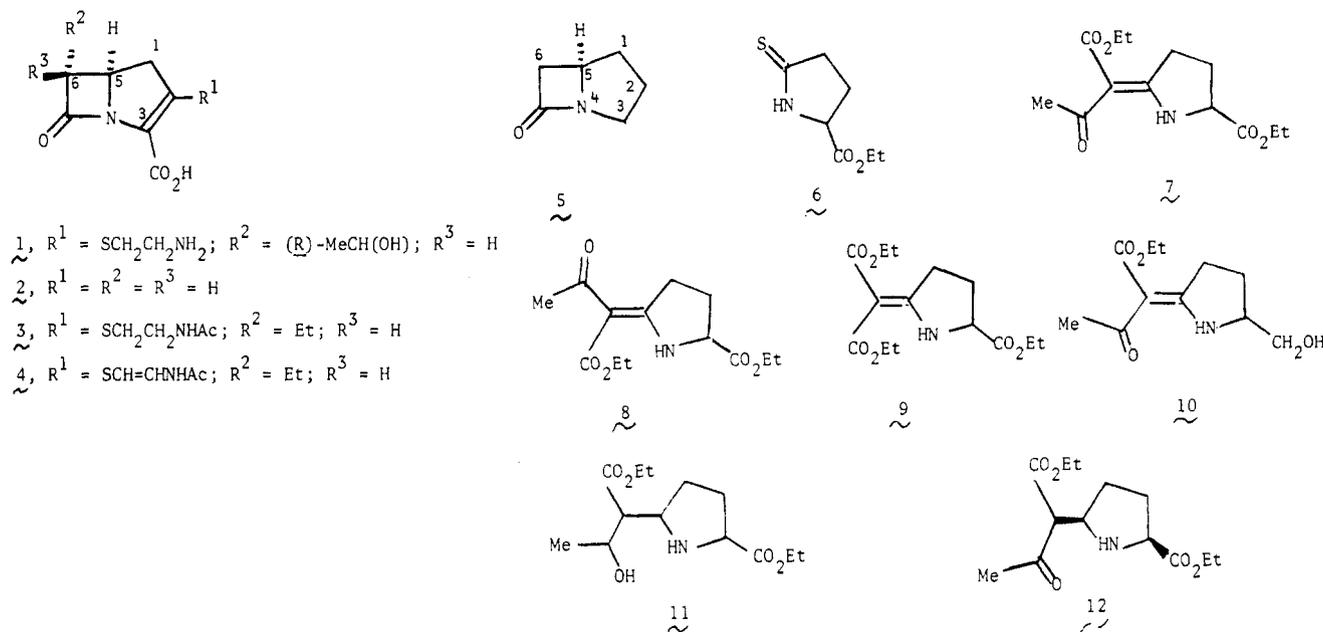
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A synthesis of carbapenam-3-carboxylates is described. Ethyl pyroglutamate was converted into the corresponding thioamide **6** which was subjected to an alkylative sulfide contraction with ethyl 2-bromoacetoacetate. The resulting enamino keto ester **7** was hydrogenated to give, after appropriate manipulations with protecting groups, the two diastereoisomeric 2-[2-(*p*-nitrobenzyloxycarbonyl)-5-pyrrolidinyl]butanoic acids **22a** and **22b**. Dehydration with 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride afforded the *p*-nitrobenzyl (±)-6β-ethyl-carbapenam-3-carboxylate **25a** and its 6α epimer **25b**.

The discovery of the highly potent and broad-spectrum antibiotic thienamycin **1**² (Chart I) was followed by the

isolation of other antibiotics deriving from the carbapenam-3-carboxylic acid **2** and bearing alkyl or hydroxyalkyl

Chart I



groups in either the 6 α or 6 β positions.³ Noteworthy in the context of the present investigation are the 6-ethyl derivatives **3** and **4**⁴ and their 6-epimers.⁵ At the outset of this work, compounds deriving from the more saturated carbapenam system **5**, which formally links the molecular backbone of the penicillin with that of the thienamycin antibiotics, were considered as unstable elusive compounds which tend to polymerize.⁶ Recently, however, a few syntheses of carbapenams including derivatives bearing a carboxylic salt or ester group at position 3 have been described.⁷ We now report a simple synthesis of *p*-nitrobenzyl 6 β -ethylcarbapenam-3 β -carboxylate (**25a**)⁸ and its 6 α -epimer **25b**⁸ by a route based on the use of glutamic acid and acetoacetic acid derivatives as starting materials.

Glutamic acid was reported as a biogenetic precursor of the carbapenam antibiotics.^{2a,3}

The construction of the target's molecular backbone from these starting materials requires the bonding of C-5 of glutamic acid with C-2 of acetoacetic acid. For this purpose ethyl (\pm)-pyroglutamate was converted into the thiolactam **6** which was treated with ethyl 2-bromoacetoacetate to give the alkylidenepyrrolidine **7**. Best results in this condensation, which involves sulfide contraction via alkylative coupling,⁹ were obtained when the reaction was performed in one pot with NaHCO_3 as a base. Use of stronger bases as K_2CO_3 , Et_3N , NaH , or EtONa resulted in a decrease of yield from 61% to 10% or less. The exocyclic double bond in the condensation product was assigned the *E* geometry as depicted in formula **7** rather than the alternative *Z* geometry shown in **8** on the grounds of its IR and NMR spectral data which indicate the existence of intramolecular hydrogen bonding between the pyrrolidine NH and the ketonic C=O groups.¹⁰ Thus, the signal for the NH proton in the NMR spectrum of the keto diester **7** appears at a lower field than the corresponding signal in the spectrum of the similar triester **9**. The IR absorption band for the α,β -unsaturated ester carbonyl in **7** appears at a frequency comparable to that of the α,β -unsaturated ester in **9**, which is not intramolecularly hydrogen bonded. Compound **9** was prepared from the thiolactam **6** and diethyl 2-bromomalonnate analogously to the described preparation of **7**.

The tetrasubstituted double bond in compound **7**, which constitutes part of cross-conjugated vinylogous amide and vinylogous urethan systems, was found to be rather resistant to reduction. Thus, treatment of **7** with sodium borohydride in boiling aqueous tetrahydrofuran resulted in the exclusive reduction of the nonconjugated ester grouping to give the hydroxy derivative **10**. The double bond in **7** also resisted catalytic hydrogenation over PtO_2 in acetic acid but was hydrogenated with the same catalyst in a solution of 20% trifluoroacetic acid in acetic acid. These strong acidic conditions precluded the employment

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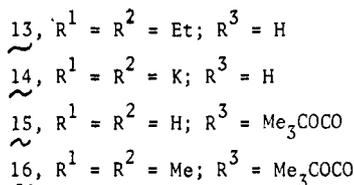
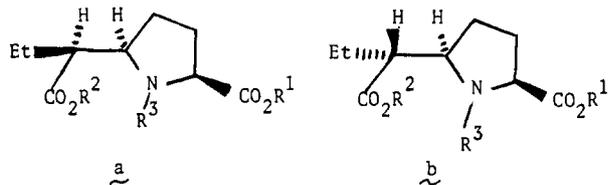
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of acid-sensitive protecting groups for the carboxylates which might have been more convenient than the ethyl groups in later steps of the synthesis. The hydrogenation product consisted of two epimers of the hydroxyethyl derivatives **11** (50%) and two epimers of the ethyl derivatives **13** (39%) which a posteriori were assigned the structures **13a** and **13b** by correlation to their corre-



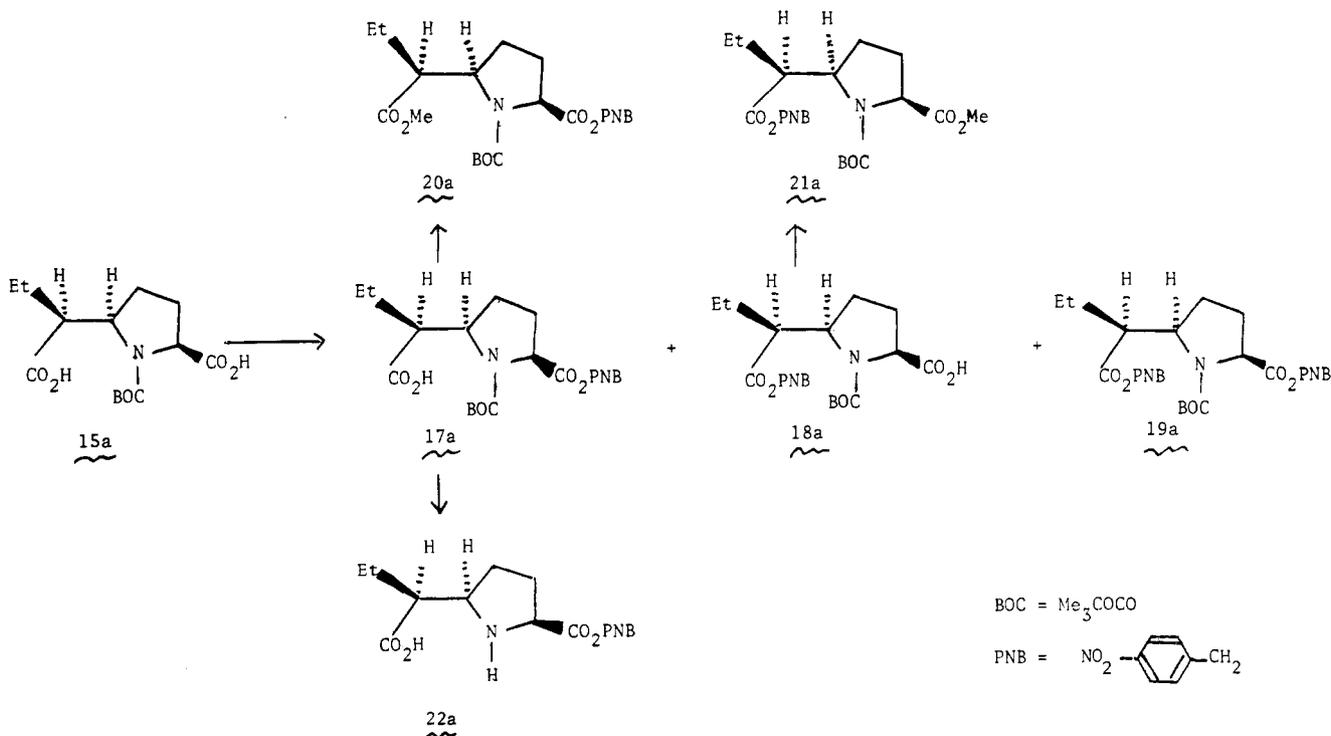
sponding transformation products **25a** and **25b**.⁸ This reduction probably involves an initial 1,4-hydrogenation of the cross-conjugated system from the opposite side the 2-ethoxycarbonyl group to give stereospecifically the saturated β -keto ester **12**, which is further reduced to the carbinols **11** or hydrogenolyzed to the ethyl derivatives **13**. A similar dual reduction of the ketonic carbonyl group has been previously observed in β -keto esters.¹¹ The failure of the carbinol **11** to undergo further hydrogenolysis to **13** when exposed to the same reaction conditions for a longer period of time indicates that it is not an intermediate in the formation of **13**.

Saponification of the diethyl esters **13** with potassium hydroxide followed by treatment of the resulting salts **14** with di-*tert*-butyl dicarbonate afforded a mixture of the

two N-protected epimeric amino dicarboxylic acids **15a** and **15b** (86%). Treatment of this mixture with diazomethane afforded the dimethyl esters **16**. Pure **15a** was obtained by partial separation of the epimeric mixture of **15**. Selective esterification of **15a** with *p*-nitrobenzyl chloride, dicyclohexylamine, and potassium iodide in dimethylformamide afforded the desired *p*-nitrobenzyl ester **17a** (29%) along with its isomer **18a** (15%) and the diester **19a** (23%) (Scheme I). The last two compounds could be recycled, after hydrogenolysis¹² of the *p*-nitrobenzyl group, together with recovered starting material **15a**. The structure assignment of **17a** and **18a** and of their methyl esters **20a** and **21a** was based on the fragmentation patterns of their high-resolution mass spectra which show the corresponding characteristic fragments **23** and/or **24** (see Chart II and Experimental Section).

Deprotection of the amino group in the pyrrolidine **17a** (HCl in EtOAc) afforded the key compound **22a** which possesses the properly protected molecular backbone. Its conversion into a penam system requires the apparently simple operation of removing the elements of water from a β -amino carboxylic acid to form the β -lactam ring. However, attempts to cyclize the amino acid **22a** by conventional methods through the activation of the carboxylic group as its corresponding chloride or *p*-nitrophenyl ester or by using dicyclohexylcarbodiimide (DCC) were rather disappointing. When DCC was used, the IR spectrum of the crude reaction product indicated the presence of a β -lactamic compound which decomposed during attempted purification. To avoid any detrimental intermolecular reactions of the formed β -lactam or of its reactive precursor, we have elaborated a procedure by which the annelation to the bicyclic β -lactam and its primary purification, were performed in a dilute solution. 1-[3-(Dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride, which is soluble in both CH₂Cl₂ and in water, was used as the dehydrating agent. This reagent and the β -amino amino acid **22a** were dissolved in CH₂Cl₂ (concentration

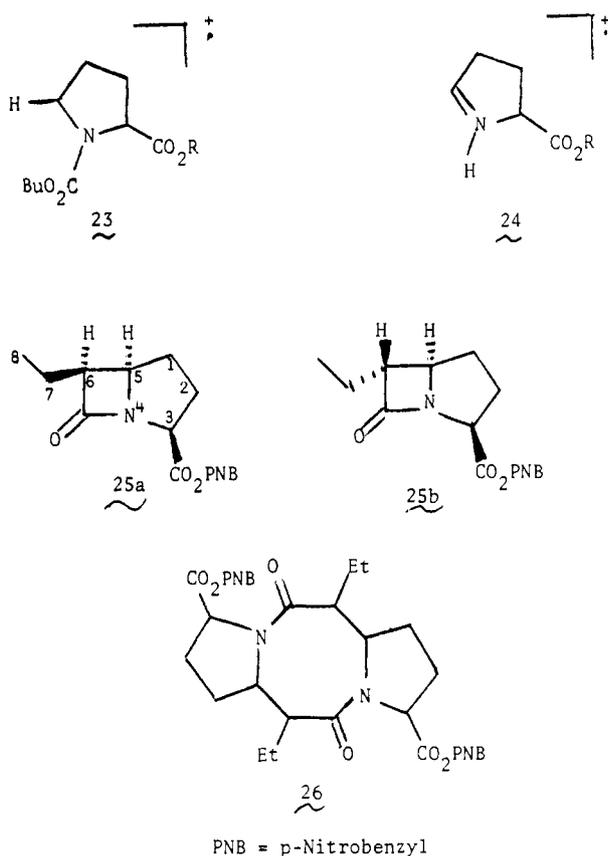
Scheme I



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Chart II



ca. 10^{-3} M), and after 22 h any excess of the carbodiimide and of any derivatives thereof, as well as any other water soluble compound, was removed from the dilute organic solution by washing with cold (0 °C) water. Flash chromatography of the crude product afforded *p*-nitrobenzyl 6 β -ethylcarbapenam-3 β -carboxylate (**25a**, 41%) and the dimer **26** (10%). When a mixture of the acids **15a** and **15b** was subjected to the sequence of reactions as described above for the case of pure **15a**, a mixture of the carbapenam **25a** and its 6 α -epimer **25b** was obtained. Attempts to deblock the esters **25** by catalytic hydrogenolysis over palladium (5% on charcoal), either in dioxane, or in an ethyl acetate-aqueous NaHCO₃ mixture,¹³ resulted in destruction of the β -lactam ring.

The assignment of the relative configuration to the three chiral centers in the carbapenams **25a**⁸ and **25b**⁸ resulted from the analysis of their 270-MHz H¹ NMR spectra which included pertinent decoupling by double irradiation. The coupling constants $J_{5,6} = 5$ Hz, typical for *cis*- β -lactams, and $J_{5,6} = 2$ Hz, typical for *trans*- β -lactams, are observed respectively in the spectra of **25a** and **25b**. The chemical shift for the C-3 protons in these compounds appear at δ 3.92 and 3.93, respectively, namely, within the range characteristic for the analogous proton in penams,¹⁴ oxapenam,¹⁵ and carbapenams,^{7a,b} having C-3 carboxylates in a *trans* relationship with the C-5 proton. In compound **25a**, which possess a C-6 α proton, a long-range coupling constant ($J_{3,6} = 1$ Hz) was observed; no similar coupling was detected in the spectrum of **25b**, where this proton occupies a β -position. It is noteworthy that in the carba-

penams **25a** and **25b** the steric nonbonding intramolecular compression is higher than in their corresponding C-3 epimers. The formation of the thermodynamically less stable isomers is a consequence of the kinetically controlled hydrogenation of the double bond in the intermediate **7**.

Experimental Section

IR spectra were recorded with a Perkin-Elmer 237 spectrophotometer. H¹ NMR data were determined on an 80-MHz Varian FT-80A, a 90MHz Bruker FT-HFX-10, or a 270-MHz Bruker WH-270 instrument. Low- and high-resolution mass spectra were recorded on a Varian MAT-731 (double focusing) spectrometer. Melting points were measured by using a Büchi apparatus and are uncorrected.

Ethyl 5-Thioxopyrrolidine-2-carboxylate (6). A solution of ethyl pyroglutamate (54.95 g, 0.35 mol) in chloroform (200 mL) was added during 15 min to a stirred suspension of P₂S₅ (77.70 g, 0.35 mol) in carbon disulfide (150 mL). The reaction mixture was stirred for 22 h at 55–60 °C and then cooled and evaporated. The residue was stirred in a water–chloroform mixture until all the P₂S₅ decomposed. The organic phase was washed with water, dried (MgSO₄), and evaporated. Chromatography on silica gel (hexane–acetone, 4:1) followed by recrystallization (ether) afforded the thiolactam **6**: 42.38 g (70%); mp 72–73 °C; IR (CHCl₃) 3400, 1735, 1485 cm⁻¹; NMR (90 MHz, CDCl₃) δ 1.31 (t, $J = 7$ Hz, OCH₂CH₃), 2.26–2.55 (m, CHCH₂CH₂), 2.80–3.06 (m, CH₂CH₂CS), 4.25 (q, $J = 7$ Hz, OCH₂CH₃), 4.51 [dd, $J = 7.5, 7$ Hz, NCH(CO₂)CH₂], 8.15 (br, NH); high-resolution mass spectrum, calcd for C₇H₁₁NO₂S m/e 173.0511, found m/e 173.0532; m/e 173 (M⁺), 100 (M⁺ – CO₂Et). Anal. Calcd for C₇H₁₁NO₂S: C, 48.55; H, 6.40; N, 8.09; S, 18.48. Found: C, 48.38; H, 6.58; N, 8.00; S, 18.20.

Ethyl 2-[2-(Ethoxycarbonyl)-5-pyrrolidinylidene]-3-oxobutyrates (7). Ethyl 2-bromo-3-oxobutyrates (41.80 g, 0.2 mol) and NaHCO₃ (33.60 g, 0.4 mol) were added to a solution of the thiolactam **6** (17.30 g, 0.1 mol) in dry CH₂Cl₂ (1400 mL). The suspension was heated under reflux for 36 h, cooled, filtered through Celite, and evaporated. Silica gel chromatography (toluene–ethyl acetate) followed by recrystallization (hexane) afforded the title compound **7**: 16.41 g (61%); mp 64–65 °C; IR (CHCl₃) 1740, 1690, 1600, 1540 cm⁻¹; NMR (80 MHz, CDCl₃) δ 1.29 (t, $J = 7$ Hz, OCH₂CH₃), 1.31 (t, $J = 7$ Hz, OCH₂CH₃), 2.07–2.40 (m, CHCH₂CH₂), 2.42 (s, CH₃CO), 3.11–3.31 (m, CH₂CH₂C), 4.22 (q, $J = 7$ Hz, 2 OCH₂CH₃), 4.50 (dd, $J = 7.5, 7$ Hz, NCH(CO₂)CH₂), 11.78 (br, NH, exchangeable with D₂O); high-resolution mass spectrum, calcd for C₁₃H₁₉NO₅ m/e 269.1263, found m/e 269.1246; m/e 269 (M⁺), 224 (M⁺ – OEt), 196 (M⁺ – CO₂Et), 150 (M⁺ – CO₂Et – C₂H₆O), 122 (M⁺ – CO₂Et – C₂H₆O – CO). Anal. Calcd for C₁₃H₁₉NO₅: C, 57.98; H, 7.11; N, 5.20. Found: C, 58.16; H, 7.21; N, 5.26.

Diethyl 2-[2-(Ethoxycarbonyl)-5-pyrrolidinylidene]-malonate (9). Diethyl 2-bromomalonate (239 mg, 1 mmol) and NaHCO₃ (168 mg, 2 mmol) were added to a solution of the thiolactam **6** (86 mg, 0.5 mmol) in CH₂Cl₂ (3 mL). The suspension was heated under reflux for 24 h, cooled, filtered through Celite, and evaporated. The residue was chromatographed on a silica gel plate (ether) to give the title compound **9**: 63 mg (42%); IR (CHCl₃) 1740, 1695 (br), 1650, 1570; NMR (80 MHz, CDCl₃) δ 1.28 (t, $J = 7$ Hz, OCH₂CH₃), 1.30 (t, $J = 7$ Hz, OCH₂CH₃), 1.31 (t, $J = 7$ Hz, OCH₂CH₃), 2.10–2.50 (m, CHCH₂CH₂), 3.06–3.36 (m, C=C CH₂CH₂), 4.18 (q, $J = 7$ Hz, OCH₂CH₃), 4.21 (q, $J = 7$ Hz, 2 OCH₂CH₃), 4.43 (dd, $J = 7.5, 7$ Hz, NCH(CO₂)CH₂), 9.64 (br, NH); high-resolution mass spectrum, calcd for C₁₁H₂₁NO₆ m/e 299.1369, found m/e 299.1392; m/e 299 (M⁺), 254 (M⁺ – OEt), 226 (M⁺ – CO₂Et), 180 (M⁺ – CO₂Et – C₂H₆O), 152 (M⁺ – CO₂Et – C₂H₆O – CO).

Ethyl 2-[2-(Hydroxymethyl)-5-pyrrolidinylidene]-3-oxobutyrates (10). To a suspension of the keto ester **7** (54 mg, 0.2 mmol) in a mixture of THF (4 mL) and water (1 mL) was added NaBH₄ (76 mg, 2 mmol). The reaction mixture was boiled under reflux for 90 min, and the THF was removed under reduced pressure. The aqueous solution was extracted with chloroform, and the extract was dried, evaporated, and chromatographed (silica gel) to give the carbinol **10**: 36 mg (80%); IR (CHCl₃) 1680, 1600, 1540 cm⁻¹; NMR (90 MHz, CDCl₃) δ 1.31 (t, $J = 7$ Hz, OCH₂CH₃), 1.50–2.30 (m, CHCH₂CH₂), 2.38 (s, COCH₃), 3.10–3.30 (m, C=

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CCH₂CH₂), 3.40–3.80 (m, CHCH₂OH), 11.70 (br, NH); high-resolution mass spectrum, calcd for C₁₁H₁₇NO₄ *m/e* 227.1157, found *m/e* 227.1167; *m/e* 227 (M⁺), 212 (M⁺ – CH₃), 196 (M⁺ – CH₂OH), 182 (M⁺ – C₂H₅O), 150 (M⁺ – CH₂OH – C₂H₆O), 122 (M⁺ – CH₂OH – C₂H₆O – CO).

Hydrogenation of 7. Platinum oxide (645 mg) was added to a solution of compound 7 (4.30 g, 16 mmol) in a mixture of acetic acid (84 mL) and trifluoroacetic acid (16 mL). The reaction mixture was shaken at room temperature under hydrogen at 4 atm, after 12 h a second portion of platinum oxide (65 mg) was added, and the hydrogenation was continued under the same conditions for an additional 12 h. Filtration through Celite followed by evaporation afforded a residue which was dissolved in cold chloroform (100 mL) and washed with cold dilute aqueous ammonia and with water (5 × 20 mL). Evaporation of the chloroform afforded a residue which was chromatographed on silica gel (hexan-ether) to give the following. (a) The diesters 13:¹⁶ 1.60 g (39%); IR (CHCl₃), 1725 cm⁻¹; NMR (80 MHz, CDCl₃) δ 0.92 (t, *J* = 7 Hz, CH₂CH₃), 1.26 (t, *J* = 7 Hz, 2 OCH₂CH₃), 1.40–2.60 (m, CH₃CH₂CH(CO₂) and CH(N)(CO₂)CH₂CH₂CH), 2.42 (br, NH, exchangeable with D₂O), 3.24 (m, CHCH(N)CH₂), 3.75 (dd, *J* = 9, 6.5 Hz, NCH(CO₂)CH₂), 4.13 (q, *J* = 7 Hz, OCH₂CH₃), 4.16 (q, *J* = 7 Hz, OCH₂CH₃); high-resolution mass spectrum, calcd for C₁₃H₂₃NO₄ *m/e* 257.1626, found *m/e* 257.1616; *m/e* 257 (M⁺), 184 (M⁺ – CO₂Et), 142 (M⁺ – CH₃CH₂CHCO₂Et), 138 (M⁺ – CO₂Et – C₂H₆O), 110 (M⁺ – CO₂Et – C₂H₆O – CO). Anal. Calcd for C₁₃H₂₃NO₄: C, 60.68; H, 9.01; N, 5.45. Found: C, 60.40; H, 8.92; N, 5.49. (b) The less polar isomer of the hydroxy diester 11: 1.80 g (25%); IR (CHCl₃) 1720 cm⁻¹; NMR (80 MHz, CDCl₃) δ 1.26 (t, *J* = 7 Hz, OCH₂CH₃), 1.28 (t, *J* = 7 Hz, OCH₂CH₃), 1.28 (d, *J* = 6.5 Hz, OCHCH₃), 1.50–2.40 (m, CHCH₂CH₂CH), 2.50 (dd, *J* = 6.5, 2.5 Hz, CHCH(CO₂)CH), 3.62 (br, NH and OH, exchangeable with D₂O), 3.50–4.00 (m, CH₃C-H(OH)CH and CHCH(N)CH₂), 4.16 (q, *J* = 7 Hz, OCH₂CH₃), 4.18 (q, *J* = 7 Hz, OCH₂CH₃), 4.18 (dd, *J* = 7, 6 Hz, NCHCO₂CH₂); high-resolution mass spectrum, calcd for C₁₃H₂₁NO₄ *m/e* 255.1470, found *m/e* 255.1494; *m/e* 255 (M⁺ – H₂O), 200 (M⁺ – CO₂Et), 182 (M⁺ – CO₂Et – H₂O), 154 (M⁺ – CO₂Et – C₂H₆O), 142 (M⁺ – CH₃CH(OH)CH(CO₂Et), 136 (M⁺ – CO₂Et – C₂H₆O – H₂O). (c) The more polar isomer of the hydroxy diester 11: 1.10 g (25%); IR 1720 cm⁻¹; NMR (80 MHz, CDCl₃) δ 1.26 (t, *J* = 7 Hz, OCH₂CH₃), 1.28 (d, *J* = 6.5 Hz, OCHCH₃), 1.30 (t, *J* = 7 Hz, OCH₂CH₃), 1.50–2.30 (m, CHCH₂CH₂CH), 2.46 (dd, *J* = 8, 3.5 Hz, CHCH(CO₂)CH), 2.90 (br, NH and OH, exchangeable with D₂O), 3.50–4.20 (m, CH₃CH(OH)CH, CHCH(N)CH₂, and NCH(CO₂)CH₂), 4.14 (q, *J* = 7 Hz, OCH₂CH₃), 4.18 (q, *J* = 7 Hz, OCH₂CH₃); mass spectrum, *m/e* 273 (M⁺), 255 (M⁺ – H₂O), 200 (M⁺ – CO₂Et), 182 (M⁺ – CO₂Et – H₂O), 154 (M⁺ – CO₂Et – C₂H₆O), 142 (M⁺ – CH₃CH(OH)CHCO₂Et), 136 (M⁺ – CO₂Et – C₂H₆O – H₂O). Anal. Calcd for C₁₃H₂₃NO₅: C, 57.12; H, 8.48. Found: C, 57.05; H, 8.60.

2-[1-(*tert*-Butoxycarbonyl)-2-carboxy-5-pyrrolidinyl]butanoic Acids 15. To a solution of the diesters 13 (257 mg, 1 mmol) in ethanol (5 mL) was added a solution of potassium hydroxide (168 mg, 3 mmol) in water (5 mL). After the mixture was stirred for 12 h at room temperature, the solvent was evaporated. The residue, which consisted of the potassium salts 14, was dissolved in a mixture of *tert*-butyl alcohol (2 mL) and water (1 mL), di-*tert*-butyl dicarbonate (240 mg, 1.1 mmol) was added, and the reaction mixture was stirred for 18 h, diluted with water, and extracted with *n*-pentane. The aqueous phase was acidified with KHSO₄ to pH 2 and then extracted with ethyl acetate. Evaporation of the solvent afforded the diacids 15: 258 mg (86%, mixture of the isomers 15a,b); NMR (90 MHz, CDCl₃) δ 0.97 (t, *J* = 7 Hz, CH₂CH₃), 1.15–2.20 (m, 3 CH₂), 1.48 (s, *t*-Bu), 3.0–3.4 (m, CH₂CH(CO₂)CH), 4.0–4.4 (m, NCH(CO₂)CH₂, and CH₂CH(N)CH), 10.19 (br, 2 CO₂H); *R_f* 0.45, 0.11 (silica gel plate, 98:1:1 Et₂O/MeOH/HOAc).

A sample of the diacids 15 was treated with diazomethane in ether to give the dimethyl esters 16 (quantitative):¹⁶ NMR (80 MHz, CDCl₃) δ 0.90 (t, *J* = 7 Hz, CH₂CH₃), 1.15–2.15 (m, 3 CH₂), 1.44 (s, *t*-Bu), 2.60 (m, CH₂CH(CO₂)CH), 3.69 (s, OMe), 3.72 (s,

OMe), 3.25–4.35 (m, NCH(CO₂)CH₂ and CH₂CH(N)CH); high-resolution mass spectrum, calcd for C₁₆H₂₇NO₆ *m/e* 329.1838, found *m/e* 329.1789; *m/e* 329 (M⁺), 273 (M⁺ – C₄H₉), 270 (M⁺ – CO₂Me), 256 (M⁺ – OBu), 228 (M⁺ – BuOCO, or M⁺ – CH₃CH₂CHCO₂Me), 170 (M⁺ – CO₂Me – C₄H₉ – CO₂), 138 (M⁺ – CO₂Me – BuOCO – CH₄O), 128 (M⁺ – CH₃CH₂CHCO₂Me – C₄H₉ – CO₂).

Chromatography of the mixture of the isomers 15a and 15b on a silica gel column (diethyl ether-methanol), afforded the diacid 15a followed by a mixture of 15a and 15b. The following analytical data were obtained for 15a: IR (CHCl₃), 1711, 1693 cm⁻¹; NMR (270 MHz, CDCl₃) δ 0.98 (t, *J* = 7 Hz, CH₂CH₃), 1.47 and 1.52 (2 s, *t*-Bu of two conformers), 1.57–1.77 (m, CH₂CH₃), 1.88–2.10 (m, NCHCH₂CH₂), 2.19–2.44 (m, NCH(CO₂)CH₂CH₂), 2.98–3.16 and 3.27–3.45 (2 m, CH₂CH(CO₂)CH of two conformers), 3.92–4.11 (m, NCH(CH)CH₂), 4.32–4.60 (m, NCH(CO₂)CH₂), 10.61 (br, 2 CO₂H); high-resolution mass spectrum, calcd for C₁₃H₂₂NO₄ *m/e* 256.1549, found *m/e* 256.1559; *m/e* 256 (M⁺ – CO₂H), 245 (M⁺ – C₄H₉), 228 (M⁺ – BuO), 214 (M⁺ – CH₃CH₂CHCO₂H), 200 (M⁺ – BuOCO) 156 (M⁺ – BuOCO – CO₂), 138 (C₈H₁₂NO⁺), 110 (C₈H₁₂NO⁺ – CO); the low-resolution mass spectrum shows a weak peak at *m/e* 301 (M⁺).

***p*-Nitrobenzyl Esters of the Dicarboxylic Acid 15a.** A solution of the diacid 15a (273 mg, 0.91 mmol), dicyclohexylamine (164 mg, 0.91 mmol), *p*-nitrobenzyl chloride (187 mg, 1.1 mmol), and potassium iodide (18 mg, 0.01 mmol) in dry DMF (20 mL) was heated under argon at 100 °C for 19 h. The reaction mixture was brought to room temperature and diluted with ethyl acetate (50 mL). The precipitated dicyclohexylammonium hydrochloride was filtered off, and the filtrate was washed with water, dried, and evaporated. The residue was chromatographed on silica gel (20 g) to give the following.

(a) The di-*p*-nitrobenzyl ester 19a: 120 mg (23%); eluted with ether-hexane (1:1); mp 111–112 °C (from CH₂Cl₂-hexane); IR (CHCl₃) 1730, 1690 cm⁻¹; NMR (270 MHz, CDCl₃) δ 0.86 (t, *J* = 7 Hz, CH₂CH₃), 1.36 and 1.47 (2 s, *t*-Bu of two conformers), 1.60–1.82 (m, CH₂CH₃), 1.84–2.10 (m, NCHCH₂CH₂ and one proton of NCH(CO₂)CH₂CH₂), 2.10–2.36 (m, one proton of NCH(CO₂)CH₂CH₂), 2.71–3.02 (m, CH₂CH(CO₂)CH), 4.01–4.28 (m, NCH(CH)CH₂), 4.28–4.53 (m, NCH(CO₂)CH₂), 5.17 and 5.25 (AB q, *J* = 14 Hz, 2 CH₂Ar), 7.51 and 8.20 (AB q, *J* = 8.8 Hz, 2 C₆H₄NO₂); high-resolution mass spectrum, calcd for C₁₄H₂₄N₃O₉ *m/e* 498.1506, found *m/e* 498.1511; *m/e* 498 (M⁺ – BuO), 470 (M⁺ – BuOCO), 391 (M⁺ – CO₂PNB), 291 (M⁺ – CO₂PNB – C₄H₉OCO), 249 (M⁺ – CH₃CH₂CHCO₂PNB – C₄H₉OCO); the low-resolution mass spectrum shows a molecular ion at *m/e* 571 (M⁺).

(b) The *p*-nitrobenzyl ester 18a: 55 mg (14%); eluted with ether-hexane (1:1); IR (CHCl₃) 1730–1690 cm⁻¹ (broad band); NMR (270 MHz, CDCl₃) δ 0.90 (t, *J* = 7 Hz, CH₂CH₃), 1.47 (s, *t*-Bu), 1.56–1.83 (m, CH₂CH₃), 1.84–2.05 (m, NCH(CH)CH₂), 2.04–2.51 (m, CH₂CH(CO₂)CH and NCH(CO₂)CH₂CH₂), 3.84–4.23 (m, NCH(CH)CH₂), 4.23–4.50 (m, NCH(CO₂)CH₂), 5.12 and 5.27 (AB q, *J* = 13.5 Hz, CH₂Ar), 7.52 and 8.21 (AB q, *J* = 8.8 Hz, C₆H₄NO₂); high-resolution mass spectrum, calcd for C₂₀H₂₇N₂O₆ *m/e* 391.1868, found *m/e* 391.1849; *m/e* 391 (M⁺ – CO₂H), 335 (M⁺ – BuOCO), 291 (M⁺ – BuOCO – CO₂), 214 (M⁺ – CH₃CH₂CHCO₂PNB), 156 (M⁺ – CO₂PNB – C₄H₉OCO), 110 (C₇H₁₂N⁺).

A sample of 18a was treated with diazomethane in ether to give the methyl ester 21a (quantitative): high-resolution mass spectrum, calcd for C₁₈H₂₁N₂O₇ *m/e* 377.1348, found *m/e* 377.1360; *m/e* 377 (M⁺ – OBu), 349 (M⁺ – BuOCO), 128 (M⁺ – CH₃CH₂CHCO₂PNB – C₄H₉OCO); the low-resolution mass spectrum shows a molecular ion at *m/e* 450.

(c) The *p*-nitrobenzyl ester 17a: 113 mg (29%); eluted with ether; mp 143–145 °C (CH₂Cl₂-hexane); NMR (270 MHz, CDCl₃) δ 0.91 (t, *J* = 7 Hz, CH₂CH₃), 1.42 (s, *t*-Bu), 1.60–1.83 (m, CH₂CH₃), 1.83–2.12 (m, NCH(CH)CH₂CH₂ and one proton of NCH(CO₂)CH₂CH₂), 2.12–2.42 (m, one proton of NCH(CO₂)CH₂CH₂), 2.48–2.85 (m, CH₂CH(CO₂)CH), 4.01–4.29 (m, NCH(CH)CH₂), 4.29–4.55 (m, NCH(CO₂)CH₂), 5.14 and 5.26 (AB q, *J* = 13.5 Hz, CH₂Ar), 7.53 and 8.22 (AB q, *J* = 8.8 Hz, C₆H₄NO₂); high-resolution mass spectrum, calcd for C₁₇H₂₀N₂O₈ *m/e* 380.1219, found *m/e* 380.1253; *m/e* 380 (M⁺ – C₄H₉), 363 (M⁺ – OBu), 349 (M⁺ – CH₃CH₂CHCO₂H), 335 (M⁺ – BuOCO), 291

(16) Epimers a and b were not differentiated in the 80-MHz NMR spectrum. In the case of 13, the existence of two epimers was deduced from their derivatives and in the case of 16 from their precursors.

(M^+ - BuOCO - CO₂), 256 (M^+ - CO₂PNB), 249 (M^+ - C₄H₈OCO - CH₃CH₂CHCO₂H), 156 (M^+ - C₄H₈OCO - CO₂PNB), 138 (C₈H₁₂NO⁺); the low-resolution mass spectrum shows a low-intensity molecular ion at m/e 436.

A sample of 17a was treated with diazomethane in ether to give the methyl ester 20a (quantitative); high-resolution mass spectrum, calcd for C₁₇H₂₁N₂O₆ m/e 349.1399, found m/e 349.1405; m/e 349 (M^+ - BuOCO), 249 (M^+ - CH₃CH₂CHCO₂CH₃ - C₄H₈OCO), 170 (M^+ - CO₂PNB - C₄H₈OCO); the low-resolution mass spectrum shows a molecular ion at m/e 450.

***p*-Nitrobenzyl (±)-6-Ethylcarbapenam-3-carboxylates 25a and 25b.** 2-[1-(*tert*-butoxycarbonyl)-2-(*p*-nitrobenzyloxy-carbonyl)-5-pyrrolidinyl]butanoic acid (17a; 140 mg, 0.32 mmol) was dissolved in 1 N HCl in EtOAc (4 mL) and stirred for 2.5 h at room temperature. The solvent was evaporated, and CCl₄ was added and evaporated again. The residue was washed with ether and dried to give the hydrochloride of 2-[2-(*p*-nitrobenzyloxy-carbonyl)-5-pyrrolidinyl]butanoic acid (22a) as a solid (119 mg, quantitative). To a cold solution (0 °C) of the hydrochloride of 22a (112 mg, 0.30 mmol) in dry CH₂Cl₂ (180 mL) were added pyridine (34 mg, 0.43 mmol) and 1-(3'-dimethylamino-propyl)-3-ethylcarbodiimide hydrochloride (154 mg, 0.80 mmol). The reaction mixture was stirred at 0 °C under argon for 1 h and then for 22 h at room temperature, and it was then washed with ice-cold water (4 × 50 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue (silica gel, 0.04–0.063 mm; ether-hexane, 2:1) afforded the carbapenam 25a: 39 mg (41%); IR (CH₂Cl₂) 1760, 1742, 1610, 1522 cm⁻¹; NMR (270 MHz, CD₂Cl₂) δ 0.95 (t, J = 7.4 Hz, 8-H₃), 1.40–1.72 (m, 7-H₂), 1.74–1.96 (m, 1-H₂), 2.30 (m, 2-H₂), 3.13 (dddd, J = 9, 8, 5, 1 Hz, 6-H), 3.78 (ddd, J = 9, 6, 5 Hz, 5-H), 3.92 (dt, J = 5, 1 Hz, 3-H), 5.21 and 5.28

(AB q, J = 13.4 Hz, CH₂Ar), 7.58 and 8.22 (aB q, J = 8.8 Hz, C₆H₄NO₂); high-resolution mass spectrum, calcd for C₁₆H₁₈N₂O₅ m/e 318.1215, found m/e 318.1211; m/e 318 (M^+), 301 (M^+ - OH), 290 (M^+ - CO), 249 (M^+ - CH₃CH₂C=C=O), 182 (M^+ - PNB), 138 (M^+ - CO₂PNB), 137 (CH₃C₆H₄NO₂⁺), 136 (PNB⁺). Further elution with ether gave the dimer 26: 10 mg (10%); IR (CH₂Cl₂) 1730, 1660 cm⁻¹; high-resolution mass spectrum, calcd for C₃₂H₃₆N₄O₁₀ m/e 636.2431, found m/e 636.2436; m/e 636 (M^+), 607 (M^+ - Et), 456 (M^+ - CO₂PNB), 428 (M^+ - CO₂PNB - CO).

When a mixture of the two isomers 15a and 15b was subjected to the sequence of reactions described above for the conversion of 15a into 25a, a mixture of the *cis*-6-ethylcarbapenam 25a and its trans isomer 25b was obtained. This mixture exhibits the same IR and high-resolution mass spectra as pure 25a. NMR of 25b, obtained by subtraction of the spectrum of 25a from that of the mixture (270 MHz, CD₂Cl₂): δ 1.00 (t, J = 7 Hz, 8-H₃), 1.30–1.75 (m, 7-H₂), 1.75–2.20 (m, 1-H₂), 2.25–2.39 (m, 2-H₂), 2.86 (ddd, J = 8, 6, 2 Hz, 6-H), 3.46 (ddd, J = 8, 5, 2 Hz, 5-H), 3.93 (t, J = 5 Hz, 3-H), 5.22 and 5.30 (AB q, J = 13.5 Hz, CH₂Ar), 7.58 and 8.21 (AB q, J = 9 Hz, C₆H₄NO₂).

Registry No. (±)-6, 84911-17-1; (±)-7, 84927-07-1; (±)-9, 84911-19-3; (±)-10, 84911-20-6; 11, 84911-22-8; (±)-13a, 84911-21-7; (±)-13b, 84984-88-3; (±)-14a, 84911-23-9; (±)-14b, 84984-89-4; (±)-15a, 84911-24-0; (±)-15b, 84984-90-7; (±)-16a, 84911-25-1; (±)-16b, 84984-91-8; (±)-17a, 84911-29-5; (±)-18a, 84911-27-3; (±)-19a, 84911-26-2; (±)-20a, 84911-30-8; (±)-21a, 84911-28-4; (±)-22a-HCl, 84911-31-9; (±)-25a, 84911-32-0; (±)-25b, 84984-92-9; 26, 84911-33-1; ethyl (±)-pyroglutamate, 66183-71-9; ethyl (±)-2-bromo-3-oxobutyrate, 84911-18-2; diethyl 2-bromomalonate, 685-87-0.

Reaction of Sydnones with Oxygen[†]

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The reaction of 3-benzyl- and 3-(*p*-chlorobenzyl)-4-phenylsydnones (1a and 1b) and of 3-benzylsydnone (1c) with oxygen at room temperature in the dark is described. Possible rationalizations for the formation of the products obtained are suggested.

In the context of our study of the decarboxylations of α -nitrosamino acids,¹ some anhydro- α -nitrosamino acids² (sydnones³) were prepared. Sydnones, unsubstituted at the 4-position, are known to undergo electrophilic aromatic substitution.³ Thus the conversion of α -nitrosamino acids to sydnones, followed by introduction of a substituent at the 4-position of the sydnone ring and regeneration of the α -nitrosamino acid, would constitute a means of obtaining *N*-nitrosamines⁴ from glycines, via the decarboxylation of *N*-nitrosoglycines.^{1,5}

During the course of the recrystallization of 3-benzyl-4-phenylsydnone (1a), a benzene-ether solution of sydnone 1a upon long standing deposited colorless crystals, identified as benzoic acid. This result was surprising in view of the fact that the chemistry of sydnones had been studied extensively and thus was believed to be understood rather

well. Yet no precedent could be found in the literature with regard to the fact that sydnones may be sensitive to oxygen. Recently George and his co-workers⁶ have reported the *photosensitized* oxidation of sydnones with *singlet* oxygen. Therefore it was decided to undertake the investigation of the reaction of sydnones with oxygen.

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(4) This would appear to be perhaps one true case of reverse *polarization* (also called *umpolung*) as the 4-position of the sydnone is the position that is flanked *both* by the nitrosamino and the carbonyl group of α -nitrosamino acids.

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[†]Taken in part from the Ph.D. Thesis of M. Nakajima, University of Massachusetts, Sept 1982. This is the 10th article dealing with the chemistry of *N*-nitrosamines and related compounds. For the previous publication, see K. Kano, C. A. Kelly, and J.-P. Anselme, *Tetrahedron Lett.*, 1427 (1982).