

Stereoselective Synthesis of L-Guluronic Acid Alginates

Jasper Dinkelaar, Leendert J. van den Bos, Wouter F. J. Hogendorf, Gerrit Lodder, Herman S. Overkleeft, Jeroen D. C. Codée,* and Gijsbert A. van der Marel*^[a]

Dedicated to Professor Jan Reedijk on the occasion of his 65th birthday and his retirement

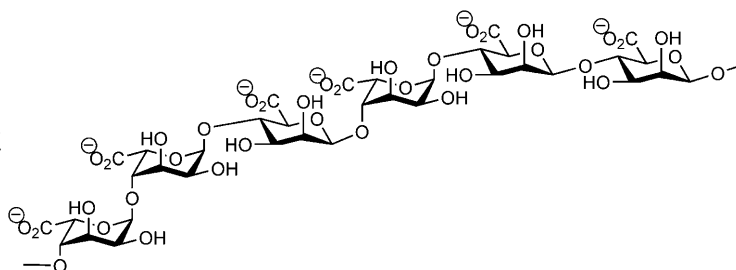
Abstract: The glycosylation properties of gulopyranosides have been mapped out, and it is shown that gulose has an intrinsic preference for the formation of 1,2-*cis*-glycosidic bonds. It is postulated that this glycosylation behaviour originates from nucleophilic attack at the oxacarbenium ion, which adopts the most favourable ³H₄ conformation. Building on the stereoselectivity of gulose, a guluronic acid alginate trisaccharide was assembled for the first time by using gulopyranosyl building blocks.

Keywords: glycosylation • gulose • oligosaccharides • oxacarbenium ions • oxidation • thioglycosides

Introduction

Alginates are naturally occurring polysaccharides composed of 1,2-*cis*-linked L-guluronic and D-mannuronic acid residues that are arranged in homopolymer (i.e., polyguluronate and polymannuronate) or heteropolymer (a mixed sequence of these residues) sections (Scheme 1). Alginate polymers, isolated from marine brown algae (Phaeophyta),^[1] are used for food, textile, and pharmaceutical purposes.^[2] Bacteria, such as *Pseudomonas aeruginosa*, also produce alginates as exopolysaccharides, and alginate oligomers appear to have cytokine-inducing activities by binding to Toll-like receptors (TLRs) 2 and 4.^[3] As part of a program to evaluate the immunomodulating properties of carbohydrate structures, we set out to develop synthetic routes towards well-defined alginate fragments.

In this framework, we recently reported the synthesis of a 1,2-*cis*-linked D-mannuronic acid trimer by the use of 1-(*S*)-phenylmannuronic acid pyranosides.^[4] The present study is focused on the development of a synthetic route to the cor-



Scheme 1. Alginate oligosaccharide.

responding 1,2-*cis*-linked L-guluronic acid oligomers. Analogously to other acidic oligosaccharides,^[5] the carboxylate function in L-guluronic acid oligomers can be introduced at the monosaccharide stage by the use of suitably protected L-guluronate ester building blocks or at the oligomer level by selective deprotection and oxidation of the incorporated orthogonally protected L-gulose residues. Which route of synthesis is more efficient relies on the glycosylation properties of the L-guluronate ester and L-gulose donors, respectively. Up until now, L-gulose donors have only been employed in the total synthesis of bleomycin,^[6] in which L-guluronate ester donors are completely unexplored.

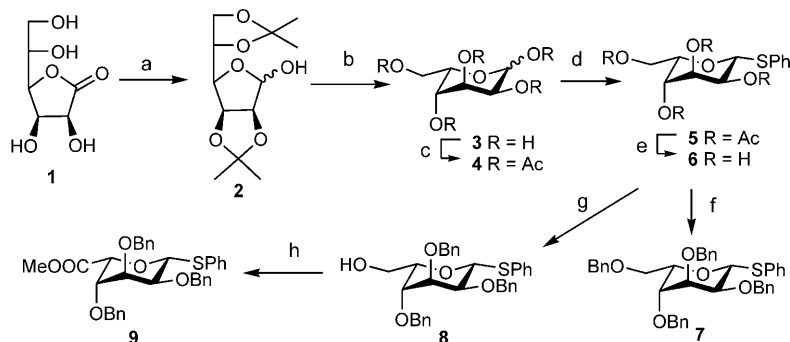
Herein, we describe an evaluation of the glycosylating properties of gulose and guluronate ester donors and the first synthesis of a guluronic acid trimer. It is revealed that gulopyranose has a very high tendency to form 1,2-*cis*-linkages without the need to incorporate stereodirecting groups, as is the case for most other pyranosides.

[a] J. Dinkelaar, Dr. L. J. van den Bos, W. F. J. Hogendorf, Dr. G. Lodder, Prof. Dr. H. S. Overkleeft, Dr. J. D. C. Codée, Prof. Dr. G. A. van der Marel
Leiden Institute of Chemistry
Leiden University, P.O. Box 9502
2300 RA Leiden (The Netherlands)
Fax: (+31) 715-274-307
E-mail: jcodee@chem.leidenuniv.nl
marel_g@chem.leidenuniv.nl

Supporting information for this article is available on the WWW under <http://dx.doi.org/10.1002/chem.200800960>.

Results and Discussion

The rare L-gulose and L-guluronic acid are C-5-epimers of D-mannose and mannuronic acid, respectively, and not readily commercially available. Therefore, our first aim was to develop a scalable route of a synthesis for L-gulopyranose, provided with an anomeric thio function for ensuing glycosylations. A practical approach was found in the use of L-gulonic acid γ -lactone (**1**), commercially available at reasonable cost. The transformation of this lactone into β -S-phenylgulopyranose **6** is depicted in Scheme 2 and started with



Scheme 2. Synthesis of β -(S)-phenyl-L-guloses **6**, **7**, and **9**. a) i. Acetone, dimethoxypropane, H_2SO_4 ; ii. toluene, DIBAL-H, 0°C (84%, over 2 steps); b) 20% TFA in H_2O (quant.); c) pyridine, Ac_2O (72%); d) i. AcOH, HBr; ii. EtOAc, Bu_4NHSO_4 , Na_2CO_3 , PhSH, H_2O (66%, over 2 steps); e) MeOH, NaOMe (cat.; 97%); f) DMF, BnBr, NaH, 0°C \rightarrow room temperature (89%); g) i. pyridine, TrCl; ii. DMF, BnBr, NaH, 0°C \rightarrow room temperature; iii. MeOH, *p*-TsOH (cat.; 88%, over 3 steps); h) i. CH_2Cl_2 , H_2O , TEMPO, BAIB; ii. DMF, K_2CO_3 , MeI (72%, over 2 steps). BAIB = [bis(acetoxy)iodo]benzene, DIBAL-H = diisobutylaluminium hydride, TEMPO = 2,2,6,6-tetramethylpiperidin-1-oxyl, TFA = trifluoroacetic acid, TrCl = triphenylmethyl chloride, *p*-TsOH = *para*-toluenesulfonic acid.

protection of the four hydroxy groups by treatment of **1** with dimethoxypropane. The lactone was reduced by using DIBAL-H in toluene to yield **2**,^[7] and acidic hydrolysis of both acetones delivered the hygroscopic L-gulose **3**, which was immediately acetylated to give per-acetyl gulose **4**. A Lewis acid-mediated introduction of the anomeric thiophenol resulted in an inseparable α/β mixture of thioglycosides. Alternatively, the transformation of **4** into the anomeric bromide moiety and ensuing treatment with PhSH under phase-transfer conditions^[8] gave thioguloside **5** as a single anomer, which was deacetylated to furnish key β -(S)-phenylgulopyranoside **6**. The sequence of reactions to furnish **6** required only one chromatographic purification and can be performed on a 500-mmol scale (100 g). Standard protecting-group manipulations on **6** delivered gulose donor **7** and guluronate ester **9**^[9] devoid of any stereodirecting protecting groups.

The glycosylating properties of these donors were examined in the condensation with acceptors **10–12**.^[10] Each donor was activated with diphenyl sulfoxide (Ph_2SO) and triflic anhydride (Trf_2O) in the presence of TTBP in dichloromethane at -78 or -45°C for donors **7** and **9**, respectively. After complete activation, the temperature was adjusted to -78°C , 1.5 equivalents of the acceptor was added,

and the reaction mixture was allowed to warm to 0°C . The condensation of primary acceptor **10** with gulose **7** almost exclusively gave the 1,2-*cis* product, whereas the reaction of guluronate ester donor **9** with **10** proceeded slightly less stereoselectively (Table 1). Glycosylation of the secondary acceptors **11** and **12** proceeded in a highly α -selective manner for both gulose and guluronate ester. Apparently, gulose and guluronate ester have an unusually strong preference for the formation of 1,2-*cis* glycosidic bonds.

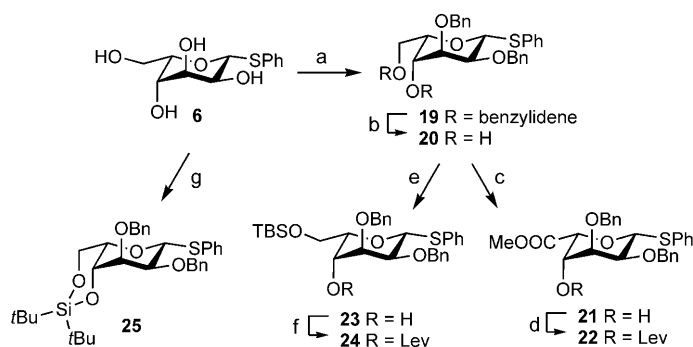
To explore the influence of different protecting groups on the α -selectivity and to find the most convenient building block for the synthesis of guluronic acid alginates, we next investigated one guluronate ester (i.e., **22**) and three differently protected gulose donors (i.e., **19**, **24**, and **25**). The conformationally constrained 4,6-*O*-benzylidene gulose **19** and 4,6-*O*-di-*tert*-butylsilylidene (DTBS) gulose **25** were selected on the basis of their reputation to influence the stereochemical outcome of glycosylations.^[11,12] Gulose donor **24** and guluronate ester donor **22**, with a selectively removable levulinoyl (Lev) group at the C-4 position, allow elongation to alginate oligomers with minimal protective-group manipulation. All

Table 1. Glycosylations of β -(S)-phenyl-L-gulose donors **7**^[a] and **9**^[b].

Entry	Donor	Acceptor	Yield [%]	(α/β)	Product
1	7 ^[a]	10	71	(13:1)	13
2	7 ^[a]	11	91	(10:1)	14
3	7 ^[a]	12	73	(>20:1)	15
4	9 ^[b]	10	73	(3:1)	16
5	9 ^[b]	11	94	(>20:1)	17
6	9 ^[b]	12	79	(10:1)	18

[a] Ph_2SO , TTBP, CH_2Cl_2 , -78°C , Trf_2O , 10 min, nucleophile, to 0°C .
[b] Ph_2SO , TTBP, CH_2Cl_2 , -45°C , Trf_2O , 10 min, then -78°C , nucleophile, to 0°C .

four donor building blocks were assembled from phenyl-1-thio- β -L-gulopyranoside (**6**; Scheme 3). The instalment of the 4,6-*O*-benzylidene group on **6** and subsequent benzylation yielded 4,6-*O*-benzylidenegulose donor **19**. Guluronate

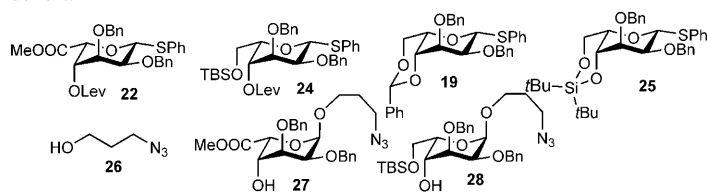


Scheme 3. Synthesis of gulose building blocks **19**, **22**, **24**, and **25**. a) i. MeCN, PhCH(OMe)₂, *p*-TsOH (cat.); ii. DMF, BnBr, NaH, 0°C → room temperature (70%, over 2 steps); b) MeOH, *p*-TsOH (cat.; quant.); c) i. CH₂Cl₂, H₂O, TEMPO, BAIB; ii. Et₂O, TMS diazomethane (49%, over 2 steps); d) pyridine, dioxane, Lev₂O (95%); e) DMF, TBDMSCl, imidazole, 0°C → room temperature (90%); f) pyridine, dioxane, Lev₂O (94%); g) i. pyridine, (*t*Bu)₂Si(OTf)₂, -20°C → room temperature; ii. DMF, BnBr, NaH, 0°C → room temperature (88%, over 2 steps). DMF = *N,N*-dimethylformamide, TBDMSCl = *tert*-butyldimethylsilyl chloride.

ester donor **22** was accessible by the acidic cleavage of the benzylidene group in **19**, regio- and chemoselective oxidation of the primary hydroxy group in diol **20** with TEMPO/BAIB, methylation of the carboxylic acid with trimethylsilyl (TMS) diazomethane, and levulinoylation of the hydroxy group at C-4 in **21**. Protection of the hydroxy group at C-6 in diol **20** with the *tert*-butyldimethylsilyl (TBDMS) group and levulinoylation of the remaining alcohol furnished donor **24**. Treatment of tetraol **6** with di-*tert*-butylsilyl bistriflate in pyridine at -20°C followed by benzylation of the crude product afforded di-*O*-*tert*-butylsilylidene gulose **25** (Scheme 3).

With the thioguloside donors (i.e., **19**, **22**, **24**, and **25**) in hand, we first focused on the glycosylating properties of thioguluronate ester **22** (Table 2). For comparative reasons, **22** was condensed with model acceptors **10** and **11**. In line with the glycosylations of donor **9**, levulinoyl donor **22** was moderately α -selective with the primary alcohol **10** and completely α -selective with secondary acceptor **11** (Table 2, entries 1 and 2, respectively). Next, donor **22** was coupled with 3-azidopropanol (**26**) and 3-azidopropylmethyl-2,3-*O*-benzyl- α -L-gulopyranoside uronate (**27**). In contrast to the gulosylations described above, the β product prevailed in the condensation

Table 2. Glycosylations of orthogonally protected β -D-phenyl-L-gulose donors.^[a,b]

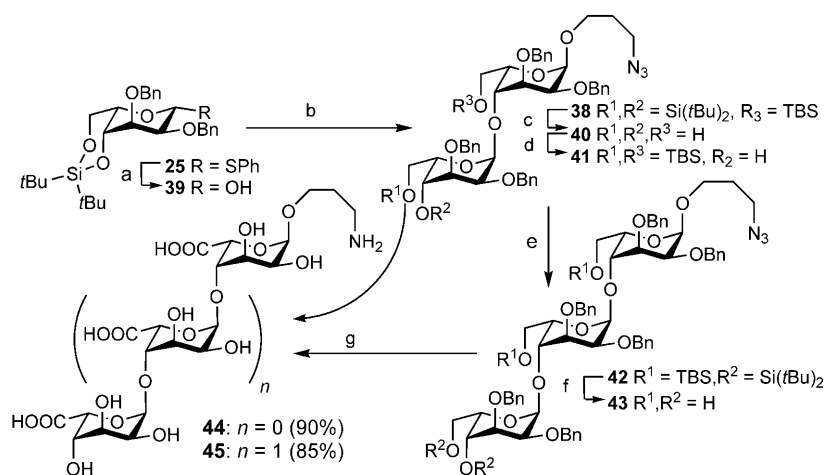


Entry	Donor	Acceptor	Yield [%]	(α/β)	Product
1	22 ^[a]	10	66	(1:3)	29
2	22 ^[a]	11	64	(>20:1)	30
3	22 ^[a]	26	77	(1:3)	31
4	22 ^[a]	27	34	(3:1)	32
5	24 ^[b]	26	86	(3:1)	33
6	24 ^[b]	28	48	(6:1)	34
7	19 ^[b]	26	88	(3:1)	35
8	19 ^[b]	28	45	(6:1)	36
9	25 ^[b]	26	75	(5:1)	37
10	25 ^[b]	28	48	(10:1)	38

[a] Ph₂SO, TTBP, CH₂Cl₂, -45°C, Tf₂O, 10 min, then -78°C, nucleophile, to 0°C. [b] Ph₂SO, TTBP, CH₂Cl₂, -78°C, Tf₂O, 10 min, nucleophile, to 0°C.

of **22** and the rather reactive primary alcohol **26** (Table 2, entry 3). Diuronate **30** was formed in low yield from **22** and **27**, again with α -selectivity (Table 2, entry 4). The gulose donors **19**, **24**, and **25** all predominantly provided the 1,2-*cis* linked products, with both the primary and secondary alcohol acceptors. Thus, the presence of the carboxylic acid ester at C-5 does not contribute favourably to the formation of the 1,2-*cis* gulosidic linkage. Because the DTBS donor **25** shows the best α -selectivity of the three thioguloses examined, this guloside was further examined in the synthesis of a guluronic alginate trimer (Scheme 4).

It is clear that the guluronate ester and hydroxy group at C-4 in gulose are poor nucleophiles, thus leading to moder-



Scheme 4. Synthesis of the alginate trisaccharide **45**. a) CH₂Cl₂, NIS, TFA (95%); b) Ph₂SO, TTBP, -60°C, Tf₂O, **28**, -60°C → room temperature (84%; α/β = 10:1); c) THF, TBAF (87%); d) DMF, TBDMSCl, imidazole (78%); e) **37**, Ph₂SO, TTBP, -60°C, Tf₂O, then **41**, -60°C → room temperature (42%; α); f) THF, TBAF (83%); g) i. CH₂Cl₂, *t*BuOH, H₂O, TEMPO, BAIB; ii. *t*BuOH, Pd/C, H₂ (**44**: 90%, **45**: 85%, over 2 steps). NIS = *N*-iodosuccinimide, TTBP = tri-*tert*-butylpyrimidine.

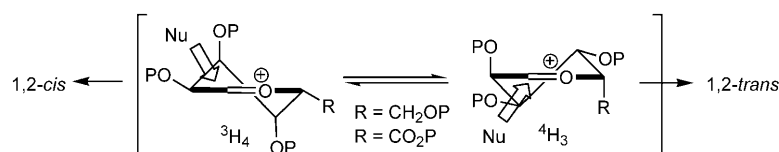
ate yields in the gulosylations (Table 2, entries 4, 6, 8, and 10). We attribute these moderate yields to a reactivity mismatch of the coupling partners^[13] because in the condensation reactions all the donor gulose was consumed, whereas some of the acceptor remained untouched. Other promoter systems were investigated to modulate the reactivity of the activated donor species. Iodonium-mediated glycosylations with NIS/trimethylsilyl trifluoromethanesulfonate (TMSOTf) and iodonium dicollidine perchlorate (IDCP)^[4,14] resulted in the same stereoselectivity, although the yield did not improve (results not shown). Next, a dehydrative condensation strategy, originally devised by Gin and co-workers,^[15] was explored. In our experience, this type of glycosylation is well suited to unreactive nucleophiles because the activated sulfoxonium species is relatively stable and survives longer at higher temperatures.^[16] Therefore, we hydrolyzed thiogulose **25** by using NIS/TFA^[17] to give hemiacetal donor **39** (Scheme 4). This lactol was activated by Ph₂SO/Tf₂O and coupled to gulose **28** to provide disaccharide **38** in a slightly improved yield (55%). Importantly, the excellent α -stereoselectivity of DTBS-protected gulose **25** was maintained and was thus shown to be independent of the activation method. Changing the donor/acceptor ratio (1.2:1 to 2:1) further increased the yield to 84%.

Next, we elongated disaccharide **38** by liberation of the hydroxy group at C-4' and subsequent glycosylation with two equivalents of **39**. Trimer **42** was obtained as a single diastereomer, albeit in a moderate 42% yield. The introduction of the carboxylate functions was achieved by desilylation of dimer **38** and trimer **42** and subsequent oxidation mediated with TEMPO/BAIB. It was observed that primary alcohols **40** and **43** were smoothly transformed into the corresponding aldehydes, but that ensuing oxidation to the acid was troublesome. The addition of *t*BuOH to homogenise the biphasic reaction mixture of dichloromethane and water enhanced the rate of aldehyde hydration, thereby allowing the efficient oxidation of the lipophilic substrates.^[18] Hydrogenolysis completed the synthesis, and the di- and triacids **44** and **45** were isolated in excellent yield over the two final steps.

Finally, it is appropriate to comment on the unusually high α -selectivity displayed by the gulopyranosides described herein. In most pyranosides, an alkoxide aglycon prefers to adopt an axial position as dictated by the anomeric effect.^[19] In gulose, however, the anomeric effect will be largely offset by the 1,3-diaxial interaction of the α -oriented aglycon and the substituent at C-3.^[20] Therefore, we dismiss the anomeric effect to be the basis of the observed selectivities. Anomeric triflates have convincingly been demonstrated to be intermediates in sulfoxonium-mediated glycosylations, as practiced herein.^[21] In most cases, the anomeric α -triflates serve as a reservoir for the actual glycosylation species, being a close ion pair (CIP) or a solvent-separated ion pair

(SSIP). In the gulosylations at hand, it is difficult to predict which anomeric triflate will be the most stable, because also in this case the anomeric effect is counterbalanced by steric interactions and both the α - and β -triflates may exist in solution. It is unlikely that the α -stereoselectivity in gulosylation arises from the selective S_N2-type displacement on the β -triflate or β -CIP.

Rather, we believe that the high 1,2-*cis* selectivity originates from the conformational preferences of the intermediate solvent-separated oxacarbenium ion. Lemieux and Huber postulated that the relative stabilities of the oxacarbenium conformers can influence the stereochemical outcome of glycosylation reactions.^[22] Recently, the conformational restriction of oxacarbenium ions was exploited in the stereoselective synthesis of β -arabinofuranoses.^[23] Computational^[24] and experimental^[25] data have established that substituents on the pyranose ring influence the stability of the oxacarbenium intermediate and thereby affect the outcome of a glycosylation reaction. It was shown that electronegative substituents at C-3 and C-4 prefer to adopt a pseudoaxial orientation, thereby minimizing their electron-withdrawing nature. Substituents at C-2 and C-5 prefer to adopt a pseudoequatorial orientation because of hyperconjugative effects for the former^[26] and steric reasons for the latter.^[25] When we apply these findings to the two likely half-chair conformations of the L-gulose oxacarbenium ion intermediate, it becomes clear that all substituents occupy their preferred orientation in the ³H₄ conformer (Scheme 5). On the contrary, all the substituents are in disfavoured positions in the ⁴H₃ conformer. An incoming nucleophile will approach the oxacarbenium ion in a pseudoaxial trajectory, with a preference for the diastereotopic face that leads to the more favourable chair-like product.^[27] If no prohibitive steric interactions evolve in the transition state, thus leading to the coupled product, the L-gulose oxacarbenium ion will react from the ³H₄ conformer to form the α product. The protecting-group pattern on the gulosides only has a marginal effect on the stereochemical outcome of the condensation reactions. Although the cyclic protecting groups in **19** and **25** restrict the conformational freedom of the gulopyranoses, they do not a priori rule out either of the two half-chair oxacarbenium ion conformers because they can be accommodated in both constellations. Participation of the acyl group at C-4 in **24**, as has been suggested in α -galactosylations,^[28] does not play a decisive role in the gulosylations described herein. In comparison with the gulose donors, the stereoselectivity of the guluronate esters is somewhat decreased. The counterproductive effect of the carboxylic ester could suggest that this electron-withdrawing group at C-5 prefers



Scheme 5. Proposed oxacarbenium ion conformers.

to adopt a pseudoaxial orientation in the oxacarbenium ion, thereby shifting the ratio of the two half-chair intermediates and the stereochemical outcome of the glycosylations.

Conclusion

In closing, we have mapped out the glycosylation properties of gulopyranosides and have shown that gulose has an intrinsic preference for the formation of a 1,2-*cis*-glycosidic bond. It is postulated that this glycosylation behaviour originates from nucleophilic attack at the oxacarbenium ion, which adopts the most favourable conformation, that is, ³H₄. Building on the stereoselectivity of gulose, a guluronic acid alginate trisaccharide has been assembled for the first time. Insight into the influence of the stereochemistry of substituents on the pyranoside ring on the stereochemical outcome of a glycosylation will aid the future development of stereoselective glycosylation reactions.

Experimental Section

Dichloromethane was heated to reflux with P₂O₅ and distilled before use. Trifluoromethanesulfonic anhydride was distilled from P₂O₅. Traces of water in the donor and acceptor glycosides, diphenyl sulfoxide, and TTBP were removed by coevaporation with toluene. All other chemicals were used as received from Acros, Fluka, Merck, and Schleicher & Schue. Column chromatography was performed on Merck silica gel 60 (0.040–0.063 mm). Analysis by TLC was conducted on HPTLC aluminum sheets (Merck; silica gel 60, F245). Compounds were visualised by UV absorption (λ =245 nm) by spraying with 20% H₂SO₄ in ethanol or with a solution of [Mo₇(NH₄)₆O₂₄] \cdot 4H₂O (25 g L⁻¹), [Ce(NH₄)₄(SO₄)₄] \cdot 2H₂O (10 g L⁻¹), or 10% H₂SO₄ in H₂O followed by charring at \pm 140°C. ¹H and ¹³C NMR spectra were recorded on Bruker AV 400 (400 and 100 MHz, respectively), AV 500 (500 and 125 MHz, respectively), or DMX 600 (600 and 150 MHz, respectively) spectrometers. NMR spectra were recorded in CDCl₃ with the chemical shift δ relative to trimethylsilane (TMS) unless stated otherwise. Optical rotations were measured on a Propol automatic polarimeter. High-resolution (HR) mass spectra were recorded on a LTO-orbitrap (thermoelectron). IR spectra were recorded on a Shimadzu FTIR-8300 spectrometer and are reported in cm⁻¹.

2,3,5,6-Di-O-isopropylidene- α -L-gulofuranose (2): H₂SO₄ (5 drops) was added to a suspension of L-gulonic acid γ -lactone (**1**; 40.0 g, 225 mmol) and 2,2-dimethoxypropane (71 mL, 788 mmol) in acetone (800 mL). The reaction mixture was stirred overnight and quenched with Et₃N until neutral pH. The acetone was removed in vacuo, and the residue was taken up in Et₂O and washed twice with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was dissolved in toluene (2.0 L) and DIBAL-H (137 mL, 1.7 M solution in toluene) was slowly added at 0°C. The reaction was quenched with EtOAc (20 mL) after 20 min, 2 M NaOH (215 mL) was added under vigorous stirring, and the reaction mixture was filtered over Hyflo Gel. The aqueous layer was extracted twice with EtOAc and the combined organic layers dried over MgSO₄ and concentrated in vacuo to afford **2** as a white solid (49 g, 84%). ¹H NMR (400 MHz, CDCl₃): δ =1.29 (s, 3H, CH₃ isoprop), 1.39 (s, 3H, CH₃ isoprop), 1.45 (s, 6H, 2 \times CH₃ isoprop), 3.44 (d, 1H, *J*=2.0 Hz, OH), 3.74 (dd, 1H, *J*=7.2, 8.4 Hz, H-6), 4.13 (dd, 1H, *J*=3.6, 8.4 Hz, H-4), 4.21 (dd, 1H, *J*=6.4, 8.4 Hz, H-6), 4.37 (dd, 1H, *J*=6.8, 8.4 Hz, H-5), 4.63 (d, 1H, *J*=5.6 Hz, H-2), 4.70 (dd, 1H, *J*=3.8, 5.8 Hz, H-3), 5.46 ppm (d, 1H, *J*=2.0 Hz, H-1); ¹³C NMR (100 MHz, CDCl₃): δ =24.7 (CH₃ isoprop), 25.4 (CH₃ isoprop), 25.9 (CH₃ isoprop), 26.7 (CH₃ isoprop), 66.0 (C-6), 75.5 (C-5), 79.8 (C-3), 82.1 (C-4), 85.6 (C-2), 101.3 (C-1), 109.7 (C_q isoprop), 112.8 ppm (C_q isoprop); HRMS: *m/z*:

calcd for C₁₂H₂₀O₆+H⁺: 261.1333; found: 261.1313; IR (neat): $\tilde{\nu}$ =1091, 1263, 1380, 1454, 2986, 3445 cm⁻¹.

α , β -L-Gulose (3): Compound **2** (75.5 g, 290 mmol) was added to H₂O (870 mL) and TFA (174 mL) at 0°C, after which the reaction mixture was allowed to warm to room temperature. After stirring for 5 h, approximately half of the volume was evaporated in vacuo. The solution was diluted with H₂O (500 mL) and evaporated to half the volume. This process was repeated twice, and the remaining TFA was quenched with Et₃N until neutral pH. The reaction mixture was concentrated in vacuo to obtain **3** as a colourless oil (52.2 g, quantitative). ¹H NMR (500 MHz, D₂O): δ =3.65 (dd, 1H, *J*=3.3, 8.3 Hz, H-2), 3.74–3.79 (m, 2H, H-6, H-6), 3.83 (d, 1H, *J*=3.5 Hz, H-4), 4.02 (t, 1H, *J*=6.0 Hz, H-5), 4.09 (t, 1H, *J*=3.3 Hz, H-3), 4.90 ppm (d, 1H, *J*=8.0 Hz, H-1); ¹³C NMR (125 MHz, CDCl₃): δ =60.56 (C-6), 68.64–73.66 (C-2, C-3, C-4, C-5), 93.39 ppm (C-1); HRMS: *m/z*: calcd for C₆H₁₂O₆+Na⁺: 203.05261; found: 203.05262.

Penta-O-acetyl- α , β -L-gulopyranose (4): Gulose (**3**; 52.2 g, 290 mmol) was dissolved in pyridine (1 L) and cooled to 0°C. After the addition of Ac₂O (200 mL), the reaction mixture was allowed to stir overnight at room temperature. The reaction was quenched with MeOH and concentrated in vacuo. The reaction mixture was diluted with EtOAc, washed with 1 M HCl (aq), NaHCO₃ (aq), and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo to yield **4** as a dark-yellow oil (96.6 g, 72%). ¹H NMR major anomer (400 MHz, CDCl₃): δ =2.05–2.17 (5 \times s, 15H, 5 \times CH₃ acetyl), 4.11 (m, 1H, H-6), 4.17 (m, 1H, H-6), 4.37 (m, 1H, H-5), 5.00 (dd, 1H, *J*=1.2, 4.0 Hz, H-4), 5.12 (dd, 1H, *J*=3.6, 8.6 Hz, H-2), 5.44 (t, 1H, *J*=3.6 Hz, H-3), 6.00 ppm (d, 1H, *J*=8.4 Hz, H-1); ¹³C NMR (100 MHz, CDCl₃): δ =20.5–20.9 (CH₃ acetyl), 61.5 (C-6), 67.2 (C-3), 67.3 (C-2), 67.5 (C-4), 71.3 (C-5), 89.9 (C-1), 168.9–170.4 ppm (CO); HRMS: *m/z*: calcd for C₁₆H₂₂O₁₁+Na⁺: 413.10543; found: 413.10544; IR (neat): $\tilde{\nu}$ =1065, 1207, 1369, 1744 cm⁻¹.

Phenyl-2,3,4,6-tetra-O-acetyl-1-thio- β -L-gulopyranoside (5): Per acetyl gulose (**4**; 83.8 g, 215 mmol) was dissolved in AcOH (86 mL) and cooled to 0°C. HBr/AcOH (33%, 102 mL) was added slowly and the reaction mixture was stirred at 0°C for 2 h. The reaction mixture was then poured into icewater and extracted with EtOAc (2 \times 250 mL). The organic layers were carefully washed with NaHCO₃ (aq) and added to a solution of Bu₄NHSO₄ (730 g, 215 mmol), Na₂CO₃ (286 g, 1.00 mol), and PhSH (26.4 mL, 258 mmol) in H₂O (1 L). When analysis by TLC showed complete consumption of the starting material, the layers were separated and the organic layer was washed with 1 M NaOH and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **5** as a slightly yellow oil (62.5 g, 66%). [α]_D²⁵=–20.6 (*c*=1, CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ =2.01–2.22 (m, 12H, CH₃ acetyl), 4.145–4.25 (m, 3H, H-5, H-6, H-6), 4.98 (dd, 1H, *J*=0.8, 4.0 Hz H-4), 5.05 (m, 2H, H-1, H-2), 5.37 (dd, 1H, *J*=2.0, 3.2 Hz, H-3), 7.26–7.33 (m, 3H, Ar-H Sph), 7.50–7.54 ppm (m, 2H, Ar-H Sph); ¹³C NMR (150 MHz, CDCl₃): δ =20.6–21.0 (CH₃ acetyl), 62.0 (C-6), 66.3 (C-2), 66.8 (C-3), 67.8 (C-4), 72.7 (C-5), 83.0 (C-1), 127.4–132.4 (Ar-CH), 132.6 (C_q Sph), 168.8–170.4 ppm (CO); HRMS: *m/z*: calcd for C₂₀H₂₄O₉S+Na⁺: 463.10332; found: 463.10311; IR (neat): $\tilde{\nu}$ =1026, 1059, 1209, 1369, 1736 cm⁻¹.

Phenyl-1-thio- β -L-gulopyranoside (6): Compound **5** (62.5 g, 142 mmol) was dissolved in MeOH (750 mL) and a catalytic amount of NaOMe was added. The reaction mixture was stirred overnight, quenched with Amberlite H⁺, and concentrated in vacuo. Crystallisation from acetone yielded **6** as white crystals (37.5 g, 97%). [α]_D²⁵=+48.6 (*c*=1, CHCl₃); ¹H NMR (400 MHz, MeOD): δ =3.67–3.79 (m, 4H, H-2, H-4, H-6, H-6), 3.92 (t, 1H, *J*=5.8 Hz, H-4), 3.97 (t, 1H, *J*=3.6 Hz, H-3), 5.02 (d, 1H, *J*=10.4 Hz, H-1), 7.19–7.29 (m, 3H, Ar-H), 7.53–7.56 ppm (m, 2H, Ar-H); ¹³C NMR (150 MHz, CDCl₃): δ =62.7 (C-6), 68.1 (C-2), 71.2 (C-5), 72.6 (C-4), 77.3 (C-3), 87.3 (C-1), 127.8, 129.8, 131.9 (Ar-CH), 136.3 ppm (C_q Sph); HRMS: *m/z*: calcd for C₁₂H₁₆O₅S+Na⁺: 295.06107; found: 295.06130; IR (neat): $\tilde{\nu}$ =974, 1034, 1051, 1412, 3234, 3440 cm⁻¹.

Phenyl-2,3,4,6-tetra-O-benzyl-1-thio- β -L-gulopyranoside (7): Thiogulose (**6**; 1.54 g, 5.66 mmol) was dissolved in DMF (56 mL) and cooled to 0°C. Respectively, BnBr (3.2 mL, 27 mmol) and NaH (1.08 g, 27 mmol, 60% dispersion in oil) were added. After stirring overnight, the reaction mixture was quenched with MeOH and concentrated in vacuo. The residue

was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **7** as a colourless oil (3.19 g, 89%). [α]_D²⁵ = +9.17 (*c* = 0.024, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): δ = 3.51 (m, 1H, H-4), 3.57–3.67 (m, 2H, H-6), 3.71 (t, 1H, *J* = 3.2 Hz, H-3), 3.75 (dd, 1H, *J* = 2.8, 10 Hz, H-2), 4.13 (t, 1H, *J* = 6.4 Hz, H-5), 4.26 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.31 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.37 (d, 1H, *J* = 10.8 Hz, CH₂ Bn), 4.40 (d, 1H, *J* = 8.8 Hz, CH₂ Bn), 4.47–4.52 (m, 2H, CH₂ Bn), 4.59 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.66 (d, 1H, *J* = 12 Hz, CH₂ Bn), 5.23 (d, 1H, *J* = 10 Hz, H-1), 7.08–7.10 (m, 2H, Ar-H), 7.12–7.35 (m, 21H, Ar-H), 7.51–7.60 ppm (m, 2H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 69.0 (C-6), 72.4 (CH₂ Bn), 72.8 (CH₂ Bn), 73.1 (C-3), 73.2 (CH₂ Bn), 73.4 (CH₂ Bn), 74.4 (C-5), 74.8 (C-2), 74.9 (C-4), 84.3 (C-1), 126.8–128.6 (Ar-CH), 131.4 (Ar-CH), 138.2 ppm (C_q Bn); HRMS: *m/z*: calcd for C₄₀H₄₀O₅S + Na⁺: 655.24887; found: 655.24860; IR (neat): $\tilde{\nu}$ = 741, 1001, 1028, 1076, 1101, 1207, 1360, 1439, 1454, 1497, 2866, 3032, 3061 cm⁻¹.

Phenyl-2,3,4-tri-*O*-benzyl-1-thio- β -L-gulopyranoside (8): Compound **6** (5.45 g, 20 mmol) was dissolved in pyridine (100 mL) and trityl chloride (8.36 g, 30 mmol) was added, and the reaction mixture was stirred for 3 days. The reaction was quenched with MeOH and concentrated in vacuo. The mixture was diluted with EtOAc, washed with 1 M HCl (aq), NaHCO₃ (aq), and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was dissolved in DMF (100 mL) and cooled to 0 °C. Respectively, BnBr (8.55 mL, 72 mmol) and NaH (2.88 g, 72 mmol, 60 % dispersion in oil) were added. After stirring overnight, the reaction mixture was quenched with MeOH and concentrated in vacuo. The residue was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was dissolved in CH₂Cl₂ (50 mL) and MeOH (200 mL) and a catalytic amount of *p*-TsOH was added. The reaction mixture was stirred overnight. The reaction mixture was neutralised with Et₃N and concentrated in vacuo. Column chromatography yielded **8** as a colourless oil (9.58 g, 88%). [α]_D²⁵ = +11.8 (*c* = 0.024, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): δ = 3.40 (m, 1H, H-3), 3.45–3.53 (m, 1H, H-6), 3.76 (m, 2H, H-2, H-4), 3.80–3.85 (m, 1H, H-6), 3.94–3.97 (m, 1H, H-5), 4.22 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.29 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.40 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.49 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.61 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.71 (d, 1H, *J* = 12 Hz, CH₂ Bn), 5.23 (d, 1H, *J* = 9.6 Hz, H-1), 7.07–7.10 (m, 2H, Ar-H), 7.23–7.36 (m, 16H, Ar-H), 7.55–7.57 ppm (m, 2H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 62.5 (C-6), 72.1 (CH₂ Bn), 72.7 (C-2 or C-4), 72.9 (CH₂ Bn), 73.4 (CH₂ Bn), 74.9 (C-2 or C-4), 75.2 (C-3), 75.8 (C-5), 84.0 (C-1), 127.0–128.8 (Ar-CH), 131.6 (Ar-CH), 134.0 (C_q Bn), 137.5 (C_q Bn), 137.8 (C_q Bn), 138.1 ppm (C_q Bn); HRMS: *m/z*: calcd for C₃₃H₃₄O₅S + H⁺: 543.21997; found: 543.22015; IR (neat): $\tilde{\nu}$ = 739, 922, 1001, 1026, 1042, 1074, 1207, 1358, 1439, 1454, 1477, 1497, 2878, 3030, 3063 cm⁻¹.

Methyl (phenyl-2,3,4-tri-*O*-benzyl-1-thio- β -D-gulopyranosyluronate) (9): Compound **8** (1.95 g, 3.6 mmol) was taken up in CH₂Cl₂ (24 mL) and H₂O (12 mL) and TEMPO (0.115 g, 0.74 mmol) and BAIB (2.89 g, 9.0 mmol) were added. The reaction mixture was stirred vigorously until analysis by TLC showed complete conversion of the starting material. Na₂S₂O₃ (50 mL, aq) was added and the resulting mixture was stirred for 15 min. The layers were separated and the aqueous phase acidified with 1 M HCl and extracted three times with CH₂Cl₂. The combined organic layers were dried over MgSO₄ and concentrated in vacuo. The residue was taken up in DMF (20 mL) and K₂CO₃ (2.49 g, 18 mmol) and MeI (0.56 mL, 9.0 mmol) was added. After 2 h, the reaction mixture was diluted with Et₂O (50 mL) and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **9** as a white solid (1.48 g, 72%). [α]_D²⁵ = +17.0 (*c* = 1, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): δ = 3.72–3.75 (m, 4H, H-3, CH₃ CO₂Me), 3.83 (dd, 1H, *J* = 3.2, *J* = 10 Hz, H-2), 3.91 (dd, 1H, *J* = 3.6, 1.6 Hz, H-4), 4.31 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.39 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.41 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.56 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.60 (s, 1H, H-5), 4.61 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.71 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 5.23 ppm (d, 1H, *J* = 10 Hz, H-1); ¹³C NMR (100 MHz, CDCl₃): δ = 52.1 (CH₃ CO₂Me), 72.5 (CH₂ Bn), 72.7 (CH₂ Bn), 72.7 (C-3), 72.2 (CH₂ Bn), 73.9 (C-2), 74.7 (C-5), 76.2 (C-4), 84.4 (C-1), 127.2–128.6 (Ar-C), 132.4 (Ar-C), 133.7 (C_q Bn), 137.4 (C_q

Bn), 137.7 (C_q Bn), 137.8 (C_q Bn), 169.1 ppm (COOMe); HRMS: *m/z*: calcd for C₃₄H₃₄O₆S + NH₄⁺: 588.24144; found: 588.24210; IR (neat): $\tilde{\nu}$ = 731, 897, 814, 939, 1026, 1074, 1126, 1209, 1265, 1304, 1358, 1420, 1439, 1454, 1477, 1497, 1734, 1765, 2876 cm⁻¹.

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-(2,3,4,6-tetra-*O*-benzyl- α,β -L-gulopyranoside)- α -D-glucopyranoside (13): A solution of donor **7** (0.127 g, 0.2 mmol), diphenyl sulfoxide (0.045 g, 0.22 mmol), and tri-*tert*-butylpyrimidine (0.124 g, 0.5 mmol) in CH₂Cl₂ (4 mL) was stirred over activated 3-Å molecular sieves (MS) for 30 min. The reaction mixture was cooled to –78 °C before triflic acid anhydride (37 μ L, 0.22 mmol) was added. The reaction mixture was stirred for 10 min at –78 °C followed by the addition of acceptor **10** (0.139 g, 0.3 mmol) in CH₂Cl₂ (3 mL). Stirring was continued and the reaction mixture was allowed to warm to 0 °C and Et₃N (0.15 mL) was added. The reaction mixture was diluted with CH₂Cl₂ and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH₂Cl₂. The collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by size exclusion and column chromatography yielded **13** as a colourless oil (71 %, 0.140 g, α/β = 13:1). Determination of the α/β ratio by ¹H NMR: δ = 3.25 (s, 3H, OCH₃ α), 3.33 ppm (s, 0.22H, OCH₃ β). α isomer: ¹H NMR (400 MHz, CDCl₃): δ = 3.25 (s, 3H, OCH₃), 3.38 (dd, 1H, *J* = 3.6, *J* = 9.6 Hz, H-2), 3.53–3.62 (m, 2H, H-6'), 3.63–3.85 (m, 6H, H-3', H-4, H-6, H-5, H-2', H-4'), 3.96 (t, 1H, *J* = 9.2 Hz, H-3), 4.01 (d, 1H, *J* = 10 Hz, H-6'), 4.33–4.55 (m, 9H, H-5', CH₂ Bn), 4.57 (d, 1H, *J* = 3.6 Hz, H-1), 4.68 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.71 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.80 (d, 1H, *J* = 10.8 Hz, CH₂ Bn), 4.83 (d, 1H, *J* = 12 Hz, CH₂ Bn), 4.93 (d, 1H, *J* = 10.8 Hz, CH₂ Bn), 4.98 ppm (d, 1H, *J* = 3.2 Hz, H-1'); ¹³C NMR (100 MHz, CDCl₃): δ = 54.8 (OCH₃), 65.5 (C-5), 67.2 (C-6'), 68.8 (C-6), 70.1 (C-4'), 71.1 (CH₂ Bn), 72.7 (C-2'), 72.7 (CH₂ Bn), 72.8 (CH₂ Bn), 73.0 (CH₂ Bn), 73.1 (CH₂ Bn), 74.2 (C-5'), 74.8 (CH₂ Bn), 75.5 (CH₂ Bn), 75.6 (C-3'), 77.9 (C-4'), 80.2 (C-2), 82.0 (C-3), 97.7 (C-1'), 97.8 (C-1), 127.1–128.3 (Ar-CH), 138.1 (C_q Bn), 138.3 (C_q Bn), 138.4 (C_q Ph), 138.5 (C_q Bn), 138.8 (C_q Bn), 139.1 (C_q Bn), 139.4 ppm (C_q Bn); HRMS: *m/z*: calcd for C₆₂H₆₆O₁₁ + NH₄⁺: 1004.49434; found: 1004.49579; IR (neat): $\tilde{\nu}$ = 733, 820, 908, 1026, 1047, 1069, 1194, 1207, 1310, 1327, 1360, 1454, 1497, 2870, 3030, 3063 cm⁻¹.

Methyl-2,3,6-tri-*O*-benzyl-4-*O*-(2,3,4,6-tetra-*O*-benzyl- α,β -L-gulopyranoside)- α -D-glucopyranoside (14): As described for the synthesis of **13** using acceptor **11** (0.139 g, 0.3 mmol). Purification by size exclusion and column chromatography yielded **14** as a colourless oil (91 %, 0.179 g, α/β = 10:1). Determination of the α/β ratio by ¹H NMR: δ = 5.11 (d, 1H, *J* = 4.0 Hz, H-1' α), 5.38 ppm (d, 0.09H, *J* = 8.0 Hz, H-1' β). α isomer: ¹H NMR (400 MHz, CDCl₃): δ = 3.34 (s, 3H, CH₃ OMe), 3.45 (m, 1H), 3.51 (dd, 1H, *J* = 3.6 Hz, 8.8 Hz), 3.60 (m, 1H), 3.68 (t, 1H, *J* = 3.2 Hz), 3.73 (m, 1H, H-2'), 3.76–3.89 (m, 4H), 4.09 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.15 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.26–4.40 (m, 7H), 4.51–4.56 (m, 3H, H-1), 4.66 (d, 1H, *J* = 8.8 Hz, CH₂ Bn), 4.68 (d, 1H, *J* = 8.8 Hz, CH₂ Bn), 4.88 (d, 1H, *J* = 11.2 Hz, CH₂ Bn), 4.95 (d, 1H, *J* = 11.2 Hz, CH₂ Bn), 5.11 (d, 1H, *J* = 4.0 Hz, H-1'), 7.08–7.30 ppm (m, 35H, Ar-CH); ¹³C NMR (100 MHz, CDCl₃): δ = 54.9 (CH₃ OMe), 65.3, 68.1, 68.9, 70.1, 71.3, 72.6, 72.7, 72.8, 72.9, 73.0, 73.1, 73.3, 73.9, 74.5, 75.8, 80.3, 80.5, 97.6 (C-1'), 97.8 (C-1), 126.8–128.7 (Ar-CH), 137.9–139.5 ppm (C_q Bn); ESI-MS: *m/z*: 987.5 [M+H⁺]; IR (neat): $\tilde{\nu}$ = 732, 909, 1027, 1454, 2866, 3030 cm⁻¹.

para-Methoxyphenyl-2-*O*-benzyl-(2,3,4,6-tetra-*O*-benzyl- α -L-gulopyranoside)-4,6-benzylidene- β -D-galactopyranoside (15): As described for the synthesis of **13** using acceptor **12** (0.139 g, 0.3 mmol). Purification by size exclusion and column chromatography yielded **15** as a colourless oil (73 %, 0.144 g). Determination of the α/β ratio by ¹H NMR: δ = 5.47 (s, 0.07H, CH benzylidene β), 5.55 ppm (s, 1H, CH benzylidene α). α anomer: ¹H NMR (400 MHz, CDCl₃): δ = 3.28 (s, 1H, H-6), 3.43 (dd, 1H, *J* = 10 Hz, H-6), 3.57–3.60 (m, 1H, H-4'), 3.62 (t, 1H, *J* = 10 Hz, H-5'), 3.68 (dd, 1H, *J* = 10, 3.6 Hz, H-3), 3.71 (s, 1H, H-6'), 3.86 (m, 1H, H-2'), 3.90 (m, 1H, H-3'), 4.03–4.10 (m, 2H, H-2, H-6), 4.30–4.49 (m, 6H, CH₂ Bn), 4.58–4.63 (m, 2H, H-4, CH₂ Bn), 4.71–4.72 (m, 1H, H-5), 4.76 (d, 1H, *J* = 11.2 Hz, CH₂ Bn), 4.87–4.90 (m, 2H, H-1, CH₂ Bn), 5.23 (d, 1H, *J* = 3.6 Hz, H-1'), 5.55 (s, 1H, CH benzylidene), 6.74–6.76 (m, 2H, Ar-H), 6.99–7.01 (m, 2H, Ar-H), 7.12–7.31 (m, 35H, Ar-H), 7.38–7.40 (m, 3H, Ar-H), 7.50–7.52 ppm (m, 2H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 55.5 (OCH₃ *p*MP (*p*-methoxyphenyl)), 66.6 (C-5), 68.5 (C-6), 70.9 (C-6'),

70.9 (C-4), 72.0 (C-3'), 72.5 (CH₂ Bn), 72.8 (CH₂ Bn), 73.8 (CH₂ Bn), 74.3 (C-2'), 75.1 (C-5'), 75.1 (CH₂ Bn), 76.8 (C-4'), 77.0 (C-2), 82.8 (C-3), 99.5 (CH benzylidene), 100.7 (C-1'), 102.7 (C-1), 114.2 (C₆MP), 118.9 (C₆MP), 125.2–128.3 (Ar-CH), 137.7 (C_q Bn), 138.9 (C_q Bn), 138.1 (C_q Bn), 138.3 (C_q Bn), 138.9 (C_q Bn), 139.0 (C_q Bn), 151.6 (C_q Bn), 155.0 ppm (C_q Bn); HRMS: *m/z*: calcd for C₆₁H₆₂O₁₂+NH₄⁺: 1004.45795; found: 1004.45946; IR (neat): $\tilde{\nu}$ = 731, 824, 872, 910, 997, 1026, 1065, 1080, 1173, 1217, 1265, 1308, 1367, 1454, 1506, 2866, 3030 cm⁻¹.

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-[methyl (2,3,4-tri-*O*-benzyl- α , β -L-gulopyranosyl)uronate]- α -D-glucopyranoside (16): A solution of donor **7** (0.114 g, 0.2 mmol), diphenyl sulfoxide (0.045 g, 0.22 mmol), and tri-*tert*-butylpyrimidine (0.124 g, 0.5 mmol) in CH₂Cl₂ (4 mL) was stirred over activated 3-Å MS for 30 min. The mixture was cooled to -60°C before triflic acid anhydride (37 μ L, 0.22 mmol) was added. The mixture was warmed to -45°C then cooled to -78°C followed by the addition of acceptor **10** (0.139 g, 0.3 mmol) in CH₂Cl₂ (3 mL). Stirring was continued and the reaction mixture was allowed to warm to 0°C and Et₃N (0.15 mL) was added. The reaction mixture was diluted with CH₂Cl₂ and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH₂Cl₂. The collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by size exclusion and column chromatography yielded **16** as a colourless oil (115 mg, 73%, α/β = 3:1). Determination of the α/β ratio by ¹H NMR (400 MHz, CDCl₃): δ = 3.60 (s, 1.15 H, CH₃ CO₂Me β), 3.64 (s, 3.62 H, CH₃ CO₂Me α), 4.96 (d, 0.33 H, *J* = 8 Hz, H-1' β), 5.08 ppm (d, 1 H, *J* = 3.6 Hz, H-1); ¹³C NMR (100 MHz, CDCl₃): δ = 97.8 (C-1 α), 98.0 (C-1' α), 100.6 ppm (C-1' β); HRMS: *m/z*: calcd for C₅₆H₆₀O₁₂+NH₄⁺: 942.44230; found: 942.44351; IR (neat): $\tilde{\nu}$ = 731, 808, 910, 1026, 1047, 1070, 1207, 1265, 1304, 1358, 1439, 1454, 1497, 1732, 1765, 2876, 3030 cm⁻¹.

Methyl-2,3,6-tri-*O*-benzyl-4-*O*-[methyl (2,3,4-tri-*O*-benzyl- α -L-gulopyranosyl)uronate]- α -D-glucopyranoside (17): As described for the synthesis of **16** using acceptor **11** (0.139 g, 0.3 mmol). Purification by size exclusion and column chromatography yielded **17** as a colourless oil (189 mg, 94%). ¹H NMR (400 MHz, CDCl₃): δ = 3.31 (s, 3 H, CH₃ CO₂Me), 3.42 (s, 3 H, CH₃ OMe), 3.63 (dd, 1 H, *J* = 3.6, 8.8 Hz, H-2), 3.72–3.75 (m, 2 H, H-6, H-3'), 3.85–3.97 (m, 5 H, H-3, H-5, H-6, H-2', H-4'), 4.35 (s, 2 H, CH₂ Bn), 4.40–4.53 (m, 4 H, CH₂ Bn), 4.57 (d, 1 H, *J* = 12 Hz, CH₂ Bn), 4.63 (d, 1 H, *J* = 3.6 Hz, H-1), 4.69 (d, 1 H, *J* = 12 Hz, CH₂ Bn), 4.83 (d, 1 H, *J* = 12 Hz, CH₂ Bn), 4.99 (m, 2 H, H-5', CH₂ Bn), 5.26 (d, 1 H, *J* = 3.6 Hz, H-1'), 7.16–7.40 ppm (m, 30 H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 51.4 (CH₃ COOMe), 54.9 (CH₃ OMe), 67.6 (C-5'), 68.3 (C-6), 70.2, 71.5, 72.2, 72.6, 72.9, 73.1, 73.2, 73.3, 74.2, 75.0, 76.6, 76.7, 77.3, 79.8, 80.1, 97.4 (C-1'), 97.8 (C-1), 126.5–128.3 (CH Bn), 137.5–139.2 (C_q Bn), 169.6 ppm (COOMe); ESI-MS: 925.4 [*M*+H]⁺; IR (neat): $\tilde{\nu}$ = 533, 732, 910, 1027, 1208, 1554, 1797, 1734, 1764, 2892, 3031 cm⁻¹.

para-Methoxyphenyl-2-*O*-benzyl-3-*O*-[methyl (2,3,4-tri-*O*-benzyl- α , β -L-galactopyranosyl)uronate]-4,6-benzylidene- β -D-galactopyranoside (18): As described for the synthesis of **13** using acceptor **12** (0.139 g, 0.3 mmol). Purification by size exclusion and column chromatography yielded **18** as a colourless oil (124 mg, 79%, α/β = 10:1). Determination of the α/β ratio by ¹H NMR (400 MHz, CDCl₃): δ = 5.21 (d, 0.11 H, *J* = 8 Hz, H-1' β), 5.34 ppm (d, 1 H, *J* = 3.6 Hz, H-1' α); ¹³C NMR (100 MHz, CDCl₃): δ = 100.2 (C-1' α), 102.6 ppm (C-1 α); HRMS: *m/z*: calcd for C₅₅H₅₆O₁₃+Na⁺: 947.36131; found: 947.36190; IR (neat): $\tilde{\nu}$ = 731, 826, 908, 997, 1026, 1065, 1078, 1175, 1217, 1265, 1306, 1366, 1439, 1454, 1506, 175, 2870, 3030 cm⁻¹.

Phenyl-2,3-di-*O*-benzyl-4,6-*O*-benzylidene-1-thio- β -L-gulopyranoside (19): Compound **6** (10.88 g, 40.0 mmol) was dissolved in MeCN (400 mL) and benzaldehyde dimethylacetal (6.32 mL, 42.0 mmol) and a catalytic amount of *p*-TsOH were added. After stirring for 15 h, the reaction mixture was quenched with Et₃N until neutral pH and concentrated in vacuo. The residue was dissolved in DMF (200 mL) and cooled to 0°C. Respectively, BnBr (11.4 mL, 96 mmol) and NaH (3.84 g, 96 mmol, 60% dispersion in oil) were added. After stirring overnight, the reaction mixture was quenched with MeOH and concentrated in vacuo. The residue was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **19** as a colourless oil (15.12 g, 70%). [α]_D²⁵ = +39.2 (*c* = 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 3.81 (dd, 1 H, *J* = 2.8,

10.0 Hz, H-2), 3.83 (s, 1 H, H-5), 3.92 (t, 1 H, *J* = 3.2 Hz), 3.97 (dd, 1 H, *J* = 1.6, 12.4 Hz, H-6), 4.05 (d, 1 H, *J* = 3.6 Hz, H-4), 4.33 (d, 1 H, *J* = 12.8 Hz, H-6), 4.36 (d, 1 H, *J* = 11.6 Hz, CH₂ Bn), 4.47 (d, 1 H, *J* = 11.2 Hz, CH₂ Bn), 4.63 (d, 1 H, *J* = 12.0 Hz, CH₂ Bn), 4.79 (d, 1 H, *J* = 12.0 Hz, CH₂ Bn), 5.27 (d, 1 H, *J* = 9.6 Hz, H-1), 5.47 (s, 1 H, CH benzylidene), 7.15–7.72 ppm (m, 20 H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 67.5 (C-5), 69.5 (C-6), 72.5 (CH₂ Bn), 73.6 (CH₂ Bn), 74.0 (C-2), 74.3 (C-3), 75.0 (C-4), 83.1 (C-1), 100.9 (CHPh), 126.4–132.7 (Ar-CH), 133.0 (C_q SPh), 137.8, 137.9, 138.1 ppm (C_q Bn, C_q benzylidene); HRMS: *m/z*: calcd for C₃₃H₃₂O₅S+H⁺: 541.20432; found: 541.20440; IR (neat): $\tilde{\nu}$ = 995, 1080, 1394, 1454, 2870, 3032 cm⁻¹.

Phenyl-2,3-*O*-benzyl-1-thio- β -L-gulopyranoside (20): A solution of **3** (10.8 g, 20 mmol) in MeOH (200 mL) with a catalytic amount of *p*-TsOH was stirred overnight at room temperature. The reaction mixture was quenched with Et₃N until neutral pH and concentrated in vacuo. Purification of the residue by column chromatography afforded **20** as a colourless oil (9.05 g, quantitative). [α]_D²² = +27.6 (*c* = 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 2.38 (bs, 1 H, 6-OH), 3.45 (d, 1 H, *J* = 3.6 Hz, 4'-OH), 3.81 (dd, 1 H, *J* = 2.8, 10.0 Hz, H-2), 3.85–3.98 (m, 5 H, H-3, H-4, H-5, H-6, H-6), 4.55 (d, 1 H, *J* = 11.6 Hz, CH₂ Bn), 4.60 (d, 1 H, *J* = 12.0 Hz, CH₂ Bn), 4.64 (d, 1 H, *J* = 11.6 Hz, CH₂ Bn), 4.77 (d, 1 H, *J* = 12.0 Hz, CH₂ Bn), 5.28 (d, 1 H, *J* = 10.0 Hz, H-1), 7.24–7.38 (m, 13 H, Ar-H), 7.52–7.54 ppm (m, 2 H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 64.0 (C-6), 70.8 (C-5), 72.7 (CH₂ Bn), 73.5 (CH₂ Bn), 74.2 (C-4), 75.0 (C-2), 75.5 (C-3), 84.7 (C-1), 127.2–131.5 (Ar-CH), 133.8 (C_q SPh), 137.9 (C_q Bn), 138.2 ppm (C_q Bn); HRMS: *m/z*: calcd for C₂₆H₂₆O₅S+H⁺: 453.17302; found: 453.17296; IR (neat): $\tilde{\nu}$ = 956, 1026, 1454, 2889, 3402 cm⁻¹.

Methyl (phenyl-2,3-*O*-benzyl-1-thio- β -L-guluronate) (21): Compound **20** (0.538 g, 1.18 mmol) was taken up in CH₂Cl₂ (10 mL) and H₂O (4 mL). TEMPO (0.038 g, 0.24 mmol) and BAIB (0.955 g, 2.96 mmol) were added to this mixture, and the reaction mixture was stirred vigorously until analysis by TLC showed complete conversion of the starting material. Na₂S₂O₃ (50 mL, aq) was added and the resulting mixture was stirred for 15 min. The layers were separated and the aqueous phase was acidified with 1 M HCl and extracted three times with CH₂Cl₂. The combined organic layers were dried over MgSO₄ and concentrated in vacuo. The resulting syrup was then dissolved in Et₂O (20 mL) and cooled to 0°C, after which TMS diazomethane (0.5 mL in Et₂O) was added until the solution turned bright yellow. AcOH was added until the yellow colour disappeared. The mixture was concentrated in vacuo. Purification by column chromatography yielded **21** as a colourless oil (0.280 g, 49%). [α]_D²² = -98.8 (*c* = 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 2.76 (bs, 1 H, 4'-OH), 3.73–3.74 (m, 1 H, H-2), 3.75 (s, 3 H, CH₃ CO₂Me), 3.95 (t, 1 H, *J* = 3.4 Hz, H-3), 4.16 (bs, 1 H, H-4), 4.51 (d, 1 H, *J* = 11.2 Hz, CH₂ Bn), 4.58–4.64 (m, 3 H, CH₂ Bn, H-5), 4.75 (d, 1 H, *J* = 12.0 Hz, CH₂ Bn), 5.21 (d, 1 H, *J* = 10.0 Hz), 7.24–7.62 ppm (m, 15 H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 52.4 (CH₃ CO₂Me), 69.4 (C-4), 72.7 (CH₂ Bn), 73.4 (CH₂ Bn), 74.3 (C-2), 74.9 (C-3), 75.1 (C-5), 85.1 (C-1), 127.5–132.3 (Ar-CH), 133.3 (C_q Bn), 137.7 (C_q Bn), 137.9 (C_q Bn), 169.4 ppm (COOMe); HRMS: *m/z*: calcd for C₂₇H₂₈O₆S+Na⁺: 503.1499; found: 503.1488; IR (neat): $\tilde{\nu}$ = 1069, 1118, 1744, 2855, 2920, 3445 cm⁻¹.

Methyl (phenyl-2,3-*O*-benzyl-4-*O*-levulinoyl-1-thio- β -L-guluronate) (22): A solution of Lev₂O in dioxane (0.5 mL, 8.0 mL, 4.0 mmol) was added to a solution of **21** (0.96 g, 2.00 mmol) in pyridine (20 mL). The reaction mixture was diluted with EtOAc after 16 h and washed with 1 M HCl (aq), NaHCO₃ (aq), and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **22** as a colourless oil (1.10 g, 95%). [α]_D²² = -105.6 (*c* = 1, CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ = 2.16 (s, 3 H, CH₃ Lev), 2.45–2.48 (m, 2 H, CH₂ Lev), 2.64–2.73 (m, 2 H, CH₂ Lev), 3.60 (dd, 1 H, *J* = 3.0, 10.2 Hz, H-2), 3.76 (s, 3 H, CH₃ CO₂Me), 3.93 (t, 1 H, *J* = 3.3 Hz, H-3), 4.47–4.76 (m, 5 H, CH₂ Bn, H-5), 5.21 (d, 1 H, *J* = 10.2 Hz, H-1), 5.29 (dd, 1 H, *J* = 1.5, 3.9 Hz, H-4), 7.26–7.66 ppm (m, 15 H, Ar-H); ¹³C NMR (150 MHz, CDCl₃): δ = 27.9 (CH₂ Lev), 29.7 (CH₃ Lev), 37.7 (CH₂ Lev), 52.3 (CH₃ CO₂Me), 70.2 (C-4), 72.4 (CH₂ Bn), 72.5 (C-3), 73.2 (C-5), 73.4 (CH₂ Bn), 73.7 (C-2), 84.6 (C-1), 127.5–132.6 (Ar-CH), 133.4 (C_q SPh), 137.5 (C_q Bn), 137.6 (C_q Bn), 167.9 (COOMe), 171.3 (COO Lev), 206.0 ppm (CO Lev); HRMS: *m/z*: calcd for C₃₂H₃₄O₈S+Na⁺: 601.1867;

found: 601.1843; IR (neat): $\bar{\nu}$ = 1026, 1064, 1111, 1250, 1713, 1740, 2873 cm⁻¹.

Phenyl-2,3-*O*-benzyl-6-*O*-tert-butyldimethylsilyl-1-thio- β -L-gulopyranoside (23): A solution of **20** (0.869 g, 1.92 mmol) in DMF (10 mL) was cooled to 0°C. Respectively, imidazole (0.136 g, 2.00 mmol) and TBDMSCl (0.301 g, 2.00 mmol) were added and the reaction mixture was warmed to room temperature. After stirring for 4 h, the reaction mixture was quenched with MeOH and concentrated in vacuo. The residue was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **23** as a colourless oil (0.978 g, 90%). [α]_D²² = +32.4 (*c* = 1, CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ = 0.08 (s, 3H, CH₃), 0.11 (s, 3H, CH₃), 0.91 (s, 9H, *t*Bu), 3.79 (dd, 1H, *J* = 3.0, 10.2 Hz, H-2), 3.82 (bs, 1H, OH), 3.87–3.91 (m, 3H, H-3, H-5, H-6), 3.97 (dd, 1H, *J* = 3.6, 10.8 Hz, H-6), 4.00 (bs, 1H, H-4), 4.46 (d, 1H, *J* = 11.4 Hz, CH₂ Bn), 4.55 (d, 1H, *J* = 11.4 Hz, CH₂ Bn), 4.57 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.74 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 5.18 (d, 1H, *J* = 10.2 Hz, H-1), 7.23–7.59 ppm (m, 15H, Ar-H); ¹³C NMR (300 MHz, CDCl₃): δ = -5.7 (CH₃), -5.5 (CH₃), 18 (C_q TBDMS), 25.8 (*t*Bu), 65.2 (C-6), 70.8 (C-4), 72.7 (CH₂ Bn), 73.5 (CH₂ Bn), 73.9 (C-5), 74.7 (C-2), 75.6 (C-3), 84.6 (C-1), 127.2–132.3 (Ar-CH), 133.8 (C_q SPh), 138.1 (C_q Bn), 138.4 ppm (C_q Bn); ESI-MS: 567.3 [*M*+H⁺]; IR (neat): $\bar{\nu}$ = 833, 1026, 1103, 1254, 2855, 2928, 3456 cm⁻¹.

Phenyl-2,3-*O*-benzyl-6-*O*-tert-butyldimethylsilyl-4-*O*-levulinoyl-1-thio- β -L-gulopyranoside (24): A solution of Lev₂O in dioxane (0.5 mL, 6.92 mL, 3.46 mmol) was added to a solution of **23** (0.978 g, 1.73 mmol) in pyridine (10 mL). The reaction mixture was diluted with EtOAc after 16 h and washed with 1 M HCl (aq), NaHCO₃ (aq), and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **24** as a colourless oil (1.08 g, 94%). [α]_D²² = +35.6 (*c* = 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = -0.01 (s, 3H, CH₃ TBDMS), -0.01 (s, 3H, CH₃ TBDMS), 0.83 (s, 9H, *t*Bu TBDMS), 2.13 (s, 3H, CH₃ Lev), 2.45–2.48 (m, 2H, CH₂ Lev), 2.64–2.67 (m, 2H, CH₂ Lev), 3.55–3.60 (m, 2H, H-2, H-6), 3.68 (dd, 1H, *J* = 6.4 Hz, 10.0 Hz, H-6), 3.90 (t, 1H, *J* = 3.6 Hz, H-3), 4.10 (t, 1H, *J* = 6.8 Hz, H-5), 4.48 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.56 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.64 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.69 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 5.06 (d, 1H, *J* = 3.6 Hz, H-4), 5.23 (d, 1H, *J* = 10.0 Hz, H-1), 7.18–7.54 ppm (m, 15H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = -5.7 (CH₃ TBDMS), -5.5 (CH₃ TBDMS), 18 (C_q TBDMS), 25.7 (*t*Bu TBDMS), 27.9 (CH₂ Lev), 29.7 (CH₃ Lev), 37.8 (CH₂ Lev), 61.0 (C-6), 68.7 (C-4), 72.2 (CH₂ Bn), 72.7 (C-3), 73.1 (CH₂ Bn), 74.4, 74.5 (C-2, C-5), 84.4 (C-1), 126.9–131.2 (Ar-CH), 134.2 (C_q SPh), 137.7 (C_q Bn), 137.8 (C_q Bn), 171.6 (COO Lev), 205.9 ppm (CO Lev); HRMS: *m/z*: calcd for C₃₇H₄₈O₇Si+H⁺: 665.29628; found: 665.29699; IR (neat): $\bar{\nu}$ = 1026, 1096, 1362, 1717, 1740, 2858, 2928 cm⁻¹.

Phenyl-2,3-di-*O*-benzyl-4,6-*O*-di-tert-butyldisilylidene-1-thio- β -L-gulopyranoside (25): (*t*Bu)₂Si(OTf)₂ (3.24 mL, 10.0 mmol) was added to a solution of **6** (2.72 g, 10.0 mmol) in pyridine (100 mL) at -20°C, after which the reaction mixture was warmed to room temperature. The reaction mixture was quenched with MeOH, concentrated in vacuo, taken up in EtOAc, and washed with 1 M HCl (aq), NaHCO₃ (aq), and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was dissolved in DMF (50 mL) and cooled to 0°C. Respectively, BnBr (2.87 mL, 24.0 mmol) and NaH (0.96 g, 24.0 mmol, 60% dispersion in oil) were added. After stirring for 15 h, the mixture was quenched with MeOH and concentrated in vacuo. The residue was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **25** as a colourless oil (5.20 g, 88%). [α]_D²² = +26.6 (*c* = 1, CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ = 1.00 (s, 9H, *t*Bu), 1.04 (s, 9H, *t*Bu), 3.74 (bs, 1H, H-5), 3.87–3.89 (m, 2H, H-2, H-3), 4.11 (dd, 1H, *J* = 2.4, 12.6 Hz, H-6), 4.15–4.18 (m, 2H, H-4, H-6), 4.59 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.66 (d, 1H, *J* = 11.4 Hz, CH₂ Bn), 4.75 (d, 1H, *J* = 11.4 Hz, CH₂ Bn), 4.84 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 5.27 (d, 1H, *J* = 9.6 Hz, H-1), 7.18–7.54 ppm (m, 15H, Ar-H); ¹³C NMR (150 MHz, CDCl₃): δ = 20.4 (C_q *t*Bu), 23.2 (C_q *t*Bu), 27.5 (CH₃ *t*Bu), 67.2 (C-6), 72.0 (C-4), 72.1 (C-5), 72.7 (CH₂ Bn), 73.7 (CH₂ Bn), 75.2 (C-2), 76.3 (C-3), 84.9 (C-1), 126.9–134.7 (Ar-CH), 134.7 (C_q SPh), 137.8 (C_q Bn), 138.4 ppm (C_q Bn); HRMS: *m/z*: calcd for

C₃₄H₄₄O₅Si+H⁺: 593.27515; found: 593.27527; IR (neat): $\bar{\nu}$ = 853, 937, 1088, 1474, 2858, 2932 cm⁻¹.

3-Azidopropyl [methyl (2,3-*O*-benzyl- α -L-gulopyranoside)uronate] (27): Hydrazine (76 μ L, 1.6 mmol) was added to a solution of **29** (178 g, 0.31 mmol) in pyridine/AcOH (4:1, 3.5 mL). The reaction mixture was stirred for 10 min, diluted with EtOAc, and washed with 1 M HCl, water, and NaHCO₃ (aq). The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography afforded **27** as a colourless oil (116 mg, 80%). ¹H NMR (400 MHz, CDCl₃): δ = 1.81–1.96 (m, 2H, CH₂ C₃H₆N₃), 2.39 (d, 1H, *J* = 5.6 Hz, 4'-OH), 3.35–3.38 (m, 2H, CH₂ C₃H₆N₃), 3.47–3.53 (m, 1H, CH₂ C₃H₆N₃), 3.78 (s, 3H, CH₃ CO₂Me), 3.82–3.88 (m, 2H, H-2, C₃H₆N₃), 3.90 (t, 1H, *J* = 3.4 Hz, H-3), 4.19 (bs, 1H, H-4), 4.57–4.67 (m, 3H, CH₂ Bn), 4.79 (d, 1H, *J* = 1.2 Hz, H-5), 4.87 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.95 (d, 1H, *J* = 3.6 Hz, H-1), 7.25–7.38 ppm (m, 10H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 28.9 (CH₂ C₃H₆N₃), 48.2 (CH₂ C₃H₆N₃), 52.4 (CH₃ CO₂Me), 65.5 (CH₂ C₃H₆N₃), 67.7 (C-5), 69.8 (C-4), 71.8 (CH₂ Bn), 72.9 (C-2), 73.1 (CH₂ Bn), 75.4 (C-3), 98.1 (C-1), 127.6–128.4 (Ar-CH), 137.9 (C_q Bn), 138.6 (C_q Bn), 170.5 ppm (COOMe); HRMS: *m/z*: calcd for C₂₄H₂₉N₃O₇+H⁺: 472.2078; found: 472.2081; IR (neat): $\bar{\nu}$ = 1034, 1088, 1450, 2095, 2870, 2924 cm⁻¹.

3-Azidopropyl-2,3-*O*-benzyl-6-*O*-tert-butyldimethylsilyl- α -L-gulopyranoside (28): A solution of **35** (0.576 g, 0.988 mmol) in THF (10 mL) was cooled to 0°C and TBAF (3 mL, 3 mmol, 1 M in THF) was added. The reaction mixture was diluted with EtOAc after 4 h and washed with water and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was taken up in DMF (5.0 mL) was cooled to 0°C. Imidazole (0.073 g, 1.08 mmol) and TBDMSCl (0.162 g, 1.08 mmol) were added and the reaction mixture was warmed to room temperature. After stirring for 4 h, the reaction mixture was quenched with MeOH and concentrated in vacuo. The residue was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **28** as a colourless oil (0.414 g, 75%). [α]_D²² = -23.0 (*c* = 1, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ = 0.09 (s, 6H, CH₃ TBDMS), 0.89 (s, 9H, *t*Bu TBDMS), 1.78–1.98 (m, 2H, CH₂ C₃H₆N₃), 3.37 (t, 2H, *J* = 6.5 Hz, 2H, CH₂ C₃H₆N₃), 3.44–3.48 (m, 1H, CH₂ C₃H₆N₃), 3.76–3.83 (m, 2H, OH, CH₂ C₃H₆N₃), 3.86–3.87 (m, 1H, H-5), 3.91–3.92 (m, 3H, H-2, H-6, H-6), 4.03–4.05 (m, 2H, H-3, H-4), 4.57–4.66 (m, 3H, CH₂ Bn), 4.86–4.89 (m, 2H, H-1, CH₂ Bn), 7.24–7.37 ppm (m, 10H, Ar-H); ¹³C NMR (125 MHz, CDCl₃): δ = -5.7 (CH₃ TBDMS), -5.6 (CH₃ TBDMS), 18.1 (C_q *t*Bu TBDMS), 25.7 (CH₃ *t*Bu TBDMS), 28.9 (CH₂ C₃H₆N₃), 48.3 (CH₂ C₃H₆N₃), 64.5 (CH₂ C₃H₆N₃), 65.1 (C-3), 65.5 (C-6), 70.9 (C-4), 71.4 (CH₂ Bn), 72.9 (CH₂ Bn), 73.5 (C-2), 75.5 (C-5), 97.9 (C-1), 127.3–128.3 (Ar-CH), 138.3 (C_q Bn), 139.0 ppm (C_q Bn); HRMS: *m/z*: calcd for C₂₉H₄₃N₃O₆Si+H⁺: 558.29939; found: 558.29985; IR (neat): $\bar{\nu}$ = 1042, 1107, 2095, 2858, 2928 cm⁻¹.

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-[methyl (2,3-di-*O*-benzyl-4-*O*-levulinoyl- α , β -L-gulopyranosyl)uronate]- α -D-glucopyranoside (29): As described for the synthesis of **13** using donor **22** (0.061 g, 0.105 mmol) and acceptor **10** (0.073 g, 0.158 mmol). Purification by size exclusion and column chromatography yielded **29** as a colourless oil (0.064 g, 66%, α/β = 3:1). Determination of the α/β ratio by ¹H NMR (400 MHz, CDCl₃): δ = 3.27 (s, 3H, OMe), 3.33 (s, 0.33H, OMe), 4.99 (d, 0.33H, *J* = 8.4 Hz, H-1' β), 5.05 ppm (bs, 1H, H-1' α); ¹³C NMR (100 MHz, CDCl₃): δ = 55.0 (CH₃ OMe $\alpha\alpha$), 55.0 (CH₃ OMe $\beta\alpha$), 97.9 (C-1' α), 100.6 ppm (C-1' β).

Methyl-2,3,6-tri-*O*-benzyl-4-*O*-[methyl (2,3-di-*O*-benzyl-4-*O*-levulinoyl- α -L-gulopyranosyl)uronate]- α -D-glucopyranoside (30): As described for the synthesis of **13** using donor **22** (0.065 g, 0.112 mmol) and acceptor **11** (0.078 g, 0.168 mmol). Purification by size exclusion and column chromatography yielded **30** as a colourless oil (0.066 g, 64%). ¹H NMR (400 MHz, CDCl₃): δ = 2.14 (s, 3H, CH₃ Lev), 2.38–2.48 (m, 2H, CH₂ Lev), 2.59–2.72 (m, 2H, CH₂ Lev), 3.25 (s, 3H, CH₃ CO₂Me), 3.36 (s, 3H, CH₃ OMe), 3.57–3.58 (m, 1H, H-2), 3.63 (bs, 1H, H-2'), 3.65 (bs, 1H, H-4), 3.88–3.91 (m, 5H, H-3, H-5, H-6, H-6, H-3'), 4.39–4.52 (m, 5H, CH₂ Bn), 4.57 (bs, 1H, H-1), 4.62 (m, 3H, CH₂ Bn), 5.01 (s, 1H, H-5'), 5.16 (bs, 1H, H-1'), 5.19 (s, 1H, H-4'), 7.11–7.43 ppm (m, 25H, Ar-H); ¹³C NMR (150 MHz, CDCl₃): δ = 27.9 (CH₂ Lev), 29.7 (CH₃ Lev), 37.7 (CH₂ Lev), 51.8 (CH₃ CO₂Me), 55.0 (CH₃ OMe), 66.0 (C-5'), 68.2 (C-6), 70.2, 70.7 (C-4'), 71.2 (CH₂ Bn), 71.8 (C-3'), 72.5 (C-2'), 73.8 (CH₂ Bn),

73.3 (CH₂ Bn), 74.2 (CH₂ Bn), 74.9, 79.7, 80.3 (C-2), 97.4 (C-1'), 97.9 (C-1), 126.5–128.4 (Ar-CH), 137.5–139.2 (C_q arom), 168.7 (CO₂Me), 171.5 (CO Lev), 206.1 ppm (COO Lev).

3-Azidopropyl [methyl(2,3-di-*O*-benzyl-4-*O*-levulinoyl- α -L-gulopyranoside)uronate] (31): A solution of **22** (0.116 g, 0.20 mmol), diphenyl sulfoxide (0.049 g, 0.24 mmol), and tri-*tert*-butylpyrimidine (0.129 g, 0.52 mmol) in CH₂Cl₂ (4 mL) was stirred over activated 3-Å MS for 30 min. The reaction mixture was cooled to –60 °C before triflic acid anhydride (40 μ L, 0.24 mmol) was added, then warmed to –45 °C. The reaction mixture was stirred for 10 min at –45 °C followed by the addition of azidopropanol (0.061 g, 0.6 mmol) in CH₂Cl₂ (1.5 mL). Stirring was continued and the reaction mixture was allowed to warm to 0 °C and Et₃N (0.14 mL) was added. The reaction mixture was diluted with CH₂Cl₂ and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH₂Cl₂. The collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **31** as a colourless oil (0.098 g, 86%, α/β = 1:3). The anomers could be separated by column chromatography; the spectral data for the α anomer are given. ¹H NMR (400 MHz, CDCl₃): δ = 1.80–1.96 (m, 2H, C₃H₆N₃), 2.17 (s, 3H, CH₃ Lev), 2.45–2.49 (m, 2H, CH₂ Lev), 2.66–2.73 (CH₂ Lev), 3.33–3.37 (m, 2H, C₃H₆N₃), 3.48–3.53 (m, 1H, C₃H₆N₃), 3.70 (t, 1H, *J* = 3.8 Hz, H-2), 3.71 (s, 3H, CO₂Me), 3.81–3.86 (m, 1H, C₃H₆N₃), 3.92 (t, 1H, *J* = 3.4 Hz, H-3), 4.52 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.62 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.72 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.82–4.86 (m, 2H, H-5, CH₂ Bn), 4.95 (d, 1H, *J* = 4.0 Hz, H-1), 5.32 (dd, 1H, *J* = 1.6 Hz, 3.2 Hz, H-4), 7.26–7.41 ppm (m, 10H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 28.0 (CH₂ Lev), 28.9 (CH₂ C₃H₆N₃), 29.7 (CH₃ Lev), 37.8 (CH₂ Lev), 48.2 (CH₂ C₃H₆N₃), 52.4 (CH₃ CO₂Me), 65.4 (C-5), 70.8 (C-4), 71.5 (CH₂ Bn), 72.5, 72.5 (C-2, C-3), 73.1 (CH₂ Bn), 97.9 (C-1), 127.6–128.4 (Ar-CH), 137.8 (C_q Bn), 138.3 (C_q Bn), 169.1 (COOMe), 171.4 (COO Lev), 206.0 ppm (CO Lev); HRMS: *m/z*: calcd for C₂₉H₃₅N₃O₉ + H⁺: 570.2446; found: 570.2447; IR (neat): $\tilde{\nu}$ = 1141, 1744, 2095, 2870, 2924 cm^{–1}.

3-Azidopropyl (methyl[2,3-di-*O*-benzyl-4-*O*-[methyl (2,3-di-*O*-benzyl-4-*O*-levulinoyl- α -L-gulopyranosyl)uronate]- α -L-gulopyranoside]uronate] (32): A solution of **22** (0.116 g, 0.20 mmol), diphenyl sulfoxide (0.049 g, 0.24 mmol), and tri-*tert*-butylpyrimidine (0.129 g, 0.52 mmol) in CH₂Cl₂ (4 mL) was stirred over activated 3-Å MS for 30 min. The reaction mixture was cooled to –60 °C before triflic acid anhydride (40 μ L, 0.24 mmol) was added, then warmed to –45 °C. The reaction mixture was stirred for 10 min at –45 °C followed by the addition of **27** (0.078 g, 0.166 mmol) in CH₂Cl₂ (1.6 mL). Stirring was continued, the reaction mixture was allowed to warm to 0 °C, and Et₃N (0.14 mL) was added. The reaction mixture was diluted with CH₂Cl₂ and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH₂Cl₂ and the collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by size exclusion and column chromatography yielded **32** as a colourless oil (0.053 g, 34%, α/β = 3:1). The anomers could be separated by column chromatography; the spectral data for the α anomer are given. [α]_D²⁵ = –64.4 (*c* = 1, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃): δ = 1.81–1.94 (m, 2H, CH₂ C₃H₆N₃), 2.17 (s, 3H, CH₃ Lev), 2.45–2.48 (m, 2H, CH₂ Lev), 2.67–2.72 (m, 2H, CH₂ Lev), 3.32–3.38 (m, 2H, CH₂ C₃H₆N₃), 3.52–3.55 (m, 1H, CH₂ C₃H₆N₃), 3.56 (s, 3H, CH₃ CO₂Me), 3.66 (t, 1H, *J* = 3.3 Hz, H-2'), 3.70–3.72 (m, 4H, H-2, CH₃ CO₂Me), 3.86–3.90 (m, 3H, H-3, H-3', CH₂ C₃H₆N₃), 4.18 (d, 1H, *J* = 12.6 Hz, CH₂ Bn), 4.27 (m, 1H, H-4), 4.30 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.47 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.56 (d, 1H, *J* = 12.0 Hz, CH₂ Bn), 4.59–4.61 (m, 3H, H-5', CH₂ Bn), 4.78–4.81 (m, 3H, H-5, CH₂ Bn), 4.97 (d, 1H, *J* = 3.6 Hz, H-1), 5.17 (d, 1H, *J* = 3.6 Hz, H-1'), 5.27 (dd, 1H, *J* = 1.8, 4.2 Hz, H-4'), 7.09–7.42 ppm (m, 20H, Ar-H); ¹³C NMR (150 MHz, CDCl₃): δ = 27.9 (CH₂ Lev), 28.9 (CH₂ C₃H₆N₃), 29.7 (CH₃ Lev), 37.8 (CH₂ Lev), 48.2 (CH₂ C₃H₆N₃), 52.1 (CH₃ CO₂Me), 52.4 (CH₃ CO₂Me), 65.3 (CH₂ C₃H₆N₃), 66.8 (C-5'), 67.1 (C-5), 70.9 (C-4'), 71.4 (CH₂ Bn), 71.6 (CH₂ Bn), 71.7 (C-3'), 72.8 (CH₂ Bn), 73.0 (CH₂ Bn), 73.1 (C-2'), 73.5 (C-2), 75.1 (C-3), 77.7 (C-4), 97.6 (C-1), 99.7 (C-1'), 127.4–128.3 (Ar-CH), 137.7 (C_q Bn), 137.9 (C_q Bn), 138.5 (C_q Bn), 138.6 (C_q Bn), 168.5, 169.9, 171.4 (COOMe, COO Lev), 206.10 ppm (CO Lev); HRMS: *m/z*: calcd for C₅₀H₅₇N₃O₁₅ + Na⁺: 962.36955; found: 962.36819; IR (neat): $\tilde{\nu}$ = 1038, 1142, 1454, 1744, 2095, 2870, 2924 cm^{–1}.

3-Azidopropyl-2,3-*O*-benzyl-6-*O*-*tert*-butyldimethylsilyl-4-*O*-levulinoyl- α , β -L-gulopyranoside (33): A solution of donor **24** (0.133 g, 0.2 mmol), diphenyl sulfoxide (0.045 g, 0.22 mmol), and tri-*tert*-butylpyrimidine (0.124 g, 0.5 mmol) in CH₂Cl₂ (4 mL) was stirred over activated 3-Å MS for 30 min. The reaction mixture was cooled to –70 °C before triflic acid anhydride (37 μ L, 0.22 mmol) was added. The reaction mixture was stirred for 10 min at –70 °C followed by the addition of azidopropanol (0.061 g, 0.6 mmol) in CH₂Cl₂ (1.5 mL). Stirring was continued, the reaction mixture was allowed to warm to 0 °C, and Et₃N (0.15 mL) was added. The reaction mixture was diluted with CH₂Cl₂ and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH₂Cl₂, and the collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **33** as a colourless oil (0.098 g, 86%, α/β = 3:1). The anomers could not be separated by column chromatography. ¹H NMR (600 MHz, CDCl₃): δ = –0.01–0.01 (m, 7.8H, CH₃ TBDMS), 0.83 (s, 9H, *t*Bu TBDMS), 0.84 (s, 3H, *t*Bu TBDMS), 1.78–1.93 (m, 2.6H, CH₂ C₃H₆N₃), 2.13 (s, 1H, CH₃ Lev), 2.14 (s, 3H, CH₃ Lev), 2.47–2.49 (m, 2.6H, CH₂ Lev), 2.61–2.73 (m, 2.6H, CH₂ Lev), 3.32–3.40 (m, 4H), 3.43–3.47 (m, 1.3H, CH₂ C₃H₆N₃), 3.51 (dd, 1H, *J* = 6.6 Hz, 9.6 Hz), 3.57–3.62 (m, 2.9H), 3.65–3.70 (m, 1.6H), 3.77–3.81 (m, 1H, CH₂ C₃H₆N₃), 3.84–3.87 (m, 1.3H), 3.93–3.96 (m, 0.3H, CH₂ C₃H₆N₃), 4.06 (t, 0.3H, *J* = 6.6 Hz), 4.24 (t, 1H, *J* = 7.2 Hz), 4.50–4.79 (m, 7.1H, H-1 α , CH₂ Bn), 4.81 (d, 0.3H, *J* = 7.8 Hz, H-1 β), 5.01 (d, 0.3H, *J* = 1.6 Hz, H-4), 5.06 (d, 1H, *J* = 1.6 Hz, H-4), 7.23–7.62 ppm (m, 13H, Ar-H); ¹³C NMR (150 MHz, CDCl₃): δ = –5.6 (CH₃ TBDMS), –5.5 (CH₃ TBDMS), 18.2 (C_q TBDMS), 25.8 (*t*Bu TBDMS), 26.7, 26.8, 28.0, 29.0, 29.3, 29.8, 31.5, 37.9, 48.5, 59.8, 60.9, 61.4, 64.8, 66.0, 66.5, 68.7, 69.2, 71.2, 72.2, 72.6, 72.9, 73.0, 73.3, 74.7, 76.2, 97.4 (C-1 α), 100.9 (C-1 β), 124.8–135.6 (Ar-CH), 138.1 (C_q Bn), 138.1 (C_q Bn), 138.6 (C_q Bn), 138.6 (C_q Bn), 171.8 (COO lev), 171.9 (COO lev), 206.1 (CO Lev), 206.2 ppm (CO Lev); HRMS: *m/z*: calcd for C₃₄H₄₉N₃O₈Si + Na⁺: 678.31811; found: 678.31810; IR (neat): $\tilde{\nu}$ = 1026, 1096, 1717, 1740, 2095, 2858, 2928 cm^{–1}.

3-Azidopropyl-2,3-*O*-benzyl-6-*O*-*tert*-butyldimethylsilyl-4-*O*-(2,3-*O*-benzyl-6-*O*-*tert*-butyldimethylsilyl-4-*O*-levulinoyl- α -L-gulopyranosyl)- α -L-gulopyranoside (34): A solution of donor **24** (0.133 g, 0.2 mmol), diphenyl sulfoxide (0.045 g, 0.22 mmol), and tri-*tert*-butylpyrimidine (0.124 g, 0.5 mmol) in CH₂Cl₂ (4 mL) was stirred over activated 3-Å MS for 30 min. The reaction mixture was cooled to –70 °C before triflic acid anhydride (37 μ L, 0.22 mmol) was added. The reaction mixture was stirred for 10 min at –70 °C followed by the addition of acceptor **28** (0.093 g, 0.166 mmol) in CH₂Cl₂ (1.6 mL). Stirring was continued, the reaction mixture was allowed to warm to 0 °C, and Et₃N (0.15 mL) was added. The reaction mixture was diluted with CH₂Cl₂ and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH₂Cl₂ and the collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by size exclusion and column chromatography yielded **34** as a colourless oil (48%, 0.089 g, α/β = 6:1). The anomers could be separated by column chromatography; the spectral data for the α anomer are given. [α]_D²⁵ = –88.0 (*c* = 0.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = –0.03 (s, 3H, CH₃ TBDMS), –0.01 (s, 3H, CH₃ TBDMS), 0.00 (s, 3H, CH₃ TBDMS), 0.01 (s, 3H, CH₃ TBDMS), 0.83 (s, 9H, *t*Bu TBDMS), 0.87 (s, 9H, *t*Bu TBDMS), 1.83–1.92 (m, 2H, CH₂ C₃H₆N₃), 2.18 (s, 3H, CH₃ Lev), 2.45–2.53 (m, 2H, CH₂ Lev), 2.67–2.73 (CH₂ Lev), 3.34–3.38 (m, 2H, CH₂ C₃H₆N₃), 3.42–3.53 (m, 3H, H-6, H-6, CH₂ C₃H₆N₃), 3.60 (t, 1H, *J* = 3.6 Hz, H-2'), 3.70–3.82 (m, 3H, H-2, H-6', CH₂ C₃H₆N₃), 3.88–3.99 (m, 4H, H-3, H-4, H-3', H-6'), 4.13 (t, *J* = 7.2 Hz, H-5'), 4.19 (t, *J* = 6.8 Hz, H-5), 4.23 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.30 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.53 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 4.59 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.64–4.677 (m, 2H, CH₂ Bn), 4.87 (d, 1H, *J* = 3.6 Hz, H-1), 4.81 (d, 1H, *J* = 11.6 Hz, CH₂ Bn), 4.86 (d, 1H, *J* = 12.4 Hz, CH₂ Bn), 5.07–5.09 (m, 2H, H-1', H-4'), 7.16–7.38 ppm (m, 20H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = –5.5 (CH₃ TBDMS), –5.4 (CH₃ TBDMS), –5.3 (CH₃ TBDMS), 18.1 (C_q *t*Bu TBDMS), 18.3 (C_q *t*Bu TBDMS), 25.9 (*t*Bu TBDMS), 28.1 (CH₂ Lev), 29.2 (CH₂ C₃H₆N₃), 29.8 (CH₃ Lev), 37.9 (CH₂ Lev), 48.6 (CH₂ C₃H₆N₃), 61.0, 61.1 (C-6, C-6'), 64.8 (CH₂ C₃H₆N₃), 66.2 (C-5), 67.0 (C-5'), 69.8 (C-4'), 71.1 (CH₂ Bn), 71.2 (CH₂ Bn), 72.0 (C-3'), 72.5 (CH₂ Bn), 72.8 (CH₂ Bn), 74.3 (C-2'), 74.3 (C-2), 74.7 (C-3), 75.8 (C-4), 97.5 (C-1), 99.6 (C-1'), 127.2–128.4 (Ar-CH), 138.2 (C_q Bn), 138.3 (C_q Bn), 139.1 (C_q Bn), 139.2 (C_q Bn), 171.8 (COO Lev), 206.1 ppm (CO

Lev); HRMS: m/z : calcd for $C_{60}H_{85}N_3O_{13}Si_2 + Na^+$: 1134.55131; found: 1134.55163; IR (neat): $\tilde{\nu}$ = 1003, 1092, 1454, 1744, 2095, 2858, 2928 cm^{-1} .

3-Azidopropyl-2,3-di-O-benzyl-4,6-O-benzylidene- α,β -L-gulopyranoside (35): As described for the synthesis of **33** using donor **19** (0.108 g, 0.20 mmol). Purification by column chromatography yielded **35** as a colourless oil (0.094 g, 88%, α/β = 3:1). The anomers could not be separated by column chromatography. 1H NMR (400 MHz, $CDCl_3$): δ = 1.84–1.96 (m, 2.6H, CH_2 $C_3H_6N_3$), 3.36–3.39 (m, 2.6H, CH_2 $C_3H_6N_3$), 3.45–3.50 (m, 1H, CH_2 $C_3H_6N_3$), 3.60–3.66 (m, 0.3H, CH_2 $C_3H_6N_3$), 3.72 (dd, 0.3H, J = 2.8, 8.0 Hz, H-2 β), 3.77 (s, 0.3H), 3.82–3.88 (m, 1H, CH_2 $C_3H_6N_3$), 3.91–3.97 (m, 3.6H), 4.00–4.05 (m, 2H), 4.10 (d, 1H, J = 2.4 Hz), 4.23 (d, 1H, J = 12.4 Hz, CH_2 Bn), 4.28 (d, 0.3H, J = 12.4 Hz, CH_2 Bn), 4.57–4.69 (m, 3.9H, CH_2 Bn), 4.83–4.88 (m, 0.6H, CH_2 Bn), 4.92 (d, 1H, J = 12.0 Hz, CH_2 Bn), 4.94 (d, 0.3H, J = 8.4 Hz, H-1 β), 4.98 (d, 1H, J = 4.0 Hz, H-1 α), 5.48 (s, 0.3H, CH benzylidene), 5.50 (s, 1H, CH benzylidene), 7.23–7.65 ppm (m, 19.5H, Ar-H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 29.0 (CH_2 $C_3H_6N_3$), 29.3 (CH_2 $C_3H_6N_3$), 48.3 (CH_2 $C_3H_6N_3$), 48.4 (CH_2 $C_3H_6N_3$), 59.8, 64.7, 65.5, 66.2, 69.4, 69.7, 71.6, 73.2, 73.5, 73.7, 73.9, 74.2, 75.3, 75.6, 76.4, 76.6, 97.8 (C-1 α), 100.7 (C-1 β), 101.0 (CHPh), 124.7–131.0 (Ar-CH), 137.7 (C_q Bn), 138.1 (C_q Bn), 138.8 ppm (C_q Bn); HRMS: m/z : calcd for $C_{30}H_{33}N_3O_6 + H^+$: 532.24212; found: 532.24421; IR (neat): $\tilde{\nu}$ = 1088, 1109, 2095, 2874 cm^{-1} .

3-Azidopropyl-2,3-O-benzyl-6-O-tert-butyldimethylsilyl-4-O-(2,3-di-O-benzyl-4,6-O-benzylidene- α -L-gulopyranosyl)- α -L-gulopyranoside (36): As described for the synthesis of **34** using donor **19** (0.108 g, 0.20 mmol). Purification by size exclusion and column chromatography yielded **36** as a colourless oil (45%, 0.074 g, α/β = 6:1). The anomers could be separated by column chromatography; the spectral data for the α anomer are given. $[a]_D^{25}$ = –78.0 (c = 0.5, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ = –0.04 (s, 3H, CH_3 TBDMS), 0.01 (s, 3H, CH_3 TBDMS), 0.86 (s, 9H, *t*Bu TBDMS), 1.86–2.01 (m, 2H, CH_2 $C_3H_6N_3$), 3.11 (bs, 1H, H-4'), 3.35–3.44 (m, 2H, CH_2 $C_3H_6N_3$), 3.55–3.58 (m, 1H, CH_2 $C_3H_6N_3$), 3.63 (t, 1H, J = 3.6 Hz, H-3'), 3.69–3.76 (m, 4H, H-2, H-6, H-6', H-6'), 3.84–3.95 (m, 4H, H-3, H-2', H-5', CH_2 $C_3H_6N_3$), 3.95–3.99 (m, 2H, H-4, H-6), 4.13 (d, 1H, J = 7.2 Hz, H-5), 4.45 (d, 1H, J = 12.8 Hz, CH_2 Bn), 4.54 (d, 1H, J = 12.0 Hz, CH_2 Bn), 4.59 (d, 1H, J = 10.8 Hz, CH_2 Bn), 4.67–4.76 (m, 3H, CH_2 Bn), 4.91 (d, 1H, J = 3.6 Hz, H-1), 5.00 (d, 1H, J = 11.2 Hz, CH_2 Bn), 5.19 (d, 1H, J = 3.6 Hz, H-1'), 5.02 (d, 1H, J = 10.8 Hz, CH_2 Bn), 5.45 (s, 1H, CH benzylidene), 6.94–7.48 ppm (m, 25H, Ar-H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = –5.4 (CH_3 TBDMS), –5.3 (CH_3 TBDMS), 18.2 (C_q *t*Bu TBDMS), 25.8 (*t*Bu TBDMS), 29.2 (CH_2 $C_3H_6N_3$), 48.6 (CH_2 $C_3H_6N_3$), 60.1 (C-4'), 61.1 (C-6), 64.8 (CH_2 $C_3H_6N_3$), 66.8 (C-5), 69.5 (C-6'), 71.5 (CH_2 Bn), 71.5 (CH_2 Bn), 72.2 (CH_2 Bn), 73.3 (CH_2 Bn), 73.4 (C-2), 73.8 (C-4), 75.1 (C-3), 75.3 (C-5'), 75.5 (C-3'), 76.1 (C-2'), 97.0 (C-1), 99.9 (C-1'), 100.9 (CH benzylidene), 126.2–129.1 (Ar-CH), 137.7–139.5 ppm (C_q arom); HRMS: m/z : calcd for $C_{56}H_{69}N_3O_{11}Si + Na^+$: 1010.45936; found: 1010.46006; IR (neat): $\tilde{\nu}$ = 1088, 1109, 2095, 2874 cm^{-1} .

3-Azidopropyl-2,3-di-O-benzyl-4,6-O-di-tert-butyldimethylsilyl- α -L-gulopyranoside (37): As described for the synthesis of **33** using donor **25** (0.118 g, 0.20 mmol). Purification by column chromatography yielded **37** as a colourless oil (0.088 g, 75%, α/β = 5:1). The anomers could be separated by column chromatography; the spectral data for the α anomer are given. $[a]_D^{25}$ = –57.6 (c = 1, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ = 0.91 (s, 9H, *t*Bu), 0.98 (s, 9H, *t*Bu), 1.82–1.85 (m, 1H, CH_2 $C_3H_6N_3$), 1.91–1.94 (m, 1H, CH_2 $C_3H_6N_3$), 3.37 (t, 2H, J = 6.8 Hz, CH_2 $C_3H_6N_3$), 3.44–3.49 (m, 1H, CH_2 $C_3H_6N_3$), 3.78–3.82 (m, 1H, CH_2 $C_3H_6N_3$), 3.83–3.88 (m, 2H, H-2, H-3), 3.93 (bs, 1H, H-5), 4.09 (dd, 1H, J = 1.6, 12.4 Hz, H-6), 4.18–4.19 (m, 2H, H-4, H-6), 4.58–4.62 (m, 2H, CH_2 Bn), 4.67 (d, 1H, J = 12.4 Hz, CH_2 Bn), 4.80 (d, 1H, J = 3.6 Hz, H-1), 4.96 (d, 1H, J = 12.0 Hz, CH_2 Bn), 7.26–7.39 ppm (m, 10H, Ar-H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 20.4 (C_q *t*Bu), 23.3 (C_q *t*Bu), 27.2 (CH_3 *t*Bu), 27.6 (CH_3 *t*Bu), 29.0 (CH_2 $C_3H_6N_3$), 48.4 (CH_2 $C_3H_6N_3$), 64.1 (C-5), 64.6 (CH_2 $C_3H_6N_3$), 67.2 (C-6), 71.3 (CH_2 Bn), 72.4 (C-4), 72.6 (C-2), 73.3 (CH_2 Bn), 76.1 (C-3), 97.8 (C-1), 127.5–128.4 (Ar-CH), 138.0 (C_q Bn), 139.1 ppm (C_q Bn); HRMS: m/z : calcd for $C_{31}H_{45}N_3O_6Si + H^+$: 584.31504; found: 584.31479; IR (neat): $\tilde{\nu}$ = 1089, 1138, 2095, 2858, 2931 cm^{-1} .

3-Azidopropyl-2,3-O-benzyl-6-O-tert-butyldimethylsilyl-4-O-(2,3-di-O-benzyl-4,6-O-di-tert-butyldimethylsilyl- α -L-gulopyranosyl)- α -L-gulopyranoside (38): Compound **38** was synthesised by two pathways: 1) As described for the synthesis of **34** using donor **25** (0.118 g, 0.20 mmol). Purification by size exclusion and column chromatography yielded **38** as a colourless oil (0.083 g, 48%, α/β = 10:1). 2) A solution of hemiacetal **39** (0.235 g, 0.47 mmol), diphenyl sulfoxide (0.237 g, 1.17 mmol), and tri-*tert*-butylpyrimidine (0.233 g, 0.939 mmol) in CH_2Cl_2 (10 mL) was stirred over activated 3-Å MS for 30 min. The reaction mixture was cooled to –60°C before triflic acid anhydride (83 μ L, 0.493 mmol) was added. The reaction mixture was warmed to –40°C and stirred for 1 h followed by the addition of acceptor **28** (0.131 g, 0.234 mmol) in CH_2Cl_2 (2.5 mL). Stirring was continued and the reaction mixture was allowed to warm to room temperature, after which Et_3N (5 equiv) was added. The reaction mixture was diluted with CH_2Cl_2 and washed with $NaHCO_3$ (aq). The aqueous layer was extracted twice with CH_2Cl_2 , and the collected organic layers were dried over $MgSO_4$ and concentrated in vacuo. Purification by size exclusion and column chromatography yielded **38** as a colourless oil (0.205 g, 84%, α/β = 10:1). The anomers could be separated by column chromatography; the spectral data for the α anomer are given. $[a]_D^{25}$ = –75.0 (c = 1, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ = –0.03 (s, 3H, CH_3 TBDMS), 0.01 (s, 3H, CH_3 TBDMS), 0.86 (s, 9H, *t*Bu silylidene), 0.91 (s, 9H, *t*Bu silylidene), 1.03 (s, 9H, *t*Bu TBDMS), 1.84–1.97 (m, 2H, CH_2 $C_3H_6N_3$), 3.29–3.40 (m, 3H, H-4', CH_2 $C_3H_6N_3$), 3.50–3.52 (m, 1H, CH_2 $C_3H_6N_3$), 3.60 (t, 1H, J = 3.6 Hz, H-3), 3.67 (t, 1H, J = 3.6 Hz, H-2), 3.71 (dd, 1H, J = 1.2, 12.4 Hz, H-6'), 3.76–3.81 (m, 3H, H-4, H-6, H-5'), 3.82–3.86 (m, 3H, H-2', H-6', CH_2 $C_3H_6N_3$), 3.89–3.95 (m, 2H, H-6, CH_2 Bn), 4.09–4.13 (m, 2H, H-5, H-3'), 4.37 (d, 1H, J = 12.4 Hz, CH_2 Bn), 4.55–4.59 (m, 2H, CH_2 Bn), 4.69 (d, 1H, J = 12.0 Hz, CH_2 Bn), 4.77 (d, 1H, J = 12.0 Hz, CH_2 Bn), 4.87 (d, 1H, J = 3.6 Hz, H-1) 4.98 (d, 1H, J = 3.6 Hz, H-1'), 5.02 (d, 1H, J = 10.8 Hz, CH_2 Bn), 6.96–7.48 ppm (m, 20H, Ar-H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = –5.4 (CH_3 TBDMS), –5.3 (CH_3 TBDMS), 18.2 (C_q *t*Bu TBDMS), 20.4 (C_q *t*Bu silylidene), 23.3 (C_q *t*Bu silylidene), 25.9 (*t*Bu TBDMS), 27.2 (CH_3 *t*Bu silylidene), 27.6 (CH_3 *t*Bu silylidene), 29.2 (CH_2 $C_3H_6N_3$), 48.6 (CH_2 $C_3H_6N_3$), 61.4 (C-6), 64.5 (C-4'), 64.7 (CH_2 $C_3H_6N_3$), 66.9 (C-6'), 67.1 (C-5), 71.2 (CH_2 Bn), 71.5 (CH_2 Bn), 72.9 (CH_2 Bn), 73.1 (C-3'), 73.2 (CH_2 Bn), 73.8 (C-2'), 73.9 (C-2), 75.5 (C-4), 75.6 (C-5'), 76.1 (C-3), 97.0 (C-1), 99.7 (C-1'), 127.3–128.5 (Ar-CH), 138.1 (C_q Bn), 138.5 (C_q Bn), 139.0 (C_q Bn), 139.7 ppm (C_q Bn); HRMS: m/z : calcd for $C_{57}H_{81}N_3O_{11}Si_2 + Na^+$: 1062.53018; found: 1062.53082; IR (neat): $\tilde{\nu}$ = 1084, 1138, 1454, 1474, 2095, 2859, 2932 cm^{-1} .

2,3-Di-O-benzyl-4,6-O-di-tert-butyldimethylsilyl- α,β -L-gulopyranoside (39): NIS (0.243 g, 1.08 mmol) and TFA (83 μ L, 1.08 mmol) were added to a solution of **25** (0.638 g, 1.08 mmol) in CH_2Cl_2 (10 mL) at 0°C. After analysis by TLC showed complete consumption of the starting material, the reaction was quenched with Et_3N . Saturated $Na_2S_2O_3$ (aq) was added to the reaction mixture, which was then stirred for 30 min. The aqueous layer was extracted twice with CH_2Cl_2 , and the combined organic layers were dried over $MgSO_4$ and concentrated in vacuo. Purification by column chromatography yielded **39** as a colourless oil (0.514 g, 95%, α/β = 3:1). 1H NMR (400 MHz, $CDCl_3$): δ = 0.91 (s, 3H, *t*Bu), 0.99 (s, 9H, *t*Bu), 3.48 (d, 0.3H, J = 5.2 Hz, OH β), 3.65 (dd, 0.3H, J = 2.8, 8.0 Hz, H-2 β), 3.80 (bs, 0.3H), 3.88–3.89 (m, 1H, H-2 α), 3.98 (m, 1.3H), 4.13–4.23 (m, 3H), 4.57–4.71 (m, 2.6H, CH_2 Bn), 4.80 (d, 0.3H, J = 12.0 Hz, CH_2 Bn), 4.85 (d, 1H, J = 11.6 Hz, CH_2 Bn), 5.16 (dd, 0.3H, J = 5.2, 2.6 Hz, H-1 β), 5.22 (m, 1H, H-1 α), 7.21–7.37 ppm (m, 10.6H, Ar-H); ^{13}C NMR (100 MHz, $CDCl_3$): δ = 20.3 (C_q *t*Bu), 20.4 (C_q *t*Bu), 23.2 (C_q *t*Bu), 27.1 (CH_3 *t*Bu), 27.3 (CH_3 *t*Bu), 27.4 (CH_3 *t*Bu), 27.8 (CH_3 *t*Bu), 62.3, 67.0, 67.1, 70.2, 70.5, 70.6, 71.6, 72.0, 72.7, 73.6, 74.6, 76.9, 77.6, 78.2, 92.9, 94.8 (C-1, α/β), 127.5–128.5 (Ar-CH), 137.2 (C_q Bn), 137.5 (C_q Bn), 138.3 (C_q Bn), 138.5 ppm (C_q Bn); HRMS: m/z : calcd for $C_{28}H_{40}O_6Si + H^+$: 501.26669; found: 501.26671; IR (neat): $\tilde{\nu}$ = 826, 1045, 1084, 1138, 1474, 1736, 2858, 2932, 3472 cm^{-1} .

3-Azidopropyl-2,3-O-benzyl-4-O-(2,3-O-benzyl- α -L-gulopyranosyl)- α -L-gulopyranoside (40): A solution of **38** (0.233 g, 0.244 mmol) in THF (2.5 mL) was cooled to 0°C and TBAF (0.74 mL, 0.74 mmol, 1M in THF) was added. The reaction mixture was diluted with $EtOAc$ after 4 h, and washed with water and brine. The organic layer was dried over $MgSO_4$ and concentrated in vacuo. Purification by column chromatogra-

phy yielded **40** as a colourless oil (0.153 g, 87%). $[\alpha]_D^{25} = -73.2$ ($c=1$, CHCl_3); $^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 1.81$ – 2.01 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.37–3.43 (m, 3H, H-6, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.49–3.54 (m, 1H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.58 (dd, 1H, $J=2.8$, 12 Hz, H-6), 3.70 (bs, 1H, OH), 3.77–3.77 (m, 5H), 3.83–3.88 (m, 5H), 4.12 (bs, 1H), 4.18 (d, 1H, $J=12.6$ Hz, CH_2 Bn), 4.27 (m, 1H, H-4), 4.33 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.37 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.43 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.54–4.61 (m, 3H, CH_2 Bn), 4.65 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.80 (d, 1H, $J=2.4$ Hz, H-1), 4.85 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.98 (d, 1H, $J=3.6$ Hz, H-1'), 7.17–7.41 ppm (m, 20H, Ar-H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): $\delta = 29.0$ (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 48.4 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 62.6, 64.3 (C-6, C-6'), 64.8 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 65.8, 65.9, 70.8, 71.3, 71.4, 72.7, 72.9, 73.3, 73.6, 74.6, 80.4, 97.1 (C-1), 99.8 (C-1'), 127.3–129.1 (Ar-CH), 137.4 (C_q Bn), 138.2 (C_q Bn), 138.3 (C_q Bn), 138.7 ppm (C_q Bn); ESI-MS: m/z : 786.4 $[M+H]^+$; IR (neat): $\tilde{\nu} = 1069$, 1109, 1454, 2095, 2878 cm^{-1} .

3-Azidopropyl-2,3-O-benzyl-6-O-tert-butylidimethylsilyl-4-O-(2,3-O-benzyl-6-O-tert-butylidimethylsilyl- α -L-gulopyranosyl)- α -L-gulopyranoside (41): A solution of **40** (0.278 g, 0.35 mmol) in DMF (5 mL) was cooled to 0°C. Imidazole (0.049 g, 0.725 mmol) and TBDMSCl (0.109 g, 0.725 mmol) were added respectively and the reaction mixture was warmed to room temperature. After stirring for 4 h, the reaction mixture was quenched with MeOH and concentrated in vacuo. The residue was taken up in Et₂O and washed three times with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Column chromatography yielded **41** as a colourless oil (0.273 g, 78%). $[\alpha]_D^{25} = -60.2$ ($c=1$, CHCl_3); $^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = -0.01$ (s, 3H, CH_3), 0.02 (s, 3H, CH_3), 0.11 (s, 3H, CH_3), 0.12 (s, 3H, CH_3), 0.89 (s, 9H, *t*Bu), 0.93 (s, 9H, *t*Bu), 1.88–1.99 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.32–3.46 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.53–3.57 (m, 1H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.60–3.66 (m, 3H, H-5', H-6', H-6'), 3.70 (t, 1H, $J=3.2$ Hz, H-3), 3.74–3.78 (m, 2H, H-2, H-6), 3.83–3.87 (m, 1H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.88–4.04 (m, 5H, H-4, H-6, H-2', H-4', CH_2 Bn), 4.09 (s, 1H, H-3'), 4.16 (t, 1H, $J=6.8$ Hz, H-5), 4.42 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.58–4.62 (m, 2H, CH_2 Bn), 4.65 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.73 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.80 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.87 (d, 1H, $J=3.6$ Hz, H-1), 5.00 (d, 1H, $J=11.2$ Hz, CH_2 Bn), 5.15 (d, 1H, $J=3.6$ Hz, H-1'), 7.04–7.49 ppm (m, 20H, Ar-H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): $\delta = -5.6$ (CH_3), -5.5 (CH_3), -5.4 (CH_3), 18.0 (C_q *t*Bu), 18.2 (C_q *t*Bu), 25.8 (*t*Bu), 29.1 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 48.5 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 61.1 (C-6), 64.6 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 65.3 (C-5'), 65.6 (C-6'), 67.0 (C-5), 71.4 (CH_2 Bn), 71.5 (CH_2 Bn), 72.5 (CH_2 Bn), 73.1 (CH_2 Bn), 72.1, 74.1, 74.9, 75.1, 75.2 (C-2, C-4, C-2', C-3', C-4'), 76.1 (C-3), 97.0 (C-1), 99.9 (C-1'), 127.0–128.3 (Ar-CH), 138.4 (C_q Bn), 138.5 (C_q Bn), 139.0 (C_q Bn), 139.6 ppm (C_q Bn); HRMS: m/z : calcd for $\text{C}_{55}\text{H}_{79}\text{N}_3\text{O}_{11}\text{Si}_2 + \text{Na}^+$: 1036.51453; found: 1036.51517; IR (neat): $\tilde{\nu} = 1003$, 1092, 1454, 2095, 2858, 2928 cm^{-1} .

3-Azidopropyl-2,3-O-benzyl-6-O-tert-butylidimethylsilyl-4-O-(2,3-O-benzyl-6-O-tert-butylidimethylsilyl-4-O-(2,3-di-O-benzyl-4,6-O-di-tert-butylsilylidene- α -L-gulopyranosyl)- α -L-gulopyranoside (42): A solution of **39** (0.162 g, 0.324 mmol), diphenyl sulfoxide (0.164 g, 0.81 mmol), and tri-*tert*-butylpyrimidine (0.504 g, 0.125 mmol) in CH_2Cl_2 (6 mL) was stirred over activated 3-Å MS for 30 min. The reaction mixture was cooled to -60°C before triflic acid anhydride (57 μL , 0.340 mmol) was added. The reaction mixture was allowed to warm to -40°C and stirred for 1 h followed by the addition of acceptor **41** (0.164 g, 0.162 mmol) in CH_2Cl_2 (1.6 mL). Stirring was continued, the reaction mixture was allowed to warm to 10°C , and Et₃N (0.1 mL) was added. The reaction mixture was diluted with CH_2Cl_2 and washed with NaHCO₃ (aq). The aqueous layer was extracted twice with CH_2Cl_2 , and the collected organic layers were dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **42** as a colourless oil (0.102 g, 42%). $[\alpha]_D^{25} = -75.8$ ($c=1$, CHCl_3); $^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = -0.02$ (s, 3H, CH_3), -0.01 (s, 3H, CH_3), 0.01 (s, 3H, CH_3), 0.06 (s, 3H, CH_3), 0.87 (s, 9H, *t*Bu), 0.93 (s, 9H, *t*Bu), 0.96 (s, 9H, *t*Bu), 1.08 (s, 9H, *t*Bu), 1.88–1.93 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.32–3.37 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.39–3.45 (m, 2H), 3.69–3.86 (m, 8H), 3.89–3.98 (m, 7H), 4.03 (t, 1H, $J=9.6$ Hz), 4.09–4.20 (m, 5H), 4.28 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.45 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.59–4.66 (m, 3H, CH_2 Bn), 4.70 (d, 1H, $J=4.0$ Hz, H-1), 4.73 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.79 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.96 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 5.03 (d, 1H, $J=$

12.0 Hz, CH_2 Bn), 5.08 (d, 1H, $J=11.2$ Hz, CH_2 Bn), 5.11 (d, 1H, $J=2.8$ Hz), 5.16 (d, 1H, $J=2.4$ Hz) (H-1', H-1''), 6.97–7.55 ppm (m, 30H, Ar-H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): $\delta = -5.4$ (CH_3), -5.3 (CH_3), -5.2 (CH_3), 18.1 (C_q *t*Bu), 18.2 (C_q *t*Bu), 20.3 (C_q *t*Bu), 23.3 (C_q *t*Bu), 25.9 (*t*Bu), 25.9 (*t*Bu), 27.2 (CH_3 *t*Bu), 27.6 (CH_3 *t*Bu), 29.1 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 48.5 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 61.0, 61.3 (C-6, C-6'), 64.5 (4''), 64.6 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 66.8 (C-6''), 67.6 (C-5, C-5'), 71.0 (CH_2 Bn), 71.1 (CH_2 Bn), 71.1 (CH_2 Bn), 72.5 (CH_2 Bn), 72.9 (CH_2 Bn), 73.3 (CH_2 Bn), 73.0, 73.6, 74.9, 75.1, 75.4, 75.7, 75.8, 76.2, 97.2 (C-1), 99.3, 99.7 (C-1', C-1''), 127.0–128.4 (Ar-CH), 138.0 (C_q Bn), 138.4 (C_q Bn), 138.6 (C_q Bn), 139.5 (C_q Bn), 139.6 (C_q Bn), 138.7 ppm (C_q Bn); HRMS: m/z : calcd for $\text{C}_{83}\text{H}_{117}\text{N}_3\text{O}_{16}\text{Si}_3 + \text{Na}^+$: 1518.76338; found: 1518.76503; IR (neat): $\tilde{\nu} = 1084$, 1138, 1454, 1474, 2095, 2859, 2932 cm^{-1} .

3-Azidopropyl-2,3-O-benzyl-4-O-(2,3-O-benzyl- α -L-gulopyranosyl)- α -L-gulopyranoside (43): A solution of **42** (0.094 g, 0.063 mmol) in THF (2 mL) was cooled to 0°C and TBAF (0.28 mL, 0.28 mmol, 1M in THF) was added. The reaction mixture was diluted with EtOAc after 4 h, and washed with water and brine. The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography yielded **43** as a colourless oil (0.067 g, 83%). $^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 1.88$ – 1.96 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.34–3.39 (m, 3H), 3.47–3.59 (m, 4H), 3.64–3.67 (m, 4H), 3.73–3.79 (m, 4H), 3.82–3.87 (m, 7H), 3.97 (bs, 1H), 4.05 (bs, 1H), 4.14 (bs, 1H), 4.23 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.32 (d, 1H, $J=11.6$ Hz, CH_2 Bn), 4.34 (d, 1H, $J=10.8$ Hz, CH_2 Bn), 4.43 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.49 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.55 (d, 1H, $J=12.4$ Hz, CH_2 Bn), 4.57–4.68 (m, 5H, CH_2 Bn), 4.70 (d, 1H, $J=3.2$ Hz, H-1), 4.91 (d, 1H, $J=12.0$ Hz, CH_2 Bn), 4.95 (d, 1H, $J=3.6$ Hz), 4.97 (d, 1H, $J=3.6$ Hz, H-1', H-1''), 7.10–7.42 ppm (m, 30H, Ar-H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): $\delta = 29.1$ (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 48.4 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 62.5, 62.8, 64.2, 64.7, 65.7, 66.0, 66.5, 70.7, 71.2, 71.3, 71.5, 72.1, 72.6, 72.7, 73.1, 73.2, 73.3, 74.0, 74.8, 79.7, 80.9, 97.3, 99.2, 99.9 (C-1, C-1', C-1''), 127.4–128.5 (Ar-CH), 137.4 (C_q Bn), 137.6 (C_q Bn), 138.2 (C_q Bn), 138.2 (C_q Bn), 138.3 (C_q Bn), 139.0 ppm (C_q Bn); ESI-MS: m/z : 1128.5 $[M+\text{Na}^+]$; IR (neat): $\tilde{\nu} = 1069$, 1109, 1454, 2095, 2878 cm^{-1} .

3-Aminopropyl [4-O-(α -L-gulopyranosyl)uronate]- α -L-gulopyranoside]-uronic acid (44): Compound **40** (0.243 g, 0.310 mmol) was taken up in CH_2Cl_2 (2 mL), *t*BuOH (2 mL), and H₂O (0.5 mL) and TEMPO (0.019 g, 0.124 mmol) and BAIB (0.496 g, 1.55 mmol) were added. The reaction mixture was stirred vigorously until analysis by TLC showed complete conversion of the starting material. Na₂S₂O₃ (10 mL, aq) was added and the resulting mixture was stirred for 15 min. The layers were separated and the aqueous phase was acidified with 1M HCl and extracted three times with CH_2Cl_2 . The combined organic layers were dried over MgSO₄ and concentrated in vacuo. The resulting syrup was then dissolved in *t*BuOH (2 mL) and H₂O (2 mL) before a catalytic amount of Pd on charcoal was added. The reaction mixture was stirred overnight in an H₂ atmosphere and filtered. Gel filtration (HW-40) afforded the desired disaccharide **44** (0.086 g, 90%). $^1\text{H NMR}$ (400 MHz, D₂O): $\delta = 1.98$ – 2.06 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.14–3.23 (m, 2H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.65–3.71 (m, 1H, CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 3.92–3.99 (m, 4H, H-2, H-2', H-3', CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 4.09–4.16 (m, 3H, H-3, H-4, H-4'), 4.41 (d, 1H, $J=1.2$ Hz, H-5'), 4.47 (s, 1H, H-5), 4.96 (d, 1H, $J=3.6$ Hz), 5.02 ppm (d, 1H, $J=4.0$ Hz) (H-1, H-1'); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): $\delta = 26.0$ (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 39.0 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 64.5, 65.1 (C-2, C-2'), 66.7 (C-5), 67.8 (CH_2 $\text{C}_3\text{H}_6\text{N}_3$), 68.1 (C-5'), 69.1, 70.3 (C-3, C-3'), 70.8 (C-4'), 80.5 (C-4), 98.4, 100.9 (C-1, C-1'), 175.7 (COOH), 176.1 ppm (COOH); HRMS: m/z : calcd for $\text{C}_{15}\text{H}_{25}\text{NO}_{13} + \text{Na}^+$: 450.12181; found: 450.12170.

3-Aminopropyl [4-O-(4-O-(α -L-gulopyranosyl)uronate)- α -L-gulopyranosyl]uronate]- α -L-gulopyranoside]-uronic acid (45): Compound **43** (0.039 g, 0.034 mmol) was taken up in CH_2Cl_2 (1 mL), *t*BuOH (1 mL), and H₂O (0.5 mL) and TEMPO (0.003 g, 0.021 mmol) and BAIB (0.082 g, 0.257 mmol) were added. The reaction mixture was stirred vigorously until analysis by TLC showed complete conversion of the starting material. Na₂S₂O₃ (2 mL, aq) was added and the resulting mixture was stirred for 15 min. The layers were separated and the aqueous phase was acidified with 1M HCl and extracted three times with CH_2Cl_2 . The combined organic layers were dried over MgSO₄ and concentrated in vacuo. The re-

sulting syrup was then dissolved in *t*BuOH (1 mL) and H₂O (1 mL) before a catalytic amount of Pd on charcoal was added. The reaction mixture was stirred overnight in an H₂ atmosphere and filtered. Gel filtration (HW-40) afforded the desired trisaccharide **45** (0.018 g, 85 %). ¹H NMR (400 MHz, D₂O): δ = 1.98–2.02 (m, 2H, CH₂ C₃H₆N₃), 3.14–3.19 (m, 2H, CH₂ C₃H₆N₃), 3.65–3.71 (m, 1H, CH₂ C₃H₆N₃), 3.91–4.00 (m, 5H, H-2, H-2', H-2'', H-3'', CH₂ C₃H₆N₃), 4.04–4.07 (m, 2H, H-3, H-3'), 4.14 (bs, 3H, H-4, H-4', H-4''), 4.41 (s, 1H), 4.46 (s, 1H), 4.47 (s, 1H, H-5, H-5', H-5''), 4.96 (d, 1H, *J* = 4.0 Hz), 5.02 (d, 1H, *J* = 3.6 Hz), 5.06 ppm (d, 1H, *J* = 4.0 Hz) (H-1, H-1', H-1''); ¹³C NMR (100 MHz, CDCl₃): δ = 26.2 (CH₂ C₃H₆N₃), 39.0 (CH₂ C₃H₆N₃), 64.6, 65.2, 65.2 (C-2, C-2', C-2''), 66.6, 67.3, 68.2 (C-5, C-5', C-5''), 67.8 (CH₂ C₃H₆N₃), 69.1, 69.2, 70.4 (C-3, C-3', C-3''), 70.8 (C-4'), 80.0, 80.5 (C-4, C-4'), 98.4, 100.7, 100.9 (C-1, C-1', C-1''), 175.5 (COOH), 175.6 (COOH), 176.1 ppm (COOH); HRMS: *m/z*: calcd for C₂₁H₃₃NO₁₉ + Na⁺: 626.15390; found: 626.15384.

Acknowledgements

This work was supported by the Council for Chemical Sciences of the Netherlands Organisation for Scientific Research (NWO).

- [1] T. H. Flo, L. Ryan, E. Latz, O. Takeuchi, B. G. Monks, E. Lien, Ø. Halaas, S. Akira, G. Skjåk-Bræk, D. T. Golenbock, T. Espevik, *J. Biol. Chem.* **2002**, 277(38), 35489–35495.
- [2] S. T. Moe, K. I. Draget, G. Skjåk-Bræk, O. Smidsrød, *Food Polysaccharides and Their Applications*, (Ed.: A. M. Stephen) Marcel Dekker, Inc., New York, **1995**, pp. 245–286.
- [3] M. Iwamoto, M. Kurachi, T. Nakashima, D. Kim, K. Yamaguchi, T. Oda, Y. Iwamoto, T. Muramatsu, *FEBS Lett.* **2005**, 579, 4423–4429.
- [4] L. J. van den Bos, J. Dinkelaar, H. S. Overkleef, G. A. van der Marel, *J. Am. Chem. Soc.* **2006**, 128, 13066–13067.
- [5] L. J. van den Bos, J. D. C. Codée, R. E. J. N. Litjens, J. Dinkelaar, H. S. Overkleef, G. A. van der Marel, *Eur. J. Org. Chem.* **2007**, 3963–3976.
- [6] a) K. Katano, H. An, Y. Aoyagi, M. Overhand, S. J. Sucheck, W. C. Stevens Jr., C. D. Hess, X. Zhou, S. M. Hecht, *J. Am. Chem. Soc.* **1998**, 120, 11285–11296; b) D. L. Boger, T. Honda, *J. Am. Chem. Soc.* **1994**, 116, 5647–5656.
- [7] I. M. García-Moreno, P. Díaz-Pérez, C. Ortiz Mellet, J. M. García Fernández, *J. Org. Chem.* **2003**, 68, 8890–8901.
- [8] K. Larsen, C. E. Olsen, M. S. Motawia, *Carbohydr. Res.* **2003**, 338, 199–202.
- [9] L. J. van den Bos, J. D. C. Codée, J. H. van Boom, H. S. Overkleef, G. A. van der Marel, *Org. Lett.* **2004**, 6, 2165–2168.
- [10] a) L. J. Van den Bos, R. E. J. N. Litjens, R. J. B. H. N. van den Berg, H. S. Overkleef, G. A. van der Marel, *Org. Lett.* **2005**, 7, 2007–2010; b) J. D. C. Codée, L. J. van den Bos, R. E. J. N. Litjens, H. S. Overkleef, J. H. van Boom, G. A. van der Marel, *Org. Lett.* **2003**, 5, 1947–1950; c) D. Crich, M. Smith, *J. Am. Chem. Soc.* **2001**, 123, 9015–9020.
- [11] D. Crich, M. de la Mora, A. U. Vinod, *J. Org. Chem.* **2003**, 68, 8142–8148.
- [12] A. Imamura, H. Ando, S. Korogi, G. Tanabe, O. Muraoka, H. Ishida, M. Kiso, *Tetrahedron Lett.* **2003**, 44, 6725–6728.
- [13] a) H. Paulsen, *Angew. Chem.* **1982**, 94, 184–201; *Angew. Chem. Int. Ed. Engl.* **1982**, 21, 155–173; B. Frazer-Reid, J. C. Lúpez, A. M. Gúmez, C. Uriel, *Eur. J. Org. Chem.* **2004**, 1387–1395; b) X. Zhu, B. Yu, Y. Hui, R. R. Schmidt, *Eur. J. Org. Chem.* **2004**, 965–973.
- [14] G. H. Veeneman, J. H. van Boom, *Tetrahedron Lett.* **1990**, 31, 275–278.
- [15] B. A. Garcia, J. L. Poole, D. Y. Gin, *J. Am. Chem. Soc.* **1997**, 119, 7597–7598.
- [16] a) J. D. C. Codée, B. Stubba, M. Schiattarella, H. S. Overkleef, C. A. A. van Boeckel, J. H. van Boom, G. A. van der Marel, *J. Am. Chem. Soc.* **2005**, 127, 3767–3773; b) D. Crich, W. Li, *Org. Lett.* **2006**, 8, 959–962.
- [17] J. Dinkelaar, M. D. Witte, L. J. van den Bos, H. S. Overkleef, G. A. van der Marel, *Carbohydr. Res.* **2006**, 341, 1723–1729.
- [18] L. Huang, N. Teumelsan, X. Huang, *Chem. Eur. J.* **2006**, 12, 5246–5252.
- [19] P. Deslongchamps, *Stereoelectronic Effects in Organic Chemistry*, Pergamon Press, Oxford, **1983**.
- [20] S. J. Angyal, *Adv. Carbohydr. Chem. Biochem.* **1984**, 42, 15–68.
- [21] a) D. Crich, S. X. Sun, *Tetrahedron* **1998**, 54, 8321–8348; b) T. Nokami, A. Shibuya, H. Tsuyama, S. Suga, A. A. Bowers, D. Crich, J. Yoshida, *J. Am. Chem. Soc.* **2007**, 129, 10922–10928.
- [22] R. U. Lemieux, G. Huber, *Chem. Eng. News Chem. Canada* **1954**, 32, 128–133.
- [23] X. Zhu, S. Kawatkar, Y. Rao, G.-J. Boons, *J. Am. Chem. Soc.* **2006**, 128, 11948–11957.
- [24] a) R. J. Woods, C. W. Andrews, J. P. Bowen, *J. Am. Chem. Soc.* **1992**, 114, 859–864; b) M. Miljković, D. Yeagley, P. Deslongchamps, Y. L. Dory, *J. Org. Chem.* **1997**, 62, 7597–7604; c) T. Nukada, A. Bérces, L. Wang, M. Z. Zgierski, D. M. Whitfield, *Carbohydr. Res.* **2005**, 340, 841–852.
- [25] a) H. H. Jensen, M. Bols, *Acc. Chem. Res.* **2006**, 39, 259–265; b) S. Chamberland, J. W. Ziller, K. A. Woerpel, *J. Am. Chem. Soc.* **2005**, 127, 5322–5323; c) C. G. Lucero, K. A. Woerpel, *J. Org. Chem.* **2006**, 71, 2641–2647.
- [26] a) L. Ayala, C. G. Lucero, J. A. C. Romero, S. A. Tabacco, K. A. Woerpel, *J. Am. Chem. Soc.* **2003**, 125, 15521–15528; b) O. Boutur-eira, M. A. Rodríguez, D. Benito, M. I. Matheu, Y. Díaz, S. Castil-lón, *Eur. J. Org. Chem.* **2007**, 3564–3572.
- [27] R. V. Stevens, *Acc. Chem. Res.* **1984**, 17, 289–296.
- [28] A. V. Demchenko, E. Rousson, G.-J. Boons, *Tetrahedron Lett.* **1999**, 40, 6523–6526.

Received: May 20, 2008

Revised: July 14, 2008

Published online: September 3, 2008