



## Kainic Acid as Conformationally Constrained Glutamic Acid Analog in Peptide Synthesis<sup>1</sup>

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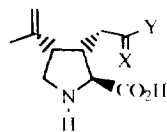
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**Abstract :** A synthetic protocol is described which allows the incorporation of the  $\gamma$ -amide (Kan) and the  $\gamma$ -benzyl ester [Kai(Bzl)] and ether [Kol(Bzl)] of kainic acid (Kai) into peptide chains as exemplified with the synthesis of the substance P (SP) analogs [Kan<sup>5</sup>]-SP<sub>5-11</sub>, [Glp<sup>5</sup>, Kan<sup>6</sup>]-SP<sub>5-11</sub>, [Kan<sup>6</sup>]-SP<sub>6-11</sub>, [Glp<sup>5</sup>, Kai(Bzl)]<sup>11</sup>-SP<sub>5-11</sub> and [Glp<sup>5</sup>, Kol(Bzl)]<sup>11</sup>-SP<sub>5-11</sub>.

We have recently indicated that (-)- $\alpha$ -kainic acid (Kai, **1**), a natural product isolated from the marine alga *Digenea simplex* with powerful neuroexcitatory activity in the mammalian central nervous system, may be used as a conformationally restricted analog of glutamic acid (Glu) in biologically important compounds.<sup>2</sup> We now wish to report on the development of a methodology which allows the incorporation of derivatives of Kai modified at the  $\gamma$ -carboxy function, such as the  $\gamma$ -amide (Kan, **2**)<sup>3</sup> and the  $\gamma$ -benzyl ester [Kai(Bzl), **3**] and ether [Kol(Bzl), **4**], into peptide chains. The C-terminal hexa (**5**)- and heptapeptides (**6**) of the mammalian tachykinin substance P (SP, **7**) were used as models in the present work. Structure-activity relationship studies have shown that the biological activity of **7** mainly depends on the amino acid sequence of peptides **5** and **6**. Moreover, replacement of N-terminal glutamine (Gln) by pyroglutamic acid (Glp) and of C-terminal methionine (Met) by Glu(Bzl) in these peptides results in analogs with higher and equipotent respectively contractile activity to that of the parent peptide **7**.<sup>4</sup> Using the synthetic protocol described below, the SP analogs **8-12** were successfully prepared. These SP analogs contain Kan and Glp in place of Gln at positions 6 or 5 and 5, respectively and Kai(Bzl) and Kol(Bzl) in place of Met at position 11. It should be pointed out that the Kai analog with a reduced  $\gamma$ -carboxy function, abbreviated here as Kol, represents a conformationally restricted analog of the hydroxyamino acid  $\delta$ -hydroxynorvaline (Hnv).<sup>5</sup>

Incorporation of compounds **2** and **3** into peptide chains and the preparation of derivatives of ether **4** suitable for use in peptide synthesis required the use of Kai-intermediates selectively protected at one of the two available carboxy functions. Such an intermediate, namely lactone **13**, had been already described and successfully used for the preparation of a series of Kai  $\gamma$ -amides.<sup>3</sup> Lactone **13**, readily available through N-protection of Kai with the *tert*-butoxycarbonyl (Boc) group, followed by iodolactonization of the resulting Boc-Kai, served in our work as the key-intermediate for the

conversion of Kai to both Kai-NH<sub>2</sub>, for use as the C-terminal amino acid, and Kan for use in the N-terminal region of the peptide chain. In both cases, the commercially available triphenylmethylamine (TrtNH<sub>2</sub>) was employed as an anhydrous and convenient source of ammonia.<sup>6</sup>

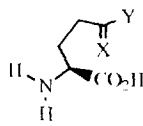


**Kai** (1) : X=O, Y=OH

**Kan** (2) : X=O, Y=NH<sub>2</sub>

**Kai(Bzl)** (3) : X=O, Y=OBzl

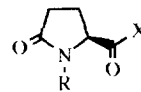
**Kol(Bzl)** (4) : X=H<sub>2</sub>, Y=OBzl



**Glu** : X=O, Y=OH

**Gln** : X=O, Y=NH<sub>2</sub>

**Hmv** : X=H<sub>2</sub>, Y=OH



**Glp** : R=H, X=OH

**Trt-Glp-OBt** : R=Trt, X=OBt

Gln-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>

**SP<sub>6-11</sub>** (5)

Kan-Gln-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>

**[Kan<sup>5</sup>]-SP<sub>5-11</sub>** (8)

Gln-Gln-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>

**SP<sub>5-11</sub>** (6)

Glp-Kan-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>

**[Glp<sup>5</sup>, Kan<sup>6</sup>]-SP<sub>5-11</sub>** (9)

Arg-Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>

**SP** (7)

Kan-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>

**[Kan<sup>6</sup>]-SP<sub>6-11</sub>** (10)

Glp-Gln-Phe-Phe-Gly-Leu-Kai(Bzl)-NH<sub>2</sub>

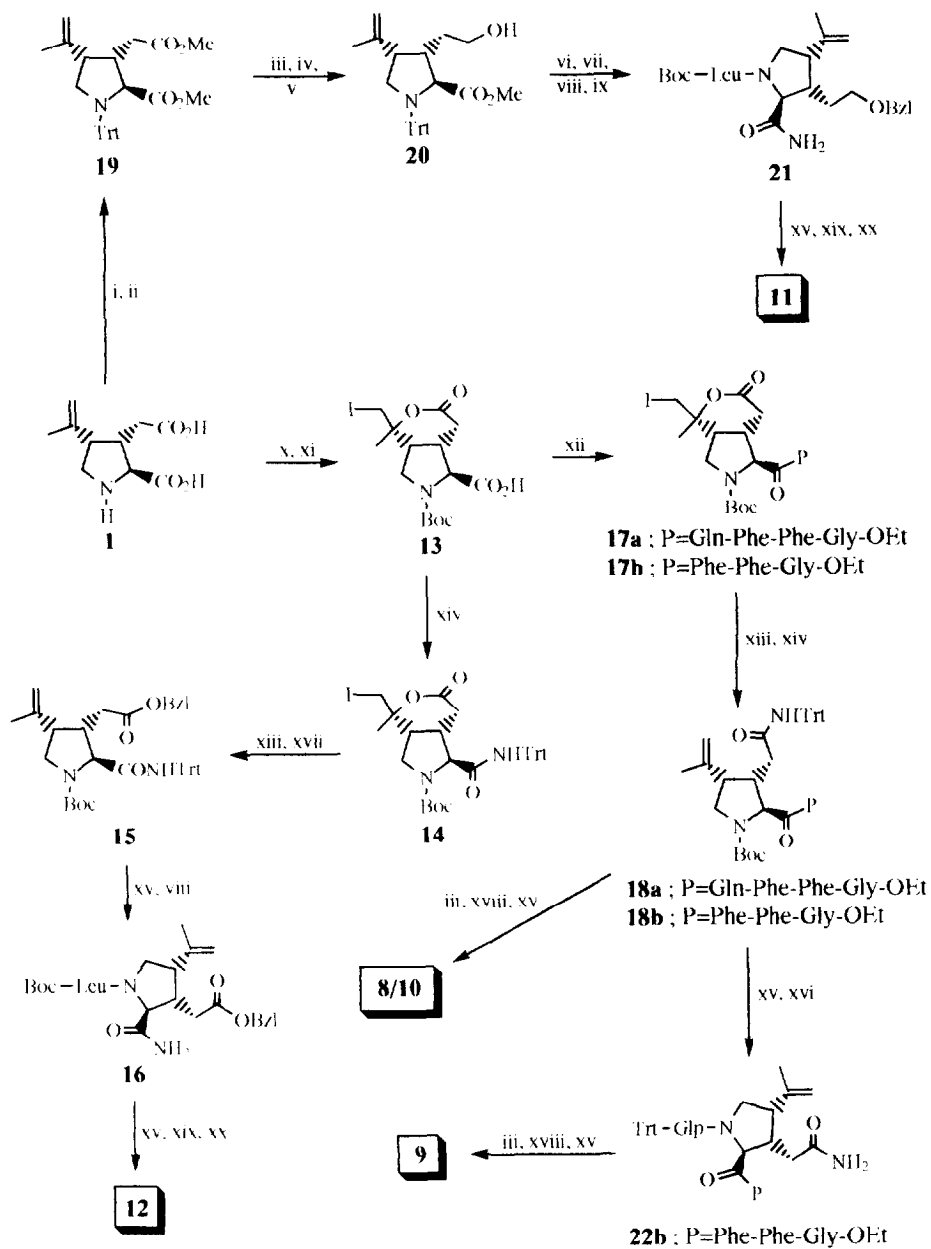
**[Glp<sup>5</sup>, Kai(Bzl)<sup>11</sup>]-SP<sub>5-11</sub>** (11)

Glp-Gln-Phe-Phe-Gly-Leu-Kol(Bzl)-NH<sub>2</sub>

**[Glp<sup>5</sup>, Kol(Bzl)<sup>11</sup>]-SP<sub>5-11</sub>** (12)

Thus, coupling lactone **13** with TrtNH<sub>2</sub> provided the corresponding tritylamide **14** which upon treatment with Zn dust in glacial (gl.) AcOH, followed by esterification with benzyl alcohol under Mitsunobu type reaction conditions, gave the  $\gamma$ -benzylester derivative **15** in 65% overall yield. It should be noted that, due to its bulk, the coupling of TrtNH<sub>2</sub> with **13** was most efficiently performed, in 83% yield, using the coupling agent PyBrOP.<sup>7</sup> Treatment of **15** with trifluoroacetic acid (TFA)/CH<sub>2</sub>Cl<sub>2</sub> (3:1) resulted in simultaneous deprotection of both amine and amide functions. The resulting trifluoroacetate salt was routinely coupled with Boc-Leu.H<sub>2</sub>O, using *N,N'*-dicyclohexylcarbodiimide (DCC) and 1-hydroxybenzotriazole (HOBt), to give the dipeptide amide **16** in 70% yield. On the other hand, coupling iodolactone **13** with the tetrapeptide ester H-Gln-Phe-Phe-Gly-OEt or the tripeptide ester H-Phe-Phe-Gly-OEt, in presence of the coupling agent BOP,<sup>8</sup> produced the corresponding iodolactones **17a** and **b** in 84-95% yields. These amino components were obtained from TFA-mediated *N*-deprotection of the corresponding peptides Boc-Gln-Phe-Phe-Gly-OEt and Boc-Phe-Phe-Gly-OEt, available in our laboratory from previous studies. Treatment of lactones **17a** and **b** with Zn/gl. AcOH, followed by coupling with TrtNH<sub>2</sub> in the presence of PyBrOP gave good yields (65-75%) of fully protected Kan peptides **18a** and **b**, respectively.

*N*-tritylation of Kai, followed by Mitsunobu type esterification gave the diester **19**, which was regiospecifically saponified at the  $\gamma$ -carboxy function.<sup>2</sup> The resulting Kai derivative was reduced at the  $\gamma$ -carboxy function in an one-pot experiment, involving activation with DCC/HOBt followed by NaBH<sub>4</sub> reduction,<sup>5</sup> to give the Kol derivative **20** in 78% yield. Alcohol **20** was sequentially benzylated at the hydroxy function with BzlBr/NaH, *N*-detritylated with TsOH.H<sub>2</sub>O in refluxing isopropanol, coupled with Boc-Leu.H<sub>2</sub>O using the system DCC/HOBt and finally ammonolysed to give the dipeptide amide **21** in



**Reagents** : i. (a)  $\text{Me}_3\text{SiCl}/\text{Et}_3\text{N}$ , (b)  $\text{TrtCl}$ , (c)  $\text{MeOH}$ ; ii,  $\text{MeOH}/\text{Ph}_3\text{P}/\text{EtO}_2\text{CN}=\text{NCO}_2\text{Et}$ ; iii,  $\text{NaOH}$ ; iv,  $\text{DCC}/\text{HOBT}$ ; v,  $\text{NaBH}_4$ ; vi,  $\text{NaH}/\text{BzI}^+\text{Br}^-$ ; vii,  $\text{TsOH}\cdot\text{H}_2\text{O}$ ; viii,  $\text{Boc-Leu}/\text{DCC}/\text{HOBT}$ ; ix,  $\text{NH}_3$ ; x,  $\text{Boc}_2\text{O}$ ; xi,  $\text{I}_2/\text{KI}/\text{NaHCO}_3$ ; xii,  $\text{BOP}/\text{H-Gln-Phe-Phe-Gly-OEt}$  or  $\text{BOP}/\text{H-Phe-Phe-Gly-OEt}$ ; xiii,  $\text{Zn}/\text{gl. AcOH}$ ; xiv,  $\text{TrtNH}_2/\text{PyBrOP}$ ; xv,  $\text{TFA}$ ; xvi,  $\text{Trt-Glp-OBt}$ ; xvii,  $\text{BzI}^+\text{OH}^-/\text{Ph}_3\text{P}/\text{EtO}_2\text{CN}=\text{NCO}_2\text{Et}$ ; xviii,  $\text{BOP}/\text{H-Leu-Met-NH}_2$ ; xix,  $\text{BOP}/\text{Trt-Glp-Gln-Phe-Phe-Gly-OH}$ ; xx, 20%  $\text{TFA}$  in  $\text{CH}_2\text{Cl}_2$ .

76% overall yield.

The dipeptides **16** and **21** and the penta- and tetrapeptides **18a** and **b**, were subsequently used for the preparation of the projected SP analogs **8-12** as follows. Saponification of **18a** and BOP-mediated coupling of the resulting pentapeptide acid with H-Leu-Met-NH<sub>2</sub> gave the corresponding heptapeptide amide in 90% yield, which upon treatment with TFA afforded the SP analog **8**. Similarly, saponification of tetrapeptide ester **18b**, followed by BOP-mediated coupling of the resulting tetrapeptide acid with H-Leu-Met-NH<sub>2</sub> gave the corresponding hexapeptide amide, in 84% yield, which was also deprotected with TFA to give the SP analog **10**. On the other hand, *N*-deprotection of **18b** with TFA and coupling of the resulting tetrapeptide ester with Trt-Glp-OBt, a useful compound for the introduction of Glp in peptide chains,<sup>9</sup> gave the pentapeptide **22b** in 82% yield. Saponification of **22b**, followed by BOP-mediated coupling with H-Leu-Met-NH<sub>2</sub> gave the corresponding heptapeptide in 84% yield which upon TFA-mediated deprotection provided the SP analog **9**. For the preparation of analogs **11** and **12**, the tetrapeptide Boc-Gln-Phe-Phe-Gly-OEt was *N*-deprotected with TFA and subsequently coupled with Trt-Glp-OBt to give Trt-Glp-Gln-Phe-Phe-Gly-OEt in 82% yield, which upon saponification provided the corresponding pentapeptide acid. BOP-mediated coupling of this acid with the trifluoroacetate salts of the dipeptides H-Leu-Kai(Bzl)-NH<sub>2</sub> and H-Leu-Kol(Bzl)-NH<sub>2</sub> gave the heptapeptides Trt-Glp-Gln-Phe-Phe-Gly-Leu-Kai(Bzl)- and Kol(Bzl)-NH<sub>2</sub>, in 88 and 85 % yield, respectively. These amides, upon treatment with 20% TFA in CH<sub>2</sub>Cl<sub>2</sub> provided the desired SP analogs **11** and **12**. Biological evaluation of the thus obtained SP analogs **8-12** is now in progress.

## REFERENCES AND NOTES

1. New compounds gave analytical and spectral (IR, <sup>1</sup>H NMR and FAB-MS) data in agreement with the proposed structures. Amino acids referred in this work were of the *S* configuration.
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