ACS Medicinal Chemistry Letters

Synthesis and Evaluation of Quinazolines as Inhibitors of the Bacterial Cell Division Protein FtsZ

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(5) Supporting Information

ABSTRACT: The bacterial cell division protein FtsZ is one of many potential targets for the development of novel antibiotics. Recently, zantrin Z3 was shown to be a cross-species inhibitor of FtsZ; however, its specific interactions with the protein are still unknown. Herein we report the synthesis of analogues that contain a more tractable core structure and an analogue with single-digit micromolar inhibition of FtsZ's GTPase activity, which represents the most potent inhibitor of *Escherichia coli* FtsZ reported to date. In addition, the zantrin Z3 core has been converted to two potential photo-cross-linking reagents for proteomic studies that could shed light on the molecular interactions between FtsZ and molecules related to zantrin Z3.



KEYWORDS: FtsZ, zantrin Z3, SAR, bacterial cell division

 ${\rm B}$ acterial infections are increasingly difficult to combat as available antibiotics become less effective and bacteria evolve resistance to current therapies.¹ With more than two million illnesses annually attributed to resistant bacteria, and the diminishing timeline between drug implementation and resistance, we face a dire need for new antibiotics.² Oftentimes, available antibiotics are modified to prolong their use for a known target,^{3,4} but a new protein or pathway offers an area of therapeutic research that has no known resistance. The bacterial divisome is a complex set of biochemical machinery that contains many proteins whose shape and function remain somewhat elusive. These proteins offer new areas of antibiotic study.^{5,6} Temperature-sensitive filamenting protein Z (FtsZ) is highly conserved among bacterial species and is essential for cell division. Therefore, FtsZ is an exciting target for developing a broad spectrum antibiotic.⁷ The homologous eukaryotic protein, tubulin, has been successfully targeted by small molecules such as taxol,^{8,9} epothilone,¹⁰ and many others to halt the division of cancer cells.¹¹⁻¹³ By analogy, an inhibitor of FtsZ could halt bacterial cell division and form the basis of a novel antibiotic.

There are many reported inhibitors of FtsZ that have shown the ability to cause filamentation in bacteria.^{14–18} However, certain inhibitors were recently shown to have irreproducible activity or showed the filamentation phenotype but were not specific to FtsZ inhibition.^{19,20} Two compounds were found to directly modulate FtsZ: PC190723 and zantrin Z3 (Figure 1). PC190723 has been shown to activate the GTPase activity of *Staphylococcus aureus* FtsZ.^{21–23} On the other hand, zantrin Z3²⁴ (ZZ3, 1) was the only compound to reliably inhibit *Bacillus subtilis* FtsZ (BsFtsZ) and *Escherichia coli* FtsZ (EcFtsZ). Although ZZ3 reliably inhibits FtsZ, its specific molecular interactions are unknown. Furthermore, very little



Figure 1. Small molecule modulators of FtsZ. (A) Activator PC190723.^{19,23} (B) Zantrin Z3 (1) and its more potent analogue, dimethyl ZZ3 (2).¹⁹ Three regions are identified in ZZ3 as synthetically accessible for further modification.

structural data is available for inhibitor-FtsZ interactions even though many crystal structures of the protein itself have been published. Despite only modest inhibitory activity in an absolute sense, zantrin Z3 remains the best known inhibitor of FtsZ's GTPase activity. Unlike tubulin, for which inhibitors act at nano- and picomolar concentrations, all known FtsZ inhibitors act in the micromolar range!

Preliminary modifications of **1** led to the discovery of **2**, which is twice as potent as its parent compound¹⁹ and provided a benchmark for further structure–activity studies. ZZ3 has three regions where modifications are synthetically accessible. Substituted anthranilic acids were used to explore the binding interactions of the benzoquinazoline core by replacing the fused

Received: December 2, 2014 Accepted: January 7, 2015

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benzene ring with functional groups of varied sterics and electronics (Figure 1, region 1), while the styryl component was replaced by functional groups thought to be isosteric in region 2. Continued exploration of the amine fragment was achieved through S_NAr reactions. Further modifications to aryl halide analogues allowed for the creation of potential photocross-linkers to be used as small molecule probes for binding site identification of ZZ3 derivatives in FtsZ.

Early changes to 1 focused on reducing its size in hopes of finding a compound with greater "ligand efficiency." This term has emerged to describe the level of activity on a per-atom (or per-Dalton) basis, thus avoiding a tendency to achieve potency at the expense of drug-likeness.²⁵ The 4-quinazalone core was synthesized in one step from commercially available substituted anthranilic acids (3a-g), subjected to acid-mediated aldol condensation conditions and subsequently chlorinated with phosphorus(V) oxychloride to give quinazolines 4a-g in moderate yields. Amination with *N*,*N*-dimethylethylenediamine and Hünig's base yielded 5a-g in high yields (Scheme 1).¹⁹





^aReagents and conditions: (a) R^2 -B(OH)₂, dppf·PdCl₂(II), CH₃CN/H₂O (1:1), μ W, 140 °C, 45 min, 12-48% yield. Compounds **5k**–I were synthesized similarly from **5g** in 24 and 20% yield, respectively.

Table 1. Region 1 Analogues of 2 and Related IC₅₀.



Compounds **5a–g** were then subjected to an enzymecoupled GTPase assay to measure their half-maximal inhibition concentration (IC_{50}) .²⁶ Methyl and bromo substitution at the 6-position (R²) exhibited the best inhibitory activity, leading to investigation of larger substituents at this position. Palladiumcatalyzed cross-coupling reactions of aryl bromide **5f** afforded **5h–k** in acceptable yields.²⁷ Cyclohexenylboronic acid was similarly coupled in order to test a nonaromatic substituent (**51**). Diminished reactivity of the alkenyl boronic ester required use of the 6-iodo precursor (**5g**) to boost the yield to an isolatable amount. None of the subsequent (**5h–k**) compounds showed better inhibitory activity compared to the 6-methyl analogue.

With the 6-position fixed as methyl, alteration and replacement of the styryl side chain (region 2) were examined. Compounds **8a**–**d** were synthesized in good yield as previously described in Scheme 1. 2-Amino-5-methylbenzoic acid was cyclized with potassium cyanate followed by sodium hydroxide and hydrochloric acid to form quinazolinedione **6**.²⁸ Chlorination and amination yielded 7 in good yield.¹⁹ Boc-protection of 7,²⁹ subsequent displacement of the chloride, and deprotection with trifluoroacetic acid resulted in benzylamine derivatives **8e** and **8f** (Figure 2).³⁰ Under the same deprotection conditions,



Figure 2. Synthesis and IC₅₀ of compounds 8a–m. (A) ^aCompounds 8a–d were made in 4 steps as previously described in Scheme 1. Compounds 8e–g: (1) Boc₂O, DMAP, Et₃N, THF, 25 ^oC, 24 h, 76%; (2) 4-RBnNH₂, DMSO, 90 ^oC, 16 h, 43–64%; (3) TFA, CH₂Cl₂, 0 ^oC to rt, 24 h, 100%. ^bUpon subjection to TFA, the 4-methoxybenzylamine derivative (not shown) yielded 8g in 40% yield. (b) Compounds 8h–l were synthesized by Suzuki couplings as described in Scheme 1 in 24–65%. (B) Region 2 analogues and their corresponding IC₅₀.

the 4-methoxybenzylamine derivative (not shown) yielded primary amine 8g. Compound 7 was also subjected to Suzuki cross-couplings with dppf·PdCl₂(II) to give biaryl derivatives 8h-1.²⁷ Unfortunately, none of the changes in region 2 demonstrated better activity than 1 or 2.

The observation that major alterations to the styryl unit diminished activity suggested that an isosteric replacement was required. Amides, often considered isosteric to alkenes, required incorporation of a pendant carboxyl group. Fisher esterification of 2-amino-5-methylbenzoic acid yielded amino ester 9.³¹ Cyclization with the ethyl variant of Mander's reagent³² followed by chlorination yielded **10**, which was subsequently aminated to provide esters **11a** and **11b** (Scheme 2). In order to directly compare the styryl moiety to an isostere,

Scheme 2. Synthesis and IC₅₀ of 11a-d^a



^aReagents and conditions: (a) **11c** (1) *n*-BuLi, 4-ClPhNH₂, DMPU THF, 0 °C, 20 min; **11a**, THF, rt to 80 °C, 1 h; 10%. (2) HCl/ dioxane, 40 °C, 3 h, 88%. (b) **11b**, *i*-PrMgCl·LiCl, THF, rt, 12 h.

11a was converted to a benzamide analogue. The dimethylamine version of the 4-chlorobenzamide (not shown) proved difficult to isolate cleanly; fortunately, concurrent exploration of the amine fragment yielded a Boc-protected amine would reveal an ammonium salt with better inhibition than the corresponding dimethylamine (Table 2). Ester **11a** was converted to benzamide **11c** in 88% yield after deprotection. 4-Chlorobenzylamide **11d** was synthesized in 25% yield providing a less rotationally hindered analogue to parallel the styrene. Neither amide modification improved inhibitory activity relative to **1** and **2** despite incorporating the ammonium side chain in **11c**.

The final modifications examined involved altering or replacing the amino ethyl side chain (region 3). Compound **4c** was subjected to S_NAr reactions with nucleophiles containing single-point changes to generate analogues with different side chains at the 4-position of the quinazoline ring. Deprotection of **13b** with TFA gave **13c** in quantitative yield. These three compounds (**13a–c**, Scheme 3) suggested a



 $\begin{array}{l} X=N,\ O;\ R^3=OCH_3,\ SCH_3,\ NHBoc,\ NH_3^\bullet TFA,\ N(CH_3)_2\\ N(CH_3)_3^\bullet I,\ CO_2t\text{-}Bu,\ NHAc \end{array}$

^aReagents and conditions: ^b13a-b were synthesized from 4c in 41% yield each. ^c13d-j were synthesized similarly from 12 in 54-82% yield. (a) Compounds 13c and 13l: TFA, CH_2Cl_2 , 0 °C to rt, 3 h, 80-100%. (b) Compound 13k: CH_3I , CH_3OH , rt, 12 h, 21%. (c) Compound 13m: Ac_2O , Et_3N , CH_2Cl_2 , rt, 45 min, 31%.





positively charged fragment is necessary for inhibition, although the ammonium substrate (13c) only showed marginal activity. Upon comparing the most potent inhibitors in the 2- and 3-ring systems (5c and 2, respectively), we prepared several other substrates (Table 2, 13d-j) by cyclizing 2-naphthoic acid and subsequently chlorinating as described in Scheme 1. Substitution with 2-(dimethylamino)ethanol resulted in 13d, which showed moderate inhibition. Further substitutions with various amines gave 13e-j. Deprotection of 13i with TFA resulted in 131.³³ Further modification through acetylation³⁴ of 13l to 13m resulted in complete loss of inhibitory activity. While a positively charged "tail" retained some inhibitory activity (13d and 13g), the quaternary ammonium side chain of compound 13k, which was formed by alkylation with $CH_3I_3^{35}$ is quite bulky and likely disrupts important binding interactions. Conversely, the benzoquinazoline ammonium substrate (13l) is now the first substrate to demonstrate activity in the single-digit micromolar range.

In addition to optimizing inhibitory activity through structure-activity relationships (SAR), knowledge of modifications that are tolerated has provided the structural basis for the design of photoaffinity reagents. A survey of potential crosslinkers revealed photoactivatable azides, benzophenones and diazirines, which upon irradiation could covalently link a ZZ3 derivative to FtsZ. Digestion and mass spectrometry analysis would lead to proteomic information that could be used to identify the binding-site of ZZ3. Benzophenones are historically reliable cross-linkers that are easily synthesized but run the risk of reduced binding due to the size of the phenyl ketone. The preceding observations suggested the phenylketone could be installed at the 6-position of the quinazoline core. A carbonylative Stille coupling with tributylphenylstannane and carbon monoxide readily converted aryl iodide 5g to the corresponding benzophenone derivative (14, Figure 3) in 67% isolated yield. Unfortunately, this photo-cross-linker showed a loss of inhibitory activity with an IC₅₀ of 85 μ M. We then turned our attention to installing an aryl azide in region 3 as it is less sterically demanding and electronically similar to an aryl chloride. Aryl iodide 15, which was synthesized from 2aminonaphthoic acid in 4 steps as described in Scheme 1, was



Figure 3. Synthesis and IC_{50} of photo-cross-linkers. (A) Benzophenone cross-linker 14. (B) Azide cross-linker 16.

subjected to copper-mediated azidonation³⁶ providing 16 in good yield. Compound 16 demonstrated similar inhibitory activity to 2, which is important for ensuring specific site selection during photoactivation.

This SAR study has revealed three important features. First, the benzo [g] quinazoline of 1 and 2 can be replaced by the smaller quinazoline, provided that a small substituent is maintained at position 6 (e.g., 5c, Table 1). Second, a smaller, and yet still positively charged, amino side chain provides the best activity to date (e.g., 13l, Table 2). Finally, the 4chlorostyryl fragment remains necessary for inhibitory activity; replacement with several different isosteres obliterates inhibitory activity. Although the IC₅₀s of the best compounds in this series are still in the micromolar range, this activity is comparable to the best inhibitors of FtsZ reported so far. More dramatic changes to the core are difficult to predict or design without direct structural data. Installing a reactive functional group for cross-linking studies may determine the binding site of this unique inhibitor. Although preliminary competition studies suggest zantrin Z3 does not bind to the GTP pocket (see Supporting Information), further illumination will require successful cross-linking studies or crystallography of the inhibitor-FtsZ complex. Although our SAR studies did not shed much light on specific interactions between the analogues and FtsZ, we discovered a derivative that could be pursued in photo-cross-linking studies as a new avenue for probing this small molecule-protein interaction. Azide 16 is under investigation as a potential photo-cross-linker.

ASSOCIATED CONTENT

S Supporting Information

Experimental details and characterization data for all synthetic products and intermediates; FtsZ GTPase assay; GTP competition experiment. This material is available free of charge via the Internet at http://pubs.acs.org.

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Funding

This work was supported by the National Institutes of Heath (NIH/NIAID, R01AI08093, R01AI08093-04S1; NIH/NIGMS, T32-GM008799). G.M.N. and T.E.O. acknowledge support in the form of predoctoral fellowships from GAANN/DOEd and the Alfred P. Sloan Foundation. L.A.E.P. thanks CNPq (Conselho Nacional de Desenvolvimento Científico Tecnológico), Brazil. F.J.S. thanks LSAMP/CAMP. Z.Y. thanks the

Arnold and Mabel Beckman Foundation. This research was partially supported by an industry/campus supported fellowship to J.T.M. under the Training Program in Biomolecular Technology (T32-GM008799) at the University of California, Davis.

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Notes

The authors declare no competing financial interest.

ABBREVIATIONS

FtsZ, temperature sensitive filamenting protein Z; ZZ3, zantrin Z3; BsFtsZ, *Bacillus subtilis* FtsZ; EcFtsZ, *Escherichia coli* FtsZ; AcOH, acetic acid; *N*,*N*-DMEDA, *N*,*N*-dimethylethylaminediamine; DIPEA, diisopropylethylamine; dppf, 1,1'-bis-(diphenylphosphino)ferrocene; μ W, microwave; IC₅₀, half-maximal inhibition concentration; μ M, micromolar; *t*-Bu, *tert*-butyl; Ph, phenyl; Bn, benzyl; Boc, *tert*-butyl carbamate; TFA, trifluoroacetic acid; *n*-BuLi, *n*-butyllithium; DMPU, *N*,*N'*-dimethylpropyleneurea; Ac₂O, acetic anhydride; DMF, dimethylformamide; EtOH, ethanol; THF, tetrahydrofuran; *N*,*N'*-DMEDA, *N*,*N'*-dimethylethylaminediamine

REFERENCES

(1) Alanis, A. J. Resistance to antibiotics: Are we in the postantibiotic era? *Arch. Med. Res.* **2005**, *36*, 697–705.

(2) Antibiotic resistance threats in the United States, 2013. http:// www.cdc.gov/drugresistance/threat-report-2013/index.html (accessed November 18, 2014).

(3) Emmerson, A. M.; Jones, A. M. The quinolones: decades of development and use. J. Antimicrob. Chemother. 2003, 51, 13–20.

(4) Appelbaum, P. C.; Hunter, P. A. The fluoroquinolone antibacterials: past, present and future perspectives. *Int. J. Antimicrob. Agents* **2000**, *16*, 5–15.

(5) Sass, P.; Brötz-Oesterhelt, H. Bacterial cell division as a target for new antibiotics. *Curr. Opin. Microbiol.* **2013**, *16*, 522–530.

(6) den Blaauwen, T.; Andreu, J. M.; Monasterio, O. Bacterial cell division proteins as antibiotic targets. *Bioorg. Chem.* 2014, 55, 27–38.
(7) Lutkenhaus, J.; Pichoff, S.; Du, S. Bacterial cytokinesis: From Z

ring to divisome. *Cytoskeleton* **2012**, *69*, 778–790. (8) Matesanz, R.; Trigili, C.; Rodríguez-Salarichs, J.; Zanardi, I.; Pera, B.; Nogales, A.; Fang, W.-S.; Jímenez-Barbero, J.; Canales, Á.;

Barasoain, I.; Ojima, I.; Díaz, J. F. Taxanes with high potency inducing tubulin assembly overcome tumoural cell resistances. *Bioorg. Med. Chem.* **2014**, *22*, 5078–5090.

(9) Kingston, D. G. I.; Snyder, J. P. The quest for a simple bioactive analog of paclitaxel as a potential anticancer agent. *Acc. Chem. Res.* **2014**, *47*, 2682–2691.

(10) Pagano, A.; Honoré, S.; Mohan, R.; Berges, R.; Akhmanova, A.; Braguer, D. Epothilone B inhibits migration of glioblastoma cells by inducing microtubule catastrophes and affecting EB1 accumulation at microtubule plus ends. *Biochem. Pharmacol.* **2012**, *84*, 432–443.

(11) Yang, Z.; Wu, W.; Wang, J.; Liu, L.; Li, L.; Yang, J.; Wang, G.; Cao, D.; Zhang, R.; Tang, M.; Wen, J.; Zhu, J.; Xiang, W.; Wang, F.; Ma, L.; Xiang, M.; You, J.; Chen, L. Synthesis and biological evaluation of novel millepachine derivatives as a new class of tubulin polymerization inhibitors. *J. Med. Chem.* **2014**, *57*, 7977–7989.

(12) Ravelli, R. B. G.; Gigant, B.; Curmi, P. A.; Jourdain, I.; Lachkar, S.; Sobel, A.; Knossow, M. Insight into tubulin regulation from a complex with colchicine and a stathmin-like domain. *Nature* **2004**, 428, 198–202.

(13) Gigant, B.; Wang, C.; Ravelli, R. B. G.; Roussi, F.; Steinmetz, M. O.; Curmi, P. A.; Sobel, A.; Knossow, M. Structural basis for the regulation of tubulin by vinblastine. *Nature* **2005**, *435*, 519–522.

(14) Ojima, I.; Kumar, K.; Awasthi, D.; Vineberg, J. G. Drug discovery targeting cell division proteins, microtubules and FtsZ. *Bioorg. Med. Chem.* **2014**, *22*, 5060–5077.

ACS Medicinal Chemistry Letters

(15) Kelley, C.; Zhang, Y.; Parhi, A.; Kaul, M.; Pilch, D. S.; LaVoie, E. J. 3-Phenyl substituted 6,7-dimethoxyisoquinoline derivatives as FtsZ-targeting antibacterial agents. *Bioorg. Med. Chem.* **2012**, *20*, 7012–7029.

(16) Ito, H.; Ura, A.; Oyamada, Y.; Tanitame, A.; Yoshida, H.; Yamada, S.; Wachi, M.; Yamagishi, J.-i A 4-aminofurazan derivative— A189—inhibits assembly of bacterial cell division protein FtsZ in vitro and in vivo. *Microbiol. Immunol.* **2006**, *50*, 759–764.

(17) Wang, J.; Galgoci, A.; Kodali, S.; Herath, K. B.; Jayasuriya, H.; Dorso, K.; Vicente, F.; González, A.; Cully, D.; Bramhill, D.; Singh, S. Discovery of a small molecule that inhibits cell division by blocking FtsZ, a novel therapeutic target of antibiotics. *J. Biol. Chem.* **2003**, *278*, 44424–44428.

(18) Läppchen, T.; Hartog, A. F.; Pinas, V. A.; Koomen, G.-J.; den Blaauwen, T. GTP Analogue Inhibits Polymerization and GTPase Activity of the Bacterial Protein FtsZ without Affecting Its Eukaryotic Homologue Tubulin[†]. *Biochemistry* **2005**, *44*, 7879–7884.

(19) Anderson, D. E.; Kim, M. B.; Moore, J. T.; O'Brien, T. E.; Sorto, N. A.; Grove, C. I.; Lackner, L. L.; Ames, J. B.; Shaw, J. T. Comparison of small molecule inhibitors of the bacterial cell division protein FtsZ and identification of a reliable cross-species inhibitor. *ACS Chem. Biol.* **2012**, *7*, 1918–1928.

(20) Foss, M. H.; Eun, Y.-J.; Grove, C. I.; Pauw, D. A.; Sorto, N. A.; Rensvold, J. W.; Pagliarini, D. J.; Shaw, J. T.; Weibel, D. B. Inhibitors of bacterial tubulin target bacterial membranes in vivo. *MedChem*-*Comm* **2013**, *4*, 112–119.

(21) Haydon, D. J.; Stokes, N. R.; Ure, R.; Galbraith, G.; Bennett, J. M.; Brown, D. R.; Baker, P. J.; Barynin, V. V.; Rice, D. W.; Sedelnikova, S. E.; Heal, J. R.; Sheridan, J. M.; Aiwale, S. T.; Chauhan, P. K.; Srivastava, A.; Taneja, A.; Collins, I.; Errington, J.; Czaplewski, L. G. An Inhibitor of FtsZ with Potent and Selective Anti-Staphylococcal Activity. *Science* **2008**, *321*, 1673–1675.

(22) Andreu, J. M.; Schaffner-Barbero, C.; Huecas, S.; Alonso, D.; Lopez-Rodriguez, M. L.; Ruiz-Avila, L. B.; Núñez-Ramírez, R.; Llorca, O.; Martín-Galiano, A. J. The antibacterial cell division inhibitor PC190723 is an FtsZ polymer-stabilizing agent that induces filament assembly and condensation. J. Biol. Chem. 2010, 285, 14239–14246.

(23) Elsen, N. L.; Lu, J.; Parthasarathy, G.; Reid, J. C.; Sharma, S.; Soisson, S. M.; Lumb, K. J. Mechanism of action of the cell-division inhibitor PC190723: modulation of FtsZ assembly cooperativity. *J. Am. Chem. Soc.* **2012**, *134*, 12342–12345.

(24) Margalit, D. N.; Romberg, L.; Mets, R. B.; Hebert, A. M.; Mitchison, T. J.; Kirschner, M. W.; RayChaudhuri, D. Targeting cell division: Small-molecule inhibitors of FtsZ GTPase perturb cytokinetic ring assembly and induce bacterial lethality. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 11821–11826.

(25) Hopkins, A. L.; Keseru, G. M.; Leeson, P. D.; Rees, D. C.; Reynolds, C. H. The role of ligand efficiency metrics in drug discovery. *Nat. Rev. Drug Discovery* **2014**, *13*, 105–121.

(26) Ingerman, E.; Nunnari, J. A Continuous, Regenerative Coupled GTPase Assay for Dynamin-Related Proteins. In *Methods in Enzymology*; William, E., Balch, C. J. D., Alan, H., Eds.; Academic Press: New York, 2005; Vol. 404, pp 611–619.

(27) Moseley, J. D.; Murray, P. M.; Turp, E. R.; Tyler, S. N. G.; Burn, R. T. A mild robust generic protocol for the Suzuki reaction using an air stable catalyst. *Tetrahedron* **2012**, *68*, 6010–6017.

(28) Okano, M.; Mito, J.; Maruyama, Y.; Masuda, H.; Niwa, T.; Nakagawa, S.-i.; Nakamura, Y.; Matsuura, A. Discovery and structure– activity relationships of 4-aminoquinazoline derivatives, a novel class of opioid receptor like-1 (ORL1) antagonists. *Bioorg. Med. Chem.* **2009**, *17*, 119–132.

(29) William, A. D.; Lee, A. C. H.; Goh, K. C.; Blanchard, S.; Poulsen, A.; Teo, E. L.; Nagaraj, H.; Lee, C. P.; Wang, H.; Williams, M.; Sun, E. T.; Hu, C.; Jayaraman, R.; Pasha, M. K.; Ethirajulu, K.; Wood, J. M.; Dymock, B. W. Discovery of kinase spectrum selective macrocycle (16E)-14-methyl-20-oxa-5,7,14,26-tetraazatetracyclo-[19.3.1.1(2,6).1(8,12)]heptacosa-1(25),2(26),3,5,8(27),9,11,16,21,23decaene (SB1317/TG02), a potent inhibitor of cyclin dependent kinases (CDKs), Janus kinase 2 (JAK2), and fms-like tyrosine kinase-3 (FLT3) for the treatment of cancer. *J. Med. Chem.* **2012**, 55, 169–196. (30) Xu, H.; Tian, H.; Zheng, L.; Liu, Q.; Wang, L.; Zhang, S. Synthesis of chiral benzimidazoles as acylating agents for kinetic resolution of racemic α -amino esters. *J. Heterocycl. Chem.* **2012**, 49, 1108–1113.

(31) Chalmers, K. H.; De Luca, E.; Hogg, N. H. M.; Kenwright, A. M.; Kuprov, I.; Parker, D.; Botta, M.; Wilson, J. I.; Blamire, A. M. Design principles and theory of paramagnetic fluorine-labelled lanthanide complexes as probes for 19F magnetic resonance: a proof-of-concept study. *Chem—Eur. J.* **2010**, *16*, 134–148.

(32) Gege, C.; Schneider, M.; Chevrier, C.; Deng, H.; Sucholeiki, I.; Gallagher, B. M., Jr.; Bosies, M.; Steeneck, C.; Wu, X.; Hochguertel, M.; Nolte, B.; Taveras, A. *Preparation of Heterobicyclic Metalloprotease Inhibitors*. WO2008063668A1, 2008.

(33) Best, M. D.; Brik, A.; Chapman, E.; Lee, L. V.; Cheng, W.-C.; Wong, C.-H. Rapid discovery of potent sulfotransferase inhibitors by diversity-oriented reaction in microplates followed by in situ screening. *ChemBioChem* **2004**, *5*, 811–819.

(34) Milkiewicz, K. L.; Aimone, L. D.; Albom, M. S.; Angeles, T. S.; Chang, H.; Grobelny, J. V.; Husten, J.; LoSardo, C.; Miknyoczki, S.; Murthy, S.; Rolon-Steele, D.; Underiner, T. L.; Weinberg, L. R.; Worrell, C. S.; Zeigler, K. S.; Dorsey, B. D. Improvement in oral bioavailability of 2,4-diaminopyrimidine c-Met inhibitors by incorporation of a 3-amidobenzazepin-2-one group. *Bioorg. Med. Chem.* **2011**, *19*, 6274–6284.

(35) Jager, W. F.; Hammink, T. S.; van den Berg, O.; Grozema, F. C. Highly sensitive water-soluble fluorescent pH sensors based on the 7-amino-1-methylquinolinium chromophore. *J. Org. Chem.* **2010**, *75*, 2169–2178.

(36) Andersen, J.; Madsen, U.; Björkling, F.; Liang, X. Rapid synthesis of aryl azides from aryl halides under mild conditions. *Synlett* **2005**, 2005, 2209–2213.