Kurzmitteilungen:

Enantiomerically Pure Aminoindolizines: Bicyclic Ergoline Analogues with Dopamine Autoreceptor Activity

Enantiomerenreine Aminoindolizine: Bizyklische Ergolin-Analoge mit Aktivität am Dopamin-Autorezeptor

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The development of drugs which stimulate selectively the prejunctional dopamine receptor has become a major challenge in recent years^{1,2)}. These autoreceptor agonists decrease synthesis and release of dopamine (DA). Thus, the symptoms of schizophrenia originated from dopamine hyperactivity in the mesolimbic system should be reduced. On the other hand, it has been shown that DA autoreceptor agonists fail to reveal classical DA antagonist side effects like catalepsy, when given to animals³⁾.

Since the highly sensitive DA autoreceptor is assumed to resemble the postsynaptic D-2 receptor^{4,5)}, we planned to develop novel autoreceptor agonists by specific structural modifications of known D-2 agonists. One



Pr: n-C₃H₇

Scheme 1

major characteristic of these compounds is an X-H moiety (X = O, N) connected to an aromatic ring, which can mimic a catechol hydroxyl group of the genuine transmitter dopamine^{4,5)}. Typical examples are (R)-apomorphine (1) and compounds with ergoline (2) or ergoline partial structure (3)^{6,7)}. An aromatic X-H substructure is also included in previous representatives of DA autoreceptor agonists (*e.g.* SND 919 (4)^{8,9})¹⁰⁾. In some cases, these groups cause a reduced bioavailability, due to a metabolic degradation or an inability to pass through the blood-brain barrier⁵⁾.

In this paper we report on the aminoindolizine regioisomers 5a and 6a, as bioisosters of DA active bicyclic ergoline analogues. One of them (5a) turned out to be the first strong and selective DA autoreceptor agonist, which is devoid of an aromatic NH or OH moiety.

Since DA receptor - ligand interactions are known to proceed stereoselectively^{4,5,9} pure enantiomers should be investigated.

The synthesis of both regioisomeric aminotetrahydroindolizines (**5a** and **6a**) was performed starting from L-asparagine, which was used likewise as an α -amino acid for the synthesis of the 7-aminoindolizine **5a**, or as a β -amino acid equivalent when the 6-amino regioisomer **6a** was approached.

For the synthesis of 5a-c N, N-dibenzylasparagine 7b, readily available from natural L-asparagine (7a), was first reduced with borane/THF to afford the diaminobutanol derivative 8. Then the pyrrole unit was introduced by a modified *Paal-Knorr* procedure to give the cyclization precursor 9. Subsequent trifluoromethanesulfonic anhydride assisted ring closure yielded 5b, which could be deprotected by catalytic



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hydrogenation (H₂, Pd(OH)₂) to afford the primary amine $5c^{11}$). The synthesis was accomplished by conversion of 5c into the dipropylamine 5a using an excess of propionic aldehyde and NaCNBH₃. Following the same reaction conditions the 6-dipropylaminoindolizine 6a was prepared from the primary amine 6b. 6b was synthesized by application of our β -amino acid methodology^{12,13}) via the azide 10 and the 1,3 diamino alcohol 11 as the key intermediates¹⁴).

Analogously, we obtained the (R)-configurated enantiomers 5d and 6c from D-asparagine (7c).



The aminotetrahydroindolizine derivatives 5a-d and 6a,c were chosen for receptor binding studies, when their ability to displace [³H]-SCH 23390, [³H]-spiroperidol and [³H]-SND 919 as specific radioligands labeling D-1, D-2, and DA-autoreceptor binding sites was investigated (Table 1). All compounds tested failed to reveal remarkable affinity to the postsynaptic D-1 and D-2 sites. However, evaluation of their autoreceptor binding showed modest to strong and highly selective affinity. The most potent derivative turned out to be 5a (IC₅₀ = 30 nmol) showing a 240 and 500-fold preference, compared to the D-1 and D-2 binding sites. The autoreceptor - ligand complexes are formed stereoselectively, indicated by a 24-fold higher IC₅₀ value of 5d. Discrimination between the enantiomers was also observed for the less active 6-amino derivatives 6a and 6c when, in contrast to 5a/5d, the eutomer (6c) has reversed configuration. These data indicate that the position of the pyrrole nitrogen, and so the contribution of the electron density at the aromatic region, is important for the receptor binding.

As expected, the IC_{50} values show that the dipropylamine is a valuable structural unit¹⁵⁾.

Table 1: Receptor Binding Data

IC ₅₀ [nM]			
compd	D-1*	D-2 b	DA-autoreceptor ^c
5a	7.100	15.000	30
5b	>100.000	>100.000	34.000
5c	33.000	>100.000	2.200
5d	33.000	22.000	730
ба	>100.000	>100.000	6.900
6c	>100.000	72.000	350

^a 3H-ligand: SCH 23390; ^b 3H-ligand: spiroperidol; ^c 3H-ligand: SND 919.

To evaluate whether the autoreceptor affinity of **5a** is connected with an intrinsic activity, the ability of **5a** to inhibit

 γ -butyrolactone (GBL) induced acceleration of DA synthesis in rat corpus striatum was examined. In fact, a 62% lower L-DOPA level was measured after injecting 10 mg/kg of 5a. This indicates DA autoreceptor agonistic effects.

Treatment of mice with 5a did not produce catalepsy (a typical extrapyramidal side effect of postsynaptic DA antagonists) nor stereotyped behavior (due to postsynaptic striatal DA receptor stimulation), up to 100 mg/kg. At 10 mg/kg reduced locomotor activity was observed. After injection of doses > 40 mg/kg of 5a excitation was shown when the mice were stimulated acoustically. Our CNS screening with mice¹⁶⁾ indicated that the primary amine 5c is also strongly CNS active, unless the affinity to the DA receptors was fairly low. The compound exhibited a strong sedative effect (extremely reduced locomotor activity, depressed pinna reflex) at doses between 5 and 50 mg/kg.. Additionally, Straub tail and mydriasis were observed. It is worthy to note, that 5c gave an ED₅₀ value of 1.8 mg/kg (1.0 - 3.3 mg/kg, p. > (0.05) in the writhing test¹⁶, indicating that 5c might have central analgesic properties.

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Experimental Part

General Remarks see ref. 13.

(S)-4-Amino-2-N,N-dibenzylaminobutanol(8)

To a solution of 7b¹³ (15.6 g, 50 mmol) in THF (400 ml) was added BH₃ x THF (300 ml of a 1 molar solution of THF) at 0°C. After 1 h at 0°C the mixture was refluxed for 4 h. Then the mixture was acidified to pH 1 with 6 N aqueous HCl and subsequently adjusted to pH 12 with 2 N aqueous NaOH. After addition of Et₂O the org. layer was dried (MgSO₄) and evaporated, and the residue was purified by flash chromatography (CHCl₃-MeOH-Et₃N 85:10:7) to give 12.2 g (86%) of pure 8 as a colorless oil; $[\alpha]_D^{23} + 32.5^{\circ}$ (c = 1.0, CHCl₃).- C₁₈H₂₄N₂O (284.4) Calcd. C 76.0 H 8.51 N 9.85 Found C 75.7 H 8.91 N 10.01.- IR (NaCl): 3280; 3240; 3030; 2930; 1600 cm⁻¹.- ¹H-NMR (CDCl₃): δ (ppm) = 1.41-1.50 (m, 1H, H-3). 1.89-1.97 (m, 1H, H-3), 2.62-2.82 (m, 3H, H-1 and H-4), 3.50 (d, J = 13.3 Hz, 2H, NCH₂Ph), 3.51 (dd, J = 10.5, 6.0 Hz, 1H, H-1), 3.61 (dd, J = 10.5, 8.0 Hz, 1H, H-1), 3.74 (d, J = 13.3 Hz, 2H, NCH₂Ph), 7.20-7.32 (m, 10 H, Ar). (R)-8($[\alpha]_D^{23} - 33^{\circ}$ (c = 1.0, CHCl₃) was prepared from (R)-7b¹³ follow-

(R)-8($[\alpha_{1D}]$ - 55° (C = 1.0, CHC₁₃) was prepared from (R)-70° rollow ing the same procedure.

(S)-2-N,N-Dibenzylamino-4-N-pyrrolylbutanol(9)

To a mixture of 8 (12.8 g, 45 mmol) and NaOAc x 3 H₂O (122 g, 900 mmol) in acetic acid (650 ml) was added dimethoxytetrahydrofuran (6.6 g, 50 mmol) at room temp. Then the temp. was slowly raised to 70°C and stirring was continued for 75 min. After the mixture was concentrated it was basified with 2 N aqueous NaOH and extracted with Et₂O. The org. layer was dried (MgSO₄) and evaporated and the residue was purified by flash chromatography (petroleum ether - EtOAc 4:1) to give 11.3 g (75%) of pure 9 as a colorless oil; $[\alpha]_D^{23}$ + 67° (c = 1.0, CHCl₃).- C₂₂H₂₆N₂O (334.5) Calcd. C 79.0 H 7.84 N 8.38 Found C 79.4 H 7.74 N 8.07.- IR (NaCl): 3600-3200; 3030; 2930; 1600 cm⁻¹.- ¹H-NMR (CDCl₃): δ (ppm) = 1.63-1.69 (m, 1H, H-3), 2.14-2.22 (m, 1H, H-3), 2.76-2.84 (m, 1H, H-2), 3.39 (d, J = 13.2 Hz, 2H, NCH₂Ph), 3.42 (dd, J = 10.8, 5.2 Hz, 1H, H-1),

3.46 (dd, J = 10.8. 10.5 Hz, 1H, H-1), 3.67 (d, J = 13.2 Hz, 2H, NCH₂Ph), 3.80-3.93 (m, 2H, H-4), 6.15-6.16 (m, 2H, NCHC<u>H</u>), 6.58-6.59 (m, 2H, NCH), 7.17-7.32 (m, 10 H, Ar).

(R)-9 ($[\alpha]_D^{23}$ - 68° (c = 1.0, CHCl₃) was prepared from (R)-8 following the same procedure.

(S)-7-N,N-Dibenzylamino-5,6,7,8-tetrahydroindolizine(5b)

To a mixture of 9 (10 g, 30 mmol) in CH_2Cl_2 (400 ml) was added trifluoromethanesulfonic anhydride (17.4 g, 61.7 mmol) at 0°C. After stirring for 16 h at room temp. saturated aqueous NaHCO3 and Et2O were added. The org. layer was dried (MgSO₄) and evaporated and the residue was purified by flash chromatography (petroleum ether - Et₂O 95:5) to give 8.9 g (94%) of pure 5b as a colorless solid; mp. 95-97°C; $[\alpha]_D^{23}$ - 50° (c = 1.0, CHCl₃).- C₂₂H₂₄N (316.4) Calcd. C 83.5 H 7.65 N 8.86 Found C 83.3 H 7.61 N 8.87.- IR (NaCl): 3030; 2960; 1600 cm⁻¹.- ¹H-NMR (CDCl₃): δ $(ppm) \approx 2.00 \, (ddd, J = 12.5, 12.5, 5.4 \, Hz, 1H, H-6), 2.13-2.18 \, (m, 1H, 1H, 1H)$ H-6), 2.85 (dd, J = 15.5, 11.8 Hz, 1H, H-8), 3.00-3.12 (m, 2H, H-7 and H-8), 3.66 (d, J = 14.0 Hz, 2H, NCH₂Ph), 3.73 (d, J = 14.0 Hz, 2H, NCH₂Ph), 3.77 (ddd, J = 12.5, 12.5, 4.4 Hz, 1H, H-5), 4.10 (ddd, J = 12.5, 12.5, 4.4 Hz, 1H, H-5), 4.10 (ddd, J = 12.5, 125.4, 2.2 Hz, 1H, H-5), 5.82-5.83 (m, 1H, H-1), 6.08-6.09 (m, 1H, H-2), 6.45-6.46 (m, 1H, H-3), 7.20 (t, J = 7.3 Hz, 2H, p-Ar), 7.28 (t, J = 7.3 Hz, 4H, m-Ar), 7.37 (d, J = 7.3 Hz, 4H, o-Ar).- 13 C-NMR (CDCl₃): δ (ppm) = 25.50 (H2C-8), 26.08 (H2C-6), 45.17 (H2C-5), 53.51 (HC-7), 53.89 (NCH₂Ph), 104.66 (HC-1), 108.24 (HC-2), 118.30 (HC-3), 126.88 (HC-Ar), 128.27 (HC-Ar), 128.58 (HC-Ar), 128.85 (C-8a); 140.14 (C-Ar).

(R)-5b ($[\alpha]_D^{23}$ + 51° (c = 1.0, CHCl₃) was prepared from (R)-9 following the same procedure.

(S)-7-Amino-5,6,7,8-tetrahydroindolizine(5c)

A mixture of **5b** (0.95 g, 3 mmol) and 20% Pd(OH)₂/C (0.70 g) in EtOAc (35 ml) and MeOH (35 ml) was stirred under a balloon of H₂ for 5 h at room temp.. The mixture was filtered through celite, the filtrate was evaporated carefully and the residue was purified by flash chromatography (CH₂Cl₂-MeOH 4:1) to give 0.35 g (85%) of pure **5c** as a colorless solid; mp. 150°C (dec.); $[\alpha]_D^{23} - 64^\circ$ (c = 0.5, MeOH).- $C_8H_{12}N_2$ (136.2) Calcd. C 70.7 H 8.88 N 20.57 Found C 70.5 H 9.12 N 20.36.- IR (NaCl): 3400-2800; 2930 cm⁻¹.- ¹H-NMR (CDCl₃): δ (ppm) = 1.83-1.92 (m, 1H, H-6), 2.10-2.17 (m, 1H, H-6), 2.56 (dd, J = 15.7, 8.8 Hz, 1H, H-8), 3.08 (dd, J = 15.7, 4.4 Hz, 1H, H-8), 3.28-3.31 (m, 1H, H-7), 3.89-3.97 (m, 1H, H-5), 4.05-4.11 (m, 1H, H-5), 5.85-5.86 (m, 1H, H-1), 6.12-6.14 (m, 1H, H-2), 6.53-6.54 (m, 1H, H-3).

(R)-5c ($[\alpha]_D^{23}$ + 64.5° (c = 0.5, MeOH) was prepared from (R)-5b following the same procedure.

(S)-7-N,N-Dipropylamino-5,6,7,8-tetrahydroindolizine(5a)

To a solution of **5c** (136 mg, 1 mmol) in MeOH (15ml) was added propionic aldehyde (581 mg, 10 mmol) and then NaCNBH₃ (126 mg, 2 mmol) at 0°C. After the mixture was stirred for 20 h at room temp. it was acidified to pH 1 with 2 N aqueous HCl and subsequently basified with saturated aqueous NaHCO₃. After addition of Et₂O the org. layer was dried (MgSO₄) and evaporated, and the residue was purified by flash chromatography (CH₂Cl₂ - MeOH 97:3) to give 158 mg (72%) of pure **5a** as a colorless oil; $[\alpha]_D^{23} - 35^\circ$ (c = 0.5, CHCl₃).- C₁₄H₂₄N₂ (220.4) Calcd. C 76.3 H 10.98 N 12.71 Found C 76.5 H 10.97 N 12.52.- IR (NaCl): 2960 cm⁻¹.- ¹H-NMR (CDCl₃): δ (ppm) = 0.88 (t, J = 7.3 Hz, 6H, NCH₂CH₂CH₃), 1.47 (sext., J = 7.3 Hz, 4H, NCH₂CH₂), 1.79-1.95 (m, 1H, H-6), 2.05-2.10 (m, 1H, H-6), 2.45-2.50 (m, 4H, NCH₂), 2.65 (dd, J = 15.4, 11.0 Hz, 1H, H-8), 2.94-3.07 (m, 2H, H-7 and H-8), 3.88 (ddd, J = 11.7, 11.7, 4.4 Hz, 1H, H-5), 4.11 (ddd, J = 11.7, 5.2, 2.0 Hz, 1H, H-5), 5.82-5.83 (m, 1H, H-1), 6.11-6.13 (m, 1H, H-2), 6.49-6.50 (m, 1H, H-3).

5d $([\alpha]_D^{23} + 34.5^\circ (c = 0.4, CHCl_3)$ was prepared from (R)-5c following the same procedure.

(S)-6-N,N-Dipropylamino-5,6,7,8-tetrahydroindolizine(6a)

6b¹⁴⁾ (136 mg, 1 mmol) in MeOH (15 ml) as well as propionic aldehyde (581 mg, 10 mmol) and NaCNBH₃ (126 mg, 2 mmol) were reacted and worked up as described above for 5a to give 150 mg (68%) of pure 6a as a colorless oil; $[\alpha]_D^{23}$ - 32° (c = 0.9, CHCl₃).- C₁₄H₂₄N₂ (220.4) Calcd. C 76.3 H 10.98 N 12.71 Found C 76.7 H 11.02 N 12.50.- IR (NaCl): 2960 cm⁻¹.- ¹H-NMR (CDCl₃): δ (ppm) = 0.88 (t, J = 7.3 Hz, 6H, NCH₂CH₂CH₃), 1.43-1.46 (m, 4H, NCH₂CH₂), 1.60-1.69 (m, 1H, H-7), 2.00-2.05 (m, 1H, H-7), 2.44-2.49 (m, 4H, NCH₂), 2.66 (ddd, J = 16.1, 16.1, 5.0 Hz, 1H, H-8), 2.95 (ddd, J = 16.1, 5.2, 3.0 Hz, 1H, H-8), 3.09-3.16 (m, 1H, H-6), 3.72 (dd, J = 11.5, 11.5 Hz, 1H, H-5), 4.23 (dd, J = 11.5, 5.9 Hz, 1H, H-5), 5.80-5.81 (m, 1H, H-1), 6.12-6.13 (m, 1H, H-2), 6.49-6.50 (m, 1H, H-3).

6c ($[\alpha]_D^{23}$ + 31.5° (c = 0.9, CHCl₃) was prepared from (R)-6b¹⁴) following the same procedure.

Receptor Binding Assay

DA receptor binding was performed as described^{17,18} using [³H]-SCH 23390 and [³H]-spiroperidol as radioligands in concentrations of 0.3 nM and 0.5 nM, respectively. In the receptor binding assay for the characterization of the DA autoreceptor, [³H]-SND 919 (51 Ci/mmol specific activity) was used in a concentration of 0.5 nM. The experimental procedure was performed in analogy to the binding assay with [³H]-spiroperidol as radioligand. For all receptor binding tests rat brain striatum was used.

Determination of DA Synthesis Rate

For the determination of the DA synthesis rate the inhibition of the GBL-induced L-DOPA accumulation was measured as described $^{19,20)}$.

In vivo Screening with Mice

The behavioural activity screening with mice was carried out according to ref. 16.

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