Synthetic Approaches To Novel Cis and Trans Dideoxynucleosides of the Apiose Family

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Abstract: Stereoselective synthesis of the complete family of optically active dideoxygenated nucleosides of the apiose family have been developed. The chiral aldodiol system 7, a key intermediate in this synthesis, was prepared from the prochiral molecule 6, through the action of the lipase from *Candida cylindracia*. Approaches to novel enantiomeric and diastereoisomeric dideoxynucleosides containing the tetrahydrofuranethanol moiety have also been discovered. A key intermediate in this approach was the optically active trans-allyllactone 61, prepared from L-glutamic acid, and its isomerization product, the corresponding cis-allylbutyrolactone 62. The methodologies developed have generality and allow synthetic access to a wide variety of new nucleosides.

INTRODUCTION

9-(B-D-Apio-D-furanosyl)adenine (1), a biologically-active, relatively non-cytotoxic nucleoside related to natural D-apiose,¹⁻³ is a regioisomer of adenosine through transposition of the C-4' hydroxymethyl to C-3'. Compounds belonging to the enantiomeric series represented by 2(3'S,5'R) and 3(3'R,5'S) are dideoxygenated analogs of D-apio-D-adenosine 1. They may also be conceptualized as dideoxynucleosides with transposed hydroxymethyl groups or transposed oxygen atoms. A few dideoxynucleosides derived from the natural nucleosides have been found to be potently active against HIV through inhibition of HIV-encoded reverse transcriptase (RT).⁴ The design and evaluation of additional novel nucleoside-based RT inhibitors are needed in order to develop analogs that exhibit increasingly more favorable toxicity profiles and are less susceptible to the development of resistant strains of HIV.⁵ One essential feature in the design of these inhibitors is retention of the 2',3'-dideoxygenation which is necessary for termination of the viral DNA chain elongation.⁶ Other modifications have included strategic substitution on the carbohydrate moiety, removal of the endocyclic heteroatom or introduction of an additional heteroatom.⁷⁻¹² An interesting new family of dideoxynucleosides involves transposition of the 5'-CH,OH or transposition of the endocyclic oxygen.¹³⁻¹⁷ These compounds are regioisomeric with respect to dideoxy analogs of the natural nucleosides and two representative classes are depicted by structures 2 and 3. We wish to report on approaches to the stereoselective synthesis of the complete family of optically active 2',3'-dideoxygenated nucleosides represented by 2 and 3. This study is supported by the observation¹⁶ that one member of Class 3 has been reported to have anti-HIV activity in MT-4 cells with no apparent toxicity. Development of additional methodologies in this area has also allowed access to novel chiral isomeric dideoxynucleosides containing a furanethanol moiety and this is described.



RESULTS AND DISCUSSION

Potential strategies for the synthesis of the carbohydrate moiety of this family of optically active regioisomeric dideoxynucleosides could utilize apiose or its deoxygenated derivatives as precursors. However, due to the multistep syntheses required to obtain these chiral precursors,^{18,19} alternative approaches were examined. Syntheses utilizing enzyme catalyzed reactions represent alternative approaches that have proven effective for the stereoselective synthesis of biologically active nucleosides.²⁰ Additionally, a divergent synthesis from a key chiral precursor would allow for an efficient approach to each enantiomeric series of these dideoxynucleosides. Such a divergent point would arise from the cyclization of a chiral derivative of the prochiral aldodiol 4 (Figure 2). Lactol formation with the *pro-S* hydroxymethyl group would allow for the formation of the dideoxygenated D-apiose [tetrahydro-3(S)-furanmethanol] nucleoside of class 3.



2-Allyl-1.3-propanediol was chosen as the starting material due to its potential ease of conversion into 4. Chiral derivatives of the aldodiol system 4 are accessible through the resolution of diastereometric urethanes,²¹ or through stereoselective enzymatic hydrolysis of a suitable derivative.^{22,23} The high stereoselectivity and relatively low cost of the latter method warranted its application in this synthetic work. 2-Allyl-1,3-propanediol was readily prepared by the reduction of diethyl allylmalonate 5. Acetylation afforded 2-(2propenyl)-1,3-propanediol diacetate (6) in an 80% overall yield (Scheme 1). Stereoselective ester hydrolysis with the lipase from candida cylindracea (Sigma, Type VII) in 30% aqueous acetone afforded the S-(-)monoacetate 7 ($[\alpha]_{D} = -7.95^{\circ}$, CHCl₂) in a 50% yield and in high optical purity (98.7% ee). Consistently high optical purity was observed with reaction times of 7 - 10 hours at carefully maintained pH values between 6.95 - 7.00. More extended reaction times (>12 hrs) or higher pH resulted in a decrease in optical purity. Silylation of the S-(-)-monoacetate 7 produced the R-(+)-8 which was chemically hydrolyzed to give, in near quantitative yields for two steps, the R-(+)-monosilyl ether 9. For a key transformation, the attempted oxidative cleavage of the olefin 9 to form the tetrahydro-3(R)-furanmethanol 10, the use of ruthenium dioxide (Aldrich) with sodium periodate²⁴ resulted in low and non-reproducible yields of the desired product. Contrary to previous reports,²³ no evidence for the formation of the corresponding lactone was observed. Ozonolysis (-78°C, dimethyl sulfide) of 9 and attempted acetylation did however produce the corresponding lactone of 10, presumably via an in situ dimethyl sulfoxide - acetic anhydride oxidation²⁵ of



Scheme 1

the ozonolysis product. DIBAL reduction of the lactone afforded 10 in 54% overall yield from 9. However, the most successful method employed for the conversion of 9 to 10 was the use of sodium periodate and catalytic osmium tetroxide (2.5 - 5.0 mole %),²⁶ which consistently afforded near quantitative yields of the desired 10. Acetylation gave the diastereomeric mixture of protected tetrahydro-5(R,S)-acetyloxy-3(R)furanmethanol 11 (5R:S = 2:1) in a combined yield of 78%.

Glycosylation of 11 using stochiometric amounts of trimethylsilyl triflate (TMSOTf) ²⁷ with silylated N⁶benzoyladenine [generated in situ with *bis*(trimethylsilyl)acetamide (BSA) in refluxing acetonitrile], afforded a 1:2 diastereometric mixture of the adenine dideoxynucleosides 12 and 13 (Scheme 2). The use of Lewis acids such as stannic chloride were precluded, due to the reported low diastereometric ratio of the desired cis isomer (cis:trans ratio = 1:4) in those glycosylations.¹⁶ Differentiation between the cis and trans isomers was readily discerned from the different ¹H NMR chemical shifts of their tetrahydrofuran H-2' protons: δ 4.20 and 3.83 for the major isomer, 13, and 4.08 and 3.98 for the minor isomer, 12. These assignments were confirmed by extensive NOE experiments. Separation of the diastereometrs by preparative layer chromatography with multiple immersions afforded pure 5'(R)-3'(R)-12 (15%) and 5'(S)-3'(R)-13 (27%). Deprotection [(i) NH₃, (ii) Et₄NF] of each isomer afforded 5(R)-(6-amino-9H-purin-9-yl)tetrahydro-3(S)furanmethanol (14) ([α]_D = -22.6°, MeOH) and the trans compound [5'(S)-3'(S)], 15 ([α]_D = +39.8°, MeOH).



Scheme 2

The anomeric stereochemical assignments for 14 and 15 were determined, in part, by ¹H NMR difference NOE data. Upon irradiation of the C-6' methylene protons a NOE of 3% was observed for both of the cis H-2' and H-4' protons of 14. Irradiation of these protons resulted in NOEs of 2% and 8%, respectively, on the adenine base H-8. Similar results with 15 show, that upon irradiation of the C-6' methylene protons, NOEs of 2% and 3% on the cis H-2' and H-4' protons were observed. Irradiation of this H-4' proton resulted in a NOE of 6% on the anomeric H-5' proton. Additional support for the anomeric assignments is the levorotatory nature of 14, which is consistent with that observed for 9-(D-apio-D-furanosyl)adenine,¹ and also for adenosine, and 2',3'-dideoxyadenosine.

Glycosylation of 11 with silylated N²-acetyl-O⁶-diphenylcarbamoylguanine²⁸ (Scheme 3) afforded a mixture of the cis-5'(R)-3'(R)-16 and trans-5'(S)-3'(R)-17 (1:2) in 21% yield. Separation of the diastereomers by preparative layer chromatography and deprotection of the individual anomers afforded 5(R)-(2-amino-1,9-dihydro-6H-purin-6-one)tetrahydro-3(S)-furanmethanol (18) and the trans-5'(S)-3'(S)-19 in 76% and 86% yields, respectively. Similarly, under the standard glycosylation conditions (TMSOTf, 0°C - 25°C, CH₃CN), silylated uracil and 11 were coupled to form the separable compounds, 20 and 21. Desilylation of the individual anomers afforded the regioisomeric dideoxyuridine nucleosides 22 and 23 in respective yields of 23% and 52%.





Additionally, 11 was coupled with silvlated cytosine and thymine (Scheme 4) to form the diastereoisomers 24, 25 and 28, 29, respectively. In each case, the lack of significant difference in R_f values between the diastereomers prevented an efficient separation. Desilvlation of the diastereoisomeric pairs also resulted in an inefficiently separable mixture of 26, 27 and 30, 31. This problem could not be surmounted through the formation of chiral esters (e.g. camphanyl or N-benzoyl-L-phenylalaninyl) as these failed to significantly enhance separation of the diastereoisomers. Separation of 26, 27, 30, and 31 could be achieved, albeit tediously, utilizing preparative layer chromatography (multiple immersion technique) and careful dissection of the diffused bands. The resulting fractions were analyzed (¹H NMR), appropriately combined, and rechromatographed. Repeating this process provided pure products.



Scheme 4

In the case of the isodideoxycytidines 26 and 27, the intricate chromatographic process was avoided through preparation of these compounds from the corresponding uridine analogs 20 and 21 in 61% and 65% yields utilizing the well established sequential phosphorous oxychloride (POCl₃)/1,2,4-triazole/ammonia methodology.²⁹ However, attempts to prepare the thymidine nucleosides 30 and 31 from the uridine nucleosides 22 and 23 using known chemical methodologies³⁰ were unsuccessful.

Entry into the enantiomeric tetrahydro-3(R)-furanmethanol class of regioisomeric dideoxynucleosides (i.e. of Class 3) was achieved through the key chiral intermediate 7 (Scheme 5). Treatment of 7 with OsO_4 and $NaIO_4$ readily afforded the lactol 32 (82%). Acetylation produced 33 in a 78% yield. Glycosylation, deprotection, separation and identification were carried out in a related manner to those of the enantiomeric



S-series to give the isomeric dideoxynucleosides 34-57.



These studies were extended to the novel homologous series represented by 58 and 59 (Fig. 3).



Approaches to the synthesis of these adenine tetrahydro-3-furanethanol nucleosides utilized the known enantiospecific formation of γ -butyrolactones from L-glutamic acid^{31,32} and the potentially high stereoselectivity of alkylations of these compounds.³³⁻³⁵ Thus, the silylated hydroxymethyl- γ -butyrolactone **60** served as the starting material for the synthesis. Treatment of **60** with LDA (1 equiv.) at -98 °C and subsequent reaction with allyl iodide afforded the separable diastereomeric allylbutyrolactones **61** and its cisisomer 62 (95:5) in a total yield of 95% (Scheme 6). The diastereoisomeric lactones 61 and 62 could be identified and differentiated by high-field ¹H NMR chemical shift data and extensive differential NOE experiments. Reduction of 61 followed by desilylation afforded the 4(R)-hydroxymethyl-6-hepten-1,2(S)-diol (63) in 90% yield ($[\alpha]_D = -19^\circ$, MeOH). The order of the reactions in this conversion of 61 to 63 is critical in order to prevent epimerization of the allyl group. Oxidative cleavage of the glycol in 63 (NaIO₄) was followed by spontaneous intramolecular hemiacetal formation of the resulting aldehyde to produce the tetrahydro-5-hydroxy-3(R)-allylfuran 64 (88%, $[\alpha]_D = +46^\circ$, MeOH). Protection of the anomeric hydroxyl group was then required for the successful conversion to the furanethanol system and to exclude the potential formation of the corresponding 4-hydroxymethylpyranoside. Thus, ozonolysis of the methyl furanoside followed by reduction of the resulting aldehyde gave the tetrahydro-3(R)-furanethanol 65 (75%, $[\alpha]_D = +29^\circ$, MeOH). Benzoylation, demethylation, and acetylation provided the glycosylation precursor 68 (56%, $[\alpha]_D =$ +10°, MeOH). Condensation of 68 in the presence of TMS triflate with silylated N⁶-benzoyladenine (35 % vield), followed by debenzoylation and separation of the anomers by fractional crystallization and by



Scheme 6

preparative layer chromatography, produced the pure *cis*-compound, 5(R)-(6-amino-9H-purin-9-yl)-3(R)furanethanol (69, 32%), and its *trans*-isomer [5'(S)-3'(R), 70, 56%].

The enantiomeric 3(S)-furanethanol adenine dideoxynucleosides were obtained from the β allylbutyrolactone **62** (Scheme 6). Treatment of the 95:5 mixture of allylbutyrolactones **61** (α) and **62** (β) with LDA, followed by quenching of the resulting enolate with water, resulted in isomerization to produce predominantly the β -allyl compound **62** (81%, **62:61** ratio = 84:16). Separation of the desired diastereoisomer **62** and subjecting it to the sequence of reactions previously described, converted **62** through the tetrahydro-3(S)-allylfuranoside **71** (71%, $[\alpha]_D = -44^\circ$, MeOH), to the 5- α , β mixture of acetylated tetrahydro-3(S)-furanethanols **74** ($[\alpha]_D = -10^\circ$, MeOH) in 31% yield. Glycosylation of the latter with N⁶benzoyladenine, deprotection and separation of the resulting diastereoisomers produced the *cis*-5(S)-(6-amino-9H-purin-9-yl)tetrahydro-3(S)-furanethanol, **75** (9%, $[\alpha]_D = +54.3^\circ$, MeOH) and its diastereoisomeric *trans*-5'(R),3'(S)-isomer, **76** (23%, $[\alpha]_D = -22.3^\circ$, MeOH). Finally, it should be mentioned that these approaches can be used to gain entry into the entire series of enantiomeric dideoxynucleosides of the tetrahydrofuranethanol family involving both purine and pyrimidine bases.

As hydrolysis (chemical and enzymatic) plays a major role in the catabolism of some potential anti-HIV nucleosides, the resistance toward enzymatic deamination and acid catalyzed glycosidic bond hydrolysis was studied. Marked resistance toward deamination by mammalian adenosine deaminase was seen for the ciscompound, 14, and its enantiomer 38 (0.12% and 0.10%, respectively, of the rate of adenosine). Their transisomers, 15 and 39, were even more resistant to deamination (0.015 % and 0.013 %, respectively, of adenosine). The corresponding tetrahydrofuranethanol compounds 69 and 75 were almost totally resistant to deamination by adenosine deaminase (< 0.005 % of the rate of adenosine). Studies of the relative rates of glycosidic bond hydrolysis ³⁶ show that the cis-enantiomers 14 and 38 were slightly more stable (84%) than 2',3'-dideoxyadenosine (100%), while the trans-enantiomers 15 and 39 were significantly more stable (50%) at pH 3. Compounds 69 and 75 exhibited similar behavior.

In summary, approaches to enantiomeric pairs of isomeric dideoxynucleosides related to D- and L-apiose have also been developed. The key precursor for the construction of the dideoxyapiose ring was a chiral aldodiol system synthesized from a pro-chiral molecule through the action of the lipase from *Candida cylindracia*. Routes to novel enantiomeric dideoxynucleosides containing the tetrahydrofuranethanol moiety have also been discovered. Key steps in this approach, among others, were the preparation of two important intermediates, a trans-allylbutyrolactone and its cis-isomer which was accessible from the ready isomerization of the trans-compound. The methodologies developed have generality and allow access to a

wide variety of new nucleosides.

Experimental Section

Melting points reported are uncorrected and were determined on a Thomas-Hoover melting point apparatus fitted with a microscope. Nuclear magnetic resonance spectra were recorded on Bruker Models AMX-600, MSL-300, and AC-300 pulse Fourier transform spectrometers. Mass spectra were determined on a VG ZAB-HF high resolution mass spectrometer with FAB capability or a VG TRIO single quadrupole GC/MS system. Ultraviolet spectra were recorded on a Mattson Cygnus 25 Fourier transform instrument. Lyophilizations were performed with a Virtis freezemobile 3 unit. Preparative layer chromatography was carried out on plates prepared with E. Merck PF₂₅₄ silica gel. Flash chromatography was carried out using glass columns packed with 230-400 mesh silica gel. High performance liquid chromatography was done at 80 psi using Altex columns packed with 40-60 m Amberlite XAD-4 resin (Rohm and Haas). Fractions were monitored by a Pharmacia UV-2 ultraviolet monitor and products were also carried out with a Waters automated 600E system with photodiode array detector and FOXY fraction collector using Delta-Pak C₁₈ and Hamilton PRP-1 columns. Elemental analyses were performed at Galbraith Laboratories, Inc., Knoxville, TN., and at the University of Iowa.

Diacetyl 2-allyl-1,3-propanediol (6). Diethyl allylmalonate (20 mL, 101.4 mmol) in anhydrous Et₂O (100 mL) was added dropwise to a stirred solution of LiAlH₄ (9.62 g, 253.5 mmol) in Et₂O (400 mL) at 0 °C over 1 hr. The reaction was then allowed to warm to room temperature and was stirred an additional 4 h. Wet Et₂O was then slowly added, followed by water, and the mixture was filtered through Celite, dried (Na₂SO₄), concentrated, and purified by flash chromatography (hexanes to 50% EtOAc/hexanes) to afford 10.13 g (87.20 mmol, 86%) of 2-allyl-1,3-propanediol^{21, 1}H NMR (CDCL₁) δ 1.9 (m, 3H), 3.4 (m, 4H), 4.0 (br s, 2H), 4.90 (m, 2H), 5.60 (m, 1H); ¹³C NMR (CDCL₁) δ 32.1, 41.7, 64.0, 116.0, 135.9. To a solution of the diol (3.50 g, 30.15 mmol) in CH₃CN (150 mL) were added DMAP (0.29 g, 2.41 mmol), Et₃N (12.6 mL, 90.40 mmol), and Ac₂O (8.53 mL, 90.41 mmol) at 0 °C. The reaction was allowed to stir at room temperature for 4 h then MeOH (10 mL) was added. The mixture was then concentrated under reduced pressure, dissolved in EtOAc, and washed with water (3 x 30 mL). The organics were then dried (Na₂SO₄) and purified by flash chromatography (hexanes) to afford 5.49 g (27.44 mmol, 91%) of **6**²³: ¹H NMR (CDCL₁) δ 1.81 (s, 6H), 1.90 (m, 3H), 3.83 (m, 4H), 4.84 (m, 2H), 5.53 (m, 1H); ¹³C NMR (CDCL₃) δ 20.1, 32.2, 36.5, 63.2, 116.7, 134.6, 170.1.

S-(-)-1-O-Acetyl-2-allyl-1,3-propanediol (7). Lipase (sigma, Type VII, from *Candida cylindracea*, 4 g) was added to a rapidly stirred solution of the diacetate 6 (2.0 g, 10 mmol) and Triton X-100 (0.48 g) in a 30% aqueous acetone solution (290 mL). NaOH (1 N) was added dropwise to maintain a pH of 7. The reaction was allowed to proceed for 7.5 h and was then diluted with EtOAc (200 mL) and filtered through Celite. The filtrate was then washed with water, dried (Na₂SO₄), concentrated and purified by flash chromatography (hexanes to 50% EtOAc/hexanes) to afford 0.98 g (4.90 mmol, 49%) of the unreacted diacetate 6 and 0.79 g (5.00 mmol, 50%) [98% conversion] of the S-(-)-monoacetate isomer 7: $[\alpha]_{\rm D} = -7.95^{\circ}$ (c = 1.21, CHCl₃), Lit.²² $[\alpha]_{\rm D} = -7.65^{\circ}$; ¹H NMR (CDCl₃) δ 1.66 (m, 1H), 1.80 (s, 3H), 1.86 (m, 2H), 3.30 (m, 2H), 3.48 (br s, 1H), 3.85 (m, 2H), 4.80 (m, 2H), 5.52 (m, 1H); ¹³C NMR (CDCl₃) δ 20.2, 31.9, 39.5, 61.3, 63.7, 116.7, 135.3, 170.9.

Tetrahydro-5(R and S)-acetyloxy-3(R)-[(((1,1-dimethylethyl)dimethylsilyl)oxy)methyl]furan (11). To a solution of 7 (1.10 g, 6.92 mmol) in CH₂Cl₂ (50 mL) was added *tert*-butyldimethylsilyl chloride (1.25 g, 8.31 mmol) and imidazole (0.57 g, 8.31 mmol) under nitrogen. The resulting heterogeneous mixture was stirred at room temperature for 24 h and then diluted with CH₂Cl₂, washed with water, dried (Na₂SO₄), concentrated, and was then flash chromatographed (hexanes) to afford 1.87 g (6.85 mmol, 99%) of R-(-)-3-O-acetyl-2-allyl-1-O-[(1,1-dimethylethyl)dimethylsilyl]-1,3-propanediol (8): $[\alpha]_p = -0.10^\circ$ (c = 2.08, CHCl₃); ¹H NMR (CDCl₃) δ 0.08 (s, 6H), 0.78 (s, 9H), 1.77 (m, 1H), 1.91 (s, 3H), 2.01 (m, 2H), 3.47 (m, 2H), 3.94 (m, 2H),

4.92 (m, 2H), 5.64 (m, 1H); ¹³C NMR (CDCl₃) δ -5.8, 18.0, 20.6, 25.6, 32.3, 39.9, 61.9, 64.0, 116.4, 135.8, 170.5.

To a solution of **8** (0.28 g, 1.04 mmol) in MeOH (10 mL) was added sodium methoxide (0.07 g, 1.24 mmol) and the solution was stirred for 45 min. The reaction was then neutralized with 1 N HCl. The volatiles were then removed under reduced pressure and the product was purified by flash chromatography (hexanes to 25% EtOAc/hexanes) to afford 0.23 g (1.00 mmol, 97%) of R-(+)-2-allyl-1-O-[(1,1-dimethylethyl)dimethyl-silyl]-1,3-propanediol (9): $[\alpha]_D = +3.73^{\circ}$ (c = 1.76, CHCl₃); ¹H NMR (CDCl₃) δ -0.02 (s, 6H), 0.81 (s, 9H), 1.70 (m, 1H), 1.97 (m, 2H), 3.0 (br s, 1H), 3.55 (m, 4H), 4.93 (m, 2H), 5.68 (m, 1H); ¹³C NMR (CDCl₃) δ -5.7, 18, 25.7, 32.2, 41.9, 64.9, 65.5, 116.1, 136.3.

Sodium metaperiodate (3.91 g, 18.3 mmol) in water (10.5 mL) was added to an etheral solution (10.5 mL) of **9** (1.41 g, 6.10 mmol) and OsO₄(0.08 g, 0.30 mmol). The solution was vigorously stirred for 12 h then cooled to 0°C and NaHSO₃ (1 g) was slowly added. The mixture was allowed to stir for an additional 10 min, diluted with EtOAc, then transferred to a separatory funnel and washed with saturated NaHSO₃ and water. The organics were then dried (Na₂SO₄) and purified by flash chromatography (hexanes to 50% EtOAc/hexanes) to afford 1.39 g (5.98 mmol, 98%) of tetrahydro-5(R and S)-hydroxy-3(R)-[(((1,1-dimethylethyl)dimethylsily))oxy)methyl]furan (10): $[\alpha]_D = +24.0^{\circ}$ (c = 1.78, CHCl₄); ¹H NMR (CDCl₃) δ -0.15 (s, 6H), -0.10 (s, 6H), 0.70 (s, 9H), 0.72 (s, 9H), 1.49 (m, 2H, H-4\alpha/\beta), 1.73 (ddd, 1H, J = 1.2, 7.9 and 13.1 Hz, H-4\alpha), 1.97 (ddd, 1H, J = 5.3, 10.1 and 13.3 Hz, H-4\beta), 2.34 (m, 1H, H-3\beta), 2.44 (m, 1H, H-3\alpha), 3.29 (dd, 1H, J = 7.5 and 9.8 Hz, H-6\alpha), 3.37 (dd, 1H, J = 6.0 and 9.9 Hz, H-6\alpha), 3.48 (m, 3H, H-6\beta and H-2\beta), 3.60 (dd, 1H, J = 6.5 and 8.6 Hz, H-2\beta), 3.80 (t, 1H, J = 8.3, H-2\alpha), 3.48 (m, 3H, H-6\beta and H-2\beta), 3.60 (dd, 1H, J = 6.5 and 8.6 (Hz, H-2\beta), 3.80 (t, 1H, J = 8.3, H-2\alpha), 3.48 (t, 1H, J = 7.9 Hz, H-2\alpha), 4.74 (d, 1H, J = 3 Hz, C(5)-OH\alpha), 4.86 (d, 1H, J = 6.3 Hz, C(5)-OH\beta), 5.25 (t, 1H, J = 5.1 Hz, H-5\beta), 5.31 (s, 1H, H-5\alpha): ¹³C NMR (CDCl₃) δ -5.8, 17.8, 17.9, 25.5, 35.7, 36.3, 39.4, 64.3, 64.6, 68.7, 97.8, 97.9: Mass Spectrum (70 eV) *m*/z (rel. Intensity) 215 (.37), 201 (1.39), 173 (6.25), 157 (20.27), 83 (100).

To a solution of **10** (1.11 g, 4.78 mmol) in CH₂CN (10 mL) containing 0.5 g of molecular sieves were added Et₃N (0.93 mL, 6.67 mmol), DMAP (0.06 g, 0.48 mmol), and Ac₂O (0.54 mL, 5.72 mmol). The solution was stirred for 1 h then diluted with Et₂O and washed with water, dried (Na₂SO₄), concentrated, and flash chromatographed (hexanes to 20% EtOAc/hexanes) to afford 1.02 g (3.73 mmol, 78%) of predominantly the 5(R)-anomeric acetate of **11**: ¹H NMR (CDCL₃) δ -0.06 (s, 6H), -0.05 (s, 6H), 0.78 (s, 9H), 0.79 (s, 9H), 1.76 (m, 2H, H-4\alpha/\beta), 1.95 (s and m, 7H, CH₃\alpha/\beta and H-4\beta), 2.14 (m, 1H, H-4\alpha), 2.41 (m, 1H, H-3\beta), 2.53 (m, 1H, H-3\alpha), 3.46 (m, 2H, H-6\alpha), 3.55 (m, 2H, H-6\beta), 3.67 (m, 2H, H-2\alpha/\beta), 3.97 (m, 2H, H-2\alpha/\beta), 6.16 (d, 2H, J = 4.7 Hz, H-5\alpha/\beta); ¹³C NMR (CDCL₁) δ -5.7, 18, 21, 25.6, 34.3, 34.7, 39.1, 39.8, 63.9, 64.2, 70.7, 71.1, 99, 169.9, 170. Anal. Calcd. for C₁₃H₂₆O₄Si: C, 56.90; H, 9.96. Found: C, 56.80; H, 9.99.

5(R)-(6-Amino-9H-purin-9-yl)tetrahydro-3(S)-furanmethanol (14). A solution of N⁶-benzoyladenine (0.68 g, 2.82 mmol) and BSA (0.91 mL, 3.67 mmol) in CH₃CN (5 mL) was refluxed for 1 hr then cooled to 0 °C. The acetylated sugar **11** (0.77 g, 2.82 mmol) in CH₃CN (6 mL) was then added, followed by TMSOTf (0.60 mL, 3.10 mmol), and the reaction mixture was stirred for 2 h at 0°C and then at room temperature for an additional 2 h. A saturated solution of NaHCO₃ (20 mL) was then added, followed by EtOAc (100 mL), and the organics were washed with water, dried (Na₂SO₄), and purified by flash chromatography (CHCl₃ to 5% MeOH/CHCl₃) and by preparative layer chromatography with multiple elutions (20% hexanes/Et₂O to Et₂O) to afford 0.19 g (0.42 mmol, 15%) of 5(R)-[(6-benzoylamino)-9H-purin-9-yl]tetrahydro-3(R)-[(((1,1-dimethylethyl)dimethylsilyl)oxy)methyl]furan **12**: U.V. (MeOH) $\lambda_{max} = 279$ nm; [α]_D = -10.5° (c = 1.0, CHCl₃); ¹H NMR (CDCl₃) δ -0.03 (s, 6H), 0.81 (s, 9H), 2.36 (m, 1H), 2.62 (m, 2H), 3.65 (d, 2H, J = 5.1 Hz), 3.98 (t, 1H, J = 8.1 Hz), 4.08 (t, 1H, J = 7.9 Hz), 6.27 (t, 1H, J = 6.2 Hz), 7.40 (t, 2H, J = 7.4 Hz), 7.47 (t, 1H, J = 7.3 Hz), 7.95 (d, 2H, J = 7.3 Hz), 8.16 (s, 1H), 8.67 (s, 1H), 9.52 (br s, 1H); ¹³C NMR (CDCl₃) δ -5.6, 18.1, 25.7, 34.4, 41.5, 62.7, 71.2, 85.7, 123.6, 127.8, 128.6, 132.5, 133.6, 140.9, 149.5, 151.5, 152.3, 164.9; Mass Spectrum (70 eV) *m/z* (rel. intensity) 453 (M⁺, 0.2), 424 (M⁺-30, 0.3), 396 (M⁺-57, 2.7); and 0.35 g (0.76 mmol, 27%) of 5(S)-[(6-benzoylamino)-9H-purin-9-yl]tetrahydro-3(R)-[(((1,1-dimethylethyl)dimethyl-silyl)oxy)methyl]furan **13**: U.V. (MeOH) $\lambda_{max} = 279$ nm; [α]_D = +20.5° (c = 1.0, CHCl₃); ¹H NMR (CDCl₃) δ -5.6, 18.0, 27.7, 0, 21 (d, 1H, J = 7.9 Hz), 6.24 (d, 1H, J = 7.8 Hz), 7.33 (t, 2H, J = 7.4 Hz), 7.43 (t, 1H, J = 7.2 Hz), 7.91 (d, 2H, J = 7.5 Hz), 7.99 (s, 1H), $\lambda_{max} = 279$ nm; [α]_D = +20.5° (c = 1.0, CHCl₃); ¹H NMR (CDCl₃) δ -5.6, 18.0, 25.6, 34.7, 4.20 (t, 1H, J = 7.9 Hz), 6.2

A solution of 12 (0.19 g, 0.42 mmol) in MeOH (100 mL) was cooled to 0 °C and saturated with NH₃. The reaction was then warmed to room temperature and allowed to proceed for 24 h. The volatiles were then

removed under reduced pressure and the resulting oil was purified by preparative TLC (5% MeOH/CHCl₃) to give 0.15 g (0.42 mmol, 99%) of the debenzoylated product: U.V. (MeOH) $\lambda_{max} = 259$ nm; ¹H NMR (CDCl₃) δ -0.03 (s, 6H), 0.80 (s, 9H), 2.31 (m, 1H), 2.61 (m, 2H), 3.63 (d, 2H, J = 5.2 Hz), 3.95 (t, 1H, J = 8.0 Hz), 4.05 (t, 1H, J = 7.8 Hz), 6.22 (t, 1H, J = 6.0 Hz), 6.62 (br s, 2H), 7.98 (s, 1H), 8.25 (s, 1H); ¹³C NMR (CDCl₃) δ -5.6, 18.1, 25.7, 34.3, 41.4, 62.8, 70.9, 85.3, 119.9, 138.1, 149.4, 152.7, 155.8. To a solution of this product (0.15 g, 0.42 mmol) in CH₂CN (10 mL) was added Et₄NF (0.09 g, 0.63 mmol) and the solution was stirred for 1 hr. The volatiles were then removed and the tan oil was purified by preparative TLC (10% MeOH/CHCl₃) followed by reverse-phase HPLC (20% EtOH/H₂O) to afford 0.07 g (0.31 mmol, 73%) of 14: M.P. (lyophilized powder) 182-184 °C; U.V. (H₂O) $\lambda_{max} = 259$ nm ($\epsilon = 14,580$); [α]_D = -22.6° (c = 0.25, MeOH); ¹H NMR (DMSO-d₄) δ 2.31 (m, 1H), 2.55 (m, 2H), 3.56 (m, 2H), 3.88 (t, 1H, J = 8.1 Hz), 3.99 (t, 1H, J = 7.7 Hz), 4.83 (t, 1H, J = 5.1 Hz), 6.23 (t, 1H, J = 6.5 Hz), 7.26 (br s, 2H), 8.14 (s, 1H), 8.32 (s, 1H); ¹³C NMR (DMSO-d₆) δ 33.7, 41.7, 61.7, 70.8, 84.3, 119.2, 139.1, 149.2, 152.5, 156. Anal. Calcd. for C₁₀H₁₃N₅O₂: C, 51.06; H, 5.57; N, 29.77. Found C, 50.91; H, 5.59; N, 29.68.

5(S)-(6-Amino-9H-purin-9-yl)tetrahydro-3(S)-furanmethanol (15). In a manner identical to that described for 14, 0.35 g (0.77 mmol) of 13 was converted to 0.27 g (0.76 mmol, 99%) of the debenzoylated product: U.V. (MeOH) $\lambda_{max} = 259$ nm; ¹H NMR (CDCl₃) δ 0.0 (s, 6H), 0.82 (s, 9H), 2.25 (m, 1H), 2.52 (m, 1H), 2.66 (m, 1H), 3.58 (m, 2H), 3.83 (t, 1H, J = 7.4 Hz), 4.22 (t, 1H, J = 7.9 Hz), 6.26 (dd, 1H, J = 2.8 and 6.3 Hz), 6.69 (br s, 2H), 7.87 (s, 1H), 8.25 (s, 1H); ¹³C NMR (CDCl₃) δ -5.6, 18.1, 25.7, 34.7, 40.2, 63.4, 71.6, 85.8, 120.0, 138.2, 149.0, 152.7, 155.8. To a solution of the debenzoylated product (0.27 g, 0.76 mmol) in CH₃CN (10 mL) was added Et₄NF (0.17 g, 1.15 mmol) and the solution was stirred for 1 hr. The volatiles were then removed under reduced pressure and the oil obtained was purified by preparative TLC and by reverse-phase HPLC to afford 0.16 g (0.67 mmol, 88%) of the 5'(S)-isomer 15: M.P. (lyophilized powder) 138-140 °C; U.V. (H,O) $\lambda_{max} = 259$ nm ($\epsilon = 14,180$); [$\alpha_{l_3} = +39.8^{\circ}$ (c = 0.25, MeOH); ¹H NMR (DMSO-d₆) δ 2.22 (m, 1H), 2.56 (m, ^mH), 2.76 (m, 1H), 3.43 (m, 2H), 3.75 (dd, 1H, J = 5.4 and 8.1 Hz), 4.17 (t, 1H, J = 7.6 Hz), 4.81 (t, 1H, J = 4.7 Hz), 6.27 (dd, 1H, J = 3.0 and 6.6 Hz), 7.24 (br s, 2H), 8.14 (s, 1H), 8.26 (s, 1H); ¹³C NMR (DMSO-d₆) δ 34.0, 40.5, 62.2, 70.9, 84.4, 119.2, 139.2, 149.0, 152.6, 156.0. Anal. Calcd. for C₁₀H₁₃N₅O₂.1/4 H₂O: C, 50.1; H, 5.68; N, 29.21. Found C, 50.19; H, 5.66; N, 29.14.

5(R)-(2-Amino-1,9-dihydro-6H-purin-6-one)tetrahydro-3(S)-furanmethanol (18). Under the standard glycosylation conditions, **11** (1.05 g, 3.82 mmol) in CH₃CN (6 mL) followed by TMSOTf (0.81 mL, 4.21 mmol) were added at 0 °C to a solution of silylated N²-acetyl-O⁶-diphenylcarbamoylguanine (1.78 g, 4.59 mmol), prepared with BSA (1.32 mL, 5.35 mmol) in CH₃CN (15 mL). The resulting solution was allowed to warm to room temperature and was stirred for 4 h. Workup, as previously described, followed by purification utilizing flash chromatography (5% MeOH/CHCl₃) and then preparative layer chromatography (25% hexanes/Et₄O) gave 0.16 g (0.27 mmol, 7.1%) of the slower migrating 5(R)-[2-(acetylamino)-6-(diphenyl-carbamoyloxy)-9H-purin-6-yl]tetrahydro-3(R)-[(((1,1-dimethylethyl)dimethylsilyl)oxy)methyl]furan (16): U.V. (MeOH) $\lambda_{max} = 276$ nm; ¹H NMR (CDCl₄) 60.01 (s, 6H), 0.84 (s, 9H), 2.30 (m, 1H), 2.46 (s, 3H), 2.55 (m, 1H), 2.61 (m, 1H), 3.66 (m, 2H), 3.96 (t, 1H, J = 8.2 Hz), 4.08 (t, 1H, J = 7.8 Hz), 6.18 (t, 1H, J = 6.2 Hz), 7.19 (t, 2H, J = 6.6 Hz), 7.30 (t, 4H, J = 7.7 Hz), 7.38 (d, 4H, J = 7.5 Hz), 8.14 (s, 1H), 8.42 (br s, 1H); ¹³C NMR (CDCl₃) δ -5.6, 18.1, 25.0, 25.7, 34.3, 41.5, 62.7, 71.0, 85.6, 121.1, 126.4, 126.8, 127.3, 127.5, 129.0, 141.6, 141.8, 150.2, 151.9, 154.4, 155.9, 170.7; and 0.30 g (0.50 mmol, 13%) of the faster migrating 5(S)-[2-(acetylamino)-6-(diphenylcarbamoyloxy)-9H-purin-6-yl]tetrahydro-3(R)-[(((1,1-dimethylethyl)dimethyl-silyl)oxy)methyl]furan (17): U.V. (MeOH) $\lambda_{max} = 276$ nm; ¹H NMR (CDCl₃) δ o.56 (m, 1H), 2.72 (m, 1H), 3.61 (m, 2H), 3.87 (dd, 1H, J = 6.1 and 8.5 Hz), 4.25 (t, 1H, J = 7.9 Hz), 6.19 (dd, 1H, J = 2.7 and 6.6 Hz), 7.18 (t, 2H, J = 7.0 Hz), 7.34.4, 40.2, 63.4, 71.7, 86.3, 121.3, 126.1, 126.8, 127.4, 127.7, 129.0, 141.6, 142.2, 150.3, 151.9, 154.1, 155.9, 170.6.

252 nm ($\varepsilon = 13,138$) 274 sh ($\varepsilon = 8778$); [α]_D = -22.3° (c = 0.25, 10% H₂O/MeOH); ¹H NMR (DMSO-d₂) δ 2.15 (m, 1H), 2.47 (m, 2H), 3.52 (m, 2H), 3.82 (t, 1H, J = 8.0 Hz), 3.93 (t, 1H, J = 7.7 Hz), 4.80 (br s, 1H), 5.98 (t, 1H, J = 6.5 Hz), 6.51 (br s, 2H), 7.87 (s, 1H), 10.68 (br s, 1H); ¹³C NMR (DMSO-d₂) δ 33.7, 41.5, 61.6, 70.4, 83.5, 116.8, 135.1, 150.9, 153.9, 157.2. Anal. Calcd. for C₁₀H₁₃N₅O₃.1/4 H₂O: C, 46.96; H, 5.32; N, 27.38. Found C, 46.94, H, 5.34; N, 27.29.

5(S)-(2-Amino-1,9-dihydro-6H-purin-6-one)tetrahydro-3(S)-furanmethanol (19). In a similar fashion as described for the lactam deblocking of 16, 0.32 g (0.54 mmol) of 17 was deblocked to give 0.18 g (0.49 mmol, 92%) of the silylated analog: U.V. (MeOH) $\lambda_{max} = 252$ nm (sh 274); 'H NMR (CDCL) δ 0.05 (s, 6H), 0.87 (s, 9H), 2.16 (m, 1H), 2.40 (m, 1H), 2.74 (m, 1H), 3.59 (m, 2H), 3.69 (dd, 1H, J = 5.4 and 7.9 Hz), 4.13 (t, 1H, J = 7.6 Hz), 6.04 (dd, 1H, J = 2.9 and 6.6 Hz), 6.47 (br s, 2H), 7.82 (s, 1H), 10.68 (br s, 1H); ¹³C NMR (DMSO-d), δ -5.4, 17.9, 25.8, 33.7, 39, 63.6, 70.3, 83.6, 116.9, 135.2, 150.6, 153.6, 156.8. This product 0.17 g (0.45 mmol) was then similarly desilylated with Et₄NF (0.14 g, 0.91 mmol) to afford 0.11 g (0.42 mmol, 93%) of **19**; M.P. (lyophilized powder) >250 °C (decomp.); U.V. (H₂O) $\lambda_{max} = 252$ nm ($\varepsilon = 12,600$) 274 sh ($\varepsilon = 8435$); [α]_p = +18.7° (c = 0.27, 10% H₂O/MeOH); ¹H NMR (DMSO-d₂) δ 2.16 (m, 1H), 2.37 (m, 1H), 2.67 (m, 1H), 3.70 (dd, 1H, J = 5.4 and 8.4 Hz), 4.12 (dd, 1H, J = 7.4 and 8.1 Hz), 4.80 (br s, 1H), 4.80 (br s, 1H); δ 2.16 (m, 1H); δ 2.16 (m, 2H) δ 2.16 (m, 2H) δ 2.16 (m, 2H); δ 2.16 (m, 2H) δ 2.16 (m, 2H); δ 2.16 (m, 2H) δ 2.16 (m, 2H); δ 2.16 (m, 2 6.03 (dd, 1H, J= 3.7 and 7.0 Hz), 6.48 (br s, 2H), 7.81 (s, 1H), 10.68 (br s, 1H); ¹³C NMR (DMSO-d₂) δ 34.1, 40.2, 62.1, 70.6, 83.6, 116.8, 135.1, 150.7, 153.6, 156.9. Anal. Calcd. for C₁₀H₁₃N₅O₃: C, 47.81; H, 5.22; N, 27.87. Found C, 47.73; H, 5.23; N, 27.79.

5(R)-[2,4(1H,3H)-Pyrimidinedione]tetrahydro-3(S)-furanmethanol (22). Under the standard glycosylation conditions, 11 (1.0 g, 3.65 mmol) in CH₂CN (6 mL) followed by TMSOTf (0.78 mL, 4.02 mmol) were added at 0 °C to a solution of silylated uracil (0.49 g, 4.38 mmol), prepared with BSA (2.38 mL, 9.64 mmol) in CH₂CN (3 mL). The resulting solution was allowed to warm to room temperature and was 9.64 mmol) in CH₂CN (5 mL). The resulting solution was allowed to warm to room temperature and was stirred for 4 h. After workup and purification using flash chromatography (CHCl₁ to 5% MeOH/CHCl₂), separation of the anomers by preparative TLC (25% hexanes/Et₂O) yielded 0.30 g (0.91 mmol, 25%) of the faster migrating 5(R)-[2,4(1H,3H)-pyrimidinedione]tetrahydro-3(R)-[(((1,1-dimethylethyl)dimethylsilyl)oxy) methyl]furan (20): U.V. (MeOH) $\lambda_{max} = 261$ nm; ¹H NMR (CDCl₂) δ 0.01 (s, 6H), 0.82 (s, 9H), 1.70 (m, 1H), 2.54 (m, 2H), 3.52 (dd, 1H, J = 5.6 and 10.1 Hz), 3.60 (dd, 1H, J = 4.4 and 10.2 Hz), 3.86 (t, 1H, J = 7.8 Hz), 4.01 (t, 1H, J = 7.0 Hz), 130 2.54 (iii, 21), 5.52 (di, 111, j = 5.0 and 10.1 Hz), 5.60 (di, 111, j = 4.4 and 10.2 Hz), 5.60 (d; 111, j = 7.6 Hz), 4.01 (t, 1H, J = 7.9 Hz), 5.70 (d, 1H, J = 8.1 Hz), 6.0 (t, 1H, J = 6.2 Hz), 7.42 (d, 1H, J = 8.1 Hz); ¹³C NMR (CDCl₃) δ -5.7, 18.0, 25.6, 34.6, 40.6, 62.6, 70.9, 86.7, 102.0, 139.2, 150.5, 163.8; and 0.68 g (2.08 mmol, 57%) of the slower migrating 5(S)-[2,4(1H,3H)-pyrimidinedione]tetrahydro-3(R)-[((((1,1-dimethylethyl)-dimethylsilyl)oxy)methyl]furan (21): U.V. (MeOH) $\lambda_{max} = 261.5$ nm; ¹H NMR (CDCl₃) δ -0.11 (s, 6H), 0.72 (s, 9H), 1.93 (m, 1H), 2.09 (m, 1H), 2.37 (m, 1H), 3.42 (m, 1H), 3.48 (m, 1H), 3.67 (t, 1H, J = 7.7 Hz), 4.09 (t, 1H, J = 7.7 Hz), 5.58 (d, 1H, J = 8.0 Hz), 5.85 (m, 1H), 7.26 (d, 1H, J = 8.0 Hz); ¹³C NMR (CDCl₃) δ -5.7, 18.0, 25.6, 35.2, 39.7, 63.0, 72.1, 87.3, 101.6, 139.2, 150.2, 163.9.

Treatment of 20 (0.17 g, 0.53 mmol) with Et₄NF (0.16 g, 1.06 mmol) in CH₂CN (50 mL) afforded, after The anterior of 20 (617) g, 63.5 minoly with E144 (616 g, 16.6 minol) with C13C14(60 mi

5(S)-[2,4(1H,3H)-Pyrimidinedione]tetrahydro-3(S)-furanmethanol (23). In an identical manner 0.45 g (1.37 mmol) of **21** was treated with Et NF (0.41 g, 2.74 mmol) to give 0.28 g (1.32 mmol, 96%) of **23**: M.P. (lyophilized powder) 93-95 °C; U.V. (H₂O) $\lambda_{max} = 262$ nm ($\epsilon = 10,100$); [α]_p = -17.7° (c = 0.48, MeOH); ¹H NMR (DMSO-d₆) δ 2.04 (m, 2H), 2.46 (m, 1H), 3.38 (m, 2H), 3.64 (dd, 1H, J = 6.5 and 8.3 Hz), 4.18 (dd, 1H, Hz) = 0.5 mmol + 0.5 m J = 7.3 and 8.2 Hz), 4.78 (br s, 1H), 5.56 (d, 1H, J = 8.1 Hz), 5.93 (dd, 1H, J = 0.3 and 0.3 Hz), 4.18 (dd, 1H, J = 7.9 Hz), 11.2 (br s, 1H); ${}^{13}C$ NMR (DMSO-d₂) δ 34.4, 39.9, 61.8, 71.6, 86.2, 101.3, 140.6, 150.4, 163.3. Anal. Calcd. for C₉H₁₂N₂O₄: C, 50.94; H, 5.70; N, 13.20. Found C, 50.87; H, 5.69; N, 13.15.

5(R)-[4-Amino-2(1H)-pyrimidinone]tetrahydro-3(S)-furanmethanol (26) and 5(S)-[4-Amino-2(1H)pyrimidinone]tetrahydro-3(S)-furanmethanol (27). Under the standard glycosylation conditions, 11 (1.03

g, 3.76 mmol) in CH₂CN (6 mL) followed by TMSOTf (0.80 mL, 4.13 mmol) were added at 0 °C to a solution of silylated cytosine (0.50 g, 4.51 mmol), prepared with BSA (2.23 mL, 9.02 mmol) in CH₂CN (3 mL). The resulting solution was allowed to warm to room temperature and was stirred for 3 h. The reaction was then worked up under standard conditions to give, after purification by flash chromatography (10% MeOH/CHCL 1.03 g (3.16 mmol, 84%) of a 1/1 anomeric mixture of the nucleosides 24: U.V. (MeOH) $\lambda_{max} = 272 \text{ nm}; {}^{1}\text{H}$

NMR (CDCl₃) δ -0.04 (s, 6H), 0.80 (s, 9H), 1.57 (m, 1H), 2.56 (m, 2H), 3.46 (m, 1H), 3.54 (dd, 1H, J = 5.0 and 10.1 Hz), 3.82 (t, 1H, J = 7.5 Hz), 4.0 (t, 1H, J = 7.4 Hz), 5.72 (d, 1H, J = 7.4 Hz), 5.96 (t, 1H, J = 5.9 Hz), 7.52 (d, 1H, J = 7.5 Hz), ¹³C NMR (CDCl₃) δ -5.6, 18.1, 25.7, 35.5, 41.0, 63.4, 71.3, 88.0, 94.8, 140.5, 156.2, 166.1; and 25: U.V. (MeOH) $\lambda_{max} = 272 \text{ nm}$.¹H NMR (CDCl₃) δ -0.01 (s, 6H), 0.83 (s, 9H), 2.05 (m, 1H), 2.17 (m, 1H), 2.40 (m, 1H), 3.50 (t, 1H, J = 7.5 Hz), 3.61 (dd, 1H, J = 5.1 and 9.8 Hz), 3.78 (t, 1H, J = 8.0 Hz), 4.17 (t, 1H, J = 7.8 Hz), 5.82 (d, 1H, J = 7.3 Hz), 5.98 (m, 1H), 7.35 (d, 1H, J = 7.3 Hz); ¹³C NMR (CDCl₃) δ -5.5, 18.2, 25.8, 35.8, 39.7, 63.4, 72.3, 88.1, 94.3, 139.8, 156.1, 166.1. To a solution of 24 and 25 (0.96 g, 2.96 mmol) in CH₂CN (50 mL) was added Et₄NF (0.88 g, 5.91 mmol). Purification utilizing flash chromatography (15% MeOH/CHCl₃) afforded 0.56 g (2.63 mmol, 89%) of the 1/1 anomeric mixture of 26 and 27. Pure anomers were isolated through multiple developments by preparative TLC (Et₂O 78%, CHCl₃ 18%, MeOH 2%, EtOH 2%) and further purified by reverse-phase HPLC (10% EtOH/H₂O) to give pure 26 and 27. Data for 26: low melting point solid; U.V. (H₂O) $\lambda_{max} = 272 \text{ nm}$ (ϵ = 8890); [α]₀ = +56.5° (c = 0.23, MeOH); ¹H NMR (DMSO-d₂) δ 1.58 (m, 1H), 2.36 (m, 1H), 2.47 (m, 1H), 3.40 (m, 2H), 3.82 (t, 1H, J = 7.8 Hz), 3.93 (t, 1H, J = 7.4 Hz), ¹³C NMR (DMSO-d₂) δ 34.9, 41.0, 62.0, 70.9, 86.5, 93.9, 140.6, 155.2, 165.7. Anal. Calcd. for C_{H13}N₃O₃: C, 51.18; H, 6.20; N, 19.85. Data for 26! M.H. J = 7.6 Hz), 4.77 (t, 1H, J = 7.4 Hz), 5.71 (d, 1H, J = 7.3 Hz), 5.90 (dd, 1H, J = 3.6 and 6.3 Hz), 7.11 (d, 2H, J = 11.2 Hz), 7.54 (d, 1H, J = 7.3 Hz), ¹³C NMR (DMSO-d₂) δ 35.2, 40.7, 61.8, 71.5, 86.6, 93.5, 140.7, 155.1, 165.7. Anal. Calcd. for C₉H₁₃N₃O₃: C, 51.18; H, 6.20; N, 19.89. Found C, 51.0; H, 6.18; N, 19.87.

From 20: To a solution of 20 (0.12 g, 0.36 mmol) in pyridine (5 mL) was added a heterogeneous solution of POCl₃ (0.07 mL, 0.71 mmol) and 1,2,4-triazole (0.20 g, 2.85 mmol) in pyridine (5 mL). The resulting solution was stirred for 1 hr and then cooled to 0 °C. NH₄OH (conc., 1.5 mL) was then added and the reaction was allowed to warm to room temperature and was stirred for 12 h. The volatiles were then removed and the brown residue was purified by flash and preparative layer chromatography (10% MeOH/CHCl₃) to afford 0.09 g (0.27 mmol, 75%) of 24. A solution of 24 (0.09 g, 0.27 mmol) in CH₃CN was treated with Et₄NF (0.12 g, 0.81 mmol) to afford, after purification by preparative layer chromatography (15% MeOH/CHCl₃) and with reverse-phase HPLC (10% EtOH/H₂O), 0.05 g (0.22 mmol, 81%) of 26.

From 21: In an identical manner, 21 (0.21 g, 0.65 mmol) was treated with POCl₃ (0.12 mL, 1.30 mmol), triazole (0.36 g, 5.22 mmol), and NH₄OH (conc., 5 mL) to afford 0.17 g (0.51 mmol, 79%) of 25. Desilylation with Et NF (0.15 g, 1.03 mmol) afforded, after final purification by reverse-phase HPLC, 0.09 g (0.42 mmol, 83%) of 27.

5(R)-[5-Methyl-2,4(1H,3H)-pyrimidinedione]tetrahydro-3(S)-furanmethanol (30) and 5(S)-[5-Methyl-2,4(1H,3H)-pyrimidinedione]tetrahydro-3(S)-furanmethanol (31). Under the standard glycosylation conditions, 11 (0.51 g, 1.84 mmol) in CH₃CN (6 mL) followed by TMSOTT (0.39 mL, 2.03 mmol) were added at 0 °C to a solution of silylated thymine (0.23 g, 1.84 mmol), prepared with BSA (1.18 mL, 4.79 mmol) in CH₃CN (3 mL). The resulting solution was allowed to warm to room temperature and stir for 4 h. Workup and purification by flash chromatography (5% MeOH/CHCL₃) yielded 0.49 g (1.45 mmol, 79%) of an anomeric mixture (1/2) of the 5'(R)-anomer 28: U.V. (MeOH) $\lambda_{max} = 267$ nm; ¹H NMR (CDCL₃) & -0.08 (s, 6H), 0.76 (s, 9H), 1.65 (m, 1H), 1.79 (s, 3H), 2.43 (m, 2H), 3.53 (m, 2H), 3.81 (t, 1H, J = 8.0 Hz), 3.92 (t, 1H, J = 8.1 Hz), 5.97 (t, 1H, J = 6.4 Hz), 7.10 (s, 1H), 10.25 (br s, 1H); ¹³C NMR (CDCL₃) & -5.8, 12.2, 17.8, 25.5, 34.2, 40.5, 62.4, 70.6, 86.3, 110.4, 134.6, 150.3, 164.1; and the 5'(S)-anomer 29: U.V. (MeOH) $\lambda_{max} = 267$ nm; ¹H NMR (CDCL₃) & -0.08 (s, 6H), 0.76 (s, 9H), 1.79 (s, 3H), 1.95 (m, 1H), 2.10 (m, 1H), 2.43 (m, 1H), 3.47 (m, 2H), 3.69 (t, 1H, J = 7.7 Hz), 4.13 (t, 1H, J = 7.9 Hz), 5.92 (dd, 1H, J = 4.3 and 6.2 Hz), 7.06 (s, 1H), 10.25 (br s, 1H); ¹³C NMR (CDCL₃) & -5.8, 12.2, 17.8, 25.5, 34.9, 39.8, 63.1, 71.8, 86.7, 110.0, 134.9, 150.5, 164.2. Et₄NF (0.50 g, 3.38 mmol) was added to a solution of 28 and 29 (0.58 g, 1.69 mmol) in CH₃CN (30 ML). The solution was allowed to stir for 1 h, concentrated, and purified by flash chromatography (10% MCOH/CHCL₃) to afford 0.34 g (1.50 mmol, 89%) of the anomeric mixture. Preparative TLC utilizing multiple elutions (ether 78%, chloroform 18%, MeOH 2%, ethanol 2%), dissection of the resulting band, and reclution of appropriately combined fractions provided, after final purification by reverse phase HPLC (20% EtOH/H₄O), pure 5'(R)-anomer (faster migrating component on normal phase silica) 30: M.P. (lyophilized powder) 134-135 °C; U.

Found C, 52.94; H, 6.27; N, 12.34.

Data for the 5'(S)-anomer 31: M.P. (lyophilized powder) 131-133 °C; U.V. (MeOH) $\lambda_{max} = 267$ nm ($\epsilon = 9100$); $[\alpha]_{D} = +2.84^{\circ}$ (c = 0.56, MeOH); ¹H NMR (CDCL₃) δ 1.98 (s, 3H), 2.19 (m, 1H), 2.35 (m, 1H), 2.68 (m, 1H), 3.73 (m, 2H), 3.91 (t, 1H, J = 7.3 Hz), 4.35 (t, 1H, J = 7.6 Hz), 6.10 (dd, J = 4.3, 6.5 Hz, 1H), 7.33 (s, 1H); ¹³C NMR (CDCL₃) δ 12.6, 35.5, 40.1, 63.3, 72.2, 87.1, 110.6, 135.3, 150.6, 164.3. Anal. Calcd. for C₁₀H₁₄N₂O₄: C, 53.09; H, 6.24; N, 12.38. Found C, 52.88; H, 6.26; N, 12.35.

Tetrahydro-5(R and S)-acetyloxy-3(S)-[(acetyloxy)methyl]furan (33). To a solution of the S-(-)-monoacetate of 2-ally-1,3-propanediol (7, 2.49 g, 15.74 mmol) in Et₂O (100 mL) and water (30 mL) was added NaIO₄ (10.10 g, 47.21 mmol). The mixture was cooled to 0 °C and OsO₄ (0.10 g, 0.39 mmol) in THF (4 mL) was then added dropwise. The reaction was warmed to room temperature and then allowed to stir for 12 h. The reaction mixture was filtered and saturated NaHSO₃ was then slowly added. The resulting solution was stirred for 10 min and Na₂SO₄ was then added and the dried organics were filtered. The remaining solids were washed with EtOAc and the combined organics were concentrated and purified by flash chromatography (hexanes to 50% EtOAc/hexanes) to give 2.07 g (12.9 mmol, 82%) of tetrahydro-5(R and S)-hydroxy-3(S)-[(acetyloxy)methyl]furan (32): $[\alpha]_{D} = -31.1^{\circ}$ (c = 0.39, MeOH); ¹H NMR (CDCl₂) δ 1.63 (m, 1H), 1.94 (s, 3H), 2.07 (m, 1H), 2.67 (m, 1H), 3.52 - 4.10 (m, 4H), 4.32 (t, 1H, J = 8.2 Hz), 5.43 (d, 1H, J = 2.2 Hz); ¹³C NMR (CDCl₃) δ 20.6, 36.2, 36.9, 65.6, 66.1, 68.9, 69.3, 98.0, 98.2, 170.9.

Molecular sieves (0.30 g) were added to a solution of tetrahydro-5(R and S)-hydroxy-3(S)-[(acetyloxy)methyl]furan (32) (2.07 g, 12.97 mmol) in CH₃CN (20 mL). DMAP (0.16 g, 1.30 mmol), Et₃N (2.71 mL, 19.45 mmol), and Ac₂O (1.47 mL, 15.56 mmol) were then added and the reaction mixture was stirred under nitrogen with exclusion of moisture for 1.5 h. The solution was cooled to 0 °C and quenched with MeOH (10 mL). The volatiles were then removed under reduced pressure (bath temperature <40 °C) and the remaining oil was purified by flash chromatography (hexanes to 30% EtOAc/hexanes) to afford 2.03 g (10.1 mmol, 78%) of the product as a clear oil: ¹H NMR (CDCl₃) δ 1.80 (m, 1H), 1.91 (s, 3H), 1.92 (s, 3H), 2.20 (m, 1H), 2.60 (m, 1H), 3.50 - 4.04 (m, 4H), 6.15 (d, 1H, J = 4.6 Hz); ¹³C NMR (CDCl₃) δ 20.3, 20.7, 34.5, 34.9, 35.6, 36.0, 64.8, 65.3, 70.3, 70.8, 98.1, 169.7, 170.2. Anal. Calcd. for C₉H₁₄O₅: C, 53.46; H, 6.98. Found: C, 53.31; H, 6.99.

5(S)-(6-Amino-9H-purin-9-yl)tetrahydro-3(R)-furanmethanol (38). Under the standard glycosylation conditions, N⁶-benzoyladenine (0.68 g, 2.83 mmol) was treated with BSA (0.82 mL, 3.33 mmol), 33 (0.52 g, 2.57 mmol), and TMSOTf (0.55 mL, 2.82 mmol). After workup and purification by flash chromatography (5% MeOH/CHCl₃) an anomeric mixture [5(S):5(R) = 2:3] of the acetylated products (0.54 g, 1.41 mmol, 55%) were isolated. The mixture (0.46 g, 1.20 mmol) was allowed to stir for 8 h and additional sodium amide (0.07 g, 1.79 mmol) was added. The resulting solution was allowed to stir for 8 h and additional sodium amide to dryness under reduced pressure, and purified by flash chromatography (CHCl₄ to 8% MeOH/CHCl₃) to afford 0.39 g (1.14 mmol, 95%) of the de-acetylated anomers. Silylation of the anomeric mixture with imidazole (0.15 g, 2.15 mmol) and *tert*-butyldimethylsilyl chloride (0.23 g, 1.53 mmol) in CH₂Cl₁ (5 mL) gave the separable anomers, 5(S)-[(6-benzoylamino)-9H-purin-9-yl]tetrahydro-3(S)-[(((1,1-dimethylethyl) dimethylsilyl)oxy)methyl]furan (36) (0.18 g, 0.39 mmol, 34%): U.V. (MeOH) $\lambda_{max} = 279$ nm; [α]_D = +14.5° (c = 1.0, MeOH); ¹H NMR (CDCl₃) δ 0.01 (s, 6H), 0.85 (s, 9H), 2.40 (m, 1H), 2.66 (m, 2H), 3.68 (m, 2H), 4.03 (t, 1H, J = 8.1 Hz), 4.13 (t, 1H, 7.9 Hz), 6.32 (t, 1H, J = 6.2 Hz), 7.46 (t, 2H, J = 7.4 Hz), 7.54 (t, 1H, J = 7.3 Hz), 8.00 (d, 2H, J = 7.2 Hz), 8.20 (s, 1H), 8.75 (s, 1H), 9.29 (br s, 1H); ¹³C NMR (CDCl₃) δ -5.5, 18.3, 25.8, 34.6, 41.6, 62.8, 71.3, 85.9, 123.7, 127.9, 128.8, 132.7, 133.7, 141.0, 149.5, 151.5, 152.5, 164.7; and 5(R)-[(6-benzoylamino)-9H-purin-9(c)]_D = -19.7° (c = 1.0, MeOH); ¹H NMR (CDCl₃) δ 0.02 (s, 6H), 0.86 (s, 9H), 2.34 (m, 1H), 2.58 (m, 1H), 2.72 (m, 1H), 3.62 (m, 2H), 3.88 (dd, 1H, J = 6.4 and 8.4 Hz), 4.26 (t, 1H, J = 7.5 Hz), 8.05 (s, 1H), 9.79 (br s, 1H); ¹³C NMR (CDCl₃) δ -5.5, 18.4, 18.2, 7.34 (z), 7.96 (d, 2H, J = 7.5 Hz), 8.05 (s, 1H), 9.79 (br s, 1H); ¹³C NMR (CDCl₃) δ -5.5, 18.2, 25.8, 34.8, 40.3, 63.4, 71

Under identical conditions as previously described, **36** (0.12 g, 0.26 mmol) was debenzoylated (NH₃/MeOH) and then treated with Et₄NF (0.08 g, 0.52 mmol) to afford, after purification by preparative TLC and reverse-phase HPLC 0.06 g (0.24 mmol, 91%) of the desired **38**: M.P. (lyophilized powder) 180-183 °C; U.V. (H₂O) $\lambda_{\text{pax}} = 259 \text{ nm} (\varepsilon = 14,250); [\alpha]_{\text{p}} = +22.3^{\circ} (c = 0.18, \text{ MeOH}); ^{1}\text{H NMR} (DMSO-d_{c}) \delta$ 2.27 (m, 1H), 2.54 (m, 2H), 3.52 (m, 2H), 3.84 (t, 1H, J = 8.0 Hz), 3.95 (t, 1H, J = 7.6 Hz), 4.78 (br s, 1H), 6.18 (t, 1H)

J = 6.4 Hz), 7.21 (br s, 2H), 8.10 (s, 1H), 8.28 (s, 1H); ¹³C NMR (DMSO-d₂) δ 33.7, 41.6, 61.6, 70.7, 84.2, 119.1, 139.0, 149.1, 152.5, 156. Anal. Calcd. for C₁₀H₁₃N₅O₂: C, 51.06; H, 5.57; N, 29.77. Found C, 50.94; H, 5.54; N, 29.70.

5(R)-(6-Amino-9H-purin-9-yl)tetrahydro-3(R)-furanmethanol (39). Under identical conditions previously described, 0.20 g (0.45 mmol) of **37** was converted to 0.10 g (0.43 mmol, 96%) of **39**: M.P. (lyophilized powder) 139-140 °C; U.V. (H₂O) $\lambda_{max} = 259$ nm ($\epsilon = 13,800$); [α]_D = -38.6° (c = 0.35, MeOH); ¹H NMR (DMSO-d₂) δ 2.21 (m, 1H), 2.54 (m, 1H), 2.76 (m, 1H), 3.44 (m, 2H), 3.75 (dd, 1H, J = 5.4 and 8.2 Hz), 4.17 (t, 1H, J = 7.7 Hz), 4.81 (br s, 1H), 6.26 (dd, 1H, J = 3.4 and 7.0 Hz), 7.24 (br s, 2H), 8.14 (s, 1H), 8.25 (s, 1H); ¹³C NMR (DMSO-d₂) δ 33.9, 40.4, 62.2, 70.8, 84.3, 119.2, 139.2, 148.9, 152.5, 156. Anal. Calcd. for C₁₀H₁₃N₅O₂: C, 51.06; H, 5.57; N, 29.77. Found C, 50.98; H, 5.59; N, 29.73.

5(S)-(2-Amino-1,9-dihydro-6H-purin-6-one)tetrahydro-3(R)-furanmethanol (42). Under the standard glycosylation conditions, N²-acetyl-O⁶-diphenylcarbamoylguanine (0.79 g, 2.04 mmol) was treated with BSA (0.92 mL, 3.73 mmol), **33** (0.34 g, 1.87 mmol), and TMSOTF (0.36 mL, 1.87 mmol) to afford, after workup and purification by preparative TLC (Et₂O 78%, CHCl₄ 19%, MeOH 1%, EtOH 2%) 0.08 g (0.15 mmol, 9%) of 5(S)-[2-(acetylamino)-6-(diphenylcarbamoyloxy)-9H-purin-6-yl]tetrahydro-3(S)-[(acetyloxy) methyl]furan (**40**): U.V. (MeOH) $\lambda_{max} = 276$ nm; ¹H NMR (CDCl₃) δ 2.00 (s, 3H), 2.35 (m, 1H), 2.43 (s, 3H), 2.61 (m, 1H), 2.74 (m, 1H), 4.11 (m, 4H), 6.11 (t, 1H, J = 6.4 Hz), 7.25 (m, 10H), 8.05 (s, 1H), 8.64 (br s, 1H); ¹³C NMR (CDCl₄) δ 20.7, 24.9, 34.4, 38.5, 64.3, 71.3, 85.9, 121.4, 125.9, 126.9, 127.6, 127.8, 129.0, 141.6, 142.4, 150.3, 151.8, 154.2, 155.9, 170.1, 170.7; and 0.11 g (0.21 mmol, 13%) of the faster migrating 5(R)-[2-(acetylamino)-6-(diphenylcarbamoyloxy)-9H-purin-6-yl]tetrahydro-3(S)-[(acetyloxy)methyl]furan (**41**): U.V. (MeOH) $\lambda_{max} = 276$ nm; ¹H NMR (CDCl₄) δ 2.07 (s, 3H), 2.30 (m, 1H), 2.46 (s, 3H), 2.77 (m, 1H), 2.98 (m, 1H), 3.86 (dd, 1H, J = 5.8 and 8.8 Hz), 4.07 (dd, 1H, 7.2 and 11.0 Hz), 4.15 (dd, 1H, J = 6.3 and 11.0 Hz), 4.34 (dd, 1H, J = 7.5 and 8.5 Hz), 6.21 (dd, 1H, J = 2.8 and 6.9 Hz), 7.25 (m, 10H), 8.00 (s, 1H), 8.66 (br s, 1H); ¹³C NMR (CDCl₄) δ 20.6, 24.8, 34.5, 37.2, 64.6, 71.7, 86.0, 121.3, 126.2, 126.7, 127.2, 127.4, 129.0, 141.5, 142.4, 150.2, 151.8, 154.0, 155.9, 170.1, 170.7.

In a manner identical to that previously described, **40** (0.10 g, 0.19 mmol) was deblocked with ethanolic ammonia to afford, after purification, 0.04 g (0.16 mmol, 86%) of **42**: M.P. (lyophilized powder) >250 °C (decomp); U.V. (H₂O) $\lambda_{max} = 252$ nm ($\epsilon = 13,160$) 274 sh ($\epsilon = 8990$); [α]_D = +23.7° (c = 0.25, 10% H₂O/MeOH); ¹H NMR (DMSO-d₂) δ 2.15 (m, 2H), 2.47 (m, 2H), 3.53 (m, 2H), 5.81 (t, 1H, J = 8.0 Hz), 3.93 (t, 1H, J = 7.7 Hz), 4.81 (br s, 1H), 5.98 (t, 1H, J = 6.5 Hz), 6.46 (br s, 2H), 7.89 (s, 1H), 10.64 (br s, 1H); ¹³C NMR (DMSO-d₂) δ 33.7, 41.5, 61.6, 70.4, 83.5, 116.8, 135.1, 150.9, 153.5, 156.8. Anal. Calcd. for C₁₀H₁₃N₅O₃.H₂O: C, 44.61; H, 5.61; N, 26.01. Found C, 44.44; H, 5.63; N, 25.99.

5(R)-(2-Amino-1,9-dihydro-6H-purin-6-one)tetrahydro-3(R)-furanmethanol (43). In an identical manner, 41 (0.11 g, 0.21 mmol) was converted to 0.05 g (0.19 mmol, 94%) of 43: M.P. (lyophilized powder) >250 °C (decomp.); U.V. (H,O) $\lambda_{max} = 252 \text{ nm} (\epsilon = 13,060) 274 \text{ sh} (\epsilon = 8880); [α]_{D} = -18.3° (c = 0.41, 10% H,O/MeOH); ¹H NMR (DMSO-d_{o}) δ 2.14 (m, 1H), 2.38 (ddd, 1H, J = 3.7, 8.0, and 13.4 Hz), 2.67 (m, 1H), 3.41 (m, 2H), 3.70 (dd, 1H, J = 5.4 and 8.4 Hz), 4.12 (dd, 1H, J = 7.4 and 8.1 Hz), 4.82 (br s, 1H), 6.03 (dd, 1H, J = 3.8 and 7.0 Hz), 6.55 (br s, 2H), 7.80 (s, 1H), 10.67 (br s, 1H); ¹³C NMR (DMSO-d_{o}) δ 34.1, 40.2, 62.1, 70.5, 83.6, 116.8, 135.0, 150.7, 153.9, 157.3. Anal. Calcd. for C₁₀H₁₃N₅O₃.H₂O: C, 44.61; H, 5.61; N, 26.01. Found C, 44.56; H, 5.59; N, 25.93.$

NMR (CDCL) δ -5.6, 18.1, 25.7, 35.4, 39.8, 63.1, 72.2, 87.5, 101.7, 139.2, 150.5, 163.9.

Compound **48** (0.06 g, 0.19 mmol) was treated with Et_4NF (0.06 g, 0.38 mmol) in CH₃CN (10 mL) and upon purification yielded 0.03 g (0.16 mmol, 83%) of **46** as a low melting lyophilized solid: U.V. (H₂O) $\lambda_{max} = 262 \text{ nm}$ ($\epsilon = 9400$); [$\alpha_{1p} = -32.9^{\circ}$ (c = 0.30, MeOH); ¹H NMR (DMSO-d₄) δ 1.72 (m, 1H), 2.36 (m, 1H), 2.48 (m, 1H), 3.44 (m, 2H), 3.82 (t, 1H, J = 8.0 Hz), 3.94 (t, 1H, J = 7.9 Hz), ⁴.77 (br s, 1H), 5.62 (d, 1H, J = 8.0 Hz), 5.94 (t, 1H, J = 6.5 Hz), 7.66 (d, 1H, J = 8.0 Hz), 11.20 (br s, 1H); ¹³C NMR (DMSO-d₆) δ 34.0, 40.9, 61.7, 71.0, 85.9, 101.6, 140.5, 150.5, 163.3. Anal. Calcd. for $C_9H_{12}N_2O_4$.1/2 H_2O : C, 48.87; H, 5.92; N, 12.66. Found C, 48.72; H, 5.94; N, 12.64.

5(R)-[2,4(1H,3H)-Pyrimidinedione]tetrahydro-3(R)-furanmethanol (47). Under identical conditions, **49** (0.13 g, 0.41 mmol) was treated with Et₄NF (0.12 g, 0.82 mmol) to afford, after purification, 0.08 g (0.39 mmol, 95%) of **47**: M.P. (lyophilized powder) 97-99 °C: U.V. (H₂O) $\lambda_{max} = 262 \text{ nm} (\epsilon = 10,190); [\alpha]_{D} = +20.2^{\circ}$ (c = 0.30, MeOH); ¹H NMR (DMSO-d₂) δ 2.06 (m, 2H), 2.48 (m, 1H), 3.39 (m, 2H), 3.65 (t, 1H, J = 7.3 Hz), 4.20 (t, 1H, J = 7.8 Hz), 4.80 (br s, 1H), 5.58 (d, 1H, J = 8.0 Hz), 5.94 (t, 1H, J = 4.9 Hz), 7.60 (d, 1H, J = 8.2 Hz), 11.2 (br s, 1H); ¹³C NMR (DMSO-d₂) δ 34.4, 39.9, 61.8, 71.6, 86.2, 101.7, 140.6, 150.4, 163.4. Anal. Calcd. for C₉H₁₂N₂O₄: C, 50.94; H, 5.70; N, 13.20. Found C, 50.84; H, 5.72; N, 13.15.

5(S)-[4-Amino-2(1H)-pyrimidinone]tetrahydro-3(R)-furanmethanol (52). Under similar conditions as described for **26**, 0.21 g (0.65 mmol) of the protected uridine **48** was treated with a solution of triazole (0.36 g, 5.16 mmol) and POCl₃ (0.12 mL, 1.29 mmol) in pyridine (15 mL). After treatment with NH₂OH (conc., 3 mL) and purification utilizing flash and preparative layer chromatography 0.14 g (0.44 mmol, 69%) of the protected cytidine analog **50** was isolated: U.V. (MeOH) $\lambda_{max} = 272 \text{ nm}$; ¹H NMR (CDCl₃) δ 0.0 (s, 6H), 0.83 (s, 9H), 1.63 (m, 1H), 2.61 (m, 1H), 3.48 (dd, 1H, J = 6.0 and 9.8 Hz), 3.57 (dd, 1H, J = 4.5 and 10.2 Hz), 3.90 (t, 1H, J = 7.8 Hz), 4.08 (t, 1H, J = 7.9 Hz), 5.76 (d, 1H, J = 7.4 Hz), 6.02 (t, 1H, J = 6.0 Hz), 7.58 (d, 1H, J = 7.4 Hz); ¹³C NMR (CDCl₃) δ -5.5, 18.2, 25.8, 35.7, 41.0, 63.2, 71.5, 88.1, 94.4, 140.5, 156.4, 166.0. Treatment with Et₄NF (0.19 g, 1.26 mmol) followed by purification afforded 0.09 g (0.42 mmol, 94%) of the desired **52** as a low melting lyophilized solid: U.V. (H₂O) $\lambda_{max} = 272 \text{ nm}$ ($\epsilon = 9290$); [α]_D = -59.9° (c = 0.40, MeOH); ¹H NMR (DMSO-d₄) δ 1.58 (m, 1H), 2.36 (m, 1H), 2.47 (m, 1H), 3.40 (m, 2H), 3.82 (t, 1H, J = 7.8 Hz), 3.94 (t, 1H, J = 7.8 Hz), 4.73 (br s, 1H), 5.73 (d, 1H, J = 7.4 Hz), 5.92 (t, 1H, J = 6.4 Hz), 7.12 (br s, 2H), 7.58 (d, 1H, J = 7.4 Hz); ¹³C NMR (DMSO-d₄) δ 34.9, 40.9, 61.9, 70.8, 86.4, 93.8, 140.6, 155.1, 165.6. Anal Calcd. for C₉H₁₃N₃O₃: C, 51.18; H, 6.20; N, 19.89. Found C, 51.16; H, 6.23; N, 19.85.

5(R)-[4-Amino-2(1H)-pyrimidinone]tetrahydro-3(R)-furanmethanol (53). Identically, **49** (0.23 g, 0.70 mmol) in pyridine (5 mL) was treated with POCL, (0.13 mL, 1.4 mmol) and triazole (0.39 g, 5.59 mmol) in pyridine (15 mL) followed by NH OH (conc. 5 mL) to yield, after purification, 0.19 g (0.57 mmol, 81%) of 51: U.V. (MeOH) $\lambda_{\text{pax}} = 272 \text{ nm}$; ⁴H NMR (CDCl₃) δ -0.01 (s, 6H), 0.83 (s, 9H), 2.05 (ddd, 1H, J = 2.4, 7.8, and 13.7 Hz), 2.19 (m, 1H), 2.40 (m, 1H), 3.50 (dd, 1H, J = 7.1 and 10.0 Hz), 3.61 (dd, 1H, J = 5.2 and 10.0 Hz), 3.79 (t, 1H, J = 8.0 Hz), 4.18 (t, 1H, J = 7.9 Hz), 5.78 (d, 1H, J = 7.4 Hz), 5.98 (dd, 1H, J = 2.6 and 6.0 Hz), 7.38 (d, 1H, J = 7.4 Hz), ¹³C NMR (CDCl₃) δ -5.5, 18.2, 25.8, 35.8, 39.7, 63.3, 72.4, 88.2, 94.3, 140.0, 156.3, 166.0. Desilylation with Et₄NF (0.17 g, 1.14 mmol) in CH₄CN (10 mL) gave after purification 0.11 g (0.54 mmol, 95%) of **53**: M.P. (lyophilized powder) 194-196 °C; U.V. (H₂O) $\lambda_{\text{pax}} = 272 \text{ nm}$ ($\epsilon = 9980$); [α]_D = +39.64° (c = 0.47, MeOH); ¹H NMR (DMSO-d₄) δ 1.91 (m, 1H), 2.04 (m, 1H), 2.39 (m, 1H), 3.40 (m, 2H), 3.64 (t, 1H, J = 7.6 Hz), 4.19 (t, 1H, J = 7.7 Hz), 4.75 (t, 1H, J = 7.4 Hz); ¹³C NMR (DMSO-d₄) δ 3.191 (m, 1H), 2.04 (m, 1H), 2.39 (m, 1H), 3.40 (m, 2H), 3.64 (t, 1H, J = 7.6 Hz), 4.19 (t, 1H, J = 1.2 Hz), 7.54 (d, 1H, J = 7.4 Hz); ¹³C NMR (DMSO-d₄) δ 35.2, 40.7, 61.8, 71.5, 86.6, 93.5, 140.7, 155.1, 165.7. Anal Calcd. for C₉H₁₃N₃O₃: C, 51.18; H, 6.20; N, 19.89. Found C, 51.15; H, 6.22; N, 19.83.

5(S)-[5-Methyl-2,4(1H,3H)-pyrimidinedione]tetrahydro-3(R)-furanmethanol (56) and 5(R)-[5-Methyl-2,4(1H,3H)-pyrimidinedione]tetrahydro-3(R)-furanmethanol (57). Under the standard glycosylation conditions, thymine (0.49 g, 3.87 mmol) was treated with BSA (2.10 mL, 8.51 mmol), 33 (0.78 g, 3.87 mmol), and TMSOTf (0.82 mL, 4.25 mmol) to afford after purification by flash chromatography 0.88 g (3.29 mmol, 85%) of an inefficiently separable mixture of the anomeric [5'(S):5'(R) = 1:2] nucleosides 54 and 55. Deprotection with methanolic sodium methoxide (0.29 g, 5.31 mmol) afforded 0.68 g (2.99 mmol, 91%) of the anomeric mixture. Preparative TLC (Et₂O 78%, CHCl₃ 18%, MeOH 2%, and EtOH 2%) utilizing multiple elutions, dissection of the resulting band, and re-elution of the appropriately combined fractions provided pure 5'(S)-anomer 56: M.P. (lyophilized powder) 135-136 °C; U.V. (H₂O) $\lambda_{max} = 266$ nm ($\epsilon = 8940$); [α]_D = -17.5° (c = 0.56, MeOH); ¹H NMR (CDCL₃) 8 1.81 (m, 1H), 1.92 (s, 3H), 2.60 (m, 2H), 3.67 (dd, 1H, J = 5.0 and 10.4 Hz), 3.96 (t, 1H, J = 7.9 Hz), 4.10 (t, 1H, J = 8.1 Hz), 6.00 (t, 1H, J = 6.5 Hz), 7.26 (s, 1H), 8.20 (br s, 1H); ¹³C NMR (CDCL₃) 8 1.25, 34.7, 40.7, 63.0, 71.2, 12.

86.9, 110.7, 135.4, 150.5, 164.1. Anal. Calcd. for $C_{10}H_{14}N_2O_4$: C, 53.09; H, 6.24; N, 12.38. Found C, 52.99; H, 6.26; N, 12.33. And pure 5'(R)-anomer **57**: M.P. (lyophilized powder) 132-134 °C; U.V. (H₂O) $\lambda_{max} = 267$ nm ($\epsilon = 9470$); [α]_D = -2.24° (c = 0.45, MeOH); ¹H NMR (CDCl₃) δ 1.90 (s, 3H), 2.09 (ddd, 1H, J = 4.2, 7.9, and 13.8 Hz), 2.27 (m, 1H), 2.59 (m, 1H), 3.63 (dd, 1H, J = 6.8 and 10.6 Hz), 3.68 (dd, 1H, J = 6.5 and 8.8 Hz), 4.27 (dd, 1H, J = 7.1 and 8.8 Hz), 6.01 (dd, 1H, J = 4.2 and 6.4 Hz), 7.29 (s, 1H), 8.20 (br s, 1H); ¹³C NMR (CDCl₃) δ 12.5, 35.4, 40.0, 63.0, 72.1, 87.0, 110.7, 135.2, 150.6, 164.4. Anal. Calcd. for $C_{10}H_{14}N_2O_4$: C, 53.09; H, 6.24; N, 12.38. Found C, 52.99; H, 6.23; N, 12.34.

(+)- α (**R**)-allyl- γ (**S**)-[(((1,1-dimethylethyl)dimethylsilyl) α y)methyl]- γ -butyrolactone (61). LDA (1.59 mL, 2.39 mmol) was added dropwise to a solution of the lactone 60^{32} (0.5 g, 2.17 mmol) in THF (2 mL) at -98 °C. The solution was allowed to stir for 40 min and then allyl iodide (0.3 mL, 3.26 mmol) was rapidly added. After stirring for an additional 30 min the reaction was quenched with a saturated ammonium chloride and then diluted with EtOAc. The organics were then washed until neutral, dried (Na,SO₄), concentrated and purified by flash chromatography (hexanes to 3% EtOAc/hexanes) to afford 0.52 g (1.93 mmol, 89%) of 61: $[\alpha]_{D} = +18.6^{\circ}$ (c = 0.84, MeOH); ¹H NMR (CDCl₃) δ 0.04 (s, 6H), 0.86 (s, 9H), 2.00 (dt, 1H, J = 8.8, 8.9 and 12.9 Hz), 2.24 (m, 2H), 2.55 (dd, 1H, 4.5, 6.4 and 14.4 Hz), 2.80 (dd, 1H, J = 4.4, 9.3 and 18.4 Hz), 3.63 (dd, 1H, J = 2.8 and 11.2 Hz), 3.80 (dd, 1H, J = 3.3 and 11.2 Hz), 4.49 (m, 1H), 5.09 (m. 2H), 5.73 (m, 1H); ¹³C NMR (CDCl₃) δ -5.6, 18.2, 25.8, 29.3, 35.3, 39.1, 65.1, 77.2, 117.6, 134.6, 179.1. Anal. Calcd. for C₁₄H₂₆O₃S: C, 62.18; H, 9.69. Found: 62.04; H, 9.72.

Compound **62** [S,S (+) isomer] was the minor product [0.03 g, 0.11 mmol, 5%]: $[\alpha]_{D} = +53.5^{\circ}$ (c = 0.99, MeOH); ¹H NMR (CDCl₂) δ 0.05 (s, 6H), 0.86 (s, 9H), 1.88 (m, 1H), 2.25 (m, 2H), 2.06 (m, 2H), 3.67 (dd, 1H, J = 3.9 and 11.4 Hz), 3.83 (dd, 1H, J = 3.5 and 11.4 Hz), 4.42 (m, 1H), 5.08 (m, 2H), 5.74 (m, 1H); ¹³C NMR (CDCl₃) δ -5.5, 18.1, 25.7, 28.8, 34.4, 39.9, 63.8, 78.4, 117.2, 134.5, 177.9. Anal. Calcd. for $C_{14}H_{26}O_{3}S$: C, 62.18; H, 9.69. Found: 62.07; H, 9.70.

Tetrahydro-5(R and S)-hydroxy-3(R)-allylfuran (64). A solution of **61** (0.21 g, 0.77 mmol) in Et₂O (5 mL) was added dropwise to LiAlH₄ (0.04 g, 1.16 mmol) in Et₂O (5 mL). The resulting mixture was stirred for 1 h and then wet ether, followed by water, was added. The mixture was then filtered through Celite, dried (Na₂SO₄) and concentrated. Desilylation with Et₄NF (0.16 g, 1.04 mmol) provided, after purification by flash chromatography (CHCl₃ to 10% MeOH/CHCl₃), 0.12 g (0.75 mmol, 98%) of 4(R)-hydroxymethyl-6-hepten-1,2(S)-diol (**63**): [α]_D = -18.9° (c = 0.82, MeOH); ¹H NMR (CDCl₃) δ 1.43 (m, 2H), 1.79 (m, 1H), 2.04 (m, 2H), 3.50 (m, 4H), 3.79 (m, 1H), 4.43 (br s, 3H), 5.00 (m, 2H), 5.73 (m, 1H); ¹³C NMR (CDCl₃) δ 35.0, 36.1, 36.5, 65.1, 66.7, 69.5, 116.6, 136.5.

Sodium periodate (0.52 g, 2.43 mmol) was added to a solution of **63** (0.26 g, 1.62 mmol) in Et₂O (10 mL) and water (4 mL). The resulting heterogeneous solution was stirred for 1 h and then filtered through Celite, dried (Na₂SO₄), concentrated and purified by flash chromatography (CHCl₃ to 5% MeOH/CHCl₃) to afford 0.18 g (1.41 mmol, 87%) of **64**; $[\alpha]_{D} = +45.5^{\circ}$ (c = 0.91, MeOH); ¹H NMR (CDCl₃) δ 1.42 (m, 1H), 2.10 (m, 3H), 2.43 (m, 1H), 3.37 (t, H-2 α , J = 7.5 Hz), 3.52 (t, H-2 β , J = 7.9 Hz), 3.83 (t, H-2 β , J = 8.0), 4.01 (t, H-2 α , J = 7.8 Hz), 4.66 (br s, 1H), 5.00 (m, 2H), 5.40 (d, 1H, J = 4.9 Hz), 5.75 (m, 1H); ¹³C NMR (CDCl₃) δ 36.1, 36.4, 37.4, 37.8, 39.2, 71.4, 71.7, 98.3, 98.7, 115.6, 115.8, 136.3, 136.5. Anal. Calcd. for C₇H₁₂O₂: C, 65.59; H, 9.44. Found: C, 65.66; H, 9.47.

Tetrahydro-5(R and S)-acetyloxy-3(R)-(2-benzoyloxyethyl)furan (68). Anhydrous methanolic HC1 (0.47 mL, 0.47 mmol) was added to a solution of **64** (1.20 g, 9.39 mmol) in MeOH (5 mL). The mixture was stirred for 20 min and then neutralized with Dowex-OH. The mixture was filtered, concentrated and then briefly dried under vacuum to yield 1.09 g (7.7 mmol, 82%) of the volatile methyl furanoside: $[\alpha]_{\rm p}$ = +17.4° (c = 0.38, MeOH); ¹H NMR (CDCl₃) δ 1.5 - 2.43 (m, 5H), 3.26 (s, 3H), 3.45 (m, 1H), 3.91 (m, 1H), 4.93 (m, 3H), 5.67 (m, 1H); ¹³C NMR (CDCl₃) δ 36.2, 36.3, 37.5, 38.0, 38.4, 38.5, 54.4, 54.7, 71.4, 71.6, 105.1, 105.5, 115.9, 136.4, 136.7. A freshly prepared solution of the methyl allylfuranoside (0.32 g, 2.25 mmol) in MeOH (20 mL) was cooled to -78 °C and ozonized (3 min), treated with dimethyl sulfide (3 mL) and allowed to stir for 3 h. The solution was then concentrated, redissolved in ethanol (10 mL) and treated with NaBH₄ (0.13 g, 3.37 mmol). The resulting solution was stirred for 1 h and then neutralized with 1 N HCl. The solution was evaporated to dryness, dissolved in CHCl₄ and filtered. The organics were concentrated and purified by flash chromatography (CHCl₄ to 5% MeOH/CHCl₄) to afford 0.30 g (2.07 mmol, 92%) of tetrahydro-5(R and S)-methoxy-3(R)-(2-hydroxyethyl)furan (65): $[\alpha]_{\rm p} = +28^{\circ}$ (c = 1.39, MeOH); ¹H NMR (CDCl₄) δ 1.59 (m, 2H), 1.65 (m, 1H), 2.01 (m, 1H), 2.26 (m, 1H), 3.27 (s, OCH₄ β), 3.29 (s, OCH₄ α), 3.48 (m, 1H), 3.59 (m, 2H), 4.02 (m, 1H), 4.96 (m, 1H); ¹³C NMR (CDCl₄) δ 3.3.9, 35.1, 35.9, 36.5, 38.7, 39.0, 54.5, 54.8, 61.5, 71.7, 72.4, 105.0, 105.5.

Benzoyl chloride (0.16 mL, 1.35 mmol) was added dropwise, at 0 °C, to a solution of **65** (0.16 g, 1.12 mmol) and Et₃N (0.31 mL, 2.24 mmol) in CH₂Cl₂ (3 mL). The resulting solution was stirred for 1 h at room temperature and then diluted with EtOAc and the organics were washed until neutral. Purification by flash chromatography afforded 0.24 g (0.96 mmol, 86%) of tetrahydro-5(R and S)-methoxy-3(R)-(2-benzoyloxyethyl)furan (**66**): $[\alpha]_{p} = +24.4^{\circ}$ (c = 0.98, MeOH); ¹H NMR (CDCl₃) δ 1.64 (m, 1H), 1.85 (m, 2H), 2.10 (m, 1H), 2.45 (m, 1H), 3.30 (s, OCH₃ α), 3.32 (s, OCH₃ β), 3.56 (m, 1H), 4.10 (t, 1H, J = 8.0 Hz), 4.30 (m, 2H), 4.99 (m, 1H), 7.4 (t, 2H, J = 7.5 Hz), 7.52 (t, 1H, J = 7.9 Hz), 8.0 (d, 2H, J = 7.5 Hz); ¹³C NMR (CDCl₃) δ 32.0, 32.6, 34.2, 35.6, 38.7, 39.0, 54.3, 54.7, 63.9, 71.4, 72.2, 104.9, 105.3, 128.2, 129.4, 130.0, 132.8, 166.3. A solution of **66** (0.24 g, 0.96 mmol) in dioxane (10 mL) and aqueous HCl (2.4 mL, 0.24 mmol) was stirred at room temperature for 16 h. The solution was then neutralized, concentrated, extracted with EtOAc, and purified by flash chromatography (hexanes to 30 % EtOAc/hexanes) to afford 0.19 g (0.80 mmol, 83%) of tetrahydro-5(R and S)-hydroxy-3(R)-(2-benzoyloxyethyl)furan (**67**): $[\alpha]_{p} = +8.4^{\circ}$ (c = 1.66, MeOH); ¹H NMR (CDCl₄) δ 1.63 (m, 1H), 1.95 (m, 2H), 2.34 (m, 1H), 2.63 (m, 1H), 3.53 (t, 1H, J = 7.9 Hz), 4.22 (t, 1H, J = 7.9 Hz), 4.33 (m, 2H), 5.54 (m, 1H), 7.41 (t, 2H, J = 7.5 Hz), 7.53 (t, 1H, J = 7.3 Hz), 8.0 (d, 2H, J = 7.3 Hz); ¹³C NMR (CDCl₄) δ 32.0, 32.3, 34.2, 35.4, 34.2, 35.4, 39.7, 39.9, 63.9, 63.9, 64.0, 71.9, 72.7, 98.5, 98.9, 128.4, 129.5, 130.1, 133, 166.5.

Acetic anhydride (0.16 mL, 1.64 mmol) was added to a solution of **67** (0.32 g, 1.37 mmol), DMAP (0.04 g, 0.34 mmol) and Et₃N (0.29 mL, 2.0 mmol) in acetonitrile (2 mL). After stirring for 1 h MeOH (3 mL) was added and the solution was concentrated, extracted (EtOAc / H₂O) and purified by flash chromatography (hexanes to 20% EtOAc/hexanes) to afford 0.30 g (1.08 mmol, 79%) of **68**: $[\alpha]_{p} = +10.1^{\circ}$ (c = 1.40, MeOH); ¹H NMR (CDCL₃) δ 1.85 (m, 2H), 1.98 (s, C(O)CH₃ α), 2.0 (s, C(O)CH₃ β), 2.19 (dd, 1H, J = 7.1 and 13.2 Hz), 2.42 (m, 1H), 2.61 (m, 1H), 3.59 (t, 1H, J = 8.1 Hz), 4.21 (t, 1H, J = 8.1 Hz), 4.32 (m, 2H), 6.25 (d, 1H, J = 4.7 Hz), 7.40 (t, 2H, J = 7.5 Hz), 7.53 (t, 1H, J = 7.4 Hz), 7.98 (d, 2H, J = 7.4 Hz); ¹³C NMR (CDCL₃) δ 21.2, 32.0, 32.1, 34.1, 38.2, 38.7, 63.8, 73.5, 73.9, 98.8, 99.0, 128.4, 129.5, 130.0, 133.0, 166.4, 170.3. Anal. Calcd. for C₁₅H₁₈O₅: C, 64.74; H, 6.52. Found: C, 64.98; H, 6.54.

5(R)-(6-Amino-9H-purin-9-yl)tetrahydro-3(R)-furanethanol (69) and 5(S)-(6-Amino-9H-purin-9-yl)tetrahydro-3(R)-furanethanol (70). Under the standard glycosylation conditions N⁶-benzoyladenine (0.36 g, 1.50 mmol), persilylated with BSA (0.62 mL, 2.51 mmol) in CH,CN (3 mL), was treated with the lactol 68 (0.35 g, 1.25 mmol), in CH₃CN (6 mL), and TMSOTF (0.27 mL, 1.38 mmol) to afford 0.20 g (0.44 mmol) in MeOH (2 mL) and sodium methoxide (0.2 g, 0.35 mmol) was stirred for 12 h and then neutralized with 0.1 N HCl. The solution was evaporated to dryness and the trans-5'(S)-3'(R) 70 was isolated by fractional crystalization from MeOH (0.02 g, 0.08 mmol, 56%). The filtrate was purified by preparative layer chromatography (10% MeOH/CHCl₃) to afford 0.01 g (0.04 mmol, 32%) of the cis-5'(R)-3'(R) 69: M.P. (lyophilized powder) 118-120 °C; U.V. (H₂O) $\lambda_{max} = 259$ nm ($\varepsilon = 13,420$); [α]_D = -55.6° (c = 0.45, MeOH); ¹H NMR (DMSO-d₀) δ 1.68 (m, 2H), 2.30 (m, 1H), 2.47 (m, 1H), 2.58 (m, 1H), 3.45 (m, 2H), 3.78 (t, 1H, J = 8.5 Hz), 4.03 (t, 1H, J = 6.9 Hz), 4.53 (br s, 1H), 6.20 (t, 1H, J = 6.3 Hz), 7.24 (br s, 2H), 8.15 (s, 1H), 8.32 (s, 1H); ¹³C NMR (DMSO-d₀) δ 34.2, 36.9, 37.0, 59.8, 73.1, 84.2, 119.3, 139.4, 149.1, 152.4, 156.0: Anal. Calcd. for C₁₁H₁₅N₅O₂: C, 53.00; H, 6.07; N, 28.1. Found C, 52.87; H, 6.09; N, 27.96.

Data for **70**: M.P. (H₂O) 248-249 °C; U.V. (H₂O) $\lambda_{max} = 259$ ($\epsilon = 13,700$); [α]_D = +23° (C = 0.2, H₂O); ¹H NMR (DMSO-d_c) δ 1.59 (m, 2H), 2.11 (m, 1H), 2.55 (m, 1H), 2.65 (m, 1H), 3.44 (m, 2H), 3.56 (t, 1H, J = 7.7 Hz), 4.28 (t, 1H, J = 7.8 Hz), 4.52 (br s, 1H), 6.25 (dd, 1H, J = 2.4 and 7.1 Hz), 7.23 (br s, 2H), 8.13 (s, 1H), 8.20 (s, 1H); ¹³C NMR (DMSO-d_c) δ 34.6, 35.5, 37.7, 59.6, 73.8, 84.2, 119.1, 138.9, 148.9, 152.4, 155.8. Anal. Calcd. for C₁₁H₁₅N₅O₂.1/2H₂O: C, 51.15; H, 6.24; N, 27.12. Found C, 51.28; H, 6.26; N, 27.01.

(+)- $\alpha(S)$ -allyl- $\gamma(S)$ -[(((1,1-dimethylethyl)dimethylsilyl)oxy)methyl]- γ -butyrolactone (62). LDA (3.54 mL, 5.31 mmol) was added to a solution of predominantly the lactone 61 (1.43 g, 5.30 mmol) in THF (3.5 mL) at -78 °C. After stirring for 1 h the solution was quenched with a saturated ammonium chloride solution, extracted with EtOAc, and purified by column chromatography (hexanes to 3% EtOAc/hexanes) to afford 0.19 g (0.69 mmol, 13%) of the $\alpha(R)$ -allyl-61 and 0.97 g (3.60 mmol, 68%) of the $\alpha(S)$ -allyl-62: for physical data see experimental for 61.

Tetrahydro-5(R and S)-hydroxy-3(S)-allylfuran (71). Identical to the method described for the conversion of **61** to **64**, 1.39 g (5.06 mmol) of **62** was treated with LiAlH₄ (0.29 g, 7.59 mmol) in Et₂O (20 mL) followed by Et₄NF (1.51 g, 10.12 mmol) in CH₃CN (20 mL) to afford after purification 0.73 g (4.54 mmol, 90%) of the 4(S)-hydroxymethyl-6-heptene-1,2(S)-diol: $[\alpha]_{\rm D} = -17.5^{\circ}$ (c = 1.58, MeOH); ¹H NMR (CDCl₄) δ 1.29 (m,

2H), 1.72 (m, 1H), 2.0 (m, 2H), 3.29 (dd, 1H, J = 7.5 and 14.6 Hz), 3.27 (dd, 1H, J = 7.8 and 12.1 Hz), 3.52 (dd, 1H, J = 3.2 and 11.2 Hz), 3.61 (dd, 1H, J = 3.9 and 11.0 Hz), 3.69 (m, 1H), 4.45 (br s, 3H), 4.98 (m, 2H), 5.72 (m, 1H); 13 C NMR (CDCl₃) δ 36.3, 37.1, 39.1, 66.2, 67.3, 71.6, 116.4, 136.6. The above product (0.81 g, 5.05 mmol) in Et₂O (40 mL) and water (5 mL) was then treated with sodium periodate (1.62 g, 7.57 mmol) to afford after purification 0.54 g (4.19 mmol, 83%) of 71: [α]_D = -43.7° (c = 1.01, MeOH); ¹H NMR (CDCl₃) δ 1.57 (m, 1H), 2.14 (m, 3H), 2.42 (m, 1H), 3.46 (m, 1H), 3.95 (m, 1H), 4.99 (m, 2H), 5.35 (m, 1H), 5.72 (m, 1H); ¹³C NMR (CDCl₄) δ 36.3, 36.4, 37.4, 37.9, 38.7, 71.7, 71.9, 98.5, 98.8, 115.7, 116.0, 136.5, 136.6. Anal. Calcd. for C₂H₂O₅: C, 65.59; H, 9.44. Found: C, 65.85; H, 9.48.

Tetrahydro-5(R and S)-acetyloxy-3(S)-(2-benzoyloxyethyl)furan (74). Using the method described for 65, compound 71 (0.72 g, 5.58 mmol) in MeOH (3 mL) was treated with anhydrous methanolic HCl (0.28 mL, 0.28 mmol). After neutralization with Dowex-OH the filtrate was ozonized (-78 °C), treated with DMS (3 mL) and allowed to stir for 3 h. The volatiles were then removed under reduced pressure and the remaining oil was redissolved in EtOH (5 mL) cooled to 0 °C and treated with NaBH₄ (0.32 g, 8.37 mmol) to afford after purification 0.43 g (2.96 mmol, 53%) of tetrahydro-5(R and S)-methoxy-3(S)-(2-hydroxyethyl)furan (72): $[\alpha]_{D} = -27.8^{\circ}$ (c = 1.50, MeOH); ¹H NMR (CDCl₃) δ 1.57 (m, 2H), 1.64 (m, 1H), 2.00 (dd, 1H, J = 7.5 and 12.8 Hz), 2.25 (m, 1H), 3.26 (s, OCH₄ β), 3.28 (s, OCH₄ α), 3.46 (m, 1H), 3.56 (m, 2H), 4.00 (m, 1H), 4.95 (m, 1H); ¹³C NMR (CDCl₃) δ 33.8, 35.2, 35.9, 36.5, 38.7, 39.0, 54.5, 54.8, 61.5, 71.7, 72.4, 105.0, 105.5.

Using the method described for the preparation of **66**, 0.43 g (2.96 mmol) of **72** was treated with Et_xN (0.83 mL, 5.92 mmol) and benzoyl chloride (0.52 mL, 4.44 mmol) in CH_xCl₂ (3 mL) to afford after purification 0.56 g (2.25 mmol, 76%) of tetrahydro-5(R and S)-methoxy-3(S)-(2-benzoyloxyethyl)furan (73): $[\alpha]_{\rm p} = -23.8^{\circ}$ (c = 1.14, MeOH); ¹H NMR (CDCl₂) δ 1.60 (m, 1H), 1.82 (m, 2H), 2.05 (m, 1H), 2.40 (m, 1H), 3.26 (s, OCH₄B), 3.29 (s, OCH₄Q), 3.52 (m, 1H), 4.08 (t, 1H, J = 8.0 Hz), 4.26 (m, 1H), 4.96 (m, 1H), 7.37 (t, 2H, J = 7.5 Hz), 7.49 (t, 1H, J = 7.4 Hz), 7.96 (d, 2H, J = 7.5 Hz); ¹³C NMR (CDCl₂) δ 32.1, 32.7, 34.4, 35.7, 38.9, 39.2, 54.5, 54.9, 63.9, 64.0, 71.6, 72.3, 105.1, 105.5, 128.3, 129.5, 130.5, 132.9, 166.5.

As described for **68**, 0.39 g (1.57 mmol) of **73** was treated with 3.92 mL of 0.1 N HCL in dioxane (14 mL) to afford, after purification, 0.37 g (1.57 mmol) of the demethylated lactol: $[\alpha]_{p} = -8.0^{\circ}$ (c = 1.41, MeOH), ¹H NMR (CDCL₃) δ 1.58 (m, 1H), 1.89 (m, 2H), 2.28 (m, 1H), 2.58 (m, 1H), 3.46 (m, 1H), 4.16 (m, 1H), 4.26 (m, 2H), 5.47 (m, 1H), 7.36 (t, 1H, J = 7.4 Hz), 7.48 (t, 1H, J = 7.3 Hz), 7.94 (d, 2H, J = 7.4 Hz); ¹³C NMR (CDCL₃) δ 31.7, 32.1, 34.1, 35.9, 39.5, 39.7, 63.5, 63.8, 71.5, 72.3, 98.2, 98.6, 128.2, 129.3, 129.9, 132.8, 166.3 Treatment with DMAP (0.05 g, 0.39 mmol), Et₄N (0.33 mL, 2.35 mmol), and Ac₂O (0.18 mL, 1.88 mmol) in CH₄CN (2 mL) afforded after purification 0.34 g (1.22 mmol, 78%) of **74**: $[\alpha]_{p} = -10.4^{\circ}$ (c = 2.26, MeOH); ¹H NMR (CDCl₃) δ 1.86 (m, 2H), 1.99 (s, 3H), 2.17 (m, 1H), 2.40 (m, 1H), 2.61 (m, 1H), 3.58 (m, 1H), 4.21 (m, 1H), 4.31 (m, 2H), 6.24 (d, 1H, J = 4.3 Hz), 7.40 (t, 2H, J = 7.5 Hz), 7.52 (t, 1H, J = 7.2 Hz), 7.99 (d, 2H, J = 7.5 Hz); ¹³C NMR (CDCl₃) δ 21.2, 31.9, 32.0, 34.1, 38.1, 38.6, 63.7, 73.4, 73.9, 98.7, 98.9, 128.3, 129.4, 129.9, 132.9, 166.3, 170.3. Anal. Calcd. for C₁₅H₁₈O₅: C, 64.74; H, 6.52. Found: C, 64.94; H, 6.53.

5(S)-(6-Amino-9H-purin-9-yl)tetrahydro-3(S)-furanethanol (75) and **5(R)-(6-Amino-9H-purin-9-yl)tetrahydro-3(S)-furanethanol** (76). Identical to the method described for 69 and 70, 74 (0.34 g, 1.22 mmol) followed by TMSOTf (0.26 mL, 1.34 mmol) were added to a solution of N⁶-benzoyladenine (0.35 g, 1.46 mmol), persilylated with BSA (0.6 mL, 2.43 mmol) in CH₃CN (3 mL). After work-up and purification 0.20 g (0.43 mmol, 35%) of the dibenzoylated anomeric mixture was isolated. Sodium methoxide (0.07 g, 1.29 mmol) deprotection in MeOH (5 mL) afforded after purification 0.027 g (0.11 mmol, 25%) of the cis-5'(R)-3'(R)-isomer, 75: M.P. (lyophilized powder) 119-121 °C; U.V. (H₂O) λ_{max} = 260 nm (ε = 13,100); [α]_p = +54.3° (c = 0.56, MeOH); ¹H NMR (DMSO-d₂) δ 1.68 (m, 2H), 2.32 (m, 1H), 2.46 (m, 1H), 2.59 (m, 1H), 3.46 (m, 2H), 3.79 (t, 1H, J = 8.7 Hz), 4.03 (t, 1H, 7.14 Hz), 4.52 (br s, 1H), 6.20 (t, 1H, J = 6.54 Hz), 7.24 (br s, 2H), 8.14 (s, 1H), 8.32 (s, 1H); ¹³C NMR (DMSO-d₂) δ 34.2, 36.9, 37.0, 59.8, 73.1, 84.2, 119.3, 139.4, 149.1, 152.4, 156.0; Anal. Calcd. for C₁₁H₁₅N₂O₂: C, 53.00; H, 6.07; N, 28.1: Found : C, 52.90; H, 6.09; N, 27.99: and 0.07 g (0.28 mmol, 65%) of 76: M.P. (H₂O) 247-249 °C; U.V. (H₂O) λ_{max} = 260 nm (ε = 13,500); [α]_p = -22.3° (c = 0.28, H₂O); ¹H NMR (DMSO-d₂) δ 1.59 (m, 2H), 2.11 (dt, 1H, J = 7.4, 8.2 and 13.2 Hz), 2.55 (m, 1H), 2.64 (m, 1H), 3.43 (m, 2H), 3.55 (t, 1H, J = 7.8 Hz), 4.28 (t, 1H, J = 7.6 Hz), 4.51 (br s, 1H), 6.25 (dd, 1H, J = 2.5 and 7.0 Hz), 7.22 (br s, 2H), 8.13 (s, 1H), 8.21 (s, 1H); ¹³C NMR (DMSO-d₂) δ 34.6, 35.4, 37.7, 59.6, 73.8, 84.2, 119.1, 138.8, 148.8, 152.4, 155.9. Anal. Calcd. for C₁₁H₁₅N₅O₂.1/2H₂O: C, 51.15; H, 6.24; N, 27.12. Found C, 51.23; H, 6.26; N, 27.05. Acknowledgment: Support of this work by the National Institutes of Health is gratefully acknowledged. We thank the University of Iowa for a Faculty Scholar Award to V.N. and the University of Iowa Biocatalysis Center for a Biocatalysis Fellowship to T. B. S.

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