The design and synthesis of antibody binding site probes: three pentasaccharide analogues of the *Brucella* A antigen prepared by activation *in situ* of thioglycosides with bromine*

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ABSTRACT

Three pentasaccharide analogues of the *Brucella* A antigen $[\rightarrow 2)$ - α -D-Rhap4NFo- $(1\rightarrow)_{n}$, each with one formamido group replaced by a hydroxyl group, have been prepared as their methyl glycosides. Monoand di-saccharide thioglycosides of D-rhamnose and 4-azido-4,6-dideoxy-D-mannose were used as glycosyl donors for the preparation of protected pentasaccharide derivatives with trisaccharides as intermediates. Glycosylations were performed by activation *in situ* of the thioglycosides with bromine in the presence of a glycosyl acceptor and silver triflate as promoter. Reduction of the azido groups with hydrogen sulfide, *N*-formylation with ethyl formate, and hydrogenolysis then gave the target pentasaccharides.

INTRODUCTION

The Brucella A and M antigens are linear homopolymers of the rare sugar 4,6-dideoxy-4-formamido-D-mannose^{1,2}. The A antigen contains only α 1,2-glycosidic linkages¹ whereas, for the M antigen, one linkage in five is an α 1,3-linkage². The A antigen, which is found predominantly in Brucella abortus, is selectively bound by monoclonal antibody YsT9-1 (ref. 3). The formamido groups of the antigen have been shown to be essential for the formation of the antibody-antigen complex⁴, a fact which can be rationalized from studies^{2,5} that revealed a helical conformation for the A-antigen, with the formamido groups protruding from the axis of the helix. Viewed along the helical axis, the angle subtended by the formamido groups of adjacent mono-saccharide residues is 216° (Fig. 1). In this conformation the three dimensional repeating unit for the antigen contains five residues⁵.

A model⁶ of the complex between the *Brucella* A antigen and the F_v fragment of monoclonal antibody YsT9-1 has been developed using computer-assisted modelling techniques, in parallel with ongoing X-ray crystallographic studies⁴ of the complex. The model⁶ indicates that at least a pentasaccharide moiety of the *Brucella* A antigen is accomodated in the groove-type binding site of the antibody, in agreement with

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Fig. 1. Proposed interactions in the model⁶ of the complex between the *Brucella* A antigen and monoclonal antibody YsT9-1 (left) and orientation of the formamido groups when the antigen is viewed along its helical axis in the minimum-energy conformation (right).

previous inhibition studies^{3,4} using synthetic di- to penta-saccharides^{7,9}. In the modelled complex⁶ the formamido groups of residues a and e of a properly docked pentasaccharide are within hydrogen-bonding distance of the guanidino groups of two arginine residues, which are located symmetrically at either end of the binding site (Fig. 1). The formamido group of residue c is then located in a pocket of the binding site, whereas formamido group b is in contact with the surrounding solution.

In order to evaluate this model of the complex, we have prepared the three pentasaccharide analogues 25, 30, and 35 of the *Brucella* A antigen, each of which has one 4,6-dideoxy-4-formamido-D-mannose residue (a, b, or c) replaced by D-rhamnose, thus substituting a formamido group by a hydroxyl group.

RESULTS AND DISCUSSION

Regioselective benzylation of methyl 4-azido-4,6-dideoxy- α -D-mannopyranoside^{7.8}, by a modification of the procedure⁸ employing an *O*-stannylene acetal as intermediate, gave the 3-*O*-benzyl ether 1 (refs. 7 and 8) (89%) which was then converted to the ethyl thioglycosides 2 (ref. 9) and 3 (ref. 9) as previously described. The rhamnose thioglycoside 5 was obtained (64%), together with the β -anomer 7 (26%), by treatment of 1,2-di-*O*-acetyl-3,4-di-*O*-benzyl- α -D-rhamnopyranose (4) in dichloromethane with ethanethiol and boron trifluoride etherate. Similar amounts of ethylthio α - and β -L-rhamnopyranosides have been obtained^{10,11} by reaction of tetra-*O*-acetyl-Lrhamnopyranose with ethanethiol under catalysis by Lewis acids. It should also be noted that boron trifluoride etherate and ethanethiol have been found to cause *O*debenzylation of benzylated carbohydrates under more drastic reaction conditions¹². Deacetylation of 5 gave the alcohol 6 (97%).

The α 1,2-linked disaccharide thioglycosides 8 (ref. 9), 9, and 10 were envisaged as



1 $R^{1} = OMe, R^{2} = H$ 2 $R^{1} = SEt, R^{2} = Ac$ 3 $R^{1} = SEt, R^{2} = H$



	Me	OA c	
BziO —	1	-10	
BzЮ	~		SEI

7

4 $R^{1} = OAc, R^{2} = Ac$ 5 $R^{1} = SEt, R^{2} = Ac$ 6 $R^{1} = SEt, R^{2} = H$





11

8 $R^1 = R^2 = N_3$ 9 $R^1 = N_3$, $R^2 = BzIO$ 10 $R^1 = BzIO$, $R^2 = N_3$





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glycosyl donors in a blockwise synthetic approach to the target pentasaccharides 25, 30, and 35. Disaccharide 8 is composed of two 4-azido-4,6-dideoxy-D-mannose residues and was used in the recent synthesis⁹ of tri- to penta-saccharide fragments of the *Brucella* A antigen. Disaccharide 9 has a D-rhamnose unit linked to O-2 of a "reducing" 4-azido-4,6-dideoxy-D-mannose residue, whereas 10 has these two residues in the reverse order.

Disaccharide 8 was prepared as previously reported by conversion of ethyl 2-O-acetyl-4-azido-3-O-benzyl-4,6-dideoxy-1-thio- α -D-mannopyranoside (2) into the glycosyl bromide and glycosylation of the alcohol 3 (75% yield when promoted by silver triflate)⁹. Treatment of the D-rhamnose thioglycoside 5 with bromine, and glycosylation of the 4-azido-4,6-dideoxy-D-mannoside 3 (ref. 9) using mercuric cyanide and mercuric bromide as promoters gave the α - and β -linked disaccharides 9 and 11 in 62 and 20% yields, respectively. When silver triflate was used as a promoter, in combination with 2,4,6-trimethylpyridine, 9 was formed in a lower yield and was contaminated by several



by-products. Surprisingly, attempts to prepare the isomeric disaccharide 10, by glycosylation of the rhamnoside 6 with the glycosyl bromide prepared from the 4-azido-4,6dideoxy-D-mannoside 2 (ref. 9), resulted in too low a yield ($\sim 17\%$) for further synthetic purposes, when mercuric cyanide and mercuric bromide or silver triflate were used as promoters. The thioglycoside 2 was one of several by-products formed in this glycosylation, presumably by attack of the glycosyl bromide generated from 2 on the ethylthio group of the glycosyl acceptor 6 (both n.m.r. spectroscopy and t.l.c. analysis showed complete conversion of 2 into the glycosyl bromide on treatment with bromine). The difference in outcome of the glycosylation giving 8, 9, and 10 was not expected, considering the close structural similarity of these disaccharides. We also note the previously reported^{9,13} empirical observation that less than one molar equivalent of bromine is required to quantitatively convert thioglycosides, such as 2 and 5, into their corresponding glycosyl bromides.

The stereochemistry of the newly formed *O*-glycosidic linkage in the disaccharides 9, 10, and 11 was determined from the ${}^{1}J_{C-1,H-1}$ coupling constant, which has been found¹⁴ to be *ca*. 170 Hz for α -glycosides and *ca*. 160 Hz for β -glycosides. The stereochemistry of the products from the glycosylations described below were determined in the same manner.

We recently reported¹³ an improved method for glycosylations by activation *in* situ of thioglycosides with bromine in the presence of a glycosyl acceptor and silver triflate as promoter. Application of this method to the glycosylation of methyl 3,4-di-O-benzyl- α -D-rhamnopyranoside with the disaccharide donor 8 (ref. 9) was found¹³ to give the trisaccharide 12 (68%) and its corresponding β -glycoside (14%). Activation *in situ* of thioglycosides with bromine avoids the use of toxic promoters such as methyl triflate in glycosylations, and provides 1,2-*trans*- or 1,2-*cis*-O-glycosides in excellent yields and with high stereochemical control¹³. Thus, glycosylation of methyl 4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside^{7,8} (1) by activation of the disaccharide thioglycoside 9 with bromine and promotion by silver triflate gave the α - and β -linked trisaccharides 14 and 16 in 76 and 7.2% yields, respectively.

Due to the low yield obtained in the synthesis of the disaccharide donor 10, the trisaccharide 19 was instead prepared from 1 (refs. 7 and 8) by a stepwise approach. Glycosylation of the alcohol 1 by activation *in situ* of the D-rhamnose thioglycoside 5 with bromine was found to give 17 in a significantly higher yield when performed at -45° instead of at room temperature (93 and 59% yields, respectively). Deacetylation of 17 (\rightarrow 18, 93%) and glycosylation with 2 (ref. 9) gave the trisaccharide 19 (87%, at -45°).

Deacetylation of the trisaccharides 12 (ref. 13), 14, and 19 then gave the glycosyl acceptors 13, 15, and 20 ($\sim 90\%$ yields) which were used in the syntheses of penta-saccharides 21, 26, and 31.

Condensation of the disaccharide donor 8 with the trisaccharide alcohol 13 by activation¹³ in situ with bromine and promotion by silver triflate gave the pentasaccharide 21 (79%), which was subsequently deacetylated to give 22 (90%). Glycosylation of the alcohols 20 and 15 with 8, in the same manner, gave pentasaccharides 26 and 31. Compounds 26 and 31 contained small amounts of impurities (5–10%), but gave pure 27 and 32 on deacetylation (79 and 60% from 20 and 15, respectively). Pentasaccharides 21 and 32 could also be prepared by glycosylation of 13 and 15 with 8 using methylsulfenyl triflate¹⁵ as promoter in 81 and 59% yields, respectively (*i.e.*, in yields almost identical to those obtained above). It is noteworthy that, in contrast to the syntheses of the trisaccharides 12 and 14, no β -linked products were obtained in the syntheses of pentsaccharides 21, 27, and 32, even though the disaccharide donor 8 has a non-participating glycosyl residue at O-2.

The pentasaccharides 22, 27, and 32 were deprotected in three steps. The azido groups were reduced with hydrogen sulfide^{9,16}, giving the amines 23, 28, and 33 which were directly converted into the formamides 24, 29, and 34 using refluxing ethyl formate containing a catalytic amount of pyridine (64–74% overall yields). Reduction of the azido groups with hydrogen sulfide was found to give higher yields in triethylamine– pyridine than in pyridine–water⁹, whereas attempted reduction with triphenylphosphine (Ph₃P)¹⁷, lithium aluminum hydride¹⁸, or a tin(II) complex¹⁹ gave either low yields (Ph₃P) or were unsuccessful. *N*-Formylation in the higher boiling point ethyl formate, instead of in methyl formate, permitted significantly shorter reaction times. Hydrogenolysis in formic acid of the benzyl ethers in 24, 29, and 34, gave complex mixtures of *O*-formates which were converted to the target pentasaccharides 25, 30, and 35 by treatment with methanolic sodium methoxide (81–89% overall yields).

A detailed analysis of the n.m.r. spectra of the O-benzylated formamides 24, 29, and 34 was not possible due to population of both rotamers for the formamido groups^{5,20}. The structures for 24, 29, and 34 were, therefore, further confirmed through chemical transformations. That complete reduction of the azido groups in 22, 27, and 32 had been achieved with hydrogen sulfide was confirmed by acetylation of the resulting amines 23, 28, and 33 to give acetamides which displayed four N-acetates each [δ (CH₃CONH): 1.93–1.86], as well as four AcNHC-4 resonances ($\delta \sim 52$). Acetylation of the formamides 24, 29, and 34 confirmed that these were indeed tetra-N-formylated as each derivative displayed only a single O-acetyl resonance [δ (CH₃CO₂): ~2.07]. Although complicated, the ¹H- and ¹³C-n.m.r. spectra of the deprotected pentasaccharides 25, 30, and 35 could be satisfactorily analysed and were consistent with the assigned structures.

The binding of the pentasaccharides 25, 30, and 35 by monoclonal antibody YsT9-1 is presently being investigated in an enzyme-linked immunosorbent assay (ELISA) and by microcalorimetry. These results, and their use for the further refinement of the model⁶ of the complex between the *Brucella* A antigen and monoclonal antibody YsT9-1, will be separately reported.

EXPERIMENTAL

General. — ¹H- and ¹³C-n.m.r. spectra were recorded with Bruker AM 200 and AM 500 spectrometers, for solutions in CDCl₃ [residual CHCl₃ ($\delta_{\rm H}$ 7.24) and CDCl₃ ($\delta_{\rm C}$ 77.0) as internal standards] or D₂O [internal acetone ($\delta_{\rm H}$ 2.225 and $\delta_{\rm C}$ 30.5)]. First-order chemical shifts and coupling constants were obtained from one-dimensional spectra, and assignments of proton resonances were based on COSY and n.O.e. experiments. Resonances for aromatic and benzylic protons and proton resonances that could not be assigned are not reported. Optical rotations were measured with a Perkin–Elmer 243 polarimeter.

T.l.c. was performed on Silica Gel-60 F_{254} (Merck) with detection by u.v. light and charring with sulfuric acid. Silica gel-60 (Merck, 230–400 mesh) and analytical reagent grade solvents (BDH) were used for column chromatography. Dichloromethane was

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dried by distillation from phosphorus pentoxide and was stored over activated molecular sieves (4 Å). Powdered molecular sieves (Aldrich, 4 Å) were used in the glycosylations. Organic solutions were dried over Na_2SO_4 .

Ethyl 2-O-acetyl-4-azido-3-O-benzyl-4,6-dideoxy-1-thio- α -D-mannopyranoside⁹ (2), ethyl 4-azido-3-O-benzyl-4,6-dideoxy-1-thio- α -D-mannopyranoside⁹ (3), ethyl 2-O-(2-O-acetyl-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside⁹ (8), and methyl O-(2-O-acetyl-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-3,4-di-O-benzyl- α -D-rhamnopyranoside¹³ (12) were prepared according to the indicated literature methods. 1,2-Di-O-acetyl-3,4-di-Obenzyl- α -D-rhamnopyranose (4) was prepared from methyl 2,3-O-isopropylidene- α -Drhamnopyranoside⁷ as described²¹ for the preparation of the L-enantiomer of 4. Compound 4 had $[\alpha]_{D}^{25} + 22^{\circ}$ (c 0.73, chloroform) [lit.²² $[\alpha]_{D}^{25} - 20^{\circ}$ (chloroform) for the L-enantiomer of 4] and ¹H- and ¹³C-n.m.r. data as reported²² for the L-enantiomer of 4.

Satisfactory elemental analyses could not be obtained for the amorphous compounds 25, 30, and 35, but their purity was established by t.l.c. and by n.m.r. spectroscopy.

Methyl 4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside^{7,8} (1). — Dibutyltin oxide (6.37 g, 25.6 mmol) was added to a solution of methyl 4-azido-4,6-dideoxy- α -D-mannopyranoside^{7,8} (5.20 g, 25.6 mmol) in anhydrous methanol (300 mL), and the mixture was boiled under reflux for 2 h. The resulting clear solution was concentrated, and the residue was dried under high vacuum for 2 h. Tetrabutylammonium chloride (7.11 g, 25.6 mmol) and benzyl bromide (3.44 mL, 28.9 mmol) were added to a solution of the residue in dry toluene (125 mL), and the solution was boiled under reflux for 1 h, then concentrated. Column chromatography (1:5 ethyl acetate-hexanes, followed by 1:3) of the residue gave 1 (6.68 g, 89%) with optical rotation and ¹H-n.m.r. spectral data as previously reported⁷.

Ethyl 2-O-acetyl-3,4-di-O-benzyl-1-thio- α - (5) and β -D-rhamnopyranoside (7). — Ethanethiol (2.46 mL, 33.2 mmol) and boron trifluoride etherate (2.46 mL, 20.0 mmol) were added at room temperature to a stirred solution of 4 (5.68 g, 13.3 mmol) in dry dichloromethane (90 mL) containing molecular sieves (4 Å, 4.0 g). After 8 h, triethyl-amine (2.97 mL, 21.3 mmol) was added dropwise, and the mixture was filtered and concentrated. Column chromatography (1:8 ethyl acetate-hexanes) of the residue gave 5 (3.69 g, 64%) and 7 (1.48 g, 26%).

Compound **5** had $[\alpha]_{D}^{25}$ + 61° (*c* 2.1, chloroform). N.m.r. data (CDCl₃): ¹H (200.13 MHz), δ 5.43 (dd, 1 H, J 3.3 and 1.6 Hz, H-2), 5.18 (d, 1 H, J 1.4 Hz, H-1), 4.08 (dq, 1 H, J 9.4 and 6.2 Hz, H-5), 3.86 (dd, 1 H, J 9.3 and 3.2 Hz, H-3), 3.45 (t, 1 H, J 9.4 Hz, H-4), 2.15 (s, 3 H, Ac), 1.32 (d, 3 H, J 6.4 Hz, H-6), and 1.26 (t, 3 H, J 7.6 Hz, SCH₂CH₃); ¹³C (50.32 MHz), δ 82.3 (¹J_{CH} 166 Hz, C-1).

Anal. Calc. for C₂₄H₃₀O₅S: C, 66.9; H, 7.02. Found: C, 66.9; H, 7.07.

Compound 7 had $[\alpha]_{D}^{25} - 80^{\circ}$ (c 1.9, chloroform). N.m.r. data (CDCl₃): ¹H (200.13 MHz), δ 5.63 (dd, 1 H, J 3.5 and 0.9 Hz, H-2), 4.63 (bs, 1 H, H-1), 3.63 (dd, 1 H, J 8.9 and 3.4 Hz, H-3), 2.19 (s, 3 H, Ac), 1.36 (bd, 3 H, J 5.6 Hz, H-6), and 1.28 (t, 1 H, J 7.6 Hz, SCH₂CH₃); ¹³C (50.32 MHz), δ 81.8 (¹J_{CH} 151 Hz, C-1).

Anal. Calc. for C₂₄H₃₀O₅S: C, 66.9; H, 7.02. Found: C, 67.0; H, 7.37.

Ethyl 3,4-di-O-*benzyl-1-thio-α*-D-*rhamnopyranoside* (6). — A solution of **5** (779 mg, 1.81 mmol) in dichloromethane-methanolic 25mM sodium methoxide (1:1, 15 mL) was stirred for 16 h at room temperature, then neutralized [Amberlite IR-120 (H⁺) resin], and concentrated. Column chromatography (1:3 ethyl acetate-hexanes) of the residue gave **6** (680 mg, 97%), $[\alpha]_{D}^{25}$ + 127° (*c* 0.56, chloroform). ¹H-N.m.r. data (200.13 MHz, CDCl₃): δ 5.28 (bs, 1 H, H-1), 3.79 (dd, 1 H, J 9.0 and 3.3 Hz, H-3), 3.47 (t, 1 H, J 9.2 Hz, H-4), 1.30 (d, 3 H, J 6.3 Hz, H-6), and 1.26 (t, 3 H, J 7.4 Hz, SCH₂CH₃).

Anal. Calc. for C₂₂H₂₈O₄S: C, 68.0; H, 7.26. Found: C, 67.8; H, 7.37.

Ethyl 2-O-(2-O-acetyl-3,4-di-O-benzyl-α- (9) and β-D-rhamnopyranosyl)-4-azido-3-O-benzyl-4,6-dideoxy-1-thio-α-D-mannopyranoside (11). — Bromine (9.8 μ L, 190 μ mol) was added to a stirred solution of 5 (125 mg, 290 μ mol) in dry dichloromethane (2.5 mL) at 0°, and after 30 min cyclohexene (29 μ L, 290 μ mol) was added. The solution was then added to a mixture of 3 (ref. 9) (75 mg, 230 μ mol), mercuric cyanide (117 mg, 464 μ mol), mercuric bromide (84 mg, 230 μ mol), and powdered molecular sieves (4 Å, 200 mg) in dry dichloromethane (2 mL) that had been stirred for 4 h at 0°. After 16 h the mixture was filtered through Celite, and the flask and Celite were washed with dichloromethane (20 mL). The solution was washed with satd. aq. sodium hydrogencarbonate (7.5 mL), water (7.5 mL), and brine (7.5 mL), then dried and concentrated. Column chromatography (1:10 ethyl acetate-hexanes, followed by 1:7) of the residue gave 9 (100 mg, 62%) and 11 (32 mg, 20%).

Compound 9 had $[\alpha]_{D}^{25}$ + 80° (*c* 1.6, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.44 (dd, 1 H, *J* 3.0 and 2.0 Hz, H-2b), 5.19 (d, 1 H, *J* 1.0 Hz, H-1a), 4.83 (d, 1 H, *J* 1.3 Hz, H-1b), 3.96 (bs, 1 H, H-2a), 3.94 (dd, 1 H, *J* 8.8 and 3.2 Hz, H-3b), 3.84 (dq, 1 H, *J* 9.6 and 6.2 Hz, H-5b), 3.81 (dq, 1 H, *J* 10.0 and 6.2 Hz, H-5a), 3.65 (dd, 1 H, *J* 9.8 and 2.9 Hz, H-3a), 3.43 (t, 1 H, *J* 9.4 Hz, H-4b), 3.38 (t, 1 H, *J* 9.9 Hz, H-4a), 2.11 (s, 3 H, Ac), 1.31 (d, 3 H, *J* 6.2 Hz, H-6b), 1.28 (d, 3 H, *J* 6.2 Hz, H-6a), and 1.25 (t, 3 H, *J* 7.4 Hz, SCH₂CH₃); ¹³C (125.76 MHz), δ 99.6 (¹J_{C,H} 170 Hz, C-1b) and 83.3 (C-1a).

Anal. Calc. for C₃₇H₄₅N₃O₈S: C, 64.2; H, 6.56; N, 6.07. Found: C, 64.3; H, 6.60; N, 6.01.

Compound 11 was characterised by n.m.r. spectroscopy (CDCl₃): ¹H (500.14 MHz), δ 5.62 (dd, 1 H, J 3.2 and 0.6 Hz, H-2b), 5.27 (d, 1 H, J 1.3 Hz, H-1a), 4.65 (d, 1 H, J 0.8 Hz, H-1b), 4.34 (dd, 1 H, J 3.1 and 1.6 Hz, H-2a), 3.80 (dq, 1 H, J 10.0 and 6.2 Hz, H-5a), 3.66 (dd, 1 H, J 9.8 and 3.1 Hz, H-3a), 3.62 (dd, 1 H, J 9.2 and 3.3 Hz, H-3b), 3.47 (t, 1 H, J 9.2 Hz, H-4b), 3.39 (t, 1 H, J 9.9 Hz, H-4a), 3.35 (dq, 1 H, J 9.3 and 6.1 Hz, H-5b), 2.01 (s, 3 H, Ac), 1.34 (d, 3 H, J 6.1 Hz, H-6b), 1.30 (d, 3 H, J 6.2 Hz, H-6a), and 1.27 (t, 3 H, J 7.4 Hz, SCH₂CH₃); ¹³C (125.76 MHz), δ 95.3 (¹J_{C,H} 156 Hz, C-1b) and 81.1 (C-1a).

Ethyl 2-O- (2-O- acetyl-4- azido- 3-O- benzyl-4,6- dideoxy- α -D-mannopyranosyl)-3,4-di-O-benzyl-1-thio- α -D-rhamnopyranoside (10). — Treatment of 2 (ref. 9) (59 mg, 160 μ mol) with bromine, and glycosylation of 6 (50 mg, 130 μ mol), as described for 9, gave after column chromatography (1:10 ethyl acetate-hexanes), 10 (15 mg) which was indicated to be ~90% pure by ¹H-n.m.r. spectroscopy. N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.45 (dd, 1 H, J 3.2 and 1.9 Hz, H-2b), 5.13 (d, 1 H, J 1.5 Hz, H-1a), 4.92 (d, 1 H, J 1.7 Hz, H-1b), 3.99 (dq, 1 H, J 9.4 and 6.2 Hz, H-5a), 3.97 (dd, 1 H, J 2.7 and 1.9 Hz, H-2a), 3.82 (dd, 1 H, J 9.9 and 3.3 Hz, H-3b), 3.77 (dd, 1 H, J 9.2 and 2.9 Hz, H-3a), 3.65 (dq, 1 H, J 10.1 and 6.2 Hz, H-5b), 3.39 (t, 1 H, J 9.3 Hz, H-4a), 3.38 (t, 1 H, J 10.0 Hz, H-4b), 2.09 (s, 3 H, Ac), 1.29 (d, 3 H, J 6.2 Hz, H-6b), 1.27 (d, 3 H, J 6.2 Hz, H-6a), and 1.24 (t, 3 H, J 7.4 Hz, SCH₂CH₃); ¹³C (125.76 MHz), δ 99.4 (¹J_{C,H} 172 Hz, C-1b) and 83.4 (C-1a).

Methyl O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-3,4-di-O-benzyl- α -Drhamnopyranoside (13). — Deacetylation of 12 (ref. 13) (123 mg, 133 μ mol) as described for **6**, and column chromatography (1:5 ethyl acetate-hexanes) of the residue gave 13 (102 mg, 87%): [α]_D²⁵ + 77° (c 2.2, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 5.05 (d, 1 H, J 1.7 Hz, H-1b), 4.93 (d, 1 H, J 1.1 Hz, H-1c), 4.52 (d, 1 H, J 1.6 Hz, H-1a), 4.01 (t, 1 H, J2.4 Hz, H-2b), 3.99 (bs, 1 H, H-2c), 3.90 (t, 1 H, J2.2 Hz, H-2a), 3.81 (dd, 1 H, J 9.4 and 2.9 Hz, H-3a), 3.76 (dd, 1 H, J 10.1 and 2.8 Hz, H-3b), 3.71 (dd, 1 H, J 9.7 and 3.2 Hz, H-3c), 3.62 (dq, 1 H, J 9.4 and 6.2 Hz, H-5a), 3.40 (t, 1 H, J 9.9 Hz, H-4c), 3.32 (t, 1 H, J 10.0 Hz, H-4b), 3.31 (t, 1 H, J 9.5 Hz, H-4a), 3.28 (s, 3 H, MeO), 1.28 (bd, 6 H, J 6.0 Hz, H-6ab), and 1.16 (d, 3 H, J 6.1 Hz, H-6c).

Anal. Calc. for $C_{47}H_{56}N_6O_{11}$: C, 64.1; H, 6.41; N, 9.54. Found: C, 64.1; H, 6.48; N, 9.34.

Methyl O-(2-O-acetyl-3,4-di-O-benzyl- α -D-rhamnopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α - (14) and β -D-mannopyranosyl)-(1 \rightarrow 2)-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside (16). — A solution of 1 (ref. 7) (246 mg, 0.840 mmol) and 9 (465 mg, 0.672 mmol) in dry dichloromethane (14 mL) containing powdered molecular sieves (4 Å, 1.2 g) was stirred for 4 h at room temperature. Silver triflate (345 mg, 1.34 mmol) was added, followed after 10 min by bromine (26 μ L, 0.50 mmol)¹³. After a further 30 min, additional silver triflate (173 mg, 0.672 mmol) was added, followed by triethylamine (280 μ L, 2.0 mmol) when the reactants had been consumed (~60 min after the addition of bromine, t.l.c.). All operations were carried out under a positive pressure of dry nitrogen and in the dark. The mixture was filtered through Celite, and the flask and Celite were washed with dichloromethane (30 mL). The solution was washed with satd. aq. sodium hydrogencarbonate (15 mL), dried, and concentrated. Column chromatography (1:6 ethyl acetate-hexanes, followed by 1:4) of the residue gave 14 (472 mg, 76%) and 16 (45 mg, 7.2%).

Compound 14 had $[\alpha]_{D}^{25} + 46^{\circ}$ (c 2.4, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.43 (dd, 1 H, J 3.2 and 1.9 Hz, H-2c), 4.96 (d, 1 H, J 1.7 Hz, H-1b), 4.83 (d, 1 H, J 1.6 Hz, H-1c), 4.53 (d, 1 H, J 2.0 Hz, H-1a), 3.91 (t, 1 H, J 2.4 Hz, H-2b), 3.89 (dd, 1 H, J 9.4 and 3.3 Hz, H-3c), 3.81 (t, 1 H, J 2.4 Hz, H-2a), 3.72 (dq, 1 H, J 9.6 and 6.3 Hz, H-5c), 3.70 (dd, 1 H, J 9.9 and 3.0 Hz, H-3b), 3.65 (dd, 1 H, J 9.9 and 2.9 Hz, H-3a), 3.50 (dq, 1 H, J 10.0 and 6.1 Hz, H-5b), 3.39 (dq, 1 H, J 10.1 and 6.1 Hz, H-5a), 3.38 (t, 1 H, J 9.5 Hz, H-4c), 3.37 (t, 1 H, J 10.0 Hz, H-4b), 3.27 (s, 3 H, MeO), 3.22 (t, 1 H, J 10.0 Hz, H-4a), 2.11 (s, 3 H, Ac), 1.27 (d, 3 H, J 6.2 Hz, H-6a), 1.25 (d, 3 H, J 7.1 Hz, H-6b), and 1.20 (d, 3 H, J 6.2 Hz, H-6c); ¹³C (125.76 MHz), δ 100.5, 99.8, and 99.2 (¹J_{C,H} 171, 169, and 171 Hz, C-1abc).

Anal. Calc. for $C_{49}H_{58}N_6O_{12}$: C, 63.8; H, 6.33; N, 9.10. Found: C, 63.4; H, 6.28; N, 8.78.

Compound 16 had $[\alpha]_{p_0}^{25} + 31^{\circ}$ (c 2.5, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.51 (dd, 1 H, J 3.3 and 1.5 Hz, H-2c), 5.37 (d, 1 H, J 1.2 Hz, H-1c), 4.64 (d, 1 H, J 1.3 Hz, H-1a), 4.47 (bs, 1 H, H-1b), 4.31 (m, 2 H, H-2ab), 4.16 (dq, 1 H, J 9.6 and 6.2 Hz, H-5c), 4.02 (dd, 1 H, J 9.5 and 3.5 Hz, H-3c), 3.72 (dd, AB-type, 1 H, J 9.9 and 3.5 Hz, H-3a), 3.67 (t, AB-type, 1 H, J 9.8 Hz, H-4a), 3.60 (t, 1 H, J 9.8 Hz, H-4b), 3.40 (dd, 1 H, J 9.8 and 2.6 Hz, H-3b), 3.38 (t, 1 H, J 9.6 Hz, H-4c), 3.31 (s, 3 H, MeO), 3.14 (dq, 1 H, J 9.8 and 6.1 Hz, H-5b), 2.07 (s, 3 H, Ac), 1.32 (d, 1 H, J 6.2 Hz, H-6c), 1.29 (d, J 6.1 Hz, H-6b), and 1.03 (d, 1 H, J 6.2 Hz, H-6a); ¹³C (125.76 MHz), δ 98.0 and 96.7 (¹J_{C,H} 167 and 172 Hz, C-1ac) and 96.6 (¹J_{C,H} 154 Hz, C-1b).

Anal. Calc. for $C_{49}H_{58}N_6O_{12}$: C, 63.8; H, 6.33; N, 9.10. Found: C, 63.7; H, 6.40; N, 8.80.

Methyl O-(3,4-di-O-benzyl- α -D-rhamnopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside (15). — Deacetylation of 14 (446 mg, 0.483 mmol) as described for 6 and column chromatography (2:7 ethyl acetate-hexanes) of the residue gave 15 (392 mg, 92%): [α]_D²⁵ + 56° (*c* 1.4, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 4.96 (d, 1 H, *J* 1.6 Hz, H-1b), 4.95 (d, 1 H, *J* 1.3 Hz, H-1c), 4.53 (d, 1 H, *J* 1.7 Hz, H-1a), 4.03 (bs, 1 H, H-2c), 3.96 (t, 1 H, *J* 2.3 Hz, H-2b), 3.81 (dd, 1 H, *J* 9.2 and 3.1 Hz, H-3c), 3.81 (t, 1 H, *J* 2.4 Hz, H-2a), 3.71 (dq, 1 H, *J* 9.5 and 6.3 Hz, H-5c), 3.70 (dd, 1 H, *J* 9.9 and 2.8 Hz, H-3b), 3.65 (dd, 1 H, *J* 9.9 and 2.9 Hz, H-3a), 3.50 (dq, 1 H, *J* 10.0 and 6.1 Hz, H-5b), 3.43 (t, 1 H, *J* 9.4 Hz, H-4c), 3.39 (dq, 1 H, *J* 10.0 Hz, H-4a), 1.27 (d, 3 H, *J* 6.2 Hz, H-6a), 1.25 (d, 3 H, *J* 6.3 Hz, H-6b), and 1.19 (d, 3 H, *J* 6.2 Hz, H-6c).

Anal. Calc. for $C_{47}H_{56}N_6O_{11}$: C, 64.1; H, 6.41; N, 9.54. Found: C, 63.8; H, 6.23; N, 9.39.

Methyl 2-O-(2-O-acetyl-3,4-di-O-benzyl-α-D-rhamnopyranosyl)-4-azido-3-Obenzyl-4,6-dideoxy-α-D-mannopyranoside (17). — Glycosylation¹³ of 1 (ref. 7) (90 mg, 0.307 mmol) with 5 (165 mg, 0.384 mmol) at -45° as described for 14 and column chromatography (1:8 ethyl acetate-hexanes) of the residue gave 17 (189 mg, 93%): $[\alpha]_{2^{5}}^{12^{5}}$ + 12° (c 1.1, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.45 (dd, 1 H, J 3.2 and 1.9 Hz, H-2b), 4.88 (d, 1 H, J 1.7 Hz, H-1b), 4.60 (bs, 1 H, H-1a), 3.92 (dd, 1 H, J9.4 and 3.4 Hz, H-3b), 3.90 (dd, 1 H, J2.9 and 2.0 Hz, H-2a), 3.79 (dq, 1 H, J9.5 and 6.2 Hz, H-5b), 3.71 (dd, 1 H, J 9.8 and 3.0 Hz, H-3a), 3.44 (dq, 1 H, J 10.0 and 6.1 Hz, H-5a), 3.41 (t, 1 H, J9.4 Hz, H-4b), 3.36 (t, 1 H, J9.9 Hz, H-4a), 3.29 (s, 3 H, MeO), 2.10 (s, 3 H, Ac), 1.31 (d, 3 H, J6.2 Hz, H-6b), and 1.29 (d, 3 H, J6.1 Hz, H-6a); ¹³C (125.76 MHz), δ 99.8 and 99.3 (¹J_{C H} 171 and 172 Hz, C-1ab).

Anal. Calc. for $C_{36}H_{43}N_3O_9$: C, 65.3; H, 6.55; N, 6.35. Found: C, 65.0; H, 6.63; N, 6.12.

Methyl 2-O-(3,4-di-O-benzyl- α -D-rhamnopyranosyl)-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside (18). — Deacetylation of 17 (88 mg, 133 μ mol) as described for 6 and column chromatography (1:3 ethyl acetate-hexanes) of the residue gave 18 (77 mg, 93%): $[\alpha]_{D}^{25}$ + 41° (c 1.2, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 4.96 (d, 1 H, J 1.5 Hz, H-1b), 4.61 (d, 1 H, J 1.7 Hz, H-1a), 4.04 (bs, 1 H, H-2b), 3.94 (t, 1 H, J 2.4 Hz, H-2a), 3.83 (dd, 1 H, J 9.2 and 3.2 Hz, H-3b), 3.78 (dq, 1 H, J 9.5 and 6.3 Hz, H-5b), 3.71 (dd, 1 H, J 9.9 and 3.0 Hz, H-3a), 3.46 (t, 1 H, J 9.4 Hz, H-4b), 3.43 (dq, 1 H, J 10.1 and 6.2 Hz, H-5a), 3.32 (t, 1 H, J 10.0 Hz, H-4a), 3.29 (s, 3 H, MeO), 1.30 (d, 3 H, J 6.2 Hz, H-6b), and 1.29 (d, 3 H, J 6.2 Hz, H-6a).

Anal. Calc. for C₃₄H₄₁N₃O₈: C, 65.9; H, 6.67; N, 6.78. Found: C, 66.0; H, 6.74; N, 6.83.

Methyl O- (2-O-acetyl-4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-O-(3,4-di-O-benzyl-α-D-rhamnopyranosyl)-(1→2)-4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranoside (19). — Glycosylation¹³ of 18 (228 mg, 0.368 mmol) with 2 (ref. 9) (168 mg, 0.460 mmol) at -45° as described for 14 and column chromatography (1:8 ethyl acetate-hexanes) of the residue gave 19 (297 mg, 87%): $[\alpha]_{D}^{25}$ + 74° (c 1.6, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.48 (dd, 1 H, J 3.1 and 1.9 Hz, H-2c), 4.97 (d, 1 H, J 1.8 Hz, H-1b), 4.95 (d, 1 H, J 1.5 Hz, H-1c), 4.57 (d, 1 H, J 1.5 Hz, H-1a), 3.98 (t, 1 H, J 2.4 Hz, H-2b), 3.91 (t, 1 H, J 2.3 Hz, H-2a), 3.85 (dd, 1 H, J 9.1 and 2.9 Hz, H-3b), 3.80 (dd, 1 H, J 9.9 and 3.3 Hz, H-3c), 3.73 (dq, 1 H, J 9.3 and 6.2 Hz, H-5b), 3.68 (dd, 1 H, J9.9 and 2.9 Hz, H-3a), 3.53 (dq, 1 H, J 10.1 and 6.1 Hz, H-5c), 3.42 (dq, 1 H, J 10.1 and 6.4 Hz, H-5a), 3.42 (t, 1 H, J 9.1 Hz, H-4b), 3.36 (t, 1 H, J 10.0 Hz, H-4c), 3.29 (s, 3 H, MeO), 3.28 (t, 1 H, J 9.9 Hz, H-4a), 2.10 (s, 3 H, Ac), 1.29 (d, 6 H, J 6.2 Hz, H-6ab), and 1.18 (d, 3 H, J 6.2 Hz, H-6c); ¹³C (125.76 MHz), δ 100.2, 99.9, and 98.7 (¹J_{CH} 173, 170, and 172 Hz, C-1abc).

Anal. Calc. for $C_{49}H_{58}N_6O_{12}$: C, 63.8; H, 6.33; N, 9.10. Found: C, 63.8; H, 6.48; N, 9.25.

Methyl O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(3. 4-di-O-benzyl- α -D-rhamnopyranosyl)-(1 \rightarrow 2)-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranoside (**20**). — Deacetylation of **19** (276 mg, 0.299 mmol) as described for **6** and column chromatography (1:6 ethyl acetate-hexanes) of the residue gave **20** (249 mg, 94%): [α]_D²⁵ + 80° (*c* 1.1, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 5.05 (d, 1 H, *J* 1.3 Hz, H-1c), 4.96 (d, 1 H, *J* 1.9 Hz, H-1b), 4.55 (d, 1 H, *J* 1.7 Hz, H-1a), 4.03 (bs, 1 H, H-2c), 3.99 (t, 1 H, *J* 2.4 Hz, H-2b), 3.88 (t, 1 H, *J* 2.3 Hz, H-2a), 3.83 (dd, 1 H, *J* 9.1 and 2.9 Hz, H-3b), 3.72 (dd, 1 H, *J* 9.6 and 3.2 Hz, H-3c), 3.71 (dq, 1 H, *J* 9.3 and 6.1 Hz, H-5b), 3.66 (dd, 1 H, *J* 9.9 and 2.9 Hz, H-3a), 3.52 (dq, 1 H, *J* 10.1 and 6.2 Hz, H-5c), 3.39 (t, 1 H, *J* 10.0 Hz, H-4c), 3.36 (t, 1 H, *J* 9.2 Hz, H-4b), 3.27 (s, 3 H, MeO), 3.25 (t, 1 H, *J* 10.0 Hz, H-4a), 1.28 (d, 3 H, *J* 6.2 Hz, H-6b), 1.26 (d, 3 H, *J* 6.2 Hz, H-6a), and 1.16 (d, 3 H, *J* 6.2 Hz, H-6c).

Anal. Calc. for $C_{47}H_{56}N_6O_{11}$: C, 64.1; H, 6.41; N, 9.54. Found: C, 63.7; H, 6.50; N, 9.55.

Methyl O-(2-O-acetyl-4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-3,4-di-O-benzyl- α -D-rhamnopyranoside (21). Glycosylation¹³ of 13 (75 mg, 85 μ mol) with 8 (ref. 9) (67 mg, 110 μ mol) as described for 14 and column chromatography (1:6 ethyl acetate-hexanes) of the residue gave 21 (97 mg, 79%): $[\alpha]_{D}^{25} + 76^{\circ}$ (*c* 1.8, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.42 (dd, 1 H, *J* 3.0 and 1.9 Hz, H-2e), 4.98, 4.96, and 4.90 (3 d, each 1 H, *J* 1.6, 1.7, and 1.3 Hz, H-1bcd), 4.52 (d, 1 H, *J* 1.4 Hz, H-1a), 3.81 (dd, 1 H, *J* 9.4 and 2.9 Hz, H-3a), 3.78 (dd, 1 H, *J* 10.0 and 3.3 Hz, H-3e), 3.62 (dq, 1 H, *J* 9.4 and 6.1 Hz, H-5a), 3.38 (t, 1 H, *J* 10.0 Hz, H-4e), 3.29 (s, 3 H, MeO), 3.28 (t, 1 H, *J* 9.4 Hz, H-4a), 2.11 (s, 3 H, Ac), 1.29 (d, 3 H, *J* 6.1 Hz, H-6a), and 1.21 (d, 3 H, *J* 6.3 Hz, H-6e); ¹³C (125.76 MHz), δ 100.2, 100.1, 100.0, 99.7, and 99.1 (¹*J*_{CH} 172, 172, 172, 170, and 174 Hz, C-1abcde).

Anal. Calc. for $C_{75}H_{88}N_{12}O_{18}$: C, 62.3; H, 6.14; N, 11.6. Found: C, 62.4; H, 6.11; N, 11.5.

Methyl O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4-azido-3-O-benzyl-4,6-dideoxy- α -D-mannopyranosyl)-(1 \rightarrow 2)-3,4-di-O-benzyl- α -D-rhamnopyranoside (22). Deacetylation of 21 (97 mg, 67 μ mol) as described for 6 and column chromatography (1:4 ethyl acetate-hexanes) of the residue gave 22 (85 mg, 90%): $[\alpha]_D^{25}$ +88° (c 1.4, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 4.97 and 4.91 (bs, 3 H and d, 1 H, J 1.2 Hz, H-1bcde), 4.52 (d, 1 H, J 1.2 Hz, H-1a), 3.99 (bs, 1 H, H-2e), 3.88 (t, 1 H, J 2.2 Hz, H-2a), 3.81 (dd, 1 H, J 9.4 and 2.8 Hz, H-3a), 3.62 (dq, 1 H, J 9.4 and 6.2 Hz, H-5a), 3.42 (t, 1 H, J 9.8 Hz, H-4e), 3.29 (s, 3 H, MeO), 3.28 (t, 1 H, J 9.9 Hz, H-4a), 1.29 (d, 3 H, J 6.4 Hz, H-6a), and 1.20 (d, 3 H, J 6.1 Hz, H-6e).

Anal. Calc. for $C_{73}H_{86}N_{12}O_{17}$: C, 62.8; H, 6.18; N, 12.0. Found: C, 63.1; H, 6.26; N, 12.4.

Methyl O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-zvl-4.6-dideoxv-4-formamido- α -D-mannopyranosyl)- $(1 \rightarrow 2)$ -O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)- (1 \rightarrow 2)-3,4-di-O-benzyl- α -D-rhamnopyranoside (24). — A solution of 22 (100 mg, 71.2 μ mol) in pyridine-triethylamine (1:1, 16 mL) was saturated with hydrogen sulfide¹⁶ for 1 h at room temperature and then stirred for 16 h. Nitrogen was passed through the solution for 1 h to remove hydrogen sulfide, the solution was concentrated, and the residual solvents were codistilled twice with toluene. Column chromatography (methanol-dichloromethane, 1:30 \rightarrow 1:10 containing 0.1% of triethylamine) of the residue gave the amine 23 which was taken up in ethyl formate-pyridine (50:1, 15 mL). The mixture was boiled under reflux for 24 h, the resulting clear solution was concentrated, and the residual solvents were codistilled with toluene. Column chromatography (1:40 methanol-dichloromethane, followed by 1:30) of the residue gave 24 (65 mg, 65%): $[\alpha]_{p}^{25} + 39^{\circ}$ (c 0.97, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 8.21–7.94 (m, 4 H, NHCHO), 3.34 (t, 1 H, J 9.5 Hz, H-4a), 3.29 (bs, 3 H, MeO), and 1.30–1.00 (m, 15 H, H-6abcde); 13 C (125.76 MHz), δ 164.9 and 161.5 (2 m, NHCHO), 100.2-99.6 (m, C-1abcde), 50.8 (m, C-4bcde-Z).

Anal. Calc. for C₇₇H₉₀N₄O₂₁: C, 65.5; H, 6.71; N, 3.97. Found: C, 65.3; H, 6.78; N, 3.62.

 $Methyl O-(4,6-dideoxy-4-formamido-\alpha-D-mannopyranosyl)-(1\rightarrow 2)-O-(4,6-dideoxy-4-formamido-\alpha-D-mannopyranosyl)-(1\rightarrow 2)-(1\rightarrow 2)-$

mannopyranosyl)- $(1 \rightarrow 2)$ -O-(4, 6-dideoxy-4-formamido- α -D-mannopyranosyl)- $(1 \rightarrow 2)$ - α -D-rhamnopyranoside (25). — A solution of 24 (76 mg, 55 μ mol) in formic acid (10 mL) was hydrogenated over Pd–C (10%, 150 mg) at 5 atm. and room temperature for 16 h. The mixture was filtered through Celite and concentrated, and the residual solvents were codistilled twice with methanol. The residue was dissolved in methanolic 25mm sodium methoxide (8 mL) and stirred for 1 h, then neutralized [Amberlite IR-120 (H⁺) resin], and concentrated. Column chromatography (water-acetic acid-pyridine, 986:4:10) of the residue on Bio-Gel P-4 gave, after freeze drying, 25 (42 mg, 89%): $[\alpha]_{c}^{25}$ $+39^{\circ}$ (c 0.69, water). ¹H-N.m.r. spectroscopy showed a Z:E-ratio of ~4.3:1 for the formamido groups. N.m.r. data (D₂O): ¹H (500.14 MHz), δ 8.20–8.18 (3 s, 3.2 H, NHCHO-Z), 8.03-8.01 (3 s, 0.8 H, NHCHO-E), 5.18, 5.16, 5.14, and 5.03 (4 bs, 4 H, H-1bcde), 4.73 (bs, 1 H, H-1a), 4.03 (dd, 1 H, J 10.6 and 2.9 Hz, H-3a), 3.67 (dg, 1 H, J 9.6 and 6.3 Hz, H-5a), 3.45 (t, 1 H, J9.7 Hz, H-4a), 3.38 (s, 3 H, MeO), 1.29 (d, 3 H, J6.2 Hz, H-6a), and 1.27–1.18 (m, 12 H, H-6bcde); 13 C (125.76 MHz), δ 168.1 (NHCHO-E), 165.2 (NHCHO-Z), 102.2 (C-1e), 100.9, 100.8, and 100.8 (C-1bcd), 99.8 (C-1a), 78.5 and 77.4 (C-2abcd), 57.1 and 56.9 (C-4bcde-E), 55.1 (MeO), 52.2, 52.1, and 51.9 (C-4bcde-Z).

Methyl O-(4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-O-(4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-O-(4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-O-(3,4-di-O-benzyl-α-D-rhamnopyranosyl)-(1→2)-4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranoside (27). — Glycosylation¹³ of 20 (200 mg, 0.227 mmol) with 8 (ref. 9) (178 mg, 0.284 mmol) as described for 14 and column chromatography (1:6 ethyl acetate-hexanes) of the residue gave 26 (290 mg) which was indicated to be 90–95% pure by ¹H-n.m.r. spectroscopy. N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.40 (dd, 1 H, J 3.2 and 1.9 Hz, H-2e), 4.97 and 4.96 (2 d, each 1 H, each J 1.7 Hz, H-1cd), 4.89 (d, 1 H, J 1.8 Hz, H-1b), 4.84 (d, 1 H, J 1.7 Hz, H-1e), 4.54 (d, 1 H, J 1.9 Hz, H-1a), 3.89 (t, 1 H, J 2.5 Hz, H-2b), 3.86 (t, 1 H, J 2.4 Hz, H-2a), 3.81 (dd, 1 H, J 9.2 and 2.8 Hz, H-3b), 3.77 (dd, 1 H, J 9.9 and 3.2 Hz, H-3e), 3.69 (dq, 1 H, J 9.3 and 6.0 Hz, H-5b), 3.65 (dd, 1 H, J 9.9 and 2.9 Hz, H-3a), 3.51 (dq, 1 H, J 1.9 Hz, H-4e), 3.29 (t, 1 H, J 9.4 Hz, H-4b), 3.27 (s, 3 H, MeO), 2.10 (s, 3 H, Ac), and 1.26 (d, 3 H, J 6.2 Hz, H-6b); ¹³C (125.76 MHz), δ 100.3, 100.1, 99.9, 99.9, and 99.0 (each ¹J_{CH} 171 Hz, C-1abcde).

Crude **26** (290 mg) was deacetylated as described for **6** and gave, after column chromatography (2:9 ethyl acetate-hexanes) of the residue, **27** (252 mg, 79% from **20**): $[\alpha]_{D}^{25} + 87^{\circ}$ (*c* 1.3, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 4.98–4.97 (m, 3 H, H-1cde), 4.90 (d, 1 H, J 1.8 Hz, H-1b), 4.54 (d, 1 H, J 1.8 Hz, H-1a), 3.99 (bs, 1 H, H-2e), 3.89 (t, 1 H, J 2.4 Hz, H-2b), 3.87 (t, 1 H, J 2.4 Hz, H-2a), 3.81 (dd, 1 H, J 9.3 and 2.8 Hz, H-3b), 3.71 (dd, 1 H, J 9.7 and 3.2 Hz, H-3e), 3.69 (1 H, H-5b), 3.66 (dd, 1 H, J 9.9 and 2.9 Hz, H-3a), 3.53 (dq, 1 H, J 10.1 and 6.2 Hz, H-5e), 3.41 (t, 1 H, J 9.8 Hz, H-4e), 3.39 (dq, 1 H, J 10.0 and 6.2 Hz, H-5a), 3.30 (t, 1 H, J 9.2 Hz, H-4b), 3.27 (s, 3 H, MeO), 3.22 (t, 1 H, J 10.0 Hz, H-4a), 1.27 (d, 3 H, J 6.2 Hz, H-6b), 1.26 (d, 3 H, J 6.2 Hz, H-6a), and 1.18 (d, 3 H, J 6.2 Hz, H-6e).

Anal. Calc. for $C_{73}H_{86}N_{12}O_{17}$: C, 62.5; H, 6.18; N, 12.0. Found: C, 62.2; H, 6.21; N, 11.8.

Methyl O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(3-O-benzyl-4,6-dideoxy-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(3,4-di-O-benzyl- α -D-rham-nopyranosyl)-(1 \rightarrow 2)-3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranoside (29). — Reduction¹⁶ of 27 (98 mg, 70 μ mol) and N-formylation of the intermediate amine 28 as described for 24 gave, after column chromatography (1:40 methanol-ethyl acetate, followed by 1:30) of the residue, 29 (63 mg, 64%): $[\alpha]_{\rm D}^{25}$ +46° (c 3.1, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 8.19–7.94 (m, 4 H, NHCHO), 3.31 (bs, 3 H, MeO), and 1.37–0.96 (m, 15 H, H-6abcde); ¹³C (125.76 MHz), δ 165.0 and 161.4 (2 m, NHCHO), 100.5–99.6 (m, C-1abcde), and 50.8 (m, C-4acde-Z).

Anal. Calc. for C₇₇H₉₀N₄O₂₁: C, 65.5; H, 6.71; N, 3.97. Found: C, 65.3; H, 6.71; N, 3.86.

Methyl O-(4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(4,6-dideoxy-4-formamido- α -Dmannopyranosyl)-(1 \rightarrow 2)-O- α -D-rhamnopyranosyl-(1 \rightarrow 2)-4,6-dideoxy-4-formamido- α -D-mannopyranoside (30). — Debenzylation of 29 (60 mg, 43 μ mol), and column chromatography of the residue on Bio-Gel P-4 as described for 25 gave, after freeze drying, 30 (31 mg, 84%): [α]_D²⁵ + 49° (c 0.73, water). ¹H-N.m.r. spectroscopy showed a Z:E-ratio of ~ 3.9:1 for the formamido groups. N.m.r. data (D₂O): ¹H (500.14 MHz), δ 8.21–8.19 (4 s, 3.2 H, NHCHO-Z), 8.03 (bs, 0.8 H, NHCHO-E), 5.20, 5.16, 5.11, and 5.05 (4 bs, each 1 H, H-1bcde), 4.82 (bs, 1 H, H-1a), 3.50 (t, 1 H, J9.7 Hz, H-4b), 3.40 (s, 3 H, MeO), and 1.31–1.19 (m, 15 H, H-6abcde); ¹³C (125.76 MHz), δ 168.1 (NHCHO-E), 165.1 (NHCHO-Z), 102.3 (C-1e), 101.0, 100.9, and 100.8 (C-1bcd), 99.8 (C-1a), 78.3, 77.5, 77.4, and 77.3 (C-2abcd), 57.1, and 57.0 (C-4acde-E), 55.2 (MeO), 52.2, 52.2, 52.1, and 51.9 (C-4acde-Z).

Methyl O-(4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-O-(4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-O-(3,4-di-O-benzyl-α-D-rhamnopyranosyl)-(1→2)-O-(4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranosyl)-(1→2)-4-azido-3-O-benzyl-4,6-dideoxy-α-D-mannopyranoside (32). — Glyco-sylation¹³ of 15 (140 mg, 0.159 mmol) with 8 (ref. 9) (125 mg, 0.199 mmol) as described for 14 and column chromatography (1:40 ethyl acetate-toluene) of the residue gave 31 (155 mg) which was indicated to be ~90% pure by ¹H-n.m.r. spectroscopy. N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 5.41 (dd, 1 H, J 3.1 and 1.9 Hz, H-2e), 5.05 and 4.89 (2 d, each 1 H, each J 1.6 Hz, H-1bd), 4.88 (d, 1 H, J 1.7 Hz, H-1c), 4.87 (d, 1 H, J 1.6 Hz, H-1e), 4.51 (d, 1 H, J 1.7 Hz, H-1a), 3.91 (t, 1 H, J 2.3 Hz, H-2c), 3.56 (dq, 1 H, J 10.1 and 6.2 Hz, H-5e), 3.39 (dq, 1 H, J 10.0 and 6.2 Hz, H-5a), 3.37 (t, 1 H, J 10.1 Hz, H-4e), 3.30 (t, 1 H, J 9.2 Hz, H-4c), 3.27 (s, 3 H, MeO), 3.19 (t, 1 H, J 10.0 Hz, H-4a), 2.09 (s, 3 H, Ac), 1.26 (d, 3 H, J 6.2 Hz, H-6a), and 1.20 (d, 3 H, J 6.2 Hz, H-6e); ¹³C (125.76 MHz), δ 100.5, 100.3, 99.8, 99.8, and 99.2 (¹J_{C,H} 174, 172, 170, 170, and 172 Hz, C-1abcde).

Crude 31 (155 mg) was deacetylated as described for 6 and gave, after column chromatography (1:4 ethyl acetate-hexanes) of the residue, 32 (133 mg, 60% from 15): $[\alpha]_D^{25} + 80^\circ$ (c 0.67, chloroform). ¹H-N.m.r. data (500.14 MHz, CDCl₃): δ 4.99 (d, 1 H, J 1.4 Hz, H-1e), 5.05 and 4.88 (d and bs, 1 H and 2 H, J 1.7 Hz, H-1bcd), 4.50 (d, 1 H,

J 1.7 Hz, H-1a), 3.90 (t, 1 H, J 2.4 Hz, H-2c), 3.79 (dd, 1 H, J 9.2 and 2.8 Hz, H-3c), 3.78 (t, 1 H, J 2.4 Hz, H-2a), 3.72 (dd, 1 H, J 9.4 and 3.2 Hz, H-3e), 3.64 (dd, 1 H, J 9.9 and 2.9 Hz, H-3a), 3.64 (H-5c), 3.56 (dq, 1 H, J 10.1 and 6.2 Hz, H-5e), 3.41 (t, 1 H, J 10.0 Hz, H-4e), 3.38 (dq, 1 H, J 10.0 and 6.1 Hz, H-5a), 3.26 (s, 3 H, MeO), 3.18 (t, 1 H, J 10.0 Hz, H-4a), 1.25 (d, 3 H, J 6.2 Hz, H-6a), 1.19 (d, 3 H, J 6.0 Hz, H-6e), and 1.16 (d, 3 H, J 6.3 Hz, H-6c).

Anal. Calc. for C₇₃H₈₆N₁₂O₁₇: C, 62.5; H, 6.18; N, 12.0. Found: C, 62.1; H, 6.09; N, 11.7.

Methyl O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-O-(3,4-di-O-benzyl- α -D-rhamnopyranosyl)-(1 \rightarrow 2)-O-(3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranosyl)-(1 \rightarrow 2)-3-O-benzyl-4,6-dideoxy-4-formamido- α -D-mannopyranoside (34). — Reduction¹⁶ of 32 (115 mg, 81.9 μ mol) and N-formylation of the intermediate amine 33 as described for 24 gave, after column chromatography (1:40 methanol-ethyl acetate, followed by 1:30) of the residue, 34 (85 mg, 74%): [α]_D²⁵ + 35° (c 0.69, chloroform). N.m.r. data (CDCl₃): ¹H (500.14 MHz), δ 8.19–7.95 (m, 4 H, NHCHO), 3.29 (bs, 3 H, MeO), and 1.25–1.02 (m, 15 H, H-6abcde); ¹³C (125.76 MHz), δ 164.8 and 161.3 (2 m, NHCHO), 101.3–99.4 (m, C-1abcde), and 51.2–50.0 (m, C-4abde-Z).

Anal. Calc. for $C_{77}H_{90}N_4O_{21}$: C, 65.5; H, 6.71; N, 3.97. Found: C, 65.5; H, 6.76; N, 4.04.

Methyl O-(4,6-dideoxy-4-formamido-α-D-mannopyranosyl)-(1→2)-O-(4,6-dideoxy-4-formamido-α-D-mannopyranosyl)-(1→2)-O-α-D-rhamnopyranosyl-(1→2)-O-(4,6-dideoxy-4-formamido-α-D-mannopyranosyl)-(1→2)-4,6-dideoxy-4-formamido-α-D-mannopyranoside (**35**). — Debenzylation of **34** (86 mg, 61 µmol) and column chromatography of the residue on Bio-Gel P-4 as described for **25** gave, after freeze drying, **35** (43 mg, 81%): $[\alpha]_D^{25}$ + 44° (c 0.61, water). ¹H-N.m.r. showed a Z: E-ratio of ~4.0:1 for the formamido groups. N.m.r. data (D₂O): ¹H (500.14 MHz), δ 8.20–8.18 (4 s, 3.2 H, NHCHO-Z), 8.03–8.00 (4 s, 0.8 H, NHCHO-E), 5.17, 5.15, 5.09, and 5.03 (4 bs, each 1 H, H-1bcde), 4.79 (bs, 1 H, H-1a), 3.47 (t, 1 H, J 9.7 Hz, H-4c), 3.39 (s, 3 H, MeO), and 1.27–1.17 (m, 15 H, H-6abcde); ¹³C (125.76 MHz), δ 168.1 (NHCHO-E), 165.1 (NHCHO-Z), 102.2 (C-1e), 101.0, 100.9, and 100.8 (C-1bcd), 99.8 (C-1a), 78.1, 77.5, 77.4, and 77.2 (C-2abcd), 57.1 and 56.9 (C-4abde-E), 55.2 (MeO), and 52.2 and 51.9 (C-4abde-Z).

REFERENCES

- 1 M. Caroff, D. R. Bundle, M. B. Perry, J. W. Cherwonogrodzky, and J. R. Duncan, Infect. Immun., 46 (1984) 384-388.
- 2 D. R. Bundle, J. W. Cherwonogrodzky, and M. B. Perry, Biochemistry, 26 (1987) 8717-8726.
- 3 D. R. Bundle, J. W. Cherwonogrodzky, M. A. J. Gidney, P. J. Meikle, M. B. Perry, and T. Peters, *Infect. Immun.*, 57 (1989) 2829–2836.
- 4 D. R. Bundle, Pure Appl. Chem., 61 (1989) 1171-1180.
- 5 T. Peters, J.-R. Brisson, and D. R. Bundle, Can. J. Chem., 68 (1990) 979-988.
- 6 R. Oomen, N. M. Young, and D. R. Bundle, in J. W. Streilein, et al. (Eds.), ICSU Short Reports, Vol. 10, Proceedings of the 1990 Miami Bio/Technology Winter Symposia, IRL Press, Oxford, p. 56; Protein Engeneering, in press.

- 7 D. R. Bundle, M. Gerken, and T. Peters, Carbohydr. Res., 174 (1988) 239-251.
- 8 M. J. Eis and B. Ganem, Carbohydr. Res., 176 (1988) 316-323.
- 9 T. Peters and D. R. Bundle, Can. J. Chem., 67 (1989) 491-496.
- 10 A. K. Ray, U. B. Maddali, A. Roy, and N. Roy, Carbohydr. Res., 197 (1990) 93-100.
- 11 F.-I. Auzanneau and D. R. Bundle, Carbohydr. Res., (1991) in press.
- 12 H. G. Fletcher, Jr. and H. W. Diehl, Carbohydr. Res., 17 (1971) 383-391.
- 13 J. O. Kihlberg, D. A. Leigh, and D. R. Bundle, J. Org. Chem., 55 (1990) 2860-2863.
- 14 K. Bock and C. Pedersen, J. Chem. Soc., Perkin Trans. 2, (1974) 293-297.
- 15 F. Dasgupta and P. J. Garegg, Carbohydr. Res., 177 (1988) c13-c17.
- 16 T. Adachi, Y. Yamada, I. Inoue, and M. Saneyoshi, Synthesis, (1977) 45-46.
- 17 M. Vaultier, N. Knouzi, and R. Carrié, Tetrahedron Lett., 24 (1983) 763-764.
- 18 J. S. Brimacombe, J. G. H. Bryan, A. Husain, M. Stacey, and M. S. Tolley, Carbohydr. Res., 3 (1967) 318-324.
- 19 M. Bartra, F. Urpi, and J. Vilarrasa, Tetrahedron Lett., 28 (1987) 5941-5944.
- 20 L. Kenne, P. Unger, and T. Wehler, J. Chem. Soc., Perkin Trans. 1, (1988) 1183-1186.
- 21 V. Pozsgay, J.-R. Brisson, and H. Jennings, Can. J. Chem., 65 (1987) 2764-2769.
- 22 V. Poszgay and A. Neszmélyi, Carbohydr. Res., 80 (1980) 196-202.