Approach to the Synthesis of Annonaceous Acetogenins from D-Glucose

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Abstract: The preparation from D-glucose of threo-trans-threo and threo-cis-threo synthons 12 and 13 for the monochiral synthesis of mono (or eventually bis) tetrahydrofuran acetogenins is described. Their structure have been proven by degradation and comparison with a racemic sample of a threo-cis-threo aldehyde prepared by permanganate oxidation of the corresponding diene.

More than thirty tetrahydrofuran acetogenins have been isolated from Annonaceae since the discovery of uvaricin 1 in 1982^1 and shown to have various potent biological activities such as cytotoxic, antitumoral, antimalarial, immunosuppressive, pesticidal or antifeedant.² This new class of compounds, with a C_{35} - C_{37} skeleton, is characterized by the presence of one or two (generally adjacent) tetrahydrofuran ring(s) and a lactone moiety, together with hydroxyl (or scarcely carbonyl) functions. Examples of such monotetrahydrofuran acetogenins are annonacin 2^3 and annonacinone $3.^4$



The determination of the relative stereochemistry of these acetogenins is difficult due to their waxy properties and to the large number of CH-O bonds. However synthetic studies of model bistetrahydrofurans by Hoye et al.,⁵ X-Ray analysis of derivatives of rolliniastatin by Pettit⁶ and annonin I (or squamocin) by Born⁷, together with extensive ¹H and ¹³C NMR analysis have demonstrated that the most common arrangement involving the tetrahydrofuran ring(s) is *threo-trans-threo* (as for 2 and 3) although *erythro*-

trans-threo, erythro-cis-threo and threo-cis-threo relative configurations are also known such as respectively in uvaricin, rolliniastatin⁸ and annonin VI.⁷ The absolute configurations of all the acetogenins are uncertain apart for the asymmetric centers of the lactone moiety determined for some acetogenins using CD spectra and degradation to (S)-lactic acid.² Recently Hoye has shown that the chirality of uvaricin is probably as depicted in 1 after having synthesized a (15,16,19,20,23,24)-hexepimer, but, as quoted by Hoye, any prediction of absolute configurations for other acetogenins is hazardous taking into account a probable polyepoxide biosynthetic pathway.9

In order to gain more information on their structures and later on their structure-activity relationships, the synthesis of monochiral acetogenins has thus been undertaken from carbohydrates such as D-glucose and preliminary results are reported here.



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The known aldehyde 410 is first reacted with the Grignard reagent of 1-bromo-3-tetradecyne11 to give an easily separable 4 to 1 mixture of 5,¹² mp 79-80°C, $[\alpha]_D$ -33 (c 1, CCl₄) and 6¹² as an oil, in agreement with similar results reported by Hanessian¹³ and Inch,¹⁴ best explained by the Cram chelation controlled model. After Lindlar reduction of 5 to the Z-alkene 7,¹² mp 60-61°C, $[\alpha]_D$ -27 (c 1, CCl₄), epoxidation (metachloroperbenzoïc acid, CH₂Cl₂) affords as expected a 1/1 unseparable mixture of the two possible epoxides leading (AcOH, CH₂Cl₂, 20°C, 12h) to the trans and cis tetrahydrofurans 8 and 9. Although these compounds can only be separated by HPLC, it was expected that further protecting groups modification will permit easier separation on a preparative scale. Thus it was found that benzoylation to 10 and 11 followed by hydrogenolysis (H₂, Pd/C) affords as oils 12, 1^{12} [α]_D -22.2 (c 1.1, CCl₄) and 13¹² [α]_D 2.6 (c 0.9, CCl₄) which are then isolated (30% each overall from 7) by flash chromatography (silica, AcOEt/hexane 30/70 V/V, 12: R_F 0.6, 13: R_F 0.4). Unfortunately at this stage ¹H and ¹³C NMR analysis does not allow an unambiguous determination of the cis or trans relationship in the newly created tetrahydrofuran ring. Therefore 12 and 13 were respectively degradated to aldehydes 14¹² and 15¹² in two steps (AcOH-H2O 4-1, 50°C, overnight then 3 eq. NaIO4, H2O, 20°C). A racemic sample of the cis aldehyde 15 was prepared from diene 16 through KMnO₄ oxidation¹⁵ to the cis tetrahydrofuran 17 (15%), protection of the primary alcohol group (tBuMe₂SiCl), benzoylation (PhCOCl, pyridine), deprotection (nBu₄N⁺F⁻) and oxidation (PCC, CH₂Cl₂). The ¹H and ¹³C NMR spectra of this latter compound are identical to those of aldehyde 15 obtained from the more polar alcohol 13 and clearly different from those of 14. A further confirmation arised from the base catalyzed hydrolysis of 14 and 15 which gave respectively a single hydroxyaldehyde 18 and a mixture of the free aldehyde 19 and hemiacetal 20.16

In conclusion, the preparation of *threo-trans-threo* and *threo-cis-threo* tetrahydrofuran derivatives 12 and 13 is easily carried out from diacetone D-glucose in 8 steps and these intermediates will be used in the synthesis of mono and bistetrahydrofuran acetogenins.¹⁷ The use of other conditions to carry out the alkene epoxidation is also considered and if $oxone^{(R)}$ gives also after acid treatment a 1/1 ratio of 8 and 9, the use of tBuOOH-VO(Acac)₂ favors, as expected,^{18,19} the *cis* isomer 9 (2/1 ratio). Finally the R alcohol 6 has been submitted to the same sequence of reactions and the two corresponding *trans* and *cis* tetrahydrofurans have also been shown to be easily separable at the same stage.

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- 11. This compound was prepared (74% overall) from 1-dodecyne in 3 steps: 1-BuLi, ethylene oxide; 2-TsCl, pyridine; 3-MgBr₂, ether.
- 12. All new compounds have been characterized by ¹H (200 MHz) and ¹³C NMR (50.3 MHz), IR, MS and HRMS or elemental analysis. Selected NMR data (CDCl₃ as solvent, TMS as internal standard, J in Hz) are given below.

5: δ 0.88 (t, 3H, J= 7), 1.48 (s, 3H), 1.60 (s, 3H), 2.11 (m, 2H), 2.33 (m, 2H), 2.83 (s, OH), 3.97 (d, 1H, J= 3.3), 4.49 (d, 1H, J= 11.8), 4.65 (d, 1H, J= 3.8), 4.72 (d, 1H, J= 11.8), 5.99 (d, 1H, J= 3.8), 7.35 (s, 5H) ppm.

6: δ 0.88 (t, 3H, J=7), 1.32 (s, 3H), 1.48 (s, 3H), 2.12 (m, 2H), 2.33 (m, 2H), 4.06 (m, 3H), 4.53 (d, 1H, J= 11.8), 4.63 (d, 1H, J= 3.8), 4.73 (d, 1H, J= 11.8), 5.96 (d, 1H, J= 3.8), 7.33 (s, 5H) ppm.

7: δ 0.88 (t, 3H, J= 7), 1.34 (s, 3H), 1.49 (s, 3H), 2.02 (m, 2H), 2.20 (m, 2H), 2.78 (s, OH), 4.46 (d, 1H, J= 11.8), 4.65 (d, 1H, J= 3.9), 4.72 (d, 1H, J= 11.8), 5.3 (m, 2H), 5.99 (d, 1H, J= 3.8), 7.33 (s, 5H) ppm.

12: $\delta 0.88$ (t, 3H, J= 7), 1.31 (s, 3H), 1.47 (s, 3H), 2.14 (m, 4H), 4.07 (s, 1H), 4.24 (s, 1H), 4.32 (q, 1H, J= 6), 4.70 (d, 1H, J= 1.6), 5.14 (q, 1H, J= 5), 5.94 (d, 1H, J= 3.6), 7.44 (t, 2H, J= 7.4), 7.56 (t, 1H, J= 7), 8.05 (d, 2H, J= 7.4) ppm; δ 14.04, 22.67, 25.41, 26.23, 26.89, 27.48, 29.28, 29.31, 29.44, 29.55, 31.26, 31.91, 75.72, 78.48, 80.12, 81.78, 85.60, 93.17, 104.81, 111.53, 128.43, 129.69, 130.51, 132.93, 166.40 ppm (77.7 masked). 13: δ 0.88 (t, 3H, J= 7), 1.31 (s, 3H), 1.47 (s, 3H), 1.88 (d, 1H, J= 7), 1.91 (d, 1H, J= 7), 2.10 (m, 2H), 4.05 (s, 1H), 5.25 (q, 1H, J= 6.2), 5.92 (d, 1H, J= 3.6), 7.43 (t, 2H, J= 7), 7.55 (t, 1H, J= 7.4), 8.08 (d, 2H, J= 7.4) ppm; δ 14.05, 22.67, 25.34, 26.28, 26.95, 27.47, 29.31, 29.46, 29.55, 29.58, 29.71, 31.18, 31.92, 75.80, 77.10, 78.53, 80.30, 82.19, 85.81, 104.95, 111.64, 128.33, 129.84, 130.62, 132.82, 166.48 ppm.

14: $\delta 0.87$ (t, 3H, J=6), 1.75 (m, 2H), 2.0 (m, 2H), 4.31 (td, 1H, J= 5 and 2.5), 4.41 (t, 1H, J= 8), 5.18 (td, 1H, J= 8 and 5), 7.45 (d, 2H, J= 7.4), 7.55 (t, 1H, J= 7), 8.05 (d, 2H, J= 7.4), 9.66 (d, 1H, J= 1.2) ppm; δ 14.05, 22.67, 25.50, 26.80, 27.34, 27.63, 29.32, 29.47, 29.56, 29.59, 31.31, 31.92, 75.65, 81.45, 83.25, 128.46, 129.76, 130.53, 132.98, 166.38, 202.20 ppm.

15: δ 0.86 (t, 3H, J= 7), 2.10 (m, 4H), 4.29 (m, 2H), 5.21 (td, 1H, J= 6.7 and 5), 7.47 (t, 2H, J= 7.4), 7.56 (t, 1H, J= 7), 8.04 (d, 2H, J= 7.4), 9.78 (d, 1H, J= 1.5) ppm; δ 14.06, 22.69, 25.50, 27.78, 28.01, 29.33, 29.48, 29.57, 29.59, 31.50, 31.94, 75.67, 81.75, 83.42, 128.50, 129.74, 130.45, 133.02, 166.38, 203.11 ppm.

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