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Synthesis of α -L-Mannopyranosyl-containing Disaccharides and Phenols as Substrates for the α -L-Mannosidase Activity of Commercial Naringinase

Sachiko Esaki, Akemi Ohishi, Akiko Katsumata, Naoko Sugiyama, and Shintaro Kamiya

Department of Food Science, School of Food and Nutritional Sciences, University of Shizuoka, Shizuoka 422, Japan Received June 7, 1993

In order to clarify the substrate specificity of the α -L-mannosidase activity of naringinase (Sigma), the following disaccharides and phenol glycosides were freshly prepared: methyl 2-O-(α -L-mannopyranosyl)- β -D-glucoside (1), methyl 3-O-(α -L-mannopyranosyl)- α -D-glucoside (2), methyl 4-O-(α -L-mannopyranosyl)- α -D-glucoside (3), methyl 5-O-(α -L-mannopyranosyl)- β -D-glucoside (4), methyl 6-O-(α -L-mannopyranosyl)- α -D-glucoside (5), 6-O-(α -L-mannopyranosyl)-D-galactose (6), *p*-nitrophenyl α -L-mannoside (7), and 4-methyl umbelliferone α -L-mannoside (8). These compounds, except for 3 and 5, were hydrolyzed with naringinase.

Naringin, whose structure is 4',5,7-trihydroxyflavanone 7-O-[2-O-(α -L-rhamnopyranosyl)- β -D-glucopyranoside], is a bitter principle of several citrus juices, and its hydrolyzing enzyme "naringinase" has been used on an industrial scale to remove the bitterness from such juices. The term "naringinase" is given to multiple enzymes which contain α -L-rhamnosidase and β -D-glucosidase activities.

Until now, several α -L-rhamnosidases have been isolated from various microorganisms, e.g., Aspergillus niger, Aspergillus oryzae and Penicillium decumbens. Michon et al.¹⁾ have recently reported the substrate specificity of naringinase, a commerical preparation obtained from Penicillium decumbens (Sigma), by using a number of synthetic α -L-rhamnooligosides and two disaccharides containing an α -L-mannopyranosidic bond as substrates. They found that both α -L-rhamonosides and, surprisingly, α -L-mannosides were hydrolyzed, indicating that the hydroxylmethyl group at C-5 of the terminal residue of each did not retard the hydrolysis of these oligomeric molecules.

On the other hand, we have reported in a previous paper²) that α -L-rhamnosidase obtained from Aspergillus niger No. 0 was inactive toward disaccharides having an α -L-mannopyranosidic bond. Since we are interested in the α -L-mannosidase activity of the foregoing commercial naringinase, several disaccharides containing non-reducing terminal α -L-mannopyranosyl residues were synthesized, and their susceptibility to enzymatic hydrolysis with naringinase was examined. Additionally, *p*-nitrophenyl α -L-mannopyranoside and umbelliferone α -L-mannopyranoside were prepared and their hydrolysis with naringinase was tested.

The following disaccharides were prepared in order to use as the enzyme substrates: methyl 2-O-(α -L-mannopyranosyl)- β -D-glucopyranoside (1), methyl 3-O-(α -L-mannopyranosyl)- α -D-glucopyranoside (2), methyl 4-O-(α -L-mannopyranosyl)- α -D-glucopyranoside (3), methyl 5-O-(α -Lmannopyranosyl)- β -D-glucopyranoside (4), methyl 6-O-(α -Lmannopyranosyl)- α -D-glucopyranoside (5) and 6-O-(α -Lmannopyranosyl)- α -D-glucopyranoside (5) and 6-O-(α -Lmannopyranosyl)-D-galactopyranose (6). The syntheses of 1-6 do not appear to have been described previously.

Treatment of 1,3,4,6-tetra-O-acetyl- α -D-glucopyranose (7)³⁾ with chloroacetyl chloride gave 1,3,4,6-tetra-O-acetyl-2-O-chloroacetyl- α -D-glucopyranose (8). The α -glycosyl bromide (9) of 8 was treated with methanol in the presence

of silver carbonate to give a corresponding methyl glycoside (10). De-O-chloroacetylation of 10 with thiourea⁴⁾ gave methyl 3,4,6-tri-O-acetyl- β -D-glucopyranoside (11). The reaction of 2,3,4,6-tetra-O-acetyl-a-L-mannopyranosyl chloride (12)⁵⁾ with 11 in the presence of mercury(II) cyanide and mercury(II) bromide³⁾ in acetonitrile gave coupling product 13 in a 23.5% yield, which, after deprotection, afforded 1. The reaction of 12 with methyl 2-O-benzoyl-4,6-O-benzylidene- α -D-glucopyranoside (14)⁶⁾ in the presence of silver triflate and tetramethylurea⁷) in dichloromethane gave coupling product 15 in a 26. 1% yield, which, after deprotection, afforded 2. Coupling 2,3,4,6tetra-O-benzoyl- α -L-mannopyranosyl bromide (16)⁸⁾ to methyl 2,3,6-tri-O-benzoyl- α -D-glucopyranoside (17)⁹ in the presence of silver triflate and 4A molecular sieves gave reaction product 18 in a 33% yield, which, after de-O-benzoylation, gave 3. Treatment of 12 with 3,6-di-Oacetyl-1,2-O-isopropylidene- α -D-glucofuranose (19)¹⁰ in the presence of mercury(II) cyanide and mercury(II) bromide gave coupling product 20 in a 42.5% yield, which, after deacetylation gave 1,2-O-isopropylidene-5-O-(α -Lmannopyranosyl)- α -D-glucofuranose (21). Compound 21, after de-O-isopropylidenation, was reacetylated with acetic anhydride and pyridine, and the resultant product (22) was further treated with hydrogen bromide to yield a bromide (23). Methanolysis of 23 in the presence of silver carbonate, gave methyl glycoside 24, which was deacetylated to yield 4. Methyl 2,3,4-tri-O-acetyl- α -D-glucopyranoside (25)¹¹⁾ was reacted with 12 in the presence of mercury(II) cyanide

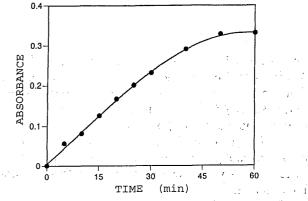
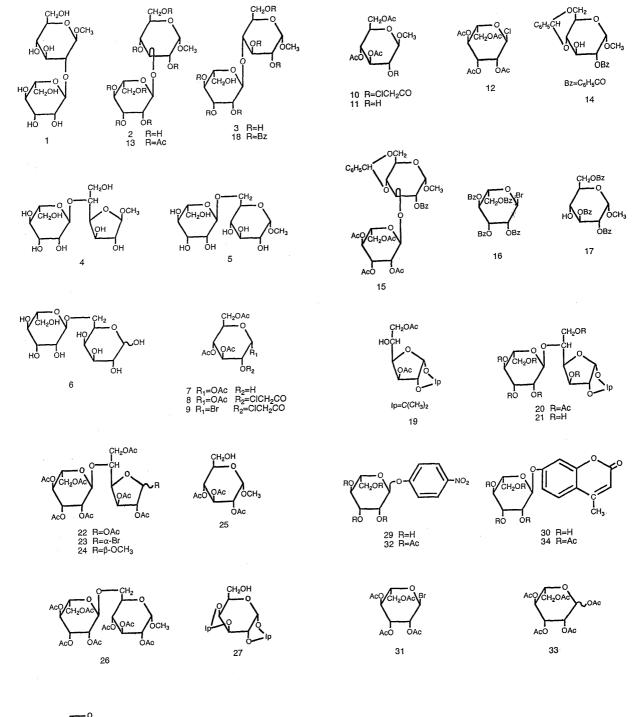
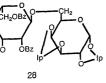


Fig. 1. Relationship between the Reaction Time and the Absorbance of Liberated *p*-Nitrophenol.





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Fig. 2.

and mercury(II) bromide to yield coupling product **26** in a 49.2% yield. Zémplen de-*O*-acetylation of **26** then gave **5**. Condensation of 1,2,3,4-di-*O*-isopropylidene- α -D-galacto-pyranose (**27**)¹²) with **16** in the presence of mercury(II) cyanide and mercury(II) bromide gave coupling product **28** in a 15.1% yield, which, after deprotection, afforded **6**. The anomeric configuration of the α -L-mannopyranoside bond in each of **1–6** was determined to be α -form by applying Klyne's rule.¹³)

p-Nitrophenyl α -L-mannopyranoside (29), 4-methyl umbelliferone α -L-mannopyranoside (30) were also synthesized.

Coupling 2,3,4-tri-O-acetyl- α -L-mannopyranosyl bromide $(31)^{14}$ to *p*-nitrophenol in aqueous sodium hydroxide¹⁵) gave the reaction product (32) in a 10.8% yield, which was subsequently deacetylated to afford 29. As an alternative method, well-mixed penta-O-acetyl-L-mannopyranose (33) and *p*-nitrophenol were heated at 100°C in the presence of *p*-toluenesulfonic acid¹⁶ to give foregoing coupling product 32 in a 24.4% yield.

Heating 33 and 4-methyl umbelliferone in an acetic acidacetic anhydride mixture in the presence of anhydrous zinc chloride gave coupling product 34 in a 15.4% yield, which was deacetylated to afford 30.

A study of the naringinase action toward compounds 1–6 was made by applying HPLC method. As a result, the $(1\rightarrow 2)$ - $(1\rightarrow 3)$ - and $(1\rightarrow 5)$ -linked methyl α -L-mannopyranosyl-D-glucosides and $(1\rightarrow 6)$ -linked α -L-mannopyranosyl-D-galactose were hydrolyzed, while, unexpectedly, the methyl $(1\rightarrow 4)$ - and $(1\rightarrow 6)$ -linked α -L-mannopyranosyl-Dglucosides did not accept the enzyme action at all.

4-Methyl umbelliferone α -L-mannopyranoside and *p*nitrophenyl α -L-mannopyranoside were hydrolyzed by the α -L-mannosidase activity of naringinase, providing 4-methyl umbelliferone and *p*-nitrophenol, respectively. Therefore, the progress of the reaction can be followed by spectrophotometrically measuring the amount of ether of these aglycones (see Fig. 1).

Experimental

General methods. Reactions were monitored by TLC on silica gel 60 (Merck), using detection by charring with 10% sulfuric acid in ethanol. Analytical high-pressure liquid chromatography (HPLC) was performed with Shimadzu LC-6S apparatus equipped with a Shim-pack (CLC-NH₂) column, using a differential refractometer as a detector and acetontrile-water (7:3) at a flow rate of 1 ml/min. Optical rotation values were measured with a Horiba SEPA-200 digital polarimeter. NMR spectra (¹H at 90 Mz and ¹³C at 24.5 MHz) were recorded with a Hitachi R-90H spectrometer, using tetramethylsilane as an internal standard for solutions in CDCl₃. Microanalyses were performed by the Microanalytical Laboratory of the School of Pharmaceutical Sciences at this university.

1,3,4,6-Tetra-O-acetyl-2-O-chloroacetyl- α -D-glucopyranose (8). Compound 7 (10 g) was dissolved in acetonitrile (430 ml), before pyridine (7 ml) was added and the solution cooled to 0°C. A solution of chloroacetyl chloride (3.2 ml) in acetonitrile (35 ml) was added dropwise, the mixture being kept at 0°C for 1 h and then at room temperature for 3 h. After diluting with chloroform, the reaction mixture was successively washed with 5% HCl, aq. NaHCO₃ and water, and dried (CaCl₂). The filtrate was concentrated, and the residue was crystallized from ethanol to provide 8 (10.8 g, 89.1%), R_f 0.27 (toluene-ethyl acetate 8:2), mp 147.5°C, $[\alpha]_D^{20}$ 91° (c=1, CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 1.98, 2.00, 2.06, 2.15 (each 3H, s, 4 CH₃CO); 3.93 (2H, s, ClCH₂CO); 4.15 (3H, m, H-5, 6); 4.95–5.60 (3H, m, H-2-4); 6.32 (1H, d, J=4 Hz, H-1). Anal. Found: C, 44.94; H, 4.79. Calcd. for C₁₆H₂₁O₁₁Cl: C, 45.23; H, 4.95%.

3,4,6-Tri-O-acetyl-2-O-chloroacetyl- α -D-glucopyranosyl bromide (9). Compound 8 (10.87g) was dissolved in chloroform (42 ml), and 30% hydrogen bromide in acetic acid (56 ml) was added at 0°C. The mixture was kept at 0°C for 1 h and then at room temperature for 1 h. After diluting with chloroform, the reaction mixture was successively washed with water (three times), aq. NaHCO₃ and water again, and then dried (CaCl₂). The filtrate was concentrated to a syrup which was crystallized from ether-petroleum ether (1:3) to yield 9 (10.5 g, 92%), R_f 0.58 (toluene-ethyl acetate 7:3), mp 107–108°C, $[\alpha]_D^{20}$ 157° (c=1, CHCl₃). Anal. Found: C, 37.49; H, 3.88. Calcd. for C₁₄H₁₈BrClO₉: C, 37.71; H, 4.04%.

Methyl 3,4,6-tri-O-acetyl-2-O-chloroacetyl- β -D-glucopyranoside (10). Compound 9 (10.49 g) was dissolved in methanol (580 ml), silver carbonate (6.5 g) was added, and the mixture was stirred for 24 h at room temperature in the dark. After filtering through a bed of Celite, the filtrate was concentrated, and the residue was chromatographed in a column of silica gel (toluene-ethyl acetate, 7:3) to afford 10 (0.76 g, 8.2%), after crystallizing from ethanol, R_f 0.37 (toluene-ethyl acetate, 7:3), mp 91–92°C, $[\alpha]_{D}^{20} - 9^{\circ}$ (c=1, CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 1.96, 2.00, 2.05 (each 3H, s, 3CH₃CO); 3.46 (3H, s, OCH₃); 3.98 (2H, s, CICH₂CO); 4.15 (2H, t, H-6); 4.40 (1H, d, J=8 Hz, H-1); 4.80–5.10 (3H, m, H-2-5). Anal. Found: C, 45.34; H, 5.17. Calcd. for C₁₅H₂₁ClO₁₀: C, 45.40; H, 5.30%.

Methyl 3,4,6-tri-O-acetyl- β -D-glucopyranoside (11). To a solution of 10 (0.76 g) in 75% aq. acetonitrile (11 ml) was added thiourea (1.36 g), and the mixture was kept at room temperature overnight while stirring. After diluting with chloroform, the reaction mixture was sufficiently washed with water and then dried (CaCl₂). The filtrate was concentrated, and the residue

was crystallized from ether-petroleum ether to afford **11** (0.35 g, 57.4%), $R_{\rm f}$ 0.22 (toluene-ethyl acetate 7:3), mp 96°C, $[\alpha]_{\rm D}^{20}$ 14° (c=1, CHCl₃); lit.¹²⁾ 97–99°C, $[\alpha]_{\rm D}^{20}$ 18° (c=1, CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 2.00 (3H), 2.04 (6H each s, 3CH₃CO); 2.60 (1H, s, OH); 3.53 (3H, s, OCH₃); 4.00–4.40, 4.90–5.20 (each m, sugar protons).

Methyl 1,3,4,6-tetra-O-acetyl-2-O-(2,3,4,6-tetra-O-acetyl- α -L-mannopyranosyl)- β -D-glucopyranoside (13). To a solution of 11 (0.25 g) in acetonitrile (3 ml) containing mercury(II) cyanide (0.1 g) and mercury(II) bromide (0.14 g) was added 12 (0.29 g). The mixture was kept at room temperature overnight, diluted with chloroform, and then filtered to remove the mercury(II) salts. The filtrate was washed three times with 1 N KBr solution, and then dried (CaCl₂). The filtrate was concentrated, and the residue was chromatographed in a column of silica gel (chloroformacetone, 15:1) to give 13 (0.12 g, 23.5%), $R_{\rm f}$ 0.30 (chloroform-acetone 9:1), $[\alpha]_D^{20} - 7^{\circ}$ (c=1, CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 1.96 (3H), 1.98 (3H), 2.00 (3H), 2.03 (6H), 2.05 (3H), 2.10 (3H each s, 7CH₃CO); 3.50 (3H, s, OCH₃); 4.00–4.40 (7H), 4.80–5.35 (7H, each m, sugar protons). Anal. Found: C, 49.95; H, 5.73. Calcd. for C₂₇H₃₈O₁₈: C, 49.85; H, 5.85%.

Methyl 2-O-(α-L-mannopyranosyl)-β-D-glucopyranoside (1). Deacetylation of 13 (0.12 g) with 0.1 N methanolic sodium methoxide (2.5 ml) in the usual manner yielded 1 (50 mg, 83.3%), $[\alpha]_D^{20} - 68^\circ$ (c = 1, CH₃OH). Anal. Found: C, 43.24; H, 6.97. Calcd. for C₁₃H₂₄O₁₁: C, 43.82; H, 6.74%. [M]_D (methyl α-L-mannoside) -15908, [M]_D (methyl β-L-mannoside) 10282, [M]_D (methyl β-D-glucoside) -6790; observed [M]_D (1) -24208, Calcd. [M]_D (1) -22698, Calcd. [M]_D (β-anomer of 1) 3492.

Methyl 2-O-benzoyl-4,6-O-benzylidene-3-O-(2,3,4,6-tetra-O-acetyl- α -L-mannopyranosyl)- α -D-glucopyranoside (15). To a stirred solution of 14 (2.79 g, 1 mmol), silver triflate (2.76 g, 1.2 mmol) and 1,1,3,3-tetramethylurea (3 ml) in dichloromethane (35 ml) was added dropwise a solution of 12 (2.97 g, 1.2 mmol) in dichloromethane (8.6 ml). After stirring for 4 h, the reaction mixture was filtered through a bed of Celite, and the filtrate was successively washed with sat. NaHCO₃ and water, before being dried (MgSO₄). After filtering, the filtrate was evaporated to a syrup, which was crystallized from ethanol to give 15 as colorless needles (1.33 g, 26.1%), mp 167–169°C, R_f 0.30 (toluene–ethyl acetate 8:2), $[\alpha]_D^{20}$ 47° (c=1, CHCl₃). Anal. Found: C, 58.38; H, 5.50. Calcd. for C₃₄G₄₀O₁₆: C, 57.95; H, 5.68%.

Methyl 3-O-(α -L-mannopyranosyl)- α -D-glucopyranoside (2). A solution of 15 (0.92 g) in 60% acetic acid (9 ml) was heated at 100°C for 30 min. The solvent was evaporated, and the residue was repeatedly distilled with toluene to remove all traces of the volatile compounds. The de-*O*benzylidene compound (0.73 g) was dissolved in 0.1 N methanolic sodium methoxide (15 ml) and left to stand for 2 h at room temperature. After neutralizing with Amberlite IR-120 (H⁺) resin, the filtrate was evaporated to give 2 as a syrup (0.38 g, 92.3%), $[\alpha]_{B}^{20}$ 27° (*c*=1, CH₃OH). The sum of the [M]_D values for methyl α -L-mannoside (-15908) and for methyl α -D-glucoside (30846) was 14938. The sum of the [M]_D values for methyl β -L-mannoside (10280) and for methyl α -D-glucoside (30840) was 41126. The observed [M]_D value for 2 was 9612; therefore, the configuration of the L-mannosidic bond of 2 was deduced to be α -form. Anal. Found: C, 43.82; H, 6.74. Calcd. for C₁₃H₂₄O₁₁ · 1/2CH₃OH: C, 43.55; H, 6.99%.

Methyl 2,3,4,6-tetra-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- α -L-mannopyranosyl)- α -D-glucopyranoside (18). To a solution of 16 (31.2 g) and 17 (24 g) in dry dichloromethane (800 ml) was added silver triflate (18.92 g) and 4A molecular sieves (105 g) while ice-cooling. The solution was stirred at 0° for 15 min and then at room temperature for 2 h in the dark. After filtering through a bed of Celite, the filtrate was successively washed with sat. NaHCO₃ and water, and dried (CaCl₂). Removal of the solvent gave a syrup (52 g), from which a 15 g sample was taken and chromatographed in a column of silica gel (toluene-ethyl acetate, 9:1) to give a syrup (14), which was crystallized from ethanol as needles (5 g, 33%), R_f 0.41 (toluene-ethyl acetate 9:1), mp 62–65°C, $[\alpha]_{D}^{20}$ 59° (c=1, CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 3.50 (3H, s, OCH₃); 3.90–6.20 (m, sugar protons); 7.20–7.65 (21H); 7.70–8.20 (14H, each, m, 7C₆H₄CO). Anal. Found: C, 66.00; H, 4.77. Calcd. for C₆₂H₅₂O₁₈·2H₂O: C, 66.42; H, 5.00%.

Methyl 4-O-(α -L-mannopyranosyl)- α -D-glucopyranoside (3). A solution of 18 (4.14 g) in 0.1 N methanolic sodium methoxide (90 ml) was left overnight at room temperature. After treating the reaction mixture as already mentioned, 3 was obtained as a syrup (1.2 g, 89.8%), $[\alpha]_D^{20} 42^\circ$

(c=1. CH₃OH). The observed [M]_D for **3** was 16732. Accordingly, the anomeric configuration of the L-mannosyl bond of **3** was deduced to be α -form. *Anal.* Found: C, 40.12; H, 6.69. Calcd. for C₁₃H₂₄O₂·H₂O: C, 39.79; H, 7.14%.

3,6-Di-O-acetyl-1,2-O-isopropylidene-5-O-(2,3,4,6-tetra-O-acetyl- α -Lmannopyranosyl)- α -D-glucofuranose (20). To a solution of Hg(CN)₂ (2.88 g) and HgBr₂ (4.11 g) in acetonitrile (87 ml) were added 19 (6.94 g) and 12 (8.37 g), and the mixture was left overnight at room temperature. After removing the solvent, the residue was dissolved in chloroform and filtered, and the filtrate was washed with aq. 1 N KBr and dried (CaCl₂). The filtrate was then concentrated, and the residual syrup was chromatographed in a column of silica gel, using benzene–ether (2:1) as the eluent, to give 20 (2.14 g, 42.5%), R_f 0.20 (chloroform–acetone 15:1), $[\alpha]_D^{20} - 30^\circ$ (c=1, CHCl₃). NMR δ_H (CDCl₃): 1.29 (3H), 1.50 (3H each s, 2CH₃); 1.95 (3H), 2.00 (3H), 2.06 (3H), 2.10 (6H), 2.12 (3H, each s, 6CH₃CO); 3.80, 4.60 (8H), 4.98 (1H, d, J=1 Hz, H-1'); 5.15–5.40 (4H); 6.83 (1H, d, J=4 Hz, H-1). Anal. Found: C, 51.05; H, 6.02. Calcd. for C₂₇H₃₈O₁₇: C, 51.10; H, 5.99%.

1,2-O-Isopropylidene-5-O-(α-L-mannopyranosyl)-α-D-glucofuranose (21). Compound 20 (2.0 g) was deacetylated with 0.1 N methanolic sodium methoxide (40 ml) as already mentioned to afford syrupy 21 (0.84 g, 69.4%), $[\alpha]_D^{20} - 48^\circ$ (c = 1, CH₃OH). NMR δ_H (DMSO- d_6): 1.20 (3H), 1.33 (3H, each s, 2CH₃); 3.10–5.10 (m, sugar protons); 5.73 (1H, d, J = 4 Hz, H-1). Anal. Found: C, 45.12; H, 6.81. Calcd. for C₁₅H₂₆O₁₁ · H₂O: C, 45.00; H, 7.00%.

Octa-O-acetyl-5-O-(α-L-mannopyranosyl)-D-glucofuranose (22). A solution of 21 (0.84 g) in 1 N HCl (40 ml) was heated at 65°C for 2 h while stirring. After neutralizying with Amberlite IRA-400 (OH⁻ form), the solution was evaporated, and 5-O-(α-L-mannopyranosyl)-D-glucofuranose was obtained in a quantitative yield, R_f 0.4 (butanol-acetic acid-water 4:1:2), $[\alpha]_D^{20} - 49^\circ$ (c=1, CH₃OH). Anal. Found: C, 39.05; H, 6.75. Calcd. for C₁₂H₂₂O₁₁·2·1/2H₂O: C, 39.02; H, 6.77%. Acetylation of the free sugar (0.89 g) with a mixture of acetic acid (10 ml) and pyridine (10 ml) in the usual manner gave 22 (1.1 g, 62.7%), R_f 0.38 (chloroform-acetone 9:1), $[\alpha]_D^{20} - 7^\circ$ (c=1, CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 1.93 (3H), 2.00 (3H), 2.05 (9H), 2.10 (9H, each s, 8CH₃CO); 3.80–6.50 (m, sugar protons); 6.03 (1H, J = 4 Hz, H-1α), 6.37 (1H, d, J = 8 Hz, H-1β). Anal. Found: C, 49.36; H, 5.71. Calcd. for C₂₈H₃₈O₁₉: C, 49.56; H, 5.60%.

2,3,6-Tri-O-acetyl-5-O-(2,3,4,6-tetra-O-acetyl- α -L-mannopyranosyl)- β -D-glucofuranoyl bromide (23). To a solution of 22 (1.1 g) in a small amount of chloroform was added a saturated solution of hydrogen bromide in acetic acid (4.4 ml) at 0°C, and the mixture was kept at the same temperature for 2 h. After diluting with chloroform, the reaction mixture was treated as already described and then concentrated to afford 23 (0.95 g, 84.4%), $[\alpha]_D^{20}$ 21° (c=1, CHCl₃). This compound was unstable, and was used immediately in the next step.

Methyl 2,3,6-*tri-O-acetyl-5-O-(2,3,4,6-tetra-O-acetyl-\alpha-L-mannopyranosyl)-\beta-D-glucofuranoside (24). Compound 23 (0.95 g) was dissolved in methanol (20 ml), siliver carbonate (0.38 g) was added, and the mixture was stirred at room temperature for 1 h in the dark. After filtering through a bed of Celite, the filtrate was concentrated and the residue was chromatographed in a column of silica gel (chloroform-acetone 20:1) to yield 24* (0.1 g, 11.3%), R_f 0.30 (chloroform-acetone 9:1), $[\alpha]_{D^0}^{20}$ -39° (c=1, CHCl₃). NMR δ_H (CDCl₃): 1.96 (3H), 2.00 (3H), 2.07 (3H), 2.09 (3H), 2.10 (6H), 2.12 (3H, each s, 7CH₃CO); 3.50 (3H, s, OCH₃): 3.85–4.40 (m, sugar protons); 4.65 (1H, d, J=8 Hz, H-1 β); 4.80 (1H, d, J=1 Hz, H-1' α); 4.90–5.40 (m, sugar protons). *Anal.* Found: C, 49.53; H, 5.78. Calcd. for C₂₇H₃₈O₁₈: C, 49.53; H, 5.78%.

Methyl 5-O-(α -L-mannopyranosyl)- β -D-glucofuranoside (4). Deacetylation of 24 (0.1 g) with methanolic sodium methoxide (2 ml) in the usual manner gave 4 in a quantitative yield, $[\alpha]_{D}^{20} - 95^{\circ}$ (c=1, CH₃OH). Anal. Found: C, 39.84; H, 7.20. Calcd. for C₁₃H₂₄O₁₁·2H₂O: C, 39.79; H, 7.14%.

Methyl 2,3,4-tri-O-acetyl-6-O-(2,3,4,6-tetra-O-acetyl- α -L-mannopyranosyl)- α -D-glucopyranoside (26). A solution of 25 (2.34 g) and 12 (2.7 g) containing mercury(II) cyanide (0.92 g) and mercury(II) bromide (1.33 g) in acetonitrile (28 ml) was kept overnight at room temperature. After processing as already described, the residual syrup was chromatographed in a column of silica gel (chloroform-acetone, 15:1) to afford 26 (1.28 g, 49.2%), $R_f 0.57$ (chloroform–acetone, 9 : 1), $[\alpha]_D^{20} 26^{\circ} (c = 1, \text{CHCl}_3)$. NMR δ_H (CDCl₃): 1.95 (3H); 2.10 (3H), 2.03 (6H), 2.06 (6H), 2.10 (3H), 2.12 (3H, each s, 8CH₃CO); 3.36 (3H, s, CH₃O); 3.60–4.40 (m), 4.60–4.58 (m, each sugar protons). *Anal.* Found: C, 49.33; H, 5.75. Calcd. for C₂₇H₃₈O₁₈: C, 49.85; H, 5.85%.

Methyl 6-O-(α -L-mannopyranosyl)- α -D-glucopyranoside (5). Deacetylation of **26** (1.28 g) with 0.1 N methanolic sodium methoxide (25 ml) in the usual manner gave 5 (0.59 g, 83.8%), $[\alpha]_D^{20}$ 36° (c = 1, CH₃OH). Anal. Found: C, 43.92; H, 7.06. Calcd. for C₁₃H₂₄O₁₁: C, 43.82; H, 6.74%. [M]_D (methyl α -L-mannoside) -15908, [M]_D (methyl β -L-mannoside) 10289, [M]_D (methyl α -D-glucoside) 30458; observed [M]_D (5) 12816, calcd. [M]_D (5) 14550, calcd. [M]_D (β -L-mannosyl anomer of 5) 40740.

6-O-(2,3,4,6-Tetra-O-benzoyl-α-L-mannopyranosyl)-1,2,3,4-di-Oisopropylidene-α-D-galactopyranose (28). To a solution of 27 (4.57 g) containing mercury(II) cyanide (2.2 g) and mercury bromide (3.16 g) was added 16 (11.58 g), and the mixture was kept overnight at room temperature. After processing as already described, the residual syrup was chromatographed in a column of silica gel (toluene-ethyl acetate, 9:1) to give 28 (2.21 g, 15.1%), R_f 0.25 (toluene-ethyl acetate, 8:1), $[\alpha]_D^{20} 8^{\circ}$ (c=1, CHCl₃). NMR δ_H (CDCl₃): 1.32 (3H), 1.36 (3H), 1.40 (3H), 1.58 (3H, each s, 4CH₃); 3.60–6.20 (m, sugar protons); 7.10–7.60 (12H), 7.70–8.15 (8H, each m, 4C₆H₅CO). Anal. Found: C, 65.35; H, 5.56. Calcd. for C₄₆H₄₂O₁₅ · 1/2H₂O: C, 65.50; H, 5.10.

6-O-(α-L-Mannopyranosyl)-α-D-galactopyranose (6). A solution of **28** (2.2 g) in 0.1 N methanolic sodium methoxide (110 ml) was left overnight at room temperature. After neutralizing with Amberlite IR-120 resin (H⁺ form), the solution was concentrated, and the residue was dissolved in water, washed with ether to remove the benzoic acid, and then concentrated. The residue was dissolved in 0.1 N H₂SO₄, and the solution was heated at 70°C for 2 h. The reaction mixture was neutralized with Amberlite IRA-400 resin (OH⁻ form), filtered and evaporated to afford 6 (0.5 g, 55.6%), R_f 0.1 (butanol-acetic acid-water 4:1:2), $[\alpha]_D^{20} - 29^\circ$ (c=1, CH₃OH). Anal. Found: C, 38.47; H, 6.34. Calcd. for C₁₂H₂₂O₁₁·2H₂O: C, 38.10; H, 6.88%. [M]_D (methyl α-L-mannoside) -15908, [M]_D (methyl β-L-mannoside) 10289, [M]_d (D-galactose) 14400. Observed [M]_D (6) 10962, calcd. [M]_D (6) 1508, calcd. [M]_D (β-L-mannosyl) anomer of 6) 24689.

p-Nitrophenyl α -L-mannopyranoside (29). To a stirred solution of *p*-nitrophenol (5.8 g) in aq. alkali (2.34 g of KOH in 50 ml of water), a solution of 31 (13.8 g) in acetone (90 ml) was added dropwise at 0°C. The mixture was stirred at room temperature for 5 h, and the acetone was distilled off *in vacuo*. The aqueous solution was extracted with chloroform, and the chloroform layer was washed with a 5% NaOH solution to remove the unreacted *p*-nitrophenol, and then with water. After drying with CaCl₂, the filtrate was concentrated, and the residue (32, 1.7 g, 10.8%) was deacetylated with 0.1 N methanolic sodium methoxide to afford 29 (0.1 g, 10%), R_f 0.58 (ethyl acetate-methanol-water 80:14:10), mp 177°C, $[\alpha]_D^{20}$ -160° (c=1, CH₃OH); lit.¹⁷ mp 180°C.

Compound **29** was also prepared according to Helferich's method. A mixture of **33** (6.1 g), *p*-nitrophenol (9 g) and *p*-toluenesulfonic acid (100 mg) was heated at 100°C for 30 min *in vacuo*. The reaction mixture was then diluted with benzene, washed with 1 N NaOH and sufficient water, and dried (CaCl₂). The filtrate was concentrated, and the residual syrup was crystallized from ethanol to give *p*-nitrophenyl 2,3,4,6-tetra-*O*-acetyl- α -L-mannopyranoside (**32**, 2.1 g, 28.8%), R_t 0.36 (toluene–ethyl acetate 7:3), mp 158°C; lit.¹⁷⁾ D-enantiomer, mp 156–157°C. Deacetylation of **32** gave **29** (0.59 g, 52.7%), mp 180–183°C, $[\alpha]_D^{20} - 165^\circ$ (c = 1,CH₃OH); lit.¹⁷⁾ D-enantiomer, $[\alpha]_D^{20}$ 161°.

4-Methyl umbelliferone 2,3,4,6-tetra-O-acetyl- α -L-mannopyranoside (34). 4-Methyl umbelliferone (2.6 g) and 33 (1.59 g) were well-mixed and melted by heating. To this melt, a mixture of acetic acid-acetic anhydride (95:5, 5 ml) and anhydrous zinc chloride (0.6 g) were added, and the resulting mixture was heated at 125–135°C for 1 h *in vacuo*, after cooling, the reaction mixture was dissolved in chloroform and filtered, and the resulting solution was successively washed with 1.5 N HCl, 1 N NaOH and water, and finally dried (MgSO₄). The filtrate was concentrated, and the residue was crystallized from ethanol to yield 34 (0.39 g, 15.4%), R_f 0.30 (toluene–ethyl acetate 7 : 3), mp 161°C, $[\alpha]_{D}^{20}$ –136° (c=1, CHCl₃); lit.¹⁸ D-enantiomer, mp 160–161°C, $[\alpha]_{D}^{20}$ 136°. NMR $\delta_{\rm H}$ (CDCl₃): 2.00 (9H), 2.19 (3H, each s, 4CH₃CO); 2.38 (3H, d, J=1 Hz, C4-CH₃); 3.87–4.32 (3H, m, H-5-6); 5.15–5.55 (3H, m, sugar H-2-4); 5.38 (1H, d, J=1 Hz, sugar H-1); 6.07 (1H, d, J=1 Hz, H-3); 6.80–7.00 (2H, m, H-6, 8); 7.40 (1H, d, J=10 Hz, H-5).

4-Methyl umbelliferone α-L-mannopyranoside (30). Deacetylation of 34 (200 mg) with 0.1 N methanolic sodium methoxide (10 ml) yielded 30 (40 mg, 30%), mp 160.8°C, $[\alpha]_D^{20} - 113^\circ$ (c = 1, CH₃OH). Anal. Found; C, 56.18; H, 5.08. Calcd. for C₁₆H₁₈O₈: C, 56.80; H, 5.33%.

Hydrolysis test on the disaccharides and phenol glycosides toward naringinase from Penicillium decumbens. To a sample (200 mg) in water (10 ml) was added an enzyme solution (1 ml), which had been prepared by dissolving 100 units of naringinase (Sigma) in water (10 ml). After 2 days of incubation at 40 °C, the reaction mixture was filtered and subjected to an HPLC analysis. Rt: methyl glucoside, 6.086; L-mannose, 8.036; D-galactose, 8.747; D-glucose, 8.275.

 α -L-Mannosidase activity assay. Naringinase (Sigma) was used in this experiment. One unit of this enzyme liberates 1 mmol of reducing sugar (measured as glucose) from naringin per min at pH 4.0 and 40°C. An enzyme solution was prepared by dissolving 1.8 units of the enzyme in 20 ml of a 0.2 M sodium acetate buffer at pH 4, *p*-nitrophenyl- α -L-mannopyranoside (PNM) being used as the substrate. The reaction mixture contained 0.8 ml of a sodium acetate buffer (pH 4) and 0.1 ml of 0.05 M PNM in a sodium acetate buffer (pH 4). After 5 min of preincubation at 40°C, the reaction was started by adding 0.05 ml of the enzyme solution. After a suitable time of incubation (5–60 min) at 40°C, the reaction was terminated by adding 4 ml of 1 M sodium carbonate, and the amount of liberated *p*-nitrophenol was measured spectrometrically at 400 nm.

References

- F. Michon, V. Pozsgay, J. Brisson, and H. J. Jennings, Carbohydr. Res., 194, 321-324 (1989).
- S. Kamiya, S. Esaki, and R. Tanaka, Agric. Biol. Chem., 49, 55–62 (1985).
- 3) B. Helferich and J. Zirner, Chem. Ber., 95, 2604–2611 (1962).
- C. P. J. Glaudemans and M. J. Bertolini, in "Methods in Carbohydrate Chemistry," Vol. VIII, ed. by R. L. Whistler and J. N. DeMiller, Academic Press, New York, 1980, pp. 271–275.
- 5) W. A. Bonner, J. Am. Chem. Soc., 80, 3372–3379 (1958).
- 6) H. Honog and H. Weidmann, Carbohydr. Res., 39, 374-379 (1975).
- 7) S. Hanessian and J. Banoub, Carbohydr. Res., 53C, 13-14 (1977).
- H. G. Fletcher, Jr., R. K. Ness, and C. S. Hudson, J. Am. Chem. Soc., 72, 2200–2205 (1934).
- 9) D. J. Bell, J. Chem. Soc., 1934, 1177-1179.
- 10) K. Freudenberg and K. V. Oertzen, *Liebigs Ann. Chem.*, **574**, 37–53 (1951).
- R. L. Whistler, L. W. Doner, and M. Kosik, in "Methods in Carbohydrate Chemistry," Vol. VI, ed. by R. L. Whistler and J. N. DeMiller, Academic Press, New York, 1972, pp. 411–412.
- 12) H. Ohle and G. Berend, Chem. Ber., 58, 2585-2589 (1925).
- 13) W. Klyne, Biochem. J., 47, XLI-XLII (1950).
- 14) W. W. Zorbach, S. Saeki, and W. Buhler, J. Med. Chem., 6, 298–300 (1963).
- 15) J. Conchie and G. A. Levy, in "Methods in Carbohydrate Chemistry," Vol. II, ed. by R. L. Whistler and M. L. Wolfrom, Academic Press, New York, 1963, pp. 335-337.
- B. Helferich and E. Schmidtz-Hillebrecht, Chem. Ber., 66, 378–383 (1933).
- 17) O. Westphal and H. Feiser, Chem. Ber., 89, 582-588 (1956).
- 18) Y. Voznyi, I. S. Kalichev, and A. A. Galoyan, *Bioorg. Khim.*, 12, 521–526 (1986).