Synthesis of 4-nitrophenyl O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 2)$ -O-(6-O-methyl- $\alpha$ -D-mannopyranosyl)- $(1 \rightarrow 6)$ - $\beta$ -D-glucopyranoside and its 4',6'-di-O-methyl analog. Potential inhibitors of N-acetylglucosaminyl-transferase V (GnT-V) \*

Shaheer H. Khan <sup>1,†</sup> and Khushi L. Matta <sup>†</sup>

Department of Gynecologic Oncology, Roswell Park Cancer Institute, Elm and Carlton Streets, Buffalo, New York, 14263 (USA)

(Received June 16th, 1992; accepted October 6th, 1992)

#### ABSTRACT

O-(2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-3,4-di-O-acetyl-6-O-methyl-  $\alpha$ -D-mannopyranosyl bromide and O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$ 2)-3-O-acetyl-4,6-di-O-methyl- $\alpha$ -D-mannopyranosyl bromide were each condensed with 4-nitrophenyl 2,3-di-O-acetyl- $\beta$ -D-glucopyranoside, and the products were deprotected to yield, respectively,  $\beta$ -D-GlcpNAc-(1  $\rightarrow$  2)-6-O-Me- $\alpha$ -D-Man p-(1  $\rightarrow$  6)- $\beta$ -D-Glcp NAc-(1  $\rightarrow$  2)-4,6-di-O-Me- $\alpha$ -D-Man p-(1  $\rightarrow$  6)- $\beta$ -D-Glcp, as their 4-nitrophenyl glycosides. These trisaccharides are expected to function as inhibitors for N-acetylglucosaminyltransferase V.

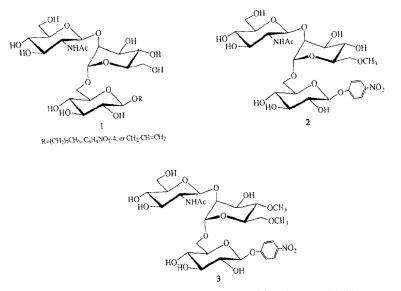
### INTRODUCTION

*N*-Acetylglucosaminyltransferase V (GnT-V, EC 2.4.1.155) transfers an *N*-acetyl- $\beta$ -D-glucosamine ( $\beta$ -D-Glc*p*NAc) unit to O-6 of the  $\alpha$ -(1  $\rightarrow$  6)-linked D-Man*p* residue that forms part of the trimannopyranosyl core of asparagine-linked *N*-glycans<sup>2</sup>. In recent years, this enzyme has been the center of great attention as a potential tumor marker because of its increased activity in cells transformed by tumor viruses<sup>3,4</sup> or oncogenes<sup>5</sup>. Furthermore, work from the laboratory of Dennis et al.<sup>6,7</sup> has suggested that a decrease in intracellular activity of GnT-V and the resulting decrease in specific cell surface structures is correlated with a reduction

<sup>\*</sup> Synthetic Studies in Carbohydrates, Part 86. For Part 85, see ref 1. This investigation was supported by grant No. AI 29326 awarded by the National Institute of Allergy and Infectious Diseases, NIH.

<sup>&</sup>lt;sup>1</sup> Present address: Applied Biosystems Inc., 850 Lincoln Centre Drive, Foster City, CA 94404, USA.

<sup>&</sup>lt;sup>†</sup> Corresponding authors.



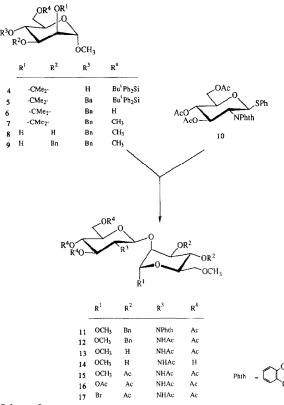
Scheme 1. Structures of the synthetic GnT-V acceptor (1) and potential inhibitors (2 and 3).

of metastatic potential of human and rodent cells. In addition these authors<sup>8</sup> have shown that, by comparison with nonmalignant tissues, increases in the expression of GnT-V activity and in the resulting cell-surface oligosaccharides are associated with a number of human carcinomas.

For the past few years, our group<sup>9-11</sup> has been actively engaged with the synthesis of acceptor substrates for GnT-V. Now we have focussed our attention on the design, synthesis, and biological evaluation of inhibitors for this particular enzyme<sup>12</sup>. Our strategy for the creation of inhibitors involved defining the specific acceptor substrate for the enzyme, then chemically synthesizing an analog having a masking group on the hydroxyl that would normally serve as the point of attachment for the transferred glycosyl unit, as shown in Scheme 1. On the basis of this rationale, we have envisioned  $\beta$ -D-GlcpNAc-(1  $\rightarrow$  2)-6-O-Me- $\alpha$ -D-Manp-(1  $\rightarrow$  6)- $\beta$ -D-Glcp-OC<sub>6</sub>H<sub>4</sub>(4-NO<sub>2</sub>) (2) as a potential inhibitor for GnT-V. A similar, presumably equally useful trisaccharide  $\beta$ -D-Glc pNAc-(1  $\rightarrow$  2)-4,6-di-O-Me- $\alpha$ -D-Man p-(1  $\rightarrow$  6)- $\beta$ -D-Glc p-OC<sub>6</sub>H<sub>4</sub>(4-NO<sub>2</sub>) (3) was also synthesized. We preferred the incorporation of a  $\beta$ -D-glucopyranose residue at the reducing terminus in place of the naturally occurring  $\beta$ -D-mannopyranose residue, since the latter is not a prerequisite<sup>13,14</sup> for recognition by GnT-V and its incorporation in the synthesis would be far from simple. Recently, the laboratories of Palcic and Hindsgaul have reported<sup>15,16</sup> the synthesis of a similar trisaccharide,  $\beta$ - $\beta$ - $\beta$ -GlcpNAc-(1  $\rightarrow$  2)-6deoxy- $\alpha$ -D-Man p-(1  $\rightarrow$  6)- $\beta$ -D-Glcp-O-(CH<sub>2</sub>)<sub>7</sub>CH<sub>3</sub>, and have shown it to be a competitive inhibitor for GnT-V. Compounds 2 and 3 also proved useful<sup>17</sup> in specificity studies of the  $\beta$ -1,4-N-acetylglucosaminyltransferase (GnT-VI') acting on the  $\alpha$ -3 and  $\alpha$ -6 arms of N-linked oligosaccharides.

# RESULTS AND DISCUSSION

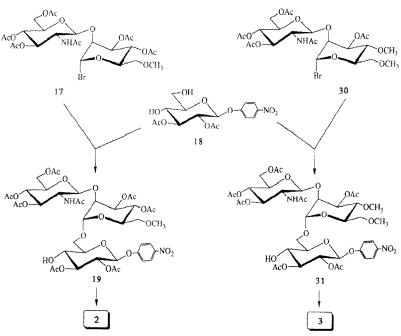
For the synthesis of the title trisaccharide 2 we employed the known 4nitrophenyl 2,3-di-O-acetyl- $\beta$ -D-glucopyranoside<sup>9</sup> 18 as a glycosyl acceptor and O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-3,4-di-Oacetyl-6-O-methyl- $\alpha$ -D-mannopyranosyl bromide 17 as a glycosyl donor. Bromide 17 was readily prepared from O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-1,3,4-tri-O-acetyl-6-O-methyl- $\alpha$ -D-mannopyranose 16. Compound 16 was obtained by condensation of phenyl 3,4,6-tri-O-acetyl-2-deoxy-2phthalimido-1-thio- $\beta$ -D-glucopyranoside<sup>11</sup> 10 with methyl 3,4-di-O-benzyl-6-Omethyl- $\alpha$ -D-mannopyranoside 9. The latter was obtained from methyl 6-O-tertbutyldiphenylsilyl-2,3-O-isopropylidene- $\alpha$ -D-mannopyranoside<sup>10</sup> 4 through a succession of chemical steps. Thus, benzylation<sup>18</sup> of compound 4 followed by cleavage of the *tert*-butyldiphenylsilyl ether group at C-6 and subsequent methylation<sup>19</sup> gave syrupy 6-O-methyl derivative 7 (77%). Deisopropylidenation of 7 gave 8 (63%) which was converted into the desired 9 by treatment of its stannylene derivative with benzyl bromide<sup>20</sup>.



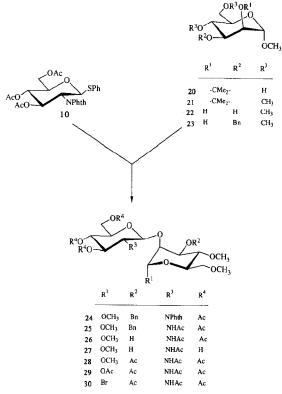
Glycosylation of 9 with thioglycoside 10 promoted by N-iodosuccinimide and trifluoromethanesulfonic (triflic) acid<sup>21,22</sup> gave 11 (86%). Treatment of the disaccharide 11 with hydrazine hydrate, followed by acetylation, gave intermediate 12, which was not characterized. Hydrogenolysis of benzyl groups of 12 gave, after chromatographic purification, 13 (91%) which was acetylated (Py-Ac<sub>2</sub>O) to afford the hexacetate 15 (94%). This compound was subjected to acetolysis to furnish 16 (89%). The <sup>1</sup>H NMR spectrum of 16 contained a low-field signal at  $\delta$  5.98 (1 H, J 2.0 Hz), suggesting that it existed almost exclusively as the  $\alpha$ -D anomer. A small portion of 13 was O-deacetylated to afford 14 (73%). Treatment of 16 with HBr in glacial acetic acid gave amorphous bromide 17 (87%).

Glycosylation of the diol 18 with bromide 17 promoted by silver trifluromethanesulfonate (triflate) and sym-collidine, gave the protected trisaccharide derivative 19 (62%). O-Deacetylation of 19 furnished the title trisaccharide 2 (86%).

The synthesis of trisaccharide 3 followed a procedure analogous to that described for the preparation of 2. Thus the reaction of thioglycoside 10 with alcohol 23 (prepared from methyl 2,3-*O*-isopropylidene- $\alpha$ -D-mannopyranoside<sup>23</sup> 20 in three steps), promoted again by *N*-iodosuccinimide and triflic acid, gave the  $\beta$ - $(1 \rightarrow 2)$ -linked disaccharide derivative 24 (76%) which was converted to the glycosyl bromide 30 in five steps, in a manner analogous to that described for conversion of



Scheme 3.



Scheme 4.

11 to 17. The free disaccharide 27 (78%) was obtained by deprotection of a small portion of 26.

Condensation of the diol 18 with the glycosyl donor 30 under conditions similar to those described for the reaction of 17 with 18 gave the partially protected trisaccharide 31 (78%), from which the acetyl groups were removed by Zemplén transesterification to afford the desired trisaccharide 3 (71%).

Preliminary evaluation of trisaccharides 2 and 3 shows that they are, as expected, potential inhibitors for GnT-V. These results will be reported in detail elsewhere.

#### EXPERIMENTAL

General methods.—Unless otherwise indicated, all reactions were carried out at ambient temperatures. Solutions were dried with  $Na_2SO_4$  and concentrated at 40–50°C/2 kPa. TLC was conducted on aluminum sheets, precoated with 0.2-mm layers of Silica Gel 60F-254 (Merck); the compounds were located by UV light and/or by charring with 5%  $H_2SO_4$ . Column chromatography was performed on silica gel (Baker Analyzed, 60–200 mesh). The following solvent systems (v/v) were

Com-	Chemical shifts ( $\delta$ ) and coupling constants (Hz)							
pound	$\overline{\text{H-1}(J_{1,2})^{h}}$	H-1' $(J_{1',2'})^{c}$	OC <i>H</i> <sub>3</sub> -1	OC <i>H</i> <sub>3</sub> -4	OC <i>H</i> <sub>3</sub> -6	NΛc	OAc	$\overline{{ m N}H\left(J_{2,{ m NH}} ight)}$
11	4.47	5.51 (8.5)	2.94		3.19		1.87 (3H), 2.04 (6H)	
13	4.66 (1.5)	4.95 (8.5)	3.36		3.40	1.96	2.04 (3H), 2.06 (3H) 2,10 (3H)	6.11 (8.5)
14	4.76	4.55 (8.3)	3.40		3.41	2.05		
15	4.63 (1.5)	5.04 (8.5)	3.36		3.39	1.94	2.00 (6H), 2.02 (6H) 2.08 (3H)	5.92 (8.5)
16	5.98 (2.0)	4.91 (8.5)			3.32	1.97	1.99 (3H), 2.01 (3H) 2.04 (3H), 2.06 (3H) 2.08 (3H), 2.13 (3H)	6.21 (8.5)
24	4.43 (2.0)	5.48 (8.5)	2.98	3.44	3.18		1.87 (3H), 2.04 (6H)	
26	4.61 (1.5)	4.98 (8.5)	3.33	3.55	3.40	1.94	2.02 (3H), 2.04 (3H) 2.10 (3H)	5.95 (8,0)
27	4.74 ( < 1)	4.55 (8.4)	3.39	3.51	3.41	2.06		
28	4.57 (2.0)	4.73 (8.5)	3.36	3.47	3.40	1.95	2.03 (3H), 2.04 (3H) 2.08 (3H), 2.10 (3H)	
29	5.94 (2.0)	4.63 (8.5)		3.48	3.37	1.95	2.01 (3H, 2.02 (3H) 2.08 (3H), 2.09 (3H) 2.11 (3H)	5.83 (9.0)

TABLE I

Selected <sup>1</sup>H NMR data for protected and unprotected disaccharides <sup>a</sup>

<sup>*a*</sup> For solutions in CDCl<sub>3</sub> at 300 MHz except for compounds **14** and **27**, which were recorded in  $D_2O$  at 500 MHz. The reference standards used are listed in the Experimental section. <sup>*b*</sup> Unprimed locants are used for protons in the reducing-end residue ( $\alpha$ -D-Man p). <sup>*c*</sup> Of the nonreducing-end residue ( $\beta$ -D-GlcpNAc).

used for chromatography: A, 2:1 hexane-CHCl<sub>3</sub>, B, 1:1 hexane-CHCl<sub>3</sub>, C, 3:1 hexane-EtOAc, D, 19:1 CHCl<sub>3</sub>-acetone, E, 4:1 CHCl<sub>3</sub>-acetone, F, 1:1 hexane-EtOAc, G, 2:1 hexane-EtOAc, H, 99:1 CHCl<sub>3</sub>-MeOH, I, 49:1 CHCl<sub>3</sub>-MeOH, J, 19:1 CHCl<sub>3</sub>-MeOH, K, 13:6:1 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O, L, 3:2 CHCl<sub>3</sub>-acetone, M, 2:1 CH<sub>2</sub>Cl<sub>2</sub>-acetone, N, 4:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH, O, 19:1  $CH_2CI_3$ -MeOH. Optical rotations were measured at  $22 + 2^\circ$  with a Perkin-Elmer 241 polarimeter. <sup>1</sup>H NMR spectra were recorded either at 90 (Varian EM-390), 300 (Bruker AM-300), or 500 MHz (Bruker AM-500) for solutions in CDCl<sub>2</sub> (internal Me<sub>4</sub>Si) or D<sub>2</sub>O (internal acetone,  $\delta$  2.225). <sup>13</sup>C NMR spectra were recorded at 50.3 (Bruker WP-200) or 75.5 MHz (Bruker AM-300) for solutions in CDCl<sub>3</sub> (internal Me<sub>4</sub>Si) or D<sub>2</sub>O (external 1% 1,4-dioxane in D<sub>2</sub>O,  $\delta$  67.4). Only partial NMR data are reported, but all values were in accord with the proposed structures. The assignments of <sup>13</sup>C chemical shifts are tentative. FAB-mass spectra were obtained using an AEI MS-9 instrument with Xe as the bombarding gas and 5:1 1,4-dithiothreitol-1,4-dithioerythritol as a matrix. Elemental analyses were performed by the Robertson Laboratory, 29 Samson Ave., Madison, NJ 08940 (USA).

Methyl 4-O-benzyl-6-O-tert-butyldiphenylsilyl-2,3-O-isopropylidene- $\alpha$ -D-mannopyranoside (5).—A mixture of 4<sup>10</sup> (5.5 g, 11.6 mmol), freshly prepared Ag<sub>2</sub>O (10

disaccharides
unprotected
d and un
otecte
is of pr
aton
its for key C
r k
fo
nal assignments
signal
NMR

TABLE II

ø

	15										Contraction of the second seco
	5	C-I	C-2	C-2′	C-6	C-6′	OCH <sub>3</sub> -1	OCH <sub>3</sub> -4	OCH <sub>3</sub> -4 OCH <sub>3</sub> -6	coch3	COCH <sub>3</sub>
11	96.75 <sup>c</sup>	97.97 °	77.91	54.76	71.06	62.36	54.50		58.89	170.65, 170.19, 169.47	20.72, 20.64, 20.48
13	98.94 <sup>d</sup>	99.72 <sup>d</sup>	78.73	55.32	72.33	61.95	54.88		59.29	171.06, 170.74, 170.67	23.30, 20.68, 20.60
										169.45	
14	98.84	100.23	76.80	55.83	72.95	61.45	56.15		59.23	175.40	23.10
15	98.44	98.44	74.05	55.93	72.08	62.19	55.07		59.33	170.86, 170.65, 170.38	23.32, 20.77, 20.69
										169.75, 169.53	20.62
16	90.91	99.23	73.12	54.78	72.04	61.99			59.37	170.95, 170.71, 170.62	23.23, 20.99, 20.76
										169.41, 169.36, 168.61	20.71, 20.64, 20.57
24	96.84 <sup>e</sup>	98.05 "	77.74	54.83	71.10	62.37	54.51	60.57	58.92	170.67, 170.20, 169.48	20.72, 20.66, 20.49
26	98.90 f	9.97 f	10.77	54.88	71.81	62.05	55.63	60.66	59.17	170.84, 170.71, 170.63	23.38, 20.72, 20.65
										169.52	
27	98.69	100.17	77.00	55.84	72.55	61.41	56.17	61.14	59.21	175.50	23.10
28	98.45 <sup>g</sup>	99.35 8	74.57	54.98	71.93	62.23	55.18	60.50	59.24	170.73, 170.68, 170.38	23.41, 21.10, 20.73
										169.53	20.70, 20.66
29	91.22	100.08	73.96	54.27	71.78	62.09		60.78	59.28	170.00, 170.66, 170.53	23.29, 21.09, 21.05
										169.42, 169.15	20.73, 20.69, 20.62

Nucleus <sup>b</sup>	Chemical shifts ( $\delta$ ) and coupling constants (Hz)						
	19	2	31	3			
H-1 (J <sub>1,2</sub> )	n.d. <sup>c</sup>	5.31 (5.4)	n.d. <sup>c</sup>	5.31 (5.5)			
H-1' $(J_{1',2'})$	4.77 ( < 1)	4.85	4.69 (2.0)	<b>n.</b> d. <sup>c</sup>			
H-1" $(J_{1'',2''})$	5.10 (9.0)	4.51 (8.4)	4.74 (8.5)	4.51 (8.4)			
$OCH_3-4$			3.38	3.40			
$OCH_{3}-6$	3.25	3,33	3.32	3.31			
NAc	1.84	2.00	1.85	2.00			
OAc	1.91, 2.02, 2.05, 2.08		2.01, 2.02, 2.17, 2.10				
	2.09, 2.11, 2.13		2.12, 2.13				
$C_6 H_4$ -NO <sub>2</sub> ( <sup>3</sup> J)	8.23 (9.0)	8.30 (9.0)	8.22 (9.0)	8.29 (9.0)			
	7.24 (9.0)	7.28 (9.0)	7.08 (9.0)	7.25 (9.0)			
C-1	97.68 <sup>d</sup>	100.02	97.86 °	99.81			
C-1'	97.38 <sup>d</sup>	97.81	97.75 °	97.57			
C-1"	98.43	100.17	99.65	100.12			
C-2'	75.37	76.80	75.70	78.22			
C-2"	55.72	56.11	55.18	56.16			
C-6	66.55	66.77	66.07	66.96			
C-6′	69.55	72.82	72.02	72.41			
C-6″	62.10	61.43	62.18	61.43			
OCH <sub>3</sub> -4			60.17	60.98			
OCH3-6	58.92	59.05	59.00	59.00			
COCH	171.45, 170.97, 170.65	175.44	171.25, 170.83, 170.72	175.47			
	170.43, 169.60, 169.45		170.64, 169.51, 169.48				
COCH <sub>3</sub>	23.14, 20.76, 20.60	23.05	23.30, 21.06, 20.87	23.06			
~*	20.30		20.71, 20.63				
$CNO_2$	161.79	162.39	161.20	162.35			
CO (phenolic)	142.88	143.42	143.11	143.25			
Aromatic	125.88, 116.41	127.01, 117.23	126.11, 116.45	127.07, 117.20			

TABLE III

Selected <sup>1</sup>H and <sup>13</sup>C NMR data for protected and unprotected trisaccharides "

<sup>a</sup> Spectra were recorded at 300 MHz (<sup>1</sup>H in CDCl<sub>3</sub>, compounds 19 and 31) or 500 MHz (<sup>1</sup>H in D<sub>2</sub>O, compounds 2 and 3) and 75.5 MHz (<sup>13</sup>C in CDCl<sub>3</sub> for compounds 19 and 31 and in D<sub>2</sub>O for 2 and 3). <sup>b</sup> Locants: unprimed,  $\beta$ -D-Glc p; single primed,  $\alpha$ -D-Man p, double primed,  $\beta$ -D-Glc pNAc. <sup>c</sup> Could not be determined due to spectral overlap. <sup>d.e</sup> Values with the same superscripts may be interchanged.

g), and benzyl bromide (10 mL) in DMF (80 mL) was stirred for 48 h at 45°C. The solids were filtered off (Celite bed), washed with DMF, and the combined filtrate was concentrated. The residue was stirred in CHCl<sub>3</sub> (200 mL), and the precipitated silver salts were filtered off and washed with CHCl<sub>3</sub>. The CHCl<sub>3</sub> solution was successively washed with water, aq NaHCO<sub>3</sub>, and water, dried, and concentrated. The crude product was chromatographed (Solvent  $A \rightarrow B$ ) to give 5 as a syrup (4.20 g, 64.1%);  $[\alpha]_D + 9.6^\circ$  (c 1.7, CHCl<sub>3</sub>);  $R_f$  0.7 (solvent B); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  7.80–7.23 (m, 15 H, arom.), 4.90 (s, 1 H, H-1), 3.33 (s, 3 H, OMe), 1.50, 1.35 (2 s, 6 H, CMe<sub>2</sub>), and 1.07 (s, 9 H, CMe<sub>3</sub>). Anal. Calcd for C<sub>33</sub>H<sub>42</sub>O<sub>6</sub>Si: C, 70.43; H, 7.52. Found: C, 70.69; H, 7.58.

Methyl 4-O-benzyl-2,3-O-isopropylidene- $\alpha$ -D-mannopyranoside (6).—To a solution of 5 (4.52 g, 8.03 mmol) in anhyd oxolane (50 mL) was added tetrabutylammo-

nium fluoride in oxolane (18 mL), and the mixture was stirred for 6 h at room temperature. After concentration the residue was chromatographed (solvent *C*), providing **6** as a syrup (2.13 g, 82%);  $[\alpha]_D + 54.3^\circ$  (*c* 1.2, CHCl<sub>3</sub>);  $R_f$  0.3 (solvent *C*); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  7.33 (br s, 5 H, arom.), 4.89 (s, 1 H, H-1), 3.31 (s, 3 H, OMe), 1.44, and 1.33 (2 s, 6 H, CMe<sub>2</sub>). *Anal.* Calcd for C<sub>17</sub>H<sub>24</sub>O<sub>6</sub>: C, 62.95; H, 7.46. Found: C, 63.03; H, 7.49.

Methyl 4-O-benzyl-2,3-O-isopropylidene-6-O-methyl- $\alpha$ -D-mannopyranoside (7).— A solution of **6** (2.1 g, 6.5 mmol) in DMF (40 mL) was stirred for 16 h at room temperature in the presence of BaO (8 g), Ba(OH)<sub>2</sub> · 8 H<sub>2</sub>O (8 g), and MeI (11 mL). The mixture was cooled (0°C) and diluted with an equal volume of CHCl<sub>3</sub>, and the insolubles were filtered off (Celite bed) and washed with CHCl<sub>3</sub>. The CHCl<sub>3</sub> solution was successively washed with water, aq Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, and water, dried, and concentrated to afford 7 as a syrup (1.69 g, 77%);  $[\alpha]_D + 44.8^\circ$  (c 1.1, CHCl<sub>3</sub>);  $R_f$  0.3 (solvent C); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  7.25 (br s, 5 H, arom.), 3.34 (s, 6 H, OMe-6 and OMe-1), 1.46, and 1.33 (2 s, 6 H, CMe<sub>2</sub>). Anal. Calcd for C<sub>18</sub>H<sub>26</sub>O<sub>6</sub>: C, 63.89; H, 7.74. Found: C, 64.19; H, 7.77.

Methyl 4-O-benzyl-6-O-methyl- $\alpha$ -D-mannopyranoside (8).—A solution of 7 (3.79 g, 11.2 mmol) in CHCl<sub>3</sub> (13 mL) containing trifluoroacetic acid (13 mL) and water (1.3 mL) was stirred for 1 h at room temperature. After concentration, and successive additions and evaporations of toluene, the crude product was chromatographed (CHCl<sub>3</sub>  $\rightarrow$  solvent D) to provide 8 as a syrup (2.11 g, 63%);  $[\alpha]_D + 75^\circ$  (c 1.1, CHCl<sub>3</sub>);  $R_f$  0.2 (solvent E); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.40–7.24 (m, 5 H, arom.), 4.71 (d, 1 H,  $J_{1,2}$  1.5 Hz, H-1), 3.36 (s, 3 H, OMe-6), and 3.33 (s, 3 H, OMe-1); <sup>13</sup>C (75.5 MHz):  $\delta$  138.42 (arom. *ipso*), 100.77 (C-1), 74.62 (PhCH<sub>2</sub>), 71.31 (C-6), 59.10 (OCH<sub>3</sub>-6), and 54.81 (OCH<sub>3</sub>-1). Anal. Calcd for C<sub>15</sub>H<sub>22</sub>O<sub>6</sub>: C, 60.39; H, 7.43. Found: C, 60.46; H, 7.37.

Methyl 3,4-di-O-benzyl-6-O-methyl- $\alpha$ -D-mannopyranoside (9).—A solution of 8 (0.9 g, 3.02 mmol) and dibutyltin oxide (0.75 g, 3.01 mmol) in MeOH (40 mL) was boiled for 1 h. The solvent was then evaporated, the residue was dissolved in DMF (10 mL), and benzyl bromide (0.5 mL, 4.21 mmol) was added. The mixture was stirred for 5 h at 110°C, then poured into water. The aqueous mixture was extracted with CHCl<sub>3</sub>, and the CHCl<sub>3</sub> solution was washed several times with water, dried, and concentrated. The residue was chromatographed (solvent *C*) to provide 9 as a syrup (0.49 g, 42%);  $[\alpha]_D + 36.1^\circ$  (*c* 1.1, CHCl<sub>3</sub>);  $R_f$  0.6 (solvent *F*); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.37–7.25 (m, 10 H, arom.), 4.76 (d, 1 H,  $J_{1,2}$  1.5 Hz, H-1), 3.38 (s, 3 H, OMe-6), and 3.34 (s, 3 H, OMe-1); <sup>13</sup>C (75.5 MHz):  $\delta$  138.39 and 137.94 (arom., *ipso*), 100.34 (C-1), 75.01 and 71.82 (PhCH<sub>2</sub>), 71.36 (C-6), 59.16 (OCH<sub>3</sub>-6), and 54.76 (OCH<sub>3</sub>-1). Anal. Calcd for C<sub>22</sub>H<sub>28</sub>O<sub>6</sub>: C, 68.02; H, 7.27. Found: C, 68.18; H,7.19.

Methyl O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 2)$ -3,4-di-O-benzyl-6-O-methyl- $\alpha$ -D-mannopyranoside (11).—Phenyl 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-1-thio- $\beta$ -D-glucopyranoside<sup>11</sup> 10 (2.64 g, 4.99 mmol) and alcohol 9 (1.57 g, 4.04 mmol) were dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (26 mL), pulverized, activated 4A molecular sieves (1.9 g) and *N*-iodosuccinimide (2.54 g, 11.3 mmol) were added, and the mixture was stirred in the dark for 30 min under Ar. After cooling (0°C; bath), a solution of trifluoromethanesulfonic acid (0.1 mL) in CH<sub>2</sub>Cl<sub>2</sub> (37 mL) was added dropwise, and the stirring was continued for 2 h. Dichloromethane (200 mL) was added, and the solids were filtered off (Celite bed) and washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate was successively washed with water, aq NaHCO<sub>3</sub>, aq Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, and water. Evaporation of the solvent and purification of the residue by chromatography (solvent  $C \rightarrow G \rightarrow F$ ) gave amorphous 11 (2.8 g, 86%);  $[\alpha]_D = 8.2^\circ$  (*c* 1.3, CHCl<sub>3</sub>);  $R_f$  0.1 (solvent *F*). Anal. Calcd for C<sub>42</sub>H<sub>47</sub>NO<sub>15</sub>: C, 62.60; H, 5.88; N, 1.74. Found: C, 62.22; H, 5.86; N, 1.86.

Methyl O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-6-O-methyl- $\alpha$ -D-mannopyranoside (13).---A mixture of 11 (2.7 g) and hydrazine hydrate (11 mL) in EtOH (37 mL) was refluxed for 3 h. The mixture was concentrated, the residue was dissolved in pyridine (50 mL), and Ac<sub>2</sub>O (25 mL) was added. After being stirred overnight at room temperature the mixture was cooled to 0°C, and the excess Ac<sub>2</sub>O was decomposed by dropwise addition of MeOH. After concentration, a solution of the residue in CHCl<sub>3</sub> (200 mL) was successively washed with water, aq NaHCO<sub>3</sub>, and water. Evaporation of the solvent and chromatography (CHCl<sub>3</sub>  $\rightarrow$  solvent *E*) gave 12 (2.0 g), which was sufficiently pure for the next step.

A mixture of **12** (2.0 g) and 10% Pd–C (2.0 g) in glacial acetic acid (30 mL) was shaken under H<sub>2</sub> at 345 kPa for 2 days at room temperature. The suspension was filtered (Celite bed), the solid was thoroughly washed with MeOH, and the filtrate and washings were combined and concentrated. The residual syrup was chromatographed (solvent  $H \rightarrow I$ ) to yield amorphous **13** (1.36 g, 91%);  $[\alpha]_D + 8.3^\circ$  (*c* 1.2, CHCl<sub>3</sub>);  $R_f$  0.1 (solvent J). Anal. Calcd for C<sub>22</sub>H<sub>35</sub>NO<sub>14</sub>: C, 49.16; H, 6.56; N, 2.61. Found: C, 48.92; H, 6.49; N, 2.73.

Methyl O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-6-O-methyl- $\alpha$ -Dmannopyranoside (14).—A solution of 13 (0.18 g) in 20 mM methanolic NaOMe (25 mL) was stirred overnight at room temperature. The base was neutralized with a few drops of glacial acetic acid, and the solution was deionized with Amberlite IR-120 (H<sup>+</sup>) cation-exchange resin. The resin was filtered off (Celite bed) and thoroughly washed with MeOH. The combined filtrate was concentrated, and a solution of the residue in water was lyophilized to give amorphous 14 (0.1 g, 73%);  $[\alpha]_D - 10.2^\circ$  (c 0.5, H<sub>2</sub>O);  $R_f$  0.3 (solvent K). FABMS: m/z 412 [11.5%, (M + 1)<sup>+</sup>] and 434 [100%, (M + Na)<sup>+</sup>]. Anal. Calcd for C<sub>16</sub>H<sub>29</sub>NO<sub>11</sub>: C, 46.71; H, 7.11; N, 3.41. Found: C, 46.35; H, 7.11; N, 3.05.

Methyl O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-3,4-di-O-acetyl-6-O-methyl- $\alpha$ -D-mannopyranoside (15).—A solution of 13 (1.11 g) in 1:2 Ac<sub>2</sub>O-pyridine (45 mL) was stirred overnight at room temperature. Methanol was added dropwise at 0°C, the solution was concentrated, and the residue was subjected to additions and evaporations of toluene to give amorphous 15 (1.2 g 93%);  $[\alpha]_D - 6.2^\circ$  (c 1.2, CHCl<sub>3</sub>);  $R_f$  0.3 (solvent J). Anal. Calcd for C<sub>26</sub>H<sub>39</sub>NO<sub>16</sub>: C, 50.24; H, 6.32; N, 2.25. Found: C, 50.02; H, 6.19; N, 1.98.

O-(2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-1,3,4-tri-O-acetyl-6-O-methyl- $\alpha$ -D-mannopyranose (16).—A solution of compound 15 (1.17 g) in Ac<sub>2</sub>O (30 mL) containing 1% (v/v) of concd. H<sub>2</sub>SO<sub>4</sub> was stirred for 17 h at room temperature. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (200 mL), successively washed with water, satd NaHCO<sub>3</sub>, and water, dried, and concentrated. The residue was dissolved in a small amount of EtOAc, and the solution diluted with ether to cause the precipitation of amorphous 16 (1.09 g, 89%); [ $\alpha$ ]<sub>D</sub> -7.6° (c 1.1, CHCl<sub>3</sub>);  $R_f$  0.3 (solvent L). Anal. Calcd for C<sub>27</sub>H<sub>39</sub>NO<sub>17</sub>: C, 49.92; H, 6.05; N, 2.16. Found: C, 49.82; H, 6.01; N, 2.07.

4-Nitrophenyl O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-O-(3,4-di-O-acetyl-6-O-methyl- $\alpha$ -D-mannopyranosyl)-(1  $\rightarrow$  6)-2,3-di-O-acetyl- $\beta$ -D-glucopyranoside (19).—To a cold (0°C, bath), stirred solution of 16 (0.97 g) in CH<sub>2</sub>Cl<sub>2</sub> (12 mL) was added a 31% solution of HBr in glacial acetic acid (12 mL), and stirring was continued for 12 h at 0°C. The mixture was then poured into ice-water and the product was extracted into CH<sub>2</sub>Cl<sub>2</sub>. The extract was successively washed with cold water, cold satd NaHCO<sub>3</sub>, and cold water, dried, and concentrated to give *O*-(2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-3,4-di-*O*-acetyl-6*O*-methyl- $\alpha$ -D-mannopyranosyl bromide (17) as an amorphous solid (0.87 g, 87%);  $[\alpha]_{\rm D}$  + 54.3° (c 0.9, CHCl<sub>3</sub>),  $R_f$  0.2 (solvent *E*); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  7.28 (d, 1 H, J 9 Hz, NH), 6.45 (s, 1 H, H-1), 3.26 (s, 3 H, OMe-6), and 2.10-1.98 (cluster of s, 18 H, 5 OAc and 1 NAc).

A solution of the glycosyl bromide **17** (0.86 g, 1.28 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) was added at 0°C to a stirred mixture of 4-nitrophenyl 2,3-di-O-acetyl- $\beta$ -D-gluco-pyranoside<sup>9</sup> (**18**, 0.47 g, 1.22 mmol), sym-collidine (0.22 mL, 1.65 mmol), silver trifluoromethanesulfonate (0.47 g, 1.83 mmol), and pulverized 4A molecular sieves (1.3 g) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL). After 6 h, CH<sub>2</sub>Cl<sub>2</sub> (200 mL) was added, the mixture was filtered (Celite bed), the solids were washed with CH<sub>2</sub>Cl<sub>2</sub> (100 mL), and the combined filtrate was concentrated. Chromatography (solvent *M*) gave unreacted **18** (0.15 g), followed by amorphous **19** (0.5 g 62%, based on **18** consumed); [ $\alpha$ ]<sub>D</sub> – 11.3° (*c* 1.1, CHCl<sub>3</sub>); *R<sub>f</sub>* 0.4 (solvent *M*). Anal. Calcd for C<sub>41</sub>H<sub>54</sub>N<sub>2</sub>O<sub>25</sub>: C, 50.51; H, 5.58; N, 2.87. Found: C, 50.69; H, 5.70; N, 2.71.

4-Nitrophenyl O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-O-(6-Omethyl- $\alpha$ -D-mannopyranosyl)-(1  $\rightarrow$  6)- $\beta$ -D-glucopyranoside (2).—Compound 19 (0.1 g) was O-deacetylated in 10 mM methanolic NaOMe (11 mL) exactly as described for the preparation of 14 to give, after freeze-drying, amorphous 2 (0.06 g, 86%);  $[\alpha]_D - 55.8^\circ$  (c 0.5, H<sub>2</sub>O);  $R_f$  0.3 (solvent K); FABMS: m/z 681 [2.5%, (M + 1)<sup>+</sup>] and 703 [1.6%, (M + Na)<sup>+</sup>]. Anal. Calcd for C<sub>27</sub>H<sub>40</sub>N<sub>2</sub>O<sub>18</sub>: C, 47.65; H, 5.92; N, 4.12. Found: C, 47.68; H, 6.12; N, 3.94.

*Methyl* 2,3-O-*isopropylidene-4*,6-*di*-O-*methyl*- $\alpha$ -D-*mannopyranoside* (21).—Methyl 2,3-O-isopropylidene- $\alpha$ -D-mannopyranoside<sup>23</sup> (20, 1.56 g, 6.66 mmol) was methylated as described for the preparation of 7 to give 21 as a syrup (1.6 g, 92%);  $[\alpha]_D$ 

+38.9° (c 3.7, CHCl<sub>3</sub>); {lit.<sup>24</sup> [ $\alpha$ ]<sub>D</sub><sup>20</sup> + 40° (c 1, CHCl<sub>3</sub>)};  $R_f$  0.3 (solvent G); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  4.86 (s, 1 H, H-1), 3.50 (s, 3 H, OMe-4), 3.40 (s, 3 H, OMe-6), 3.36 (s, 3 H, OMe-1), 1.52, and 1.33 (2 s, 6 H, CMe<sub>2</sub>). Anal. Calcd for C<sub>12</sub>H<sub>22</sub>O<sub>6</sub>: C, 54.95; H, 8.45. Found: C, 55.06; H, 8.52.

*Methyl* 4,6-*di*-O-*methyl*- $\alpha$ -D-*mannopyranoside* (22).—Deacetonation of 21 (8.16 g) as described for the preparation of **8**, gave after chromatography (solvent *E*), compound 22 as a syrup (5.5 g, 80%);  $[\alpha]_D$  + 79.2° (*c* 1.1, CHCl<sub>3</sub>); {lit.<sup>24</sup>  $[\alpha]_D^{20}$  + 99° (*c* 2, CH<sub>3</sub>OH)};  $R_f$  0.1 (solvent *E*); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  4.52 (s, 1 H, H-1), 3.55 (s, 3 H, OMe-4), 3.43 (s, 3 H, OMe-6), 3.35 (s, 3 H, OMe-1); <sup>13</sup>C (50.3 MHz):  $\delta$  101.12 (C-1), 60.53 (OMe-4), 59.18 (OMe-6), and 54.97 (OMe-1). *Anal.* Calcd for C<sub>9</sub>H<sub>18</sub>O<sub>6</sub>: C, 48.64; H, 8.16. Found: C, 48.59; H, 8.32.

*Methyl* 3-O-*benzyl-4,6-di*-O-*methyl-* $\alpha$ -D-*mannopyranoside* (23).—Benzylation of 22 (4.32 g, 19.0 mmol) as described for the preparation of 9 gave, after chromatogaphy (solvent *C*), compound 23 as a syrup (3.9 g, 66%);  $[\alpha]_D$  +45.7° (*c* 1.3, CHCl<sub>3</sub>);  $R_f$  0.2 (solvent *F*); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  7.43–7.07 (m, 5 H, arom.), 4.51 (s, 1 H, H-1), 3.48 (s, 3 H, OMe-4), 3.35 (s, 3 H, OMe-6), 3.28 (s, 3 H, OMe-1); <sup>13</sup>C (50.3 MHz):  $\delta$  100.51 (C-1), 60.79 (OMe-4), 59.29 (OMe-6), and 54.99 (OMe-1). *Anal.* Calcd for C<sub>16</sub>H<sub>24</sub>O<sub>6</sub>: C, 61.52; H, 7.74. Found: C, 61.59; H, 7.68.

Methyl O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-( $1 \rightarrow 2$ )-3-O-benzyl-4,6-di-O-methyl-α-D-mannopyranoside (24).—A reaction of alcohol 23 (1.72 g, 5.51 mmol) with the thioglycoside<sup>11</sup> 10 (3.68 g, 6.96 mmol) as described for the preparation of 11 gave, after chromatography (solvent  $C \rightarrow G$ ), disaccharide 24 as an amorphous solid (3.06 g, 76%);  $[\alpha]_D + 4.1^\circ$  (c 2.7, CHCl<sub>3</sub>);  $R_f$  0.2 (solvent F). Anal. Calcd for C<sub>36</sub>H<sub>44</sub>NO<sub>15</sub>: C, 59.17; H, 6.07; N, 1.92. Found: C, 59.06; H, 6.11; N, 2.04.

Methyl O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-4,6-di-O-methyl- $\alpha$ -D-mannopyranoside (26).—Compound 24 (1.97 g) was treated with hydrazine hydrate and then acetylated as described for the conversion of 11 into 12 to give, after chromatography (CHCl<sub>3</sub>  $\rightarrow$  solvent *E*), the amorphous disaccharide 25 (1.37 g);  $R_f$  0.1 (solvent *E*), which was sufficiently pure for the next step.

Hydrogenolysis of **25** (1.31 g), as described for the preparation of **13**, gave, after chromatography (CHCl<sub>3</sub>  $\rightarrow$  solvent *H*), the amorphous disaccharide **26** (1.0 g, 89%);  $[\alpha]_D + 17.5^\circ$  (c 2.1, CHCl<sub>3</sub>),  $R_f$  0.2 (solvent *J*). Anal. Calcd for C<sub>23</sub>H<sub>37</sub>NO<sub>14</sub>: C, 50.09; H, 6.76; N, 2.54. Found: C, 49.89, H, 6.53, N, 2.39.

Methyl O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)-( $1 \rightarrow 2$ )-4,6-di-O-methyl- $\alpha$ -D-mannopyranoside (27).—Deacetylation of 26 (0.2 g) and lyophilization of the resulting product as described for the preparation of 14 gave the amorphous disaccharide 27 (0.12 g, 78%);  $[\alpha]_D - 7.6^\circ$  (c 1.1, H<sub>2</sub>O);  $R_f$  0.3 (solvent N); FABMS: m/z 426 [20%, (M + 1)<sup>+</sup>] and 448 [100%, (M + Na)<sup>+</sup>]. Anal. Calcd for  $C_{17}H_{31}NO_{11}$ : C, 47.99; H, 7.35; N, 3.29. Found: C, 47.62; H, 7.72; N, 2.96.

Methyl O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-3-O-acetyl-4,6-di-O-methyl- $\alpha$ -D-mannopyranoside (**28**).—Compound **26** (0.36 g) was acetylated as described for the preparation of 15 to give the amorphous disaccharide 28 (0.38 g, 98%);  $[\alpha]_D - 14.3^\circ$  (c 0.9, CHCl<sub>3</sub>);  $R_f$  0.3 (solvent J). Anal. Calcd for C<sub>25</sub>H<sub>39</sub>NO<sub>15</sub>: C, 50.59; H, 6.62; N, 2.36. Found: C, 50.43; H, 6.59; N, 2.22.

O-(2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-glucopyranosyl)-(1 → 2)-1,3-di-Oacetyl-4,6-di-O-methyl-α-D-mannopyranose (29).—Acetolysis of 28 (0.45 g) as described for the preparation of 16 gave the amorphous disaccharide 29 (0.38 g, 81%);  $[\alpha]_D$  = 11.1° (c 1.1, CHCl<sub>3</sub>),  $R_f$  0.2 (solvent E). Anal. Calcd for C<sub>26</sub>H<sub>39</sub>NO<sub>16</sub>: C, 50.24; H, 6.32; N, 2.25. Found: C, 50.20; H, 6.19; N, 2.17.

4-Nitrophenyl O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-O-(3-O-acetyl-4,6-di-O-methyl- $\alpha$ -D-mannopyranosyl)-(1  $\rightarrow$  6)-2,3-di-O-acetyl- $\beta$ -D-glucopyranoside (31).—Reaction of 29 (0.3 g) with HBr as described for the preparation of 17 gave O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-3-O-acetyl-4,6-di-O-methyl- $\alpha$ -D-mannopyranosyl bromide 30 as an amorphous solid (0.28 g, 90%);  $[\alpha]_D$  + 13.6° (c 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>):  $\delta$  8.03 (d, 1 H, J 9 Hz, NH), 6.36 (d, 1 H, J < 1 Hz, H-1), 3.46 (s, 3 H, OMe-4), 3.35 (s, 3 H, OMe-6), 2.07 (s, 9 H, 3 OAc), and 1.98 (s, 6 H, OAc and NAc).

A reaction of the alcohol<sup>9</sup> **18** (0.13 g, 0.34 mmol) with glycosyl donor **30** (0.27 g, 0.42 mmol) as described for the preparation of **19** gave, after chromatography (solvent *J*), the amorphous trisaccharide **31** (0.25 g, 78%);  $[\alpha]_D - 13.9^\circ$  (c 1.1, CHCl<sub>3</sub>);  $R_f$  0.2 (solvent *O*). Anal. Calcd for C<sub>40</sub>H<sub>54</sub>N<sub>2</sub>O<sub>24</sub>: C, 50.74; H, 5.75; N, 2.96. Found: C, 51.04; H, 5.59; N, 2.68.

4-Nitrophenyl O-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)-(1  $\rightarrow$  2)-O-(4,6-di-Omethyl- $\alpha$ -D-mannopyranosyl)-(1  $\rightarrow$  6)- $\beta$ -D-glucopyranoside (3).—Deacetylation of 31 (0.27 g) as described for the preparation of 2 gave, after chromatography (solvent K), the amorphous trisaccharide 3 (0.14 g, 71%);  $[\alpha]_D - 61.5^\circ$  (c 0.6, H<sub>2</sub>O);  $R_f$  0.5 (solvent K). FABMS: m/z 717 [17%, (M + Na)<sup>+</sup>]. Anal. Calcd for C<sub>28</sub>H<sub>42</sub>N<sub>2</sub>O<sub>18</sub>: C, 48.41; H, 6.09; N, 4.03. Found: C, 48.33; H, 6.19; N, 3.89.

## ACKNOWLEDGMENTS

The authors gratefully acknowledge Suzane C. Crawly's (University of Alberta) preliminary evaluation of 2 and 3 as inhibitors for GnT-V. They are also thankful to Mr. C.F. Piskorz and Mr. R. Locke, Jr. for their valuable technical assistance. One of us (S.H.K.) thanks Professor O. Hindsgaul for providing the facilities for the preparation of this manuscript.

### REFERENCES

- 1 R.K. Jain, C.F. Piskorz, and K.L. Matta, Carbohydr. Res., (1993) in press.
- 2 R.D. Cummings, I.S. Trowbridge, and S. Kornfeld, J. Biol. Chem., 257 (1982) 13421-13427.
- 3 K. Yamashita, Y. Tachibana, T. Ohkura, and A. Kobata, J. Biol. Chem., 260 (1985) 3963-3969.
- 4 J. Arango and M. Pierce, J. Cell. Biochem., 37 (1988) 225-231.
- 5 J.W. Dennis, K. Kosh, D.-M. Bryce, and M.L. Breitman, Oncogene, 4 (1987) 853-860.

- 6 J.W. Dennis, S. Laferte, C. Waghorne, M.L. Breitman, and R.S. Kerbel, *Science*, 236 (1987) 582-585.
- 7 J.W. Dennis, Cancer Surveys, 7 (1988) 573-594.
- 8 J.W. Dennis, Cancer Res., 49 (1989) 945-950,
- 9 S.H. Khan, S.A. Abbas, and K.L. Matta, Carbohydr. Res., 193 (1989) 125-139.
- 10 S.H. Khan, S.A. Abbas, and K.L. Matta, Carbohydr. Res., 205 (1990) 385-397.
- 11 S.H. Khan and K.L. Matta, J. Carbohydr. Chem., in press.
- 12 S.H. Khan and K.L. Matta, in H.J. Allen and E.C. Kisailus (Eds.), *Glycoconjugates: Composition, Structure, and Function*, Marcel-Dekker, New York, 1992, pp 361–378.
- 13 O.P. Srivastava, O. Hindsgaul, M. Shoreibah, and M. Pierce, Carbohydr. Res., 179 (1988) 137-161.
- 14 I. Brockhausen, S.H. Khan, and K.L. Matta, unpublished results.
- 15 M.M. Palcic, J. Ripka, K.J. Kaur, M. Shorcibah, O. Hindsgaul, and M. Pierce, J. Biol. Chem., 265 (1990) 6759–6769.
- 16 O. Hindsgaul, K.J. Kaur, G. Srivastava, M. Blaszczyk-Thurin, S.C. Crawley, L.D. Heerze, and M.M. Palcic, J. Biol. Chem., 266 (1991) 17858–17862.
- 17 I. Brockhausen, G. Moller, J-M. Yang, S.H. Khan, K.L. Matta, H. Paulsen, A.A. Grey, R.N. Shah, and H. Schachter, *Carbohydr. Res.*, 236 (1992) 281–299.
- 18 S.A. Abbas, C.F. Piskorz, and K.L. Matta, Carbohydr. Res., 167 (1987) 131-141.
- 19 R. Kuhn, H.H. Bear, and A. Seeliger, *Liebigs Ann. Chem.*, 611 (1958) 236–241; R. Kuhn and H. Trischmann, *Chem. Ber.*, 96 (1963) 184–187.
- 20 M.A. Nashed and L. Anderson. Tetrahedron Lett., 39 (1976) 3503–3506; M.A. Nashed, Carbohydr. Res., 60 (1978) 200–205.
- 21 P. Konradsson, D.R. Mootoo, R.E. McDevitt, and B. Fraser-Reid, J. Chem. Soc., Chem. Commun., (1990) 270–272.
- 22 V.H. Veeneman, S.H. van Leeuwen, and J.H. van Boom, Tetrahedron Lett., 31 (1990) 1331-1334.
- 23 M.E. Evens and F.W. Parrish, Carbohydr. Res., 54 (1977) 105-14.
- 24 R.G. Ault, W.N. Haworth, and E.L. Hirst, J. Chem. Soc., (1935) 1012-1020.