

Pergamon

PII: S0960-894X(97)00057-7

Peptidyl α -Keto Thiazole as Potent Thrombin Inhibitors

Yoshihisa Akiyama, Seiji Tsutsumi, Emiko Hatsushiba, Shoukichi Ohuchi, Tsuneo Okonogi

Pharmaceutical Research Laboratory, Meiji Seika Kaisha, Ltd., 760 Morooka-cho, Kohoku-ku, Yokohama 222, Japan

Abstract: We report the synthesis and evaluation of α -keto thiazole derivatives such as D-Phe-Pro-Argthiazole **9** as a novel type of thrombin inhibitor. Tripeptidyl α -keto thiazole **9** exhibited the inhibitory activity of thrombin at nanomolar levels and showed a more potent prolongation effect on clotting time than argatroban at a dose of 3 mg/kg intravenously. © 1997 Elsevier Science Ltd. All rights reserved.

Introduction

Thrombin (EC 3.4.21.5) is a serine protease and the key enzyme in the blood coagulation cascade.¹ Thrombin catalyzes the conversion of fibrinogen to fibrin, which then polymerizes to form a hemostatic plug. Thus, there has been considerable interest in research for thrombin inhibitors as anticoagulant agents.²

An approach to the design of the serine protease inhibitor has been the replacement of the scissile amide bond by an electron-withdrawing carbonyl group.³ Edwards *et al.* originally reported that α -keto benzoxazole derivative was a mechanism-based elastase inhibitor in which the nitrogen atom of the benzoxazole interacted with the histidine residue of the catalytic triad in serine protease.⁴ Recently, we elucidated that α -keto heterocyclic compounds possessed potent inhibitory activity for prolyl endopeptidase (PEP).⁵ In the course of our PEP inhibitors study, we substituted the amino acid residue in the P₁ position of phenylbutanoyl-Pro-Athiazole **1~5** (A: Pro, Ala, Val, Lys, and Arg) (Table 1). The basic residue of Arg or Lys was indispensable for the inhibitory activity of thrombin, but not PEP.

		Inhibitory Activity IC ₅₀ (µM)			
No	A (amino acid)	PEP	Elastase Thrombin		Trypsin
1	Proline	0.0044	>1000	>1000	>1000
2	Alanine	0.0050	>1000	>1000	>1000
3	Valine	7.7	>1000	>1000	>1000
4	Lysine	7.8	Not determined	46	2.4
5	Arginine	28	>1000	2.0	0.23

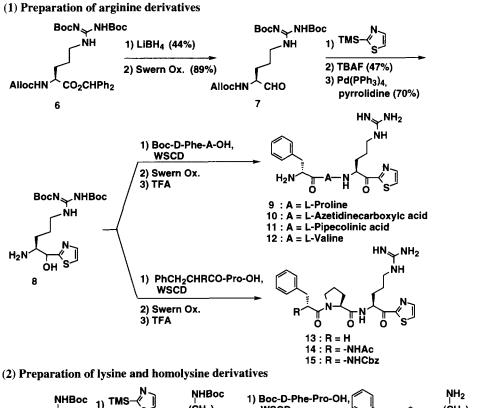
Table 1. Structure and inhibitory potencies of α -keto thiazole derivatives.

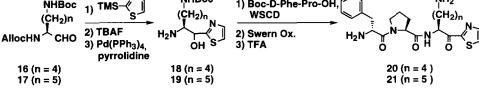
Ph(CH₂)₃CO-Pro-A-thiazole

These results also indicated that α -keto heterocyclic compound was useful for research on other serine protease inhibitors. Tripeptide aldehyde and chloromethyl ketone analogs of D-Phe-Pro-Arg sequence are reported to be high effective and reversible inhibitors of thrombin.² Considering these factors for inhibitor design, we examined the structure-activity relationship of the tripeptide α -keto thiazole compounds in the P₁, P₂ and P₃ positions.

Chemistry: We synthesized a series of tripeptide arginine, lysine, and homolysine derivatives with α -keto thiazole moiety. All compounds were obtained in a convergent strategy as shown in Scheme I. The arginine aldehyde 7 was prepared by the reduction of N-Allyloxycarbonyl(Alloc) arginine ester 6 followed by Swern oxidation.⁶ The reaction of aldehyde 7 with 2-(trimethylsilyl)thiazole gave the 2-thiazole derivative, which was converted to amino alcohol 8 by deprotection of the Alloc group. Condensation of the amino alcohol 8 with Boc-D-Phe-Pro-OH followed by Swern oxidation gave the tripeptide. The desired α -keto thiazole compound 9 was prepared by the TFA deprotection of the Boc protective group. In a similar manner, the corresponding arginine derivatives 10~15, lysine derivative 20, and homolysine derivative⁷ 21 were synthesized.⁶

Scheme I





Enzyme assay: Human thrombin was purchased from Sigma Chemical Company Ltd. Thrombin and trypsin assays were performed as described by Kawabata.⁸ PEP and elastase assays were carried out as described by Walter⁹ and Bieth¹⁰, respectively. Urokinase, plasmin, and plasma kallikrein assays were performed by the method used by Morita.¹¹ Experiments were conducted in 96-well plates, and the rates of hydrolysis were measured fluorometrically with excitation at 380 nm and emission at 440 nm or spectrophotometrically at 405 nm.

Ex vivo anticoagulant studies: Five minutes after intravenous administration of test compound **9** and argatroban¹², the prolonged activated partial thromboplastin time (APTT) and prothrombin time (PT) were determined by the method of Takamiya¹³ and Suzuki¹⁴, respectively.

Results & Discussion: Replacement of the arginine in 9 with a lysine or homolysine residue at the P_1 position resulted in a marked decrease in the inhibitory potency of thrombin, although the thrombin to trypsin ratios remained unchanged (Table 2). Unexpectedly, the homolysine derivative 21, which possesses carbon chains of the same length as arginine, was less potent than lysine derivative 20. These results showed that the arginine in the P_1 position was most suitable for the thrombin inhibitor. The α -keto thiazole inhibitor 9 exhibited a 40-fold increase in potency to inhibit thrombin as compared to argatroban.

P₃ P₂ P₁ P₁, Inhibitory Activity IC₅₀ (µM) Selectivity No R Thrombin Trypsin Trypsin/Thrombin -(CH₂)₃-N-≪^{NH} H NH₂ 9 0.0015 0.0042 2.8 -(CH₂)₄-NH₂ 0.26 20 1.2 4.6 -(CH₂)₅-NH₂ 21 14 16 1.1 Argatroban 0.060 not determined

Table 2. Effect of modifications in the P₁ position on thrombin inhibitors

The proline residue at the P₂ position in 9 was replaced by azetidine-2-carboxylic acid 10, pipecolinic acid 11, or valine 12 in order to investigate the influence of conformationally constrained amino acid (Table 3). While the azetidine-2-carboxylic acid derivative 10 maintained potency for the inhibition of thrombin, the pipecolinic acid derivative 11 was 480-times less active than 9. Compound 10 exhibited the same selectivity for thrombin over trypsin as 9. These results coincided with the results of Shuman et al.¹⁵ Replacement of proline with valine did not significantly change the potency for the inhibition of thrombin, although the valine derivative 12 exhibited remarkable selectivity for thrombin over trypsin (trypsin/thrombin=21). This selectivity would be due to a conformational difference between the S₂ subsite of thrombin and trypsin.



535

$ \begin{array}{c} $								
N	Inhibitory Activity IC ₅₀ (µM) Selectivity							
No	A	Thrombin	Trypsin	Trypsin/Thrombin				
10	-N2 0	0.003	0.009	3.0				
9	-N O	0.0015	0.0042	2.8				
11		0.73	41	56				
12	Me Me -N H O	0.0086	0.18	21				

Table 3. Effect of modifications in the P₂ position on thrombin inhibitors

536

The substitution of the D-phenylalanine residue in 9 changed the inhibitory potency of thrombin (Table 4). Deletion of an amino group at the P_3 position showed the marked decrease in potency (9 vs 13). The importance of amino group was further investigated by acylation of compound 9. A difference in inhibitory activity between the acetyl and benzyloxycarbonyl(Cbz) derivatives (14 vs 15) was observed. The acetyl compound 14 exhibited a 1000-fold loss in potency for the inhibition of thrombin as compared to compound 15. The addition of the acetyl group in 9 led to a 20-fold decrease in selectivity. In this study, the modification at the P_3 position demonstrated no improvement in selectivity for thrombin over trypsin. On the other hand, the aldehyde derivative incorporating lactam sulfonamide moiety are reported to display potent and selective inhibitor.¹⁶ The sulfonamide at the P_3 position might be a useful moiety for thrombin inhibitor.

Η

Table 4. Effect of modifications in the P₃ position on thrombin inhibitors

No	R	Inhibitory Activity IC ₅₀ (µM)		Selectivity
	K	Thrombin	Trypsin	Trypsin/Thrombi
13	-н	0.82	0.23	0.3
9	-NH ₂	0.0015	0.0042	2.8
14	-NHAc	2.3	0.23	0.1
15	NHCbz	0.002	0.0016	0.8

Recently, Costanzo et al. reported that α -keto benzothiazole inhibitor displays selectivity for thrombin compared with trypsin or plasmin.¹⁷ The moiety of α -keto benzothiazole in the P₁ position would also provide a novel interaction with thrombin, but not trypsin.

The compound **9** with D-Phe-Pro-Arg motif showed the most potent inhibitory activity for thrombin (IC₅₀ value: 1.5 nM). IC₅₀ values (μ M) for other serine proteases were as follows: trypsin, 0.0042; urokinase, 15.3; plasmin, >100; plasma kallikrein, >100.¹² These results suggested that the selectivity for thrombin over other proteases could be attributed to the different recognition of a D-Phe-Pro-Arg motif.

Finally, we evaluated the antithrombotic potency of compound 9 in comparison with argatroban for the ability to prolong prothrombin time (PT) and activated partial thromboplastin time (APTT) in rats (Table 5). Five minutes after intravenous administration of compound 9, PT was prolonged 1.5 fold over the control value at 1 mg/kg and 12 fold at 3 mg/kg. A similar inhibitory potency was observed following intravenous administration of 1 mg/kg and 3 mg/kg with 2.0 fold and 8.3 fold APTT elevation, respectively. Although the α -keto thiazole compound 9 showed a 40-fold increase in potency to inhibit the amidolytic activity of thrombin as compared to argatroban, the antithrombotic activity of compound 9 was about 2-fold more potent than that of argatroban. A possible explanation for the decreased activity of compound 9 in coagulation assays is that α -keto thiazole compound might be a slow binding inhibitor and/or was less stable in vivo than argatroban.

Compound	Dose (mg/kg)	PT inhibitor / PT control	APPT inhibitor / APPT control
9	1	1.5	2.0
9	3	12	8.3
Argatroban	3	6.2	4.8

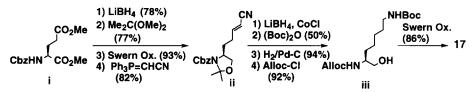
Table 5. Antithrombotic effect of compound 9 on intravenous administration

Conclusion: We extended the utility of α -keto heterocyclic compound to thrombin inhibitors. Tripeptidyl α -keto thiazoles exhibited the inhibitory activity of thrombin at nanomolar levels. The modification at the P₂ position altered the selectivity for thrombin over trypsin. Compound **9** exhibited a more potent prolongation effect on the clotting time than argatroban at a dose of 3mg/kg intravenously. The tripeptide derivative with α -keto heterocycle represented a promising new thrombin inhibitor.

References and Notes

- 1. Stubbs, M. T.; Bode, W. Trends. Biochem. Sci. 1995, 20, 23-28.
- Tapparelli, C.; Metternich, R.; Ehrhardt, C.; Cook, N. S.; *Trends. Pharmacol. Sci.* 1993, 14, 366-376. Stone, S. R. *Trends. Cardiovasc. Med.* 1996, 5, 134-140.
 Edmunds, J. J.; Rapundalo, S. T. *Annu. Rep. Med. Chem.* 1996, 31, 51-60.
- 3. Wiley, R. A.; Rich, D. H. Med. Res. Rev. 1993, 13, 327-384.
- Edwards, P. D.; Meyer, E. F., Jr.; Vijayalakshmi, J.; Tuthill, P. A.; Andisik, D. A.; Gomes, B.; Strimpler, A. J. Am. Chem. Soc. 1992, 114, 1854-1863.

- Tsutsumi, S.; Okonogi, T.; Shibahara, S.; Patchett, A. A.; Christensen, B. G.; Bioorg. Med. Chem. Lett. 1994, 4, 831-834.
 Tsutsumi, S.; Okonogi, T.; Shibahara, S.; Ohuchi, S.; Hatsushiba, E.;.Patchett, A. A.; Christensen, B. G. J. Med. Chem. 1994, 37, 3492-3502.
- 6. Meiji Seika Kaisha, Ltd. JP08020597-A
- 7. We prepared 17 from the glutamic acid ester i by using the methods shown below.



- Kawabata, S.; Miura, T.; Morita, T.; Kato, H.; Fujikawa, K.; Iwanaga, S.; Takada, K.; Kimura, T.; Sakakibara, S. *Eur. J. Biochem.* 1988, 172, 17-25.
- 9. Walter, R.; Simmons, W. H.; Yoshimoto, T. Molecular & Cellular Biochemistry 1980, 30, 111-127.
- 10. Bieth, J.; Spiess, B.; Wermuth, C. G. Biochemical Medicine 1974, 11, 350-357.
- Morita, T.; Kato, H.; Iwanaga, S.; Takada, K.; Kimura, T.; Sakakibara, S. J. Biochem. 1977, 82, 1495-1498.
- 12. Kikumoto, R.; Tamao, Y.; Tezuka, T.; Tonomura, S.; Hara, H.; Ninomiya, K.; Hijikata. A.; Okamoto, S. *Biochemistry*. **1984**, 23, 85-90.
- 13. Takamiya, O. Modern Medical Laboratory 1991, 19, 187-190.
- 14. Suzuki, S. Modern Medical Laboratory 1991, 19, 191-195.
- Shuman, R. T.; Rothenberger, R. B.; Campbell, C. S.; Smith, G. F.; Gifford-Moore, D. S.; Paschal, J. W.; Gesellchen, P. D. J. Med. Chem. 1995, 38, 4446-4453.
- Semple, J. E.; Rowley, D. C.; Brunck, T. K.; Ha-Uong, T.; Minami, N. K.; Owens, T. D.; Tamura, S. Y.; Goldman, E. A.; Siev, D. V.; Ardecky, R. J.; Carpenter, S. H.; e, Y.; Richard, B. M. Nolan, T. G.; Hakanson, K.; Tulinsky, A.; Nutt, R. F.; Ripka, W. C. J. Med. Chem. 1996, 39, 4531-4536.
- Costanzo, M. J.; Maryanoff, B. E.; Hecker, L. R.; Schott, M. R.; Yabut, S. C.; Zhang, H.-C.; Andrade-Gordon, R.; Kauffman, J. A.; Lewis, J. M.; Krishnan, R.; Tulinsky, A. J. Med. Chem. 1996, 39, 3039-3043.

(Received in Japan 15 November 1996; accepted 20 January 1997)