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TETRAHEDRON

Synthesis of Deoxy Phosphatidylinositol Analogues and Phosphonate Isosters of Ins(1,4,5)P₃

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> This paper is dedicated to the memory of Dr. S. D. Gero. Received 31 March 1999; accepted 8 September 1999

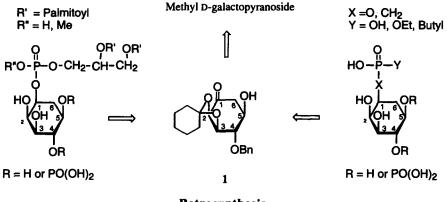
Abstract: The synthesis of phosphatidylinositol analogues, 6-deoxy Ins 1-(1,2-di-O-palmitoyl-sn-glycero)phosphate and 4,5-bisphosphate derivatives is presented. Two series of phosphonate isosters, 6-deoxy Ins(1)-butylphosphonate and 6-deoxy Ins(1)-Cmethylenephosphonate as well as its 4,5-bisphosphate analogue were also prepared. All phosphoinositide analogues were obtained from cyclohexanone polyol derived from the D-galactose. Modification of charge distribution at position 1 of PtdIns and InsP derivatives, by replacement of a P-OH group by an alkyl substitution or a P-C bond, resistant to cleavage by lipases, could induce inhibition of activity at further strategic enzymatic levels of the inositide cascade. © 1999 Elsevier Science Ltd. All rights reserved.

Agonist stimulated hydrolysis of phosphatidylinositol-4,5-bisphosphate [PtdIns(4,5)P₂] is the first step in the transmembrane signalling mechanism when cells respond to external stimuli. Under control of activated phospholipase C (PLC) via G-protein, two second messengers D-myo-inositol-1,4,5-trisphosphate $[Ins(1,4,5)P_3]$ and diacylglycerol (DG) are released into the cell.¹ From Ins(1,4,5)P_3, enzymatic process under phosphatases or kinases control affords subsequent inositol phosphate metabolites with still controversial biological role. During the last decade, the synthesis of modified inositol phosphate derivatives has been strongly investigated^{2,3} and phosphatidylinositol analogues are now prone to evaluation.⁴ Structural mimics of phosphatidylinositols might be anticipated to interfere with enzyme kinases generating PtdIns phosphate metabolites from PtdIns, or with phospholipases at the early steps of phosphoinositide cascade.⁵ Phospholipases are an important class of enzymes for growth factors and oncogene intracellular signalling.⁶ Along this line, we envisioned the synthesis of 6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-snglycero)phosphates and 4,5-bisphosphate derivatives. Deoxy analogues of PtdIns have already shown promising antitumoral activity.7 Complementarily, the synthesis of deoxy-myo-inositol phosphonates, has been considered as an interesting tool to interfere with the phosphoinositol transduction pathway, or mimicking the original PtdIns precursors. Two series of phosphonate isosters, 6-deoxy-D-myo-inositol-1-(butyl)phosphonate and 6-deoxy-D-myo-inositol-1-(C-methylene)phosphonate as well as their 4,5-bisphosphate analogues, were then prepared. 6-Deoxy-Ins(1,4,5)P₃ has already been shown to be a full agonist for Ca²⁺ release in

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permeabilized SH-SY5Y human neuroblastoma cells, a relatively potent $Ins(1,4,5)P_3$ 5-phosphatase inhibitor and a weak substrate for $Ins(1,4,5)P_3$ 3-kinase.⁸ Modifications of $Ins(1,4,5)P_3$ into lipophilic derivatives have undergone promising property with the aim of incorporating the membrane of intact cells.^{3a} Therefore, C-1 phosphonate analogues of Ins(1)P and $Ins(1,4,5)P_3$, associated with the lack of the 6-OH function, might affect the enzymatic dephosphorylation processes releasing inositol phosphate metabolites from $Ins(1,4,5)P_3$. Specific $Ins(1,4,5)P_3$ receptor's deactivation by 6-deoxy-Ins(1)-C-methylenephosphonate- $(4,5)P_2$ and consequently influence on the Ca²⁺ mobilization from endoplasmic reticulum could also be expected.

In continuation of our study on the synthesis of deoxy inositol ring from D-galactose, the inosose 1, resulting from the Ferrier rearrangement of hexenogalactopyranoside precursor,⁹ has been regarded as the suitable intermediate to accede to the proposed PtdIns analogues and phosphonate isosters of 6-deoxy InsP.

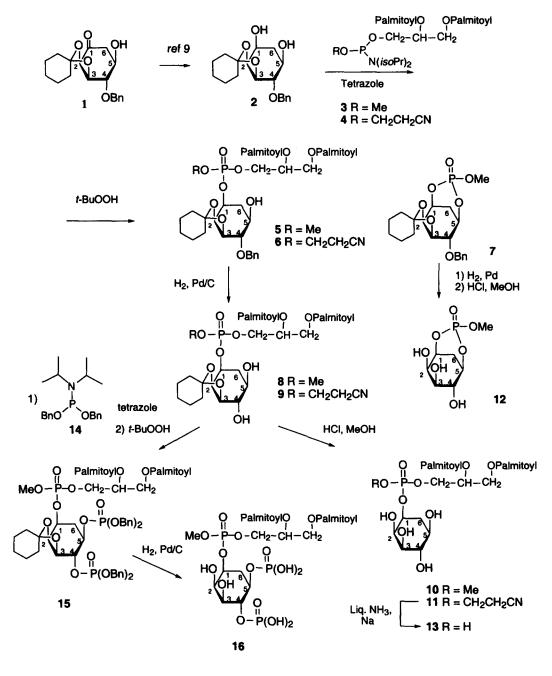


Retrosynthesis



As described recently, the protected 6-deoxy Ins(1,5) diol 2 is easily available by the stereoselective reduction of ketone 1 (Scheme 1). Phosphite triester chemistry,^{10,7} selected to minimize problems arising from steric hindrance, was applied to compound 2 to produce 6-deoxy PtdIns analogues. Thus, activated phosphites 3 and 4, bearing a protected 1,2-di-O-palmitoyl-sn-glycerol, were first prepared from diisopropylaminochloro(methoxy)phosphine and corresponding cyanoethoxy analogue, respectively. Selective reaction of the diol 2 was attempted in CH₂Cl₂ in the presence of tetrazole and the resulting phosphite intermediates were oxidized by t-butyl hydroperoxide (t-BuOOH) to give the protected phosphatidylinositols 5 and 6, respectively, in 40% overall yields. Surprisingly, the reaction involving the methoxyphosphite reagent 3 led to the concomitant formation of the Ins(1,5)-cyclic phosphate 7 in moderate 11% yield. The catalytic hydrogenolysis of the intermediates 5 and 6, using 10% Pd/C in EtOH under hydrogen pressure, furnished quantitatively the 4,5-diols 8 and 9, respectively. Acidic removal of the cyclohexylidene group from 8 and 9, with methanolic HCl solution, gave the methyl and cyanoethyl 6-deoxy PtdIns 10 and 11 over 90% yield in both cases. A similar hydrogenolysis-hydrolysis reaction sequence was applied to the cyclic phosphate 7 leading to the 6-deoxy-Ins(1,5)-methylcyclic phosphate 12 in 90% yield. Compound 12 could be regarded as an original InsIP analogue directed upon inhibition of monophosphatases. Finally, the access to the 6-deoxy-Dmyo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero)phosphate 13 was attempted from the phosphodiester

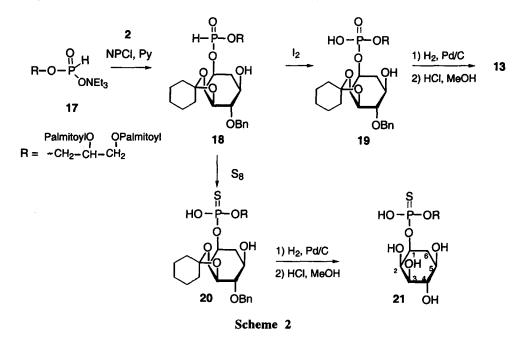
intermediate 11, by deprotection of the cyanoethoxy group using liquid ammonia treatment in the presence of a catalytic amount of sodium, but unsuitable transesterification remained difficult to avoid.





Alternatively, the phosphatidyl diol 8 was phosphorylated following (4,5)-bisphosphitylation procedure using the (dibenzyloxy)diisopropylaminophosphite reagent 14 and tetrazole. Subsequent *t*-BuOOH oxidation gave the corresponding protected trisphosphate 15 in 60% overall yield. Deprotection of the trisphosphates 15 by hydrogenolysis afforded the desired 6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-snglycero)methylphosphate-4,5-bisphosphate 16 isolated as tetra TRIS salt.

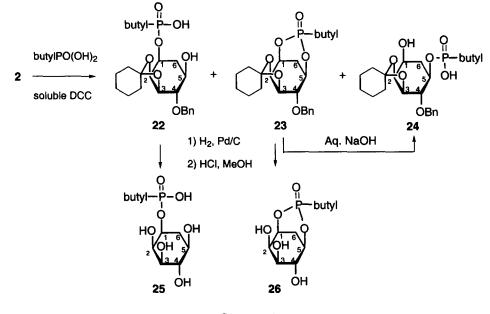
The access to the deoxy phosphatidyl analogue 13 was envisioned more efficiently using the *H*-phosphonate method, developed by Stawinski et al.,^{11,4c} involving the (1,2-di-*O*-palmitoyl-*sn*-glycero)-*H*-phosphonate reagent 17 (Scheme 2). The glycerophosphonate 17 was first prepared following the described procedure by the reaction of 1,2-dipalmitoyl-*sn*-glycerol in toluene with PCl₃/imidazole complex, in the presence of Et₃N.



Condensation of 17 with the *myo*-inositol diol 2 was carried out in the presence of 5,5-dimethyl-2-oxo-2-chloro-1,3,2-dioxaphosphorinan (NPCl) in dry pyridine. After neutralization of the medium with triethylammonium bicarbonate and extraction with CH_2Cl_2 , the *H*-phosphonate 18 was isolated as a syrup. Subsequent oxidation of intermediate 18, by addition of iodine in a pyridine/toluene solution, furnished the phosphodiester 19 in 50% overall yield. The use of sulfur (S₈) for the oxidation of compound 18 allowed the access to the corresponding phosphorothioate 20. Consequently, following a similar hydrogenolysishydrolysis sequence as describe previously, the 6-deoxy-D-*myo*-inositol-1-(1,2-di-*O*-palmitoyl-*sn*glycero)phosphate 13 and its corresponding phosphorothioate 21 (no poisoning of the catalyst was observed) were obtained from their phosphodiester precursors 19 and 20 in 90% overall yields.

During the course of our research on modified phosphate analogues of inositol metabolites, the synthesis of phosphonate isosters was investigated from the inosose 1 or its corresponding reduced derivative 2. The condensation of *n*-butyl-phosphonic acid with the 6-deoxy-Ins(1,5)diol 2, in the presence of

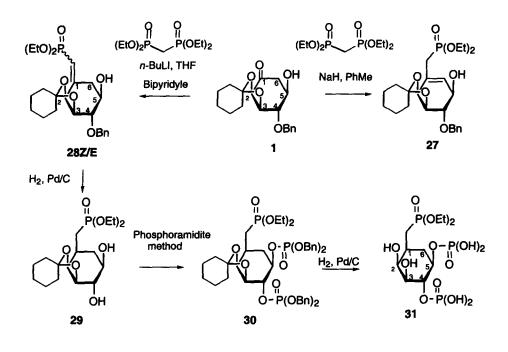
trichloroacetonitrile and pyridine, led to the formation of 6-deoxy-Ins(1)-butylphosphonate 22, (1,5)-butylcyclicphosphonate 23 and (5)-butylphosphonate 24 in 6/1/1 ratio (50% yield) (Scheme 3).



Scheme 3

The proportion in the mixture 22/23/24 of the target butylphosphonate 22 was improved to 10/1/2 using N-(dimethylaminopropyl)N'-ethylcarbodiimide (soluble DCC) as coupling agent in CH₂Cl₂ at reflux. In this latter case, the global yield of the reaction was also increased to 65%. Interestingly, the hydrolysis of the cyclic phosphonate 23, under basic condition (aq. NaOH), afforded selectively the (5)-phosphonate 24 in quantitative manner. Alternatively, the deprotection of the intermediates 22 and 23 led to the corresponding 6-deoxy Ins(1)-butylphosphonate 25, isolated as sodium salt in 90% yield, and the (1,5)-butylcyclic phosphonate 26 in 86% yield.

Finally, the synthesis of 1-C-(methylene)phosphonate analogues of $Ins(1,4,5)P_3$ was attempted by a Wittig-Horner reaction from the inosose 1 (Scheme 4). The treatment of the cyclohexanone 1 with tetraethyl methylenediphosphonate and *n*-butyllithium, in THF, furnished a mixture of two exovinyl phosphonates 28, in 70% yield (28E/Z: 3.5/1.5). The use of sodium hydride as base, instead of *n*-butyllithium, induced a subsequent migration of the unsaturation into the cycle leading to the cyclohex-1,6-enol 27 in 60% yield. Surprisingly, we were unable to reduce the endocyclic double bond of 27, whereas, the reduction of the exocyclic vinyl derivatives 28 was achieved under usual hydrogenation conditions (H₂, 10% Pd/C). The reaction applied on the diastereomeric mixture 28 (E/Z) provided stereoselectively the 1-C-methylene diol 29 with subsequent loss of the benzyl ether protection. Thus, phosphorylation of diol 29 was then achieved by phosphoramidite method, using the phosphite reagent 14, tetrazole and subsequent oxidation with *t*-BuOOH, to give the bis(dibenzyl)phosphate 30 in 70% overall yield from 28. The hydrogenolysis of the intermediate 30 furnished the 6-deoxy-Ins(1)-C-methylenediethylphosphonate-4,5-bisphosphate 31, isolated as tetra-TRIS salt.



Scheme 4

CONCLUSION

Our contribution in the elaboration of inositol metabolite analogues,^{3,9} allowed the synthesis of 6-deoxy PtdIns derivatives and some phosphonate isosters of 6-deoxy-Ins(1)P and 6-deoxy-Ins(1,4,5)P₃. Modification of charge distribution at the position 1 of PtdIns and InsP derivatives, by replacement of a P-OH function by a P-OR group or a P-C bond, resistant to cleavage by lipases, should help to modulate the phosphoinositide cascade turn-over. These analogues are of interest as inhibitors of several enzymatic activation or deactivation processes and appear as new precursors for the synthesis of modified phosphatidylinositol derivatives. As argued in a previous publication,^{3a} the transformation of these hydrophilic compounds into lipophilic derivatives, aimed to incorporate the lipidic cell membrane, should be easily envisionned to allow *in vivo* experiments. Therefore, we believe that the synthetic pathway developed from D-galactose, for the stereoselective synthesis of a variety of deoxy *myo*- and *chiro*-inositol precursors of polyphosphate and phosphatidyl derivatives,^{3,9} could be extended to the preparation of analogues of inositolphosphoglycan mediators (IPG) or glycosylphosphatidylinositol precursors (GIPs).¹²

Acknowledgements : This work was funded by Bayer-AG Geschäftsbereich Pharma und Entwicklung Institut. We thank Dr. E. Bischoff for biological investigations and Professor B. V. L. Potter for fruitful discussions.

EXPERIMENTAL PART

¹H, ¹³C and ³¹P NMR spectra were recorded on Bruker spectrometra WP 200, AC 200, AC 250, WM 400 or ARX 400; chemical shifts are expressed in parts per million (ppm) referenced to TMS or H₃PO₄. Coupling constants (J) are given in hertz (Hz). Multiplicities are recorded as s (singlet), sl (large singulet), d (doublet), t (triplet), q (quartet), and m (multiplet or complex). The $[\alpha]_D$ were recorded on Perkin-Elmer 241-MC sodium absorption at 20°C. Mass spectra (m/z % base peak) were recorded on Atlas CH₄ or AEI MS9 spectrometra. Melting points were determined on a C. REICHERT microscope apparatus and are uncorrected. Elemental analyses were carried out at the "Laboratoire de Microanalyse de l'I.C.S.N." (CNRS, Gif/Y vette). All solvents were freshly distilled prior to use by standard methods¹³. Flash chromatography was performed on silica-gel Merck 60 (230-400 mesh) or licoprep RP-8 resin (15-25µm Merck). Thin layer chromatography was performed on precoated plates of silica gel PF₂₅₄ neutralized with sodium bicarbonate.

4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-snglycero)methyl phosphate (5) and 4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-myoinositol-1,5-cyclic(methyl)phosphate (7).

A solution of methyl diisopropylaminochlorophosphite 3 (77 mg, 0.39 mmol), Et₃N (0.1 ml) and sn-1,2-di-O-palmitoyl glycerol (221 mg, 0.39 mmol) in dry CH₂Cl₂ (15 ml) was stirred for 2h at r.t.. Diol 2 (100 mg, 0.3 mmol) and tetrazole (42 mg, 0.6 mmol) were added and the mixture was stirred for a further 6h. The phosphite intermediate obtained was oxidized by addition of *t*-butyl hydroperoxide (*t*-BuOOH, 0.1 ml). The solution was treated by aq. sodium thiosulfate solution and extracted with CH₂Cl₂. The organic layer was evaporated to dryness and the residue was submitted to flash chromatography on silicagel (AcOEt, hexane) to give the phosphatidylinositol 5 (40% yield) and the cyclic phosphate analogue 7 (11% yield) both crystallized from MeOH. Compounds 5 and 7 was isolated under the form of two isomers at the phosphorus atom. Evidence is only provided for this statement by the observation of ¹H NMR singlet type signals of the OCH₃ group.

Compound 5: m.p. 39-41 °C; M.S. (C.I.; isobutanol; m/z): 979 [MH]⁺, 551 [CH₂CH(OR)CH₂OR]⁺, 313 [551-CH₃(CH₂)₁₄C=O + H]⁺, 239 [CH₃(CH₂)₁₄C=O]⁺ with R = CH₃(CH₂)₁₄C=O; ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.20 (m, 1H, CHCH₂OP), 4.80 (2d, 2H, CH₂Ph, Jgem= 10), 4.65 (m, 1H, H-1), 4.45 (t, 1H, H-2, J₂₋₁=J₂₋₃=3), 4.41 to 4.0 (m, 5H, H-3, CH₂OP, CH₂OCO), 3.76 and 3.84 (2s, 3H, OCH₃), 3.51 (m, 2H, H-4, H-5), 2.42 to 2.0 (m, 6H, H-6e, H-6a, 2 CH₂CO), 0.85 (m, 6H, 2CH₃); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 173.1, 172.2 (C=O), 81.8 (C-4), 78.8 (C-3), 74.7 (C-2), 73.4 (CH₂Ph), 71.4 (C-1), 69.3 (C-5, CHCH₂OP), 65.6, 61.7 (CH₂OP, CH₂OC=O), 54.6 (OCH₃), 37.4 (C-6); ³¹P NMR (81 MHz, CDCl₃) δ ppm: -0.27; (Found: C, 67.18; H, 10.01; P, 3.68; C₅₅H9₅O₁₂P requires C, 67.45; H, 9.78; P, 3.16 %). Compound 7: m.p. 165-167 °C; M.S. (C.I.; isobutanol; m/z): 411 [MH]⁺; ¹H NMR (250 MHz, CDCl₃) δ ppm: 4.65 (2d, 2H, CH₂Ph, Jgem= 11), 4.55 (m, 2H, H-1, H-5), 4.35 (t, 1H, H-3, J₃₋₄=J₃₋₂=6), 4.25 (sl, 1H, H-2), 4.10 (t, 1H, H-4, J₄₋₃=J₄₋₅=6), 3.30 and 3.20 (2s, 3H, OCH₃), 2.72 (m, 1H, H-6a, J_{6a-6e}=15), 2.0 (m, 1H, H-6e); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 75.4 (C-4), 73.8, 73.6, 73.3, 73.2 (C-1, C-2, C-3, C-5), 73.3 (CH₂Ph), 54.1 (OCH₃), 25.3 (C-6); ³¹P NMR (81 MHz, CDCl₃) δ ppm: -9.99; (Found: C, 58.12; H, 6.57; P, 7.52; C₂₀H₂₇O₇P requires C, 58.52; H, 6.62; P, 7.55 %).

4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-snglycero)cyanoethyl phosphate (6).

Following the same experimental conditions used for the preparation of compound **5**, using cyanoethyl diisopropylaminochlorophosphite **4** instead of reagent **3**, the phosphatidyl intermediate **6** was isolated in 40% yield from **2**. m.p. 41-43 °C; ¹H NMR (200 MHz, CDCl₃) δ ppm: 5.20 (m, 1H, CHCH₂OP), 4.75 (m, 2H, CH₂Ph), 4.65 (m, 1H, H-1), 4.50 (sl, 1H, H-2), 4.45 to 3.90 (m, 9H, H-3, CH₂CH₂CN, CH₂OP, CH₂OC=O), 3.60 (m, 2H, H-4, H-5), 2.80 to 2.0 (m, 10H, H-6a, H-6e, 2 CH₂CH₂C=O), 0.85 (m, 6H, 2CH₃); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 173.2, 172.8 (C=O), 116.0 (CN), 82.1 (C-4), 78.9 (C-3), 74.7 (C-2), 73.5 (CH₂Ph) 72.0, (C-1), 69.4, 69.3 (C-5, CHCH₂OP), 66.1, 62.2, 61.7 (CHCH₂OP, CH₂OC=O, CH₂CH₂CN), 37.5 (C-6), 19.5 (CH₂CN), 14.1 (CH₃); ³¹P NMR (81 MHz, CDCl₃) δ ppm: -2.0; (Found: C, 66.85; H, 9.56; N, 1.65; P, 3.13; C₅₇H₉₆O₁₂NP requires C, 67.23; H, 9.50; N, 1.38; P, 3.04 %).

2,3-O-Cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero)methyl phosphate (8).

The phosphate 5 (98 mg, 0.1 mmol) dissolved in EtOH (5 ml) was hydrogenated for 4h. at r.t. in the presence of palladium on carbon 10% (Pd/C) at 75 psi. The catalyst was removed by filtration and the diol **8** was isolated after evaporation of the organic solvents under reduced pressure and crystallization from MeOH (quantitative yield). m.p. 46-47 °C; ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.25 (m, 1H, CHCH₂OP), 4.65 (m, 1H, H-1), 4.40 (sl, 1H, H-2), 4.35 to 4.05 (m, 6H, H-3, H-5, CH₂OP, CH₂OC=O), 4.0 (t, 1H, H-4, J_{4.3}=J_{4.5}=9), 3.70 (sl, 3H, OCH₃), 2.50 to 2.0 (m, 6H, H-6a, H-6e, 2 CH₂C=O), 0.85 (m, 6H, 2CH₃); ¹³C NMR (63 MHz, CDCl₃) δ ppm: 173.3, 173.1 (C=O), 111.6 (O-C-O), 78.7 (C-4), 75.9 (C-3), 74.5 (C-2), 71.6 (C-1), 69.5 (C-5, CHCH₂OP), 54.8 (OCH₃), 37.8 (C-6), 14.2 (CH₃); ³¹P NMR (81 MHz, CDCl₃) δ ppm: -0.73; (Found: C, 65.27; H, 10.10; P, 3.87; C₄₈H₈₉O₁₂P requires C, 64.83; H, 10.09; P, 3.48 %).

2,3-O-Cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero) cyanoethyl phosphate (9).

Following the same experimental conditions used for the preparation of compound **8**, the phosphatidylinositol **9** was isolated in quantitative yield from **6**. m.p. 41-43 °C; ¹H NMR (200 MHz, CDCl₃) δ ppm: 5.22 (m, 1H, CHCH₂OP); 4.60 (m, 1H, H-1); 4.47 (sl, 1H, H-2); 4.45 to 3.85 (m, 9H, H-3, CH₂CH₂CN; CH₂OP et CH₂OC=O); 3.62 (m, 2H, H-4, H-5); 2.81 to 2.10 (m, 10H, H-6e, H-6a, 2CH₂CH₂C=O); 0. 82 (m, 6H, 2CH₃); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 173.0, 172.5 (2C=O); 115.4 (CN); 82.2 (C-4); 79.1 (C-3); 74.6 (C-2); 71.8 (C-1); 69.5, 69.1 (C-5, CHCH₂OP); 66.0, 62.4, 61.9 (CHCH₂OP; CH₂OC=O; CH₂CH₂CN); 36.9 (C-6); 19.5 (CH₂C N); 14.3 (CH₃).

6-Deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero)methyl phosphate (10).

The ketal **8** (90 mg, 0.1 mmol), dissolved in methanol (10ml)-conc. HCl (2ml) solution, was stirred for 3h. at r.t.. After neutralization of the solution by sodium bicarbonate, filtration on Celite with AcOEt and partial concentration of the filtrate, the tetrol **10** was crystallized from MeOH (90% yield). m.p. 70-73 °C; ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.25 (m, 1H, CHCH₂OP), 4.50 to 4.0 (m, 7H, H-1, H-2, H-3, CH₂OP, CH₂OC=O), 3.80 (m, 5H, H-4, H-5, OCH₃), 2.35 (m, 6H, H-6a, H-6e, 2 CH₂C=O), 0.90 (m, 6H, 2CH₃); ¹³C NMR (63 MHz, CDCl₃) δ ppm: 173.6, 173.3 (C=O), 73.0, 72.3, 71.2, 69.6, 68.5 (C-1, C-2, C-3, C-4, C-5, CHCH₂OP), 65.2, 61.6 (CH₂OP, CH₂OC=O), 55.0 (OCH₃), 34.3 (C-6), 14.2 (CH₃); ³¹P NMR

(81 MHz, C₅D₅N) δ ppm: +0.9; (Found: C, 61.76; H, 10.12; C₄₂H₈₁O₁₂P.1/2H₂O requires C, 61.66; H, 10.10%

6-Deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero)cyanoethyl phosphate (11) and 6-Deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero) phosphate (13).

Following the same experimental conditions used for the preparation of compound 10, the phosphatidyl tetrol 11 was obtained in quantitative yield from precursor 9. The tetrol 11 (crude product) was dissolved in saturated liquid ammonia condensed at -60°C in a solution of methanol. A little amount of sodium metal was then added and the reaction was followed by TLC. After concentration to dryness the phosphoinositide analogue 13 was obtained as sodium salt (70% yield). Compound 13 was also prepared by deprotection of 19 by the same procedure used to prepare compound 21 from 20, in 88% yield. Compound 13: ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.20 (m, 1H, CHCH₂OP), 4.30 (m, 1H, H-1), 4.31-4.0 (m, 6H, H-2, H-3, CH₂OP, CH₂OC=O), 3.60 (m, 2H, H-4, H-5), 2.40-2.0 (m, 6H, H-6a, H-6e, 2 CH₂C=O), 0.90 (m, 6H, 2CH₃); ¹3C NMR (63 MHz, CDCl₃) δ ppm: 173.4 (C=O), 78.9 (C-4), 72.1 (C-2, C-3), 69.3, 68.2 (C-1, CHCH₂OP), 65.0 (C-5), 62.5 (CH₂OP, CH₂OC=O), 34.0 (C-6), 14.1 (CH₃); ³1P NMR (81 MHz, CDCl₃) δ ppm: -1.47; (Found: C, 60.16; H, 9.92; P, 3.53; C4₁H₇₉O₁₂P.1H₂O requires C, 60.56; H, 9.79; P3.81 %).

6-Deoxy-D-myo-inositol-1,5-cyclic (methyl)phosphate (12).

To a solution of cyclic phosphate derivative 7 (80mg, 0.2mmol) in EtOH (10 ml) was added 10% Pd/C (150 mg) and the mixture was stirred for 2h under hydrogen pressure (75 psi). After filtration of the solid on Celite (EtOH) and concentration to dryness of the filtrat, the residue was dissolved in MeOH (10ml) and conc. HCl (1 ml) was then introduced. The solution was stirred for further 3h at r.t. and the organic layer was concentrated. The residue was chromatographed on RP8 (MeOH/H₂O) to give the triol **12** (90% yield). ¹H NMR (250 MHz, CDCl₃) δ ppm: 4.65 (m, 1H, H-4), 4.50 (sl, 1H, H-3), 4.20 (sl, 1H, H-2), 4.05 (sl, 1H, H-5), 3.78 to 3.72 (m, 4H, H-1, OCH₃), 2.70 (m, 1H, H-6a), 2.25 (m, 1H, H-6e); ¹³C NMR (63 MHz, CDCl₃) δ ppm: 80.9 (C-4), 78.0 (C-3), 73.5 (C-2), 71.4 (C-5), 69.2 (C-1), 54.2 (OCH₃), 25.5 (C-6); ³¹P NMR (81 MHz, CD₃OD) δ ppm: -6.46; (Found: C, 33.49; H, 5.68; C₇H₁₃O₇P.1/2H₂O requires C, 33.74; H, 5.66 %).

2,3-O-Cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero)methyl phosphate-4,5-bis(dibenzyl) phosphate (15).

Treatment of diol 8 by phosphoramidite method, using the (dibenzyl)diisopropylaminophosphite reagent 14, tetrazole and oxidation by *t*-BuOOH, gave the (dibenzyl)phosphate inositol 15 which crystallized from MeOH (60% yield). m.p. 40-42 °C; ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.25 (m, 1H, CHCH₂OP), 5.12 (m, 8H, CH₂Ph), 4.61 (m, 2H, H-1, H-4), 4.50 (t, 1H, H-2, J₂₋₁=J₂₋₃=5), 4.43 to 4.0 (m, 6H, H-3, H-5, CH₂OP, CH₂OC=O), 3.81 (sl, 3H, OCH₃), 2.49 (m, 1H, H-6e), 2.32 (m, 5H, H-6a, 2 CH₂C=O), 0.85 (m, 6H, 2CH₃); ¹³C NMR (63 MHz, CDCl₃) δ ppm: 173.3, 172.8 (C=O), 112.1 (OCO), 81.3 (C-4), 74.7 (C-3), 73.5 (C-2), 71.1 (C-1), 69.5 (C-5, CHCH₂OP, CH₂Ph), 65.9, 61.8 (CH₂OP, CH₂OC=O), 54.9 (OCH₃), 37.5 (C-6), 14.2 (CH₃); ³¹P NMR (81 MHz, CDCl₃) δ ppm: -0.95; -0.45; (Found: C, 64.02; H, 8.42; P, 6.63; Cr₆H₁₁₅O₁₈P₃.1/2H₂O requires C, 63.94; H, 8.26; P, 6.51 %).

6-Deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero)methylphosphate-4,5-bis phosphate (16).

The phosphate 15 (70 mg, 0.05 mmol) dissolved in EtOH (5 ml) was hydrogenated for 2h. at r.t. in the presence of 10% Pd/C (100mg) at 75 psi. The catalyst was removed by filtration on Whatman paper (H₂O) and the trisphosphate analogue 16 was isolated by lyophilization, after concentration of the solvents and addition of TRIS salt (24 mg). ¹H NMR (250 MHz, D₂O) δ ppm: 5.19 (m, 1H, CHCH₂OP), 4.21-3.34 (m, 48H, H-1, H-2, H-3, H-4, H-5, CH₂OP, CH₂OC=O, CH₂OH, OCH₃), 2.22 (m, 6H, H-6e, H-6a, 2 CH₂C=O), 1.49 (m, 4H, 2CH₂CH₂C=O), 1.18 (m, 48H, 2CH₂, 12CH₃), 0.82 (m, 6H, 2CH₃); ¹³C NMR (50 MHz, CD₃OD) δ ppm: 81.3 (C-4), 74.2, 73.7, 72.6 (C-1, C-2, C-3), 70.6 (C-5, CHCH₂OP), 67.2, 62.8 (CH₂OP, CH₂OC=O), 62.5 (CH₂OH), 54.9 (OCH₃), 35.0 (C-6), 14.4 (CH₃).

4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-snglycero)H-phosphonate (18); 4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero) phosphate (19) and 4-O-Benzyl-2,3-O-cyclohexylidene-6deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero) phosphorotioate (20).

The diol 2 (200 mg, 1.8 mmol) and 5,5-dimethyl-2-oxo-2-chloro-1,3,2-dioxaphosphorinane (NPCl, 330 mg, 1.8 mmol) was added to a solution of triethylammonium (1,2-di-O-palmitoyl-sn-glycero)Hphosphonate compound 17 (480 mg, 0.66 mmol) in pyridine (5 ml). The mixture was stirred for 1h. at r.t. before neutralization with aq. triethylammonium bicarbonate solution (TEAB, 0.1M). Extraction with CH₂Cl₂ and evaporation of the organic layer gave the H-phosphonate intermediate 18 which was used without further purification. For the preparation of compound 19, the H-phosphonate intermediate 18 was dissolved in aqueous-pyridine solution (10 ml; 98/2, v/v) and iodide (335 mg, 1.32 mmol) was added. The mixture was stirred for 30 mn. at r.t. before addition of an aqueous solution containing 5% of sodium bisulfite (20 ml). After extraction with CH₂Cl₂ and evaporation of the organic layer, the residue was purified by chromatography on silicagel (AcOEt, CH₃OH) to give the phosphatidyl inositol 19 as solid (50% yield). m.p. 170-172 °C; ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.17 (m, 1H, CHCH₂OP), 4.81 (2d, 2H, CH₂Ph, Jgem= 10), 4.53 (m, 1H, H-1), 4.35 (sl, 1H, H-2), 4.02 (m, 5H, H-3, CH2OP, CH2OCO), 3.55 (m, 1H, H-5), 3.46 (t, 1H, H-4, J₄₋₃=J₄₋₅=9Hz), 2.24 (m, 5H, H-6e, 2 CH₂CO), 1.90 (m, 1H, H-6a), 0.85 (m, 6H, 2CH₃); ¹³C NMR (63 MHz, CDCl₃) & ppm: 173.9, 173.6 (C=O), 110.7 (OCO), 85.1 (C-4), 79.7 (C-3), 76.0 (C-2), 73.6 (CH₂Ph), 70.7 (C-1), 70.1 (CHCH2OP), 68.2 (C-5), 64.1, 63.1 (CH2OP, CH2OC=O), 34.4 (C-6), 14.2 (CH3); ³¹P NMR (81 MHz, CDCl₃) δ ppm: -4.11; (Found: C, 64.90; H, 9.76; P, 3.28; C₅₄H₉₃O₁₂P.2H₂O requires C, 64.77; H, 9.76; P, 3.10 %). For the preparation of compound 20: the H-phosphonate intermediate 18 was then dissolved in pyridine-toluene solution (10 ml; 1/1, v/v) and sulfur (48 mg, 1.5 mmol) was added. The mixture was stirred for 8h. at r.t. After extraction with CH₂Cl₂ and evaporation of the organic layer, the residue was purified by chromatography on silicagel (AcOEt, CH₃OH) to give the phosphorothioate analogue 20 (47% yield). ¹H NMR(250 MHz, CDCl₃) 8 ppm: 5.25 (m, 1H, CHCH₂OP), 4.78 (2d, 2H, CH₂Ph, Jgem= 10), 4.46 (sl, 1H, H-2), 4.42 (m, 1H, H-1), 4.11 (m, 5H, H-3, CH₂OP, CH₂OCO), 3.50 (m, 4H, H-4, H-5, 20H), 2.33 (m, 4H, CH₂C=O), 2.21 to 1.91 (m, 2H, H-6a, H-6e), 0.89 (m, 6H, 2CH₃); ¹³C NMR (63 MHz, CDCl₃, D₂O) δ ppm: 173.9 (2C=O), 110.9 (OCO), 84.8 (C-4), 79.5 (C-3), 76.0 (C-2), 73.3 (CH₂Ph), 70.8 (C-1), 70.5 (CHCH₂OP), 68.1 (C-5), 64.4, 62.9 (CH₂OP, CH₂OC=O), 37.7 (C6), 14.2 (CH₃); (Found: C, 65.89; H, 9.85; P, 2.81; S, 2.96; C₅₄H₉₃O₁₁PS requires C, 66.09; H, 9.55; P, 3.16; S, 3.27 %).

6-Deoxy-D-myo-inositol-1-(1,2-di-O-palmitoyl-sn-glycero) phosphorotioate (21).

To a solution of phosphorothioate intermediate **20** (98 mg, 0.1mmol) in EtOH (10 ml) was added 10% Pd/C (100 mg) and the mixture was stirred for 2h. under hydrogen pressure (75 psi). After filtration of the solid on Celite (EtOH) and evaporation to dryness, the residue was dissolved in MeOH (10ml) and conc. HCl (1 ml) was then introduced. The solution was stirred for further 3h at r.t., neutralized with NaHCO₃ and concentrated to dryness. The residue was chromatographed on RP-8 (MeOH/H₂O) to give the tetrol **21** (90% yield) isolated as sodium salt. ¹H NMR (250 MHz, CDCl₃) δ ppm: 5.20 (m, 1H, CHCH₂OP), 4.52 to 4.10 (m, 7H, H-2, H-3, CH₂OP, CH₂OC=O), 3.62 (m, 2H, H-4, H-5), 2.38 to 2.05 (m, 6H, H-6a, H-6e, 2CH₂C=O), 0.91 (m, 6H, 2CH₃); ¹³C NMR (63 MHz, CDCl₃) δ ppm: 173.9, 173.5 (C=O), 77.8 (C-4), 72.3 (C-2, C-3), 68.4 (C-1, CHCH₂OP), 65.1 (C-5), 62.9 (CH₂OP, CH₂OC=O), 34.2 (C-6), 14.1 (CH₃); ³¹P NMR (81 MHz, CDCl₃) δ ppm: +56; (Found: C, 61.10; H, 9.60; P, 3.96; S, 3.58; C₄₁H₇₉O₁₁PS requires C, 60.71; H, 9.81; P, 3.82; S, 3.95 %).

4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-*myo*-inositol-1-(*n*-butyl) phosphonate (22) ; 4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-*myo*-inositol-1,5-cyclic(*n*-butyl) phosphonate (23) and 4-O-Benzyl-2,3-O-cyclohexylidene-6-deoxy-D-*myo*-inositol-5-(*n*-butyl) phosphonate (24).

To a solution of diol 2 (300 mg, 0.9 mmol) dissolved in dry CH₂Cl₂ (20 ml) under argon, was added n-butylphosphonic acid (400 mg, 1.08 mmol), DCC (860 mg, 4.5 mmol) and dimethylaminopyridine (20 mg, 0.16 mmol). The mixture was stirred for 72h. at reflux, and extracted with CH₂Cl₂. After concentration to dryness of the solution, the residue was chromatographed on silicagel (AcOEt/Heptane then AcOEt/CH₃OH). The cyclic phosphonate derivative 23 was first eluded (AcOEt/Heptane) and isolated as solid (6% yield). Then the mixture of phosphonate compounds 22 and 24 was recovered using (AcOEt/CH₃OH) as eluent. The solution was evaporated and the residue was purified by preparative thin-layer chromatography on RP8 (CH₃OH/H₂O). The 1-phosphonate analogue 22 and its 5-isomer 24 was isolated (38% and 6% yield respectively). Compound 22: ¹H NMR (250 MHz, CD₃OD) δ ppm: 4.78 (2d, 2H, CH₂Ph, Jgem= 11), 4.54 (m, 1H, H-1), 4.42 (sl, 1H, H-2), 4.10 (t, 1H, H-3, J₃₋₂=J₃₋₄=6), 3.56 (m, 1H, H-5), 3.44 (dd, 1H, H-4, $J_{4-5}=10, J_{4-3}=6), 2.10 (m, 1H, H-6e), 2.02 (q, 1H, H-6a, J_{6a-6e}=J_{6a-1}=J_{6a-5}=12), 0.90 (t, 3H, CH_3, J = 6.5);$ ¹³C NMR (63 MHz, CD₃OD) δ ppm: 111.5 (OCO), 86.6 (C-4), 81.0 (C-3), 77.8 (C-2), 74.5 (CH₂Ph), 69.8 (C-5), 69.4 (C-1), 35.7 (C-6), 14.2 (CH₃); ³¹P NMR (81 MHz, CDCl₃) δ ppm: +28.33; (Found: C, 56.11; H, 8.20; P, 6.06; C₂₃H₃₅O₇P.2H₂O requires C, 56.31; H, 8.01; P, 6.31 %). Compound 23: m.p. 135-138 °C; ¹H NMR (250 MHz, CDCl₃) δ ppm: 4.70 (s, 2H, CH₂Ph), 4.55 (m, 2H, H-1, H-5), 4.28 (m, 1H, H-3), 4.11 (sl, 2H, H-2, H-4), 3.18 (m, 1H, H-6e), 1.91 (m, 1H, H-6a), 0.88 (t, 3H, CH₃, J = 6.5); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 110.2 (OCO), 76.5 (C-4), 74.7, 73.8, 73.1 (C-2, C-3, C-5), 70.5 (CH₂Ph), 68.1 (C-1), 34.4 (C-6), 13.6 (CH₃); ³¹P NMR(81 MHz, CDCl₃) δ ppm: +27.85; (Found: C, 63.46; H, 7.55; P, 6.84; C₂₀H₂₇O₇P requires C, 63.28; H, 7.62; P, 7.10 %). Compound **24**: ¹H NMR (250 MHz, CD₃OD) δ ppm: 4.76 (dd, 2H, CH₂Ph, Jgem= 11), 4.28(sl, 1H, H-2), 4.07 (t, 1H, H-3, J₃₋₂=J₃₋₄=6), 3.95 (m, 2H, H-1, H-5), 3.51 (dd, 1H, H-4, J₄₋₃=7, J₄₋₅=10), 2.25 (m, 1H, H-6e, J_{6e-6a}=12), 2.04 (m, 3H, H-6a, CH₂P), 0.81 (t, 3H, CH₃, J =6.5); ¹³C NMR (63 MHz, CD₃OD) δ ppm: 111.3 (OCO), 85.3 (C-4), 80.8 (C-3), 78.2 (C-2), 74.9 (CH₂Ph), 73.2 (C-5), 66.5 (C-1), 36.4 (C-6), 14.1 (CH₃); ³¹P NMR (81 MHz, CD₃OD) δ ppm: +29.70.

6-Deoxy-D-myo-inositol-1-(n-butyl) phosphonate (25).

To a solution of compound **22** (46 mg, 0.1mmol) in EtOH (10 ml) was added 10% Pd/C (100 mg) and the mixture was stirred for 2h under hydrogen pressure (75psi). After filtration of the solid on celite (EtOH) and evaporation to dryness, the residue was dissolved in MeOH (10ml) and conc. HCl (1 ml) was then introduced. The solution was stirred for further 3h at r.t., neutralized with NaHCO₃ and concentrated to dryness. The residue was chromatographed on RP8 (MeOH/H₂O) to give the tetrol **25** (90% yield) isolated as sodium salt. M.S. (C.I.; isobutanol; m/z): 307 [MH]⁺, ¹H NMR (250 MHz, D₂O) δ ppm: 4.0 (m, 2H, H-2, H-5), 3.78 (m, 1H, H-1, J_{1-6a}=12), 3.60 (t, 1H, H-4, J₄₋₃=J₄₋₅=10), 3.44 (dd, 1H, H-3, J₃₋₂=3, J₃₋₄=10), 2.13 (m, 1H, H-6e, J_{6e-6a}=12), 1.88 (q, 1H, H-6a, J_{6a-1}=J_{6a-5}=J_{6e-6a}=12), 0.86 (t, 3H, CH₃, J =6.5); ¹³C NMR (50 MHz, C₅D₅N) δ ppm: 73.6 (C-2, C-4, C-5), 73.2 (C-3), 67.3 (C-1), 35.4 (C-6), 13.4 (CH₃); ³¹P NMR (81 MHz, D₂O) δ ppm: +28.58; (Found: C, 35.66; H, 6.58; P, 8.90; C₁₀H₂₀O₇PNa.3/2H₂O requires C, 36.03; H, 6.95; P, 9.30 %).

6-Deoxy-D-myo-inositol-1,5-cyclic (n-butyl)phosphonate (26).

Using the same experimental procedure used for the preparation of **25**, the 1,5-cyclic phosphonate **26** was isolated as solid from **23** (86% yield). m.p. 128-132 °C; ¹H NMR (400 MHz, CD₃OD) δ ppm: 4.56 (sl, 1H, H-4), 4.41 (sl, 1H, H-3), 4.15 (sl, 1H, H-2), 4.01 (sl, 1H, H-5), 3.73 (sl, 1H, H-1), 2.90 (m, 1H, H-6e), 2.28 (m, 1H, H-6a), 0.92 (t, 3H, CH₃, J =6.5); ¹³C NMR (63 MHz, CD₃OD) δ ppm: 77.4 (C-4), 74.6 (C-3), 73.1 (C-2), 72.1 (C-5), 69.2 (C-1), 34.1 (C-6), 13.8 (CH₃); ³¹P NMR (81 MHz, CD₃OD) δ ppm: +35.44; (Found: C, 43.96; H, 7.41; C₁₀H₁₉O₆P.1/2H₂O requires C, 43.64; H, 7.33 %).

2-D-(2,3,5/4)-4-O-Benzyl-2,3-O-cyclohexylidene-2,3,4,5-cyclohex-1,6-enetetrol-1-Cmethylene(diethyl) phosphonate (27).

To a solution of tetraethyl methylene diphosphonate (1.73 ml, 11.6 mmol) dissolved in dry THF (20 ml) was added NaH (334 mg, 13.92 mmol). The solution was stirred 15 mn. at r.t. before addition of the cyclohexanone 1 (1.9 g, 5.8 mmol) previously dissolved in dry THF (20 ml). The reaction was controlled by TLC and quenched by addition of ice containing ammonium chloride. After neutralization with aq. acetic acid and extraction with AcOEt, the organic layer was concentrated under reduced pressure. The residue was chromatographed on florisil (AcOEt/Hexane then AcOEt) to give the 1-C-phosphonate analogue 27 (60% yield). $[\alpha]_D + 34^{\circ}$ (c = 1.13, CHCl₃); ¹H NMR (200 MHz, CDCl₃) δ ppm: 5.76 (d, 1H, H-6, J₅₋₆=4), 4.77 (d, 1H, H-2, J₂₋₃=7); 4.53 (t, 1H, H-3, J₃₋₂=J₃₋₄=7); 4.0 (dd, 1H, H-5, J₅₋₄=7, J₆₋₅=4), 3.57 (t, 1H, H-4, J₄₋₃=J₄₋₅=8), 2.20 and 2.56 (d, 2H, H-7, H-7', J_{7-7'}=16); (Found: C, 61.69; H, 7.80; P, 6.61; C₂₄H₃₅O₇P requires C, 61.79; H, 7.56; P, 6.64 %).

(Z / E), 4-O-Benzyl-2,3-O-cyclohexylidene-1,6-dideoxy-D-myo-inositol-1-vinyl(diethyl) phosphonate (28).

To a solution of tetraethyl methylene diphosphonate (1 ml, 4 mmol) dissolved in dry THF (10 ml), one crystal of bipyridyle and *n*-BuLi 1.4 N (2.9 ml) was added dropwise at 0°C until the appearance of a red color. The solution was stirred 30 mn. before addition of the cyclohexanone 1 (664 mg, 2 mmol) previously dissolved in dry THF (10 ml). The reaction was followed by TLC and quenched by addition of ice containing ammonium chloride. After neutralization with aq. acetic acid and extraction with AcOEt, the organic layer was concentrated under reduced pressure. The residue was chromatographed on florisil (AcOEt/Hexane) to give the phosphonate

analogue **28** (70% yield, **E/Z** ratio = 3.5/1.5). **28E**: ¹H NMR (200 MHz, CDCl₃) δ ppm: 5.98 (s, 1H, H-7), 4.53 (d, 1H, H-2, J₂₋₃=4), 4.26 (dd, 1H, H-3, J₃₋₂=4, J₃₋₄=6), 3.91 (m, 1H, H-5), 3.60 (dd, 1H, H-4, J₄₋₃=6, J₄₋₅=7), 2.62 and 3.09 (m, 2H, H-6a, H-6e); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 152.6 (C-1), 123.3 (C-7), 85.1 (C-3), 80.6 (C-4), 72.5 (C-2), 69.9(C-5), 39.3 (C-6); (Found: C, 61.69; H, 7.80; P, 6.61; C₂₄H₃₅O₇P requires C, 61.79; H, 7.56; P, 6.64 %). **28Z**: ¹H NMR (200 MHz, CDCl₃) δ ppm: 5.82 (s, 1H, H-7), 5.54 (d, 1H, H-2, J₂₋₃=5), 4.15 (dd, 1H, H-3, J₃₋₂=5, J₃₋₄=6), 3.46 (m, 2H, H-4, H-5), 2.20 and 2.56 (m, 2H, H-6a, H-6e); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 156.5 (C-1), 117.7 (C-7), 81.0 (C-3), 79.1 (C-4), 76.6 (C-2), 74.7 (C-5), 34.5 (C-6); (Found: C, 61.92; H, 7.35; P, 6.80; C₂₄H₃₅O₇P requires C, 61.79; H, 7.56; P, 6.64 %).

2,3-O-Cyclohexylidene-1,6-dideoxy-D-myo-inositol-1-C-methylene(diethyl) phosphonate (29) and 2,3-O-Cyclohexylidene-1,6-dideoxy-D-myo-inositol-1-C-methylene(diethyl)phosphonate-4,5-bis(dibenzyl) phosphate (30).

To a solution containing a mixture of derivative **28** (**Z**/E) (500 mg, 1.07 mmol) in AcOEt/EtOH (20 ml, 1/1 v/v) was added 5% Pd/C (500 mg) and the mixture was stirred for 2h under hydrogen pressure (75 psi). After filtration of the solid on Whatman paper (EtOH) and concentration to dryness of the filtrat, the crude product of the reaction **29** was submitted to the phosphorylation procedure using the phosphoramidite method to give the inositol phosphonate bis(dibenzyl)phosphate **30** (70% overall yield). $[\alpha]_D + 3.75^\circ$ (c = 0.8, CHCl₃); ¹H NMR (200 MHz, CDCl₃) δ ppm: 4.5 (dd, 1H, H-4, J₄₋₃=8, J₄₋₅=9), 4.23 (m, 2H, H-2, H-5), 4.00 (dd, 1H, H-3, J₃₋₂=5, J₃₋₄=8), 2.06 (m, 2H, H-7, H-7'), 1.7 and 2.23 (m, 2H, H-6a, H-6e), 1.7 (m, 1H, H-1); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 82.3 (C-4), 78.5 (C-3), 75.7 (C-2), 75.4 (C-5), 32.0 (C-6), 29.4 (C-7), 29.4 (C-1); (Found: C, 60.38; H, 6.50; P, 10.53; C₄₅H₅₇O₁₃P₃ requires C, 60.12; H, 6.39; P, 10.33 %).

1,6-dideoxy-D-myo-inositol-1-C-methylene(diethyl)phosphonate-4,5-bisphosphate (31).

To a solution containing the intermediate **30** (200 mg, 0.22 mmol) in EtOH (2 ml, 1/1 v/v) was added 10% Pd/C (200 mg) and the mixture was stirred for 1h under hydrogen pressure (75 psi). After filtration of the solid on Whatman paper (H₂O), the filtrate was concentrated and TRIS salt (27 mg) was added in two instalments. The inositol analogue **31** was isolated by lyophilization as tetra-TRIS salt. ¹H NMR (400 MHz, D₂O): δ ppm: 4.23 (m, 1H, H-4), 4.15-4.05 (m, 5H, H-5, CH₂), 3.74 (sl, 1H, H-2), 3.62 (m, 1H, H-3), 2.14 to 1.97 (m, 3H, H-7'; H-6e; H-1), 1.9 (m, 1H, H-7), 1.67 (m, 1H, H-6a), 1.28 (m, 6H, 2CH₃); 1³C NMR 50 (MHz, D₂O) δ ppm: 81.0 (C-4), 77.0 (C-5), 74.8 (C-2),73.2 (C-3), 64.7,64.5 (CH₂), 32.9 (C-6), 32.1 (C-1), 27.3 (C-7),16.9, 16.8 (CH₃).

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