ISOFLAVONES FROM XANTHOCERCIS ZAMBESIACA

STANLEY H. HARPER, DIANNE B. SHIRLEY and DERYCK A. TAYLOR* Department of Chemistry, University of Rhodesia, Box MP 167, Salisbury, Rhodesia

(Received 3 November 1975)

Key Word Index—Xanthocercis zambesiaca; Leguminosae; isoflavones; 7,3'-dihydroxy-8,4'-dimethoxyisoflavone.

Abstract—In addition to the previously described 7-hydroxy-8,4'-dimethoxy- and 7,3'-dihydroxy-8,4'-dimethoxyisoflavone the heartwood of *Xanthocercis zambesiaca* has been shown to contain 7-hydroxy-8,3',4'-trimethoxy-, 3',4'-dimethoxy-6,7-methylenedioxy- and 8,3',4'-trimethoxy-6,7-methylenedioxy-isoflavone. A technique of determining isoflavone hydroxylation patterns by deuterium labelling is described.

Xanthocercis zambesiaca (Bak.) Dumaz-le-Grand, also known as *Pseudocadia zambesiaca* (Bak.) Harms [1,2] belongs to the tribe Sophoreae of the subfamily Lotoideae [3] of the Leguminosae. It occurs only in south tropical Africa as an evergreen tree [2,4], and is one of the two known species of Xanthocercis, the other, X. madagascariensis, occurring in Malagasy [2]. On exposure to the atmosphere, the pale grey-brown heartwood darkens markedly. The timber has been known to cause severe nose and throat irritation [4,5].

Milled heartwood on extraction with hexane gave 0.011% of 7-hydroxy-8,4'-dimethoxyisoflavone (8-O-methylretusin)[6] (1a), which together with its methylated and acetylated derivatives was identical (mmp, IR) with authentic samples. Mild base hydrolysis of 1a gave the deoxybenzoin 3a, which on alkali fusion gave 2-O-methylpyrogallol and 4-methoxyphenylacetic acid, identified by PC. The structure of 1a was confirmed by a novel application of deuteration and MS (see below).

After extraction with Et₂O followed by EtOAc, which each yielded 0.3% material not further investigated, 6.8% as a red gum was obtained by MeOH extraction. After extensive PLC, five isoflavones including **1a** were isolated from this. One of these, 7,3'-dihydroxy-8,4'-dimethoxyisoflavone (3'-hydroxyretusin 8-methyl ether) [7] (**1b**) was independently and concurrently isolated, together with **1a**, from another source by Thomson [7].

The least polar isoflavone fraction was a ca 1:1 mixture of two compounds (0.07%) of dry weight extracted) as a powder. Fractional crystallisation gave a compound $C_{18}H_{14}O_6$, mp 258-9°, the M⁺ of which was 30 mu lower than the highest peak in the spectrum of the mixture. UV and IR were typical of an isoflavone with no free hydroxyl. The intensity of Band 1 in the UV suggested 6,7-dioxygenation [8-11]. Mild base hydrolysis of the mixture gave two separable deoxybenzoins. The NMR of the one corresponding to the isoflavonc obtained pure showed a methylenedioxy and two methoxyl groups, para coupled (J = 1 Hz) protons at δ 7.05 and 6.35, suggesting a 2,4,5-trisubstituted phenacyl ring. A three-proton multiplet at δ 6.71 is consistent with a 3,4-disubstituted phenyl ring. The methylenedioxy group is assigned to the phenacyl ring, since cleavage α to the carbonyl group gives two appropriate fragments at m/e165 and 151.

The absence of any $M^+ - 31$ peak from the spectrum of the isoflavone suggested [12] that neither of the methoxyl groups is at C-2'. From biogenetic considerations [13,14] one methoxyl is likely to be at C-4', thus locating the other at C-3'. A comparison (mp, IR, NMR, UV) with 6,7-dimethoxy-3',4'-methylenedioxyisoflavone [12] showed the two compounds to be different. Accordingly, the isoflavone is 3',4'-dimethoxy-6,7-methylenedioxyisoflavone (2a) and its deoxybenzoin 4a.

The second deoxybenzoin contained an additional methoxyl group and lacked the NMR signal for H-3, relative to 4a, but the spectra were otherwise similar. Accordingly, the dioxybenzoin is assigned the structure 4b, confirmed by fragments at m/e 195, and as before, 151, and the parent isoflavone, $C_{19}H_{16}O_7$ is thus 8,3',4'-trimethoxy-6,7-methylenedioxyisoflavone (2b). An alternative structure, 6,3',4'-trimethoxy-7,8-methylenedioxyisoflavone cannot be excluded rigorously, but the



co-occurrence of three other 8-methoxylated isoflavones suggests the former structure is more likely.

An isoflavone fraction of intermediate polarity was a mixture of two compounds (0.40%) of dry weight extracted), as a powder. Fractional crystallisation gave **1a**, and the mother liquors provided a compound $C_{18}H_{16}O_6$, mp 185–7°, with the spectral characteristics of an isoflavone. The NMR showed the presence of three methoxyl groups and appropriate signals for H-2 and *ortho*-coupled H-5 at δ 7.88 (*s*) and 7.87 (*d*, J = 9 Hz) respectively. The IR absorption at 3140 cm⁻¹ suggested a monohydroxytrimethoxyisoflavone, unsubstituted at C-5 and C-6. Acetylation and methylation gave a monoacetate and tetramethyl ether respectively.

A bathochromic shift was observed in the UV of the isoflavone on addition of sodium acetate, locating the hydroxyl at C-7 [8a,15]. Thus ring B could be di- or tri-methoxylated, depending on whether ring A bore a methoxyl at C-8. Isoflavones trioxygenated in the A ring absorb in the region 265-70 nm [8a]. The absorption at 256 nm in the present case compares with that of 1a (257 nm) and favours a dioxygenated A ring. The presence of a peak at m/e 162 for the retro-Diels-Alder fragment [16] confirms the presence of a B ring with two methoxyl substituents. A methoxyl group can be excluded from C-2' on the same grounds argued in the case of 2a—lack of an M⁺ – 31 peak. Thus the structure 7hydroxy-8,3',4'-trimethoxyisoflavone (1c) can be assigned to the compound. As expected, methylation gave a compound identical (mp, IR) with synthetic 7,8.3',4'-tetramethoxyisoflavone (see below).

This assignment was confirmed by an unusual method. The crude mixture of **1a** and **1c** was heated under basic conditions with D_2O . affording after acidification deoxybenzoins **3a** and **3c** in which the methylene and unsubstituted ring positions *ortho* and *para* to hydroxyl are deuterium-enriched. MS of the mixture showed M⁺ ions at m/e 291 and 321 (the latter accurately determined as $C_{17}H_{15}D_3O_6$), confirming that both deoxybenzoins were trideuterated. A 7-methoxy-8-hydroxyisoflavone would be expected to provide a tetradeuterated deoxybenzoin.

The most polar isoflavone (0.32%) of dry weight extracted), $C_{17}H_{14}O_6$, mp 212--3°, was obtained pure by PLC of the crude methanol extract, and showed spectral characteristics similar to those of **1c**. NMR indicated two methoxyl groups and five aromatic protons. one of which, H-5, [8b] occurred as an *ortho*-coupled doublet (J = 9 Hz) at δ 7.67, showing that C-6 was unsubstituted. Absorptions for the remaining aromatic protons suggested a 3'.4'-, 2'.5'- or 2'.4'-oxygenation pattern. Acetylation gave a diacetate.

A hydroxyl group at C-7 was inferred from the UV shifts with sodium acetate (cf. 1c). Lack of a shift with NaOAc/B(OH)₃. AlCl₃ or AlCl₃/HCl[8a] indicated that the second hydroxyl group was probably at C-8. Comparison with the spectra of 1a and 1c suggested the compound was a 7-hydroxy-8-methoxyisoflavone, leaving one hydroxyl and one methoxyl group to be placed in ring B. The absence of an $M^+ - 31$ peak excluded a 2'-methoxyl, so that the remaining possibilities for the oxygenation pattern of the B ring were 2'-hydroxy-4'-methoxy, 2'-hydroxy-5'-methoxy, 3'-hydroxy-4'-methoxy or 4'-hydroxy-3'-methoxy. Of these, the first would correspond to an isoflavone previously synthesised [17] and can be excluded on the basis of a wide mp discrepancy.

Alkali fusion of the deoxybenzoin derived from the isoflavone gave 2-O-methylpyrogallol as expected, but the phenylacetic acid fragment could not be positively identified owing to its close similarity on TLC to 2-hydroxy-5-methoxy-, 3-hydroxy-4-methoxy- and 4-hydroxy-3-methoxy-phenylacetic acids. Partial methylation of the isoflavone gave a trimethyl ether the spectrum of which no longer shifted with NaOAc. implying that the 7-hydroxyl had been methylated and the B-ring hydroxyl was unreactive, possibly because of deactivation through H-bonding to the carbonyl, which would support placement at C-2'. However, it has been suggested [18] that the order of reactivity to methylation of the hydroxyl groups in the flavonol **5a** is 3' < 4' < 7. The monomethyl derivative 5b was isolated using conditions identical to those employed for the partial methylation of the present isoflavone, If the 7-hydroxyl is most reactive even in such a case, where the B ring is activated by conjugation to the carbonyl, there is reason to suppose the same might be true for an isoflavone where the B ring is not so activated. It was also claimed [18] that this order of methylation is reversed using Me₂SO₄-Na₂CO₃, but it is not apparent why this should be so. A similar situation arose on acetylation of the isoflavone: a monoacetate with a free 7-hydroxyl resulted, so that arguments related to hydroxyl activity are untenable for structural assignment.

Permethylation of the isoflavone gave a tetramethyl ether whose mp was identical to that reported for synthetic 7,8,3',4'-tetramethoxyisoflavone [19]. This compound was synthesised from trimethylpyrogallol and homoveratric acid. Selective monodemethylation of the deoxybenzoin obtained from their condensation was effected with AlCl₃ in refluxing ether to provide **3d**. This was converted to the isoflavone by the usual formylation with HC(OEt)₃/base followed by acid-catalysed dehydration.

The B ring of the natural isoflavone is thus 3'.4'-dioxygenated. Our observation that two of the B ring protons in the NMR of the diacetate are shifted more than the third, and to approximately equal extents [7,8b] suggested the presence of a 3'-hydroxyl. This was finally established using the deuteration procedure (see above) which gave a pentadeuterated deoxybenzoin, $C_{16}H_{11}D_5O_6$, showing only two aromatic one-proton singlets (H-5' and H-6, cf. 3b) in the NMR. Had the isoflavone B ring pattern been 2'-hydroxy-5'-methoxy or 4'-hydroxy-3'-methoxy, a tetradeuterated deoxybenzoin would be expected. Accordingly, the structure of the isoflavone is 7,3'-dihydroxy-8,4'-dimethoxyisoflavone (1b). The evidence for this structure extends that adduced by Thomson [7].

EXPERIMENTAL

Spectral data, except where otherwise indicated, are measured and presented as in Table 1. NMR spectra are run in CDCl₃ except where otherwise specified. Solvent petrol is a 50 75° boiling fraction. All PLC refers to SiO₂ plates.

Extraction. Dried, finely-milled X. *zambesiaca* heartwood (2.2 kg) was extracted (Soxhlet) successively with hexane, Et_2O , EtOAc and MeOH. On evaporation the hexane fraction gave 18.6 g oil from which a solid (300 mg) crystallised. The methanol fraction gave 150 g gum.

Purification. The crystals from the hexane extract were purified by PLC (CHCl₃-6% MeOH) yielding **1a** (244 mg). The MeOH extract (18.6 g) on PLC (CHCl₃-8% MeOH) gave 7

bands quenching 254 nm radiation. Of these, sequentially numbered, bands 1, 3 and 5, R_f 's 0.98, 0.74 and 0.56, contained the most significant amounts of solid material on workup (2.86, 1.95 and 1.96 g respectively). Further PLC gave a mixture of **2a/2b** (200 mg, R_f 0.43, CHCl₃) from band 1, **1a/1c** (1.09 g, R_f 0.43, CHCl₃–3% MeOH) from band 2, and **1b** (880 mg, R_f 0.13, CHCl₃–3% MeOH) from band 3, as powders.

General procedures. Acetylation was carried with Ac2O-pyridine for 18 hr at 15°. Methylation was with $Me_2SO_4-K_2CO_3$ NMe₂CO for 4 hr. Mild base hydrolysis was achieved by refluxing isoflavones for 30 min in 50% aq. EtOH containing 10% NaOH (0.5-1 ml). For alkali fusion, the deoxybenzoins obtained as above were heated for 1.5 min with KOH (2 molten pellets), the reaction air-cooled, acidified (6 N H₂SO₄), extracted (Et₂O) and separated into acidic and phenolic fractions [20]. These fractions were examined by PC using the organic phases of C_6H_6 -EtCO₂H-H₂O (2/2/1) Am OH-xylene-H₂O-AcOH (4/6/5/4) and BuⁿOH-AcOH-H₂O (4/1/5). The papers were developed with diazotised sulphanilic acid [21]. For deuterium exchange, the isoflavone (50 mg), KOBu^t (0.75 eq per active OH) and D₂O (0.4 ml) were heated in a sealed tube at 100° for 4 days. The mixture was cooled, acidified, extracted (Et₂O) and the deuterated deoxybenzoin purified by PLC (CHCl₃-6% MeOH).

3'.4'-Dimethox y-6,7-methylenediox yisoflavone (2a) and 8,3',4'-trimethoxy-6,7-methylenedioxyisoflavone (2b). Fractional crystallisation of the PLC fraction containing both compounds gave microcrystalline 2a (3 mg) mp 258-9° from CHCl₃-petrol (Found; M⁺ 326.0792. C₁₈H₁₄O₆ requires 326.0790), m/e 326, 311, 283, 240, 163, 147, 119 (100, 17, 7, 7, 13, 7, 7%), NMR, IR, UV-see Table 1. Mild base hydrolysis of the crude mixture (100 mg) gave 2-hydroxy-4,5-methylenedioxyphenyl-3',4'-dimethoxybenzyl ketone (4a). (13 mg), mp 148-51° from EtOAc-petrol (Found; M^+ 316.0951. $C_{17}H_{16}O_6$ requires 316.0947), λ_{max} 236, 281, 349 nm (4.19, 3.90, 3.89), ν_{max} 1620, 1600, 1530, 1515, 940 cm⁻¹, δ 3.80 (6H, s, 2 × OMe), 4.02 (2H, s, ArCH₂), 5.87 (2H, s, CH₂O₂), 6.35 (1H, d, J = 1, H-3), 6.71 (3H, m, H-2',5',6'), 7.05 (1H, d, J = 1, H-6), 12.95 (1H, bs, OH), m/e 316 (M⁺), 166, 165, 152, 151, 107 (23, 13, 100, 17, 12, 10%), R_f (in CHCl₃-3% MeOH) 0.67, and 2-hydroxy-3-methoxy-4,5-methylenedioxyphenyl-3',4'-dimethoxybenzyl ketone (**4b**). (29 mg), mp 162–3° from EtOAc-petrol (Found; C, 62.26; H, 4.91. $C_{18}H_{18}O_7$ requires C, 62.42; H, 5.24%), λ_{max} 225, 289, 350(*sh*) nm (4.39, 4.03, 3.85), ν_{max} 1615, 1595, 1525, 1510, 963 cm⁻¹, δ 3.82 (6H, *s*, MeO-4,5), 3.98 (3H, *s*, MeO-3), 4.05 (2H, *s*, ArCH₂), 5.91 (2H, *s*, CH₂O₂), 6.75 (3H, bs, H-2',5',6'), 6.88 (1H, *s*, H-6), 13.09 (1H, bs, OH), *m/e* 346 (M⁺), 196, 195, 180, 152, 151, 107, (33, 28, 100, 20, 9, 21, 8%), R_f 0.59.

7-Hydroxy-8,4'-dimethoxyisoflavone (1a). Fractional crystallisation of the PLC fraction containing la/lc gave prisms of 1a (200 mg) mp 229-32°, identical with an authentic sample [6,7]. The following data extends that already published: λ_{max} 257, 306(*sh*) nm (4.49, 3.90) (shifting to 269, 324(*sh*), 348 nm on addition of NaOMe and 267, 320(sh), 348 nm on addition of NaOAc), λ_{max} 3260, 1640, 1605, 1580, 1520 cm⁻¹, m/e 298 (M⁺), 283, 255, 166, 138, 132, 123, 117 (100, 9, 10, 9, 17, 11, 7, 5%). Monoacetate, $\lambda_{max} 256$, 310(sh) nm (4.52, 3.80), $\nu_{max} 3260$, 1640, 1605, 1580, 1520 cm⁻¹. Permethyl derivative, λ_{\max} 255, 305(*sh*) nm (4.44, 3.87), ν_{\max} 1650, 1620, 1510, 1570, 1520 cm^{-1} , m/e 312 (M⁺), 297, 180, 156, 152, 137, 132, 117, (100, 5, 5, 9, 7, 7, 8, 4%). Mild base hydrolysis of 1a (56 mg) gave 2,4-dihydroxy-3-methoxyphenyl-4'-methoxybenzyl ketone (3a) (47 mg, 86%), mp 140–2° from EtOAc/petrol, $\lambda_{max} 238(sh)$, 287, 321(sh) nm (4.01, 4.19, 3.89), v_{max} 3200, 1630, 1590, 1520 cm⁻¹, δ 3.70 (3H, s, MeO-4'), 3.85 (3H, s, MeO-3), 4.05 $(2H, s, ArCH_2), 6.39 (1H, d, J = 9, H-5), 6.74 (2H, d, J = 9, H-5), 6.74 (2H, d, J = 9)$ H-3', 5'), 7.09 (2H, d, J = 9, H-2', 6'), 7.44 (1H, d, J = 9, H-6), 12.73 (2H, hs, OH-2.4), m/e 288 (M⁺), 168, 167, 152, 121 (15, 43, 100, 62, 44%). Microfusion of 3a gave material with the same R_f 's as 2-O-methylpyrogallol [22] and 4-methoxyphenylacetic acid (see below) on PC.

7-Hydroxy-8,3',4'-trimethoxyisoflavone (1c). Evaporation of the mother liquors from the crystallisation of 1a and recrystallisation of the residue gave colourless needles of 1c. mp 185-7° from EtOAc (Found; C, 65.47, H, 4.58. $C_{18}H_{16}O_6$ requires C, 65.85; H, 4.91%), m/e 328 (M⁺), 313, 285, 166, 164, 162, 138, 123, 119 (100, 15, 5, 4, 6, 4, 12, 4, 5%), NMR, IR, UV—see Table 1. Acetate, mp 190-2° from EtOAc (Found; C, 64.55; H, 4.83. $C_{20}H_{18}O_7$ requires C, 64.86; H, 4.90%), λ_{max} 257,

Table 1. Spectral data of *X* anthocercis isoflavones IR spectra (KBr discs, cm^{-1})

Compound	OH		С=0	C=C				CH_2O_2
1b	3550,	3120	1630	1620,	1600,	1520		
1c	3140		1633	1623,	1605,	1575,	1528,	
2 a			1635	1615,	1585,	1525,	1510	945

Compound	Band 2	Band 1	Shifts		
1b	254, 289	306(sh)	269, 292(sh), 350(sh) (NaOMe)		
	(4.34), 4.05)	(3.93)	258(sh), 265, 292(sh), 348(sh) (NaOAc		
1c	256, 290(sh)	304(sh)	267, 341(sh) (NaOMe)		
	(4.48, 4.15)	(4.02)	264, 344(sh) (NaOAc)		
2a	264, 286(sh)	323, 330(sh)			
	(4.35, 4.13)	(4.05, 4.02)			

NMR Spectra 60 MHz, δ (relative to TMS). All signals singlets except where indicated. A J value signifies a doublet. 1b is run in CDCl₃-50% CD₃OD, 1c and 2a in CDCl₃

Compound	H-2	5	6	8	2'	5'	6′	CH ₂ O ₂	МеО
1b	7.97	7.67 (J = 9)	6.84 (J = 9)		6.81 (bs)	6.85 (J = 8)	6.87 (J = 8)		3.87, 3.87
1c	7.88	7.87 (J = 9)	6.94 (J = 9)		6.90 (<i>m</i>)	6.90 (m)	6.90 (m)		3.97 (8) 3.84, (3',4')
2a	7.85	7.51 (bs)	· _ /	6.78 (bs)	7.15 (m)	7.15 (m)	7.15 (m)	6.02	3.87, 3.87

285(sh) nm (4.39, 4.08), v_{max} 1750, 1650, 1610, 1590, 1530, 1505 cm⁻¹, δ 2.37 (3H, s, Ac), 3.85 (3H, s, MeO-4'), 3.86 (3H, s, MeO-3'), 3.97 (3H, s, MeO-8), 6.88 (1H, bs, H-5'), 6.91 (1H, bs, H-6'), 7.02 (1H, d, J = 9, H-6), 7.09 (1H, bs, H-2'), 7.92 (1H, d, J = 9, H-5), 7.96 (1H, s, H-2). Permethyl derivative mp 160-5° from PLC, identical with 7,8,3',4'-tetramethoxyisoflavone (see below). Deuterated deoxybenzoins. The crude mixture of 1a/1c (50 mg) was deuterated to give a mixture of inseparable deoxybenzoins, $C_{16}H_{13}D_3O_5$ (Found; M⁺ 291.1699, Requires 291.1186) and $C_{17}H_{15}D_3O_6$ (Found; M⁺ 321.1283. Requires 321.1292) corresponding to trideuterated 3a and 3c respectively. The NMR of the mixture showed an absence of ArCH₂ and H-5 signals (cf. 3a), whilst H-6 for both compounds appeared as δ 7.36 (s). The position of the other signals in the mixture was approximately the same as in 3a, except that a diminished integral for H-3' was accompanied by a corresponding increase for MeO-4', on which MeO-3' of 3c was superimposed.

7,3'-Dihydroxy-8.4'-dimethoxyisoflavone (1b). Recrystallisation of the pure PLC fraction containing 1b gave material mp 212-3° from FtOH (208-10°) [7], m/e 314 (M⁺), 299, 271, 167, 166, 148, 138, 133, 123, 105 (100, 23, 16, 13, 5, 5, 13, 10, 7, 10%), NMR, IR, UV-see Table 1. Monoacetate, (7-hydroxy-3'-acetoxy-8,4'-dimethoxyisoflavone) mp $205-6^+$ from EtOAc/petrol (Found; M⁺ 356.0972. $C_{19}H_{16}O_7$ requires 356.0896), λ_{max} 255, 305(*sh*) nm (4.54, 4.04) (shifting to 274, 354 nm on addition of NaOMe, and 273, 352 nm on addition of NaOAc). v_{max} 3350, 1735, 1630, 1595, 1520 cm⁻¹. δ (CDCl3-CD3OD 1/1) 2.30 (3H, s. Ac), 3.84 (3H, s, MeO-4'), 3.97 (3H, s, MeO-8), 6.97 (1H, d, J = 9, H-6), 7.01 (1H, d, J = 8, H-5'), 7.27 (1H, bs, H-2'), 7.37 (1H, d, J = 8, H-6'), 7.80 (1H, d, J = 9, H-5), 8.03 (1H, s, H-2). Vigorous acetylation of the monoacetate gave the diacetate, mp 173-4.5° from EtOAc-petrol (163-5°) [7]. Monomethyl derivative. (3'-hydroxy-7,8,4'-trimethoxyisoflavone), mp 149-52° from EtOH (Found: C, 64.52; H, 4.93. $C_{18}H_{16}O_6$ requires C, 64.85; H. 4.91), λ_{max} 224, 254, 290 nm (4.47, 4.53, 4.23) (shifting to 250, 298 nm on addition of NaOMe), v_{max} 3320, 1620, 1600, 1595, 1570, 1530, 1520 cm⁻¹. δ 3.87 (3H, s. MeO-4'), 3.96 (6H, s, MeO-7,8), 6.94 (1H, d, J = 5, H-5'), 6.97 (1H, d, J = 9, H-6), 7.01 (1H, d, J)J = 5, H-6'), 7.08 (111, bs, H-2'), 7.92 (1H, s, H-2), 7.96 (1H, d. J = 9, H-5), m/e 328 (M⁺), 313, 285, 181, 152, 148, 133, 105 (100, 35, 24, 6, 4, 6, 6, 5%). Permethyl derivative mp 167-9° from EtOH, identical with 7,8,3',4'-tetramethoxyisoflavone (see below). Mild base hydrolysis of 1b (102 mg) gave. 2,4dihydroxy-3-methoxyphenyl-3'-hydroxy-4'-methoxybenzyl ketone (3b) (70 mg, 71%), mp 127-9° from EtOAc (Found; C, 63.45; H, 5.14. C₁₆H₁₆O₆ requires C, 63.15; H, 5.30%), λ_{max}^{*} 231(sh), 289. 315(sh) nm (4.16, 4.21, 3.91), v_{max} 3375, 1620, 1593, 1520 cm⁻¹, δ 3.78 (3H, s, MeO-4'), 3.82 (3H, s, MeO-3), 4.03 (2H, s, ArCH₂), 6.72 (3H, bs, H-2',5'.6'), 6.38 (1H, d, J = 9, H-5), 7.39 (1H, d, J = 9, H-6), 12.80 (3H, hs, OH-2,3',4), m/e304 (M⁺), 167, 152, 149, 137, 113 (41, 100, 39, 78, 12, 29%), R_f 0.20. Microfusion of **3b** gave material with the same R_f 's as 2-O-methylpyrogallol [22] and a variety of phenylacetic acids (see text and below). Deuterated deoxybenzoin. 1b (50 mg) was deuterated to give the deoxybenzoin $C_{16}H_{11}D_5O_6$ (40 mg, 81%), (Found; M⁺ 309.1263. Requires 309.1261), differing from 3b as follows: δ 3.78 (3H, s, MeO-4'), 3.80 (3H, s, MeO-3), 6.74 (1H, s, H-5'), 7.45 (1H, s, H-6), 12.73 (3H, bs, OH-2,4,3'). m/e 310, 309, (M⁺), 308, 169, 168, 167, 153, 141, (9, 27, 12, 40, 100, 19, 32, 15%).

Arylacetic acids. Veratric acid (2.0 g) was refluxed with SOCl₂ (6 ml) for 2 hr, the excess reagent evaporated under reduced pressure, the residue dissolved in dry C_6H_6 (25 ml) and added dropwise to an excess of CH_2N_2 in Et₂O at 0°. After 20 hr at 0°, the solvent was evaporated, the residue dissolved in dry C_6H_6 (50 ml), absolute MeOH (3 ml) added and the solution irradiated with a high-pressure Hg lamp for 21 hr. Evaporation of solvent gave homoveratric acid methyl ester, purified by PLC (C_6H_6) and saponified by reflux with 10% aq. KOH (25 ml). The mixture was acidified, extracted (Et₂O), the extract washed with 10% aq NaHCO₃. the washings acidi

fied and homoveratric acid (3,4-*dimethoxyphenylacetic acid*) recovered on extraction (Et₂O) and workup, giving 1.0 g (46%) mp 88–92° (99–100) [23]. By similar Arndt-Eistert homologation utilising photo-Wolff rearrangement were prepared homovanillic acid (4-*hydroxy*-3-*methoxyphenylacetic acid*), mp 140–2° (142–3°) [24] (32% from acetyl vanillic acid), homoisovanillic acid (3-*hydroxy*-4-*methoxyphenylacetic acid*) mp 125–9° (131.5–2°) [25] (35% from acetyl isovanillic acid), and 4-*methoxyphenylacetic acid*, mp 80–4 (83–4°) [26] (46% from anisic acid).

7,8,3',4'-Tetramethoxyisoflavone. Tri-O-methylpyrogallol [27] (290 mg), homoveratric acid (340 mg, 1 eq) and polyphosphoric acid (12 g) were heated at 95° for 30 min, the mixture poured into iced water (75 ml) and extracted (Et₂O). The organic phase was dried and evaporated, and the residue purified by PLC (C₆H₆-40% EtOAc) to give 2,3,4-trimethoxyphenyl-3',4'dimethoxybenzyl ketone (240 mg, 40%) as an oil, v_{max} (film) 1673, 1593, 1520, 1500 cm⁻¹, δ 3.78 (12H, bs, 4 × OMe), 3.87 (3H, s, MeO-2), 4.12 (2H, s, ArCH₂), 6.57 (1H, d, J = 9, H-5),6.69 (3H, m, H-2'.5', 6'). 7.33 (1H, d, J = 9, H-6). m/e 345 (M⁺), 195, 180, 165 (10, 100, 11, 30%). This material (235 mg) was refluxed 1 hr with anhydrous AlCl₃ in Et₂O, the reaction poured into iced dil HCl, and extracted (CHCl₃). The organic phase was dried and evaporated and the residue recrystallised (EtOAc) to give 2-hydroxy-3,4-dimethoxyphenyl-3',4'-dimethoxybenzyl ketone (3d), mp 135-7° (134°) [19] (1150 mg, 67%). This material (150 mg) was refluxed 11 hr with HC(OEt)₃ (2 ml), Py (10 ml) and piperidine (10 drops). The reaction mixture was worked up as for 3d to give 7,8,3',4'-tetramethoxyisoflavone, mp 164-6° from EtOH (168-70°)[19] (164-5°)[7], (110 mg, 71%). The following data extends that already published: [7,19] λ_{max} 256, 287(sh), 306(sh) nm (4.44, 4.15, 3.96), ν_{max} 1638, 1618, 1598, 1568, 1520 cm⁻¹. m/e 342 (M⁻), 327, 299, 180, 171, 162, 152, 147, 137, 119 (100, 23, 7, 5, 5, 4, 8, 6, 6, 10^{°/}₂₀).

Acknowledgements—The authors are grateful to Messrs. F. L. Orpen (Forestry Commission, Salisbury), P. Guy (National Parks and Wildlife Department, Salisbury) and R. B. Drummond (Government Herbarium, Salisbury) for collection and identification of the wood, and to Dr. L. Jurd (Western Regional Research Laboratory, Berkeley, California) for samples of **1a**, its acetate and methyl ether.

REFERENCES

- 1. Drummond, R. B. (1972) Kirkia 8, 209.
- 2. Hutchinson, J. (1964) *The Genera of Flowering Plants*, Vol. I, p. 37, Oxford University Press, London.
- Harborne, J. B., Boulter, D. and Turner, L. B. (1971) Chemotaxonomy of the Leguminosae, pp. 14, 23, Academic Press, London.
- 4. Van Wyk. P. (1972) Trees of the Kruger National Park, Vol., I, p. 221-2, 204-5, Purnell, Cape Town.
- 5. Watt, J. M. and Breyer-Brandwijk. M. G. (1962) Medicinal and Poisonous Plants of Southern and Eastern Africa, 2nd. edn., p. 641, Livingston Ltd., Edinburgh.
- (a) Jurd, L., Stevens, K. and Manners, G. (1972) Phytochemistry 11, 2535; (b) idem (1972) Tetrahedron Letters 21, 2149.
- 7. Thomson, R. H. and Hayashi. T. (1974) *Phytochemistry* 13, 1943.
- (a) Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) *The Systematic Identification of Flavonoids*, p. 165, Springer-Verlag, New York; (b) *ibid*. p. 261.
- Harborne, J. B., Gottlieb, O. R. and Magalhães, M. T. (1963) J. Org. Chem. 28, 881.
- Dyke, S. F., Ollis, W. D., Sainsbury, M. Schwarz, J. S. P. (1964) *Tetrahedron* 20, 1331.
- Markham, K. R., Swift, W. T. and Mabry, T. J. (1968) J. Org. Chem. 33, 462.
- Campbell, R. V. M., Harper, S. H. and Kemp, A. D. (1969) J. Chem. Soc. (C) 1787.

- 13. Pelter, A. (1968) Tetrahedron Letters 897.
- 14. Pelter, A., Bradshaw, J. and Warren, R. F. (1971) Phytochemistry 10, 835.
- 15. Jurd, L. in *The Chemistry of Flavonoid Compounds* (1962) (Geissman, T. A., ed.), p. 147, Pergamon Press, London.
- 16. Audier, H. (1966) Bull. Soc. Chim. France 9, 2892.
- Kalra, V. K., Kukla, A. S. and Seshadri, T. R. (1967) Tetrahedron 23, 3221.
- Simpson, T. H. (1956) Sci. Proc. Roy. Dublin Soc. 27, 11 (Chem. Abstr. 51, 8081).
- 19. Kukla, A. S. and Seshadri, T. R. (1962) Tetrahedron 18, 1443.
- 20. Roux, J. (1958) J. Am. Leather Chemists' Assoc., 387.

- 21. Randerath, K. (1966) Thin Layer Chromatography 2nd ed. p. 209, Academic Press, New York.
- 22. Geissman, T. A. and Moje, W. (1951) J. Am. Chem. Soc. 73, 5765.
- 23. Kindler, K. (1941) Chem. Ber. 74, 315.
- 24. Fisher, H. E. and Hibbert, H. (1947) J. Am. Chem. Soc. 69, 1208.
- 25. Späth, E., Kromp, K. and Liebherr, F. (1941) Chem. Ber. 74, 1789.
- King, J. A. and McMillan, F. H. (1946) J. Am. Chem. Soc. 68, 2335.
- 27. Kuroda, C. (1930) J. Chem. Soc. 765.