Chem. Pharm. Bull. 29(6)1554—1560(1981)

Convenient Synthesis of 1,4-Thiazane-3-carboxylic Acid Derivatives¹⁾

KAZUO SAKAI* and NAOTO YONEDA

Research Laboratory of Applied Biochemistry, Tanabe Seiyaku Co., Ltd., 16-89 Kashima-3-chome, Yodogawa-ku, Osaka 532, Japan

(Received December 1, 1980)

A convenient method to synthesize 1,4-thiazane-3-carboxylic acid derivatives was established. Condensation of L-cysteine methyl ester (2a) with monochloroacetone (3a) followed by reduction with sodium borohydride yielded methyl (3R,5S)-5-methyl-1,4-thiazane-3-carboxylate (6a) and its (5R)-methyl isomer (7a) in a ratio of 3.1:1. The use of cysteine isopropyl ester (2c) instead of methyl ester (2a) gave the corresponding (5S)-methyl isomer (6e) more stereoselectively. The reaction of chloromethyl ethyl ketone (3b) or α -bromoacetophenone (3c) with 2a gave the corresponding 5-substituted-1,4-thiazane-3-carboxylates. Hydrolysis and oxidation of 6a yielded cycloalliin (1a).

Keywords—1,4-thiazane-3-carboxylic acid; cycloalliin; L-cysteine; α -halogeno-ketone; chondrine; cyclization; reduction

In 1956 Virtanen and Matikkala isolated²⁾ cycloalliin (1a), a sulfur-containing amino acid, in a deoxy form from onion, and confirmed³⁾ the structure to be 5-methyl-1,4-thiazane-3-carboxylic acid 1-oxide. The X-ray analysis of 1a hydrochloride hydrate proved the absolute configuration to be (1S, 3R, 5S)-5-methyl-1,4-thiazane-3-carboxylic acid 1-oxide, in which both carboxyl and methyl groups were equatorial and sulfoxide was axial.⁴⁾

Two synthetic methods for **1a** have been reported. Virtanen and Matikkala found, by a paper chromatographic study, that **1a** can be synthesized by the addition of hydrogen bromide to S-allyl-L-cysteine followed by ring closure and oxidation.³⁾ Carson and Boggs synthesized **1a** and its diastereomer by the ring closure of *cis*-S-(1-propenyl)-L-cysteine S-oxide in dilute base.⁵⁾ However, the latter method requires a long reaction time and complicated procedure

to isolate the product, resulting in a low yield, from 10% to 16%. Carson *et al.* synthesized several analogs of **1a** by using similar methods, but deoxycycloalliin and its analogs have never been synthesized directly.

Recently, the finding of a fibrinolytic activity¹¹⁾ of **1a** has led to particular interest in its analogs from a pharmacological viewpoint.

In the present paper, we describe a new, simple and more stereoselective method for the synthesis of 1,4-thiazane-3-carboxylic acid derivatives.

(3R)-1,4-Thiazane-3-carboxylic Acid Derivatives

Methyl 5-methyl-1,4-thiazane-3-carboxylates (6a and 7a) were prepared as a mixture of two diastereomers by condensation of L-cysteine methyl ester (2a) and monochloroacetone (3a) in the presence of potassium hydroxide, followed by cyclization and reduction with sodium borohydride. Its structure was determined by elemental analysis and spectral data.

The gas chromatographic analysis proved that the formation ratio of the diastereomers was 3.1:1. The main product (α -isomer) was isolated from the reaction mixture as the hydrochloride, mp $211-213^{\circ}$ (dec.), in 47% yield. From the mother liquor, the minor product (β -isomer) was isolated as the oxalate, mp $150-151^{\circ}$ (dec.), in 15% yield.

No. 6

The configurations of α - and β -isomers were established on the basis of the reaction mechanism and nuclear magnetic resonance (NMR) spectra.

The α -isomer was determined to be methyl (3R, 5S)-5-methyl-1,4-thiazane-3-carboxylate (6a), $^{12)}$ in which the methyl and the ester groups are in *cis*-configuration, while the β -isomer was presumed to be the (5R)-methyl analog (7a) having *trans*-configuration. Periodical analysis of the reaction mixture by thin-layer chromatography (TLC) suggested that prior to the cyclization the sulfide (4) was formed between 2a and 3a at the first step of the reaction, followed by the formation of a cyclic imine (5) in an acidic reaction medium. Subsequent reduction of 5 by sodium borohydride would proceed stereoselectively from the less hindered side, so that the formation of 6a (*cis*-configuration) would predominate over that of 7a (*trans*-configuration).

Carson et al. have already established the configurations and ring conformations of the two isomers by the NMR analysis of (3R)-5-methyl-1,4-thiazane-3-carboxylic acid 1,1-dioxide. In the NMR spectra of the α - and β -isomers of methyl (3R)-5-methyl-1,4-thiazane-3-carboxylate, the C-3 proton appeared as a double doublet at δ 3.68 ppm with the α -isomer whereas the C-3 proton appeared as a triplet at δ 3.89 ppm in the case of the β -isomer. This result, as compared with the result obtained by Carson et al., supported the view that the α - and β -isomers correspond to δa and δa , respectively. Further evidence of the cis-configuration of δa was provided by the finding that cycloalliin (δa) (cis-configuration) was derived exclusively from δa as described later.

In order to obtain the *cis*-isomer (6) more selectively, the effect of the ester group of 2a was examined. L-Cysteine ethyl ester (2b) or isopropyl ester (2c) was allowed to react with 3a under the same reaction condition as in the case of 2a. Though the use of the ethyl group

TABLE I. The Retention Times and Formation Ratios of Diastereomers of (3R)-5-Methyl-1,4-thiazane-3-carboxylates (6a, d, e and 7a, d, e)

Compds.	Retention time (min)	Ratio 6/7
6a	28	3.1 : 1
7a	31	
6d	29	2.9:1
7d	32	
6e	29	4.3:1
7e	32	

1556 Vol. 29 (1981)

gave nearly the same ratio of cis (6d) to trans (7d) as in the case of the methyl group, the use of the isopropyl group markedly improved the ratio (6e to 7e) to 4.3: 1, as expected, as shown in Table I. However, purification of isopropyl ester derivatives was difficult, e.g., 6e was isolated as the oxalate in 32% yield, while 7e was not isolated in a pure form.

The synthesis of methyl (3R)-5-substituted-1,4-thiazane-3-carboxylates having substituents other than a methyl group at the 5-position was then attempted. Chloromethyl ethyl ketone (3b) or α -bromoacetophenone (3c) was allowed to react with 2a to afford methyl (3R)-5-ethyl-1,4-thiazane-3-carboxylates (6b) and (7b) or the 5-phenyl analogs (6c) and (6c) and (6c) respectively. The configuration of the products (6b), (6c) and (6c) was determined by NMR analysis.

Chondrine, (3R)-1,4-thiazane-3-carboxylic acid 1-oxide, was isolated from the red alga *Chondria crassicaulis*^{13,14)} and brown alga *Undaria pinnatifida*,¹⁵⁾ and synthesized by Carson and Wong¹⁶⁾ and Dabrits and Virtanen.¹⁷⁾

In order to synthesize deoxychondrine using the same method as described above, 2a was allowed to react with chloroacetoaldehyde (8) under the same reaction conditions. However, the yield of methyl (3R)-1,4-thiazane-3-carboxylate (9) was only 12%. In this procedure, the ring closure to form 5 (R^1 =H) would be difficult and a by-product, S-(2-hydroxyethyl)-L-cysteine methyl ester (10), was formed simultaneously during sodium borohydride reduction.

Cycloalliin and Its Analogs

Derivation of the above synthesized methyl (3R)-5-substituted-1,4-thiazane-3-carboxylates into cycloalliin (1a) and its analogs was attempted.

As shown in Chart 3, hydrolysis of **6a** with sodium hydroxide gave (3R, 5S)-5-methyl-1,4thiazane-3-carboxylic acid (11a) in satisfactory yield. The physical constants of 11a agreed with those of the reference compound⁴⁾ obtained by reduction of 1a with hydriodic acid. Since Carson et al. reported¹⁸⁾ that oxidation of 11a with sodium metaperiodate or hydrogen peroxide gave 1a exclusively, we also performed the oxidation of 11a by the same method to afford 1a in good yield. On the other hand, oxidation of 6a with sodium metaperiodate in aqueous methanol gave the corresponding S-oxide, methyl (3R, 5S)-5-methyl-1,4-thiazane-3-carboxylate 1-oxide (12), in high yield. The TLC results and NMR spectrum showed that this compound (12) was a single isomer which gave 1a in good yield after hydrolysis. This finding proved that oxidation of **6a** took place stereoselectively to give only the (1S)-oxide. The structure of **1a** thus obtained from both 11a and 12 was confirmed by comparison of the physicochemical properties with those of the reference compound.⁵⁾ Similarly, hydrolysis of **6b** and **6c** gave the corresponding carboxylic acids, (3R, 5S)-5-ethyl-1,4-thiazane-3-carboxylic acid (11b) and its (5S)-phenyl analog (11c), respectively. Oxidation of 11a and 11c with sodium metaperiodate gave (1S, 3R, 5S)-5-ethyl-1,4-thiazane-3-carboxylic acid 1-oxide (1b) and its (5S)-phenyl analog (1c), respectively, in high yields. The structures of 11b and 1b were elucidated by the comparison of physical constants with those of the standard substances, vi while the structures of 11c and 1c were confirmed by the results of elemental and spectral analyses. Hydrolysis of the trans-isomer (7a) with sodium hydroxide gave (3R, 5R)-5-methyl-1,4-thiazane-3-carboxylic acid (13).5) Oxidation of 13 with sodium metaperiodate in accordance with the method of Carson et al. 18) yielded the corresponding S-oxides as a mixture of two diastereomers.

Thus, we succeeded in developing a convenient synthetic method for 5-substituted-1,4-thiazane-3-carboxylic acid derivatives by condensation of a cysteine ester with an α -halogenoketone, followed by reduction with sodium borohydride. Using this method, we were able to synthesize cycloalliin and its analogs.

Experimental

Melting points are uncorrected. IR spectra were recorded with a Shimadzu IR-27G infrared spectro-photometer. NMR spectra were obtained by the use of a Hitachi R-20A high resolution NMR spectrometer with tetramethylsilane or sodium 3-(trimethylsilyl)-propanesulfonate as an internal standard. Optical rotation was measured with a Perkin-Elmer 243 polarimeter. Gas-liquid chromatography was performed on a Shimadzu GC-4B gas chromatograph fitted with the SP-80 column (2m).

Starting Materials—Hydrochlorides of L-cysteine methyl ester (2a), ethyl ester (2b) and isopropyl ester (2c) were prepared by esterification of L-cysteine according to the method of Bergmann and Michalis¹⁹ [2a·HCl: Yield 78%, mp 140—141°, $[\alpha]_{b}^{25}$ –2.8° (c=1, MeOH). 2b·HCl: Yield 80%, mp 125—126°, $[\alpha]_{b}^{25}$ –11.8° (c=1, MeOH). 2c·HCl: Yield 71%, mp 154—155°, $[\alpha]_{b}^{25}$ –18.6° (c=1, MeOH)]. Chloromethyl ethyl ketone (3b) was prepared from propionyl chloride and diazomethane according to the method of Catch et al. ²⁰)

Typical Procedure for the Preparation of (3R)-5-Substituted-1,4-thiazane-3-carboxylic Acid Esters (6a—e and 7a—c)——L-Cysteine methyl ester (2a)·HCl (3.4 g, 20 mmol) was added portionwise to a stirred solution of KOH (85%, 2.7 g, 41 mmol) in MeOH (50 ml) at 0—5°. To the mixture, a solution of monochloroacetone (3a) (1.9 g, 20 mmol) in MeOH (10 ml) was added dropwise with stirring and the stirring was continued for 1 hr at the same temperature. Subsequently, the reaction mixture was acidified with 30% methanolic HCl (ca. 4 ml) at 0—5°. After additional stirring for 1 hr at the same temperature, sodium borohydride (1.5 g, 40 mmol) was added in small portions to the mixture while the temperature was kept below 5° by ice cooling. The mixture was stirred for 30 min, then excess sodium borohydride was destroyed by adding dilute HCl. The mixture was made alkaline by adding aqueous NaHCO₃, then extracted with CHCl₃. The organic layer was washed with H₂O, dried over MgSO₄ and concentrated in vacuo, yielding an oily residue which was dissolved in dry Et₂O (50 ml). Hydrogen chloride was bubbled into the ethereal solution to afford colorless crystals which were washed with a mixture of MeOH–Et₂O (1:10) yielding 6a·HCl (1.6 g). The mother liquor was concentrated to remove the solvent and the resulting residue was shaken with aqueous Na₂CO₃ and CHCl₃. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residual oil (a mixture

of **6a** and **7a**) was subjected to silica gel (Kieselgel 60, 0.063—0.200 mm, Merck) column chromatography using CHCl₃-MeOH (20:1) as an eluent. From the first fraction an additional amount (0.3 g) of **6a** was obtained to give a total yield of 47%. From the second fraction, **7a** (0.7 g) was obtained as a colorless syrup which crystallized as an oxalate (0.8 g, 15%).

6a·HCl: Colorless needles (from MeOH–Et₂O), mp 211—213° (dec.). Anal. Calcd for C₇H₁₃NO₂S·HCl: C, 39.71; H, 6.67; N, 6.62; S, 15.15; Cl, 16.75. Found: C, 83.78; H, 6.73; N, 6.66; S, 15.00; Cl, 16.48. [α]_b²⁵ – 13.7° (c=1, MeOH). IR $\nu_{\max}^{\text{Nuloi}}$ cm⁻¹: 2800—2400, 1730. 6a free base: NMR (CDCl₃) δ: 1.14 (3H, d, J=6.5 Hz, 5-CH₃), 2.00 (1H, s, NH), 2.35 (2H, d, J=6.0 Hz, 6-H₂), 2.65 (1H, d, J=10.0 Hz, 2-H ax.), 2.70 (1H, d, J=4.0 Hz, 2-H eq.), 2.9—3.1 (1H, m, 5-H), 3.68 (1H, d.d, J=4.0, 10.0 Hz, 3-H), 3.72 (3H, s, CO₂CH₃).

7a·Oxalate: Colorless needles (from MeOH–Et₂O), mp 150—151° (dec.). Anal. Calcd for C₇H₁₃NO₂S·C₂H₂O₄: C, 40.75; H, 5.70; N, 5.28; S, 12.09. Found: C, 40.58; H, 5.67; N, 5.29; S, 12.09. $[\alpha]_{\rm D}^{25}$ – 36.4° (c=1, MeOH). IR $\nu_{\rm max}^{\rm Nujol}$ cm⁻¹: 1745. 7a free base: NMR (CDCl₃) δ : 1.11 (3H, d, J=6.5 Hz, 5-CH₃), 2.23 (1H, s, NH), 2.38 (2H, d, J=6.5 Hz, 6-H₂), 2.92 (2H, d, J=4.0 Hz, 2-H₂), 3.05—3.45 (1H, m, 5-H), 3.77 (3H, s, CO₂CH₃), 3.89 (1H, t, J=4.0 Hz, 3-H).

In the same manner, other (3R)-1,4-thiazane-3-carboxylates (6b-e) and 7b-d) were obtained. Yields and physicochemical data of the products were as follows.

6b·HCl: Yield 35%. Colorless needles (from MeOH-Et₂O). mp 210—212° (dec.). Anal. Calcd for $C_8H_{15}NO_2S$ ·HCl: C, 42.56; H, 7.14; N, 6.21; S, 14.20; Cl, 15.71. Found: C, 42.51; H, 7.08; N, 6.16; S, 14.23; Cl, 15.39. [α]²⁵ -22.8° (c=1, MeOH). IR ν_{max}^{Nujol} cm⁻¹: 2800—2400, 1740. NMR (D₂O) δ : 1.01 (3H, t, J=7.5 Hz, CH₃CH₂), 1.4—2.1 (2H, m, CH₃CH₂), 2.5—3.5 (5H, m, 2-H₂, 5-H, 6-H₂), 3.86 (3H, s, CO₂CH₃), 4.33 (1H, d.d, J=4.0, 10.0 Hz, 3-H).

7b·Oxalate: Yield 13%. Colorless needles (from MeOH-Et₂O), mp 142—143° (dec.). Anal. Calcd for $C_8H_{15}NO_2S \cdot C_2H_2O_4$: C, 43.00; H, 6.13; N, 5.02; S, 11.48. Found: C, 42.99; H, 6.15; N, 4.98; S, 11.33. $[\alpha]_D^{25}$ -58.9° (c=1, MeOH). IR ν_{\max}^{Nulol} cm⁻¹: 1750. 7b free base: NMR (CDCl₃) δ : 0.94 (3H, t, J=6.0 Hz, C \underline{H}_3 CH₂), 1.2—1.7 (2H, m, CH₃C \underline{H}_2), 2.23 (1H, s, NH), 2.35 (1H, d, J=8.0 Hz, 6-H ax.), 2.42 (1H, d, J=4.0 Hz, 6-H eq.), 2.94 (2H, d, J=4.0 Hz, 2-H₂), 3.78 (3H, s, CO₂CH₃), 3.89 (1H, t, J=4.0 Hz, 3-H).

6c: Yield 29%. Colorless prisms (from Et₂O-*n*-hexane), mp 81—82.5°. *Anal.* Calcd for C₁₂H₁₅NO₂S: C, 60.73; H, 6.37; N, 5.90; S, 13.51. Found: C, 60.54; H, 6.32; N, 5.79; S, 13.35. $[\alpha]_D^{25}$ +27.2° (c=1, MeOH). IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3224, 1740. NMR (CDCl₃) δ : 2.32 (1H, s, NH), 2.45—3.00 (4H, m, 2-H₂, 6-H₂), 3.75 (3H, s, CO₂CH₃), 3.7—4.2 (2H, m, 3-H, 5-H), 7.35 (5H, s, arom. H).

7c: Yield 9.4%. Colorless prisms (from Et₂O-n-hexane), mp 102—103°. Anal. Calcd for C₁₂H₁₅NO₂S: C, 60.73; H, 6.37; N, 5.90; S, 13.51. Found: C, 60.81; H, 6.35; N, 5.99; S, 13.44. [α]_p = -105.2° (c=1, MeOH). IR p_{max} cm⁻¹: 3225, 1730. NMR (CDCl₃) δ : 2.31 (1H, s, NH), 2.4—3.2 (4H, m, 2-H₂, 6-H₂), 3.82 (3H, s, CO₂CH₃), 3.9—4.5 (2H, m, 3-H, 5-H), 7.34 (5H, s, arom. H).

6d·HCl: Yield 42%. Colorless needles (from MeOH–Et₂O), mp 221—223° (dec.). Anal. Calcd for $C_8H_{15}NO_2S$ ·HCl: C, 42.56; H, 7.14; N, 6.21; S, 14.20; Cl, 15.71. Found: C, 42.45; H, 7.20; N, 6.16; S, 14.15, Cl, 15.64. [α]_D²⁵ -14.7° (c=1, MeOH). IR ν _{max}^{Nujol} cm⁻¹: 2800—2400, 1750. NMR (D₂O) δ : 1.28 (3H, t, J=7.0 Hz, CH₃CH₂), 1.45 (3H, d, J=6.0 Hz, 5-CH₃), 2.81 (2H, d, J=7.0 Hz, 6-H₂), 3.07 (1H, d, J=10.5 Hz, 2-H ax.), 3.12 (1H, d, J=3.5 Hz, 2-H eq.), 3.3—3.9 (1H, m, 5-H), 4.32 (2H, q, J=7.0 Hz, CH₃CH₂), 4.32 (1H, d.d, J=3.5, 10.5 Hz, 3-H).

(1H, d.d, J=3.5, 10.5 Hz, 3-H). 7d: Yield 16%. Colorless syrup. IR $\nu_{\rm max}^{\rm flim}$ cm⁻¹: 3340, 1740. NMR (CDCl₃) δ : 1.12 (3H, d, J=7.0 Hz, 5-CH₃), 1.30 (3H, t, J=6.0 Hz, CH₃CH₂), 2.32 (1H, s, NH), 2.38 (2H, d, J=7.0 Hz, 6-H₂), 2.92 (2H, d, J=3.5 Hz, 2-H₂), 3.1—3.5 (1H, m, 5-H), 3.88 (1H, t, J=3.5 Hz, 3-H), 4.24 (2H, q, J=7.0 Hz, CH₃CH₂).

6e·Oxalate: Yield 32%. Colorless prisms (from MeOH-Et₂O), mp 166—167°. Anal. Calcd for C_9H_{17} -NO₂S·C₂H₂O₄: C, 45.04; H, 6.53; N, 4.78; S, 10.93. Found: C, 45.00; H, 6.48; N, 4.81; S, 10.79. $[\alpha]_D^{25}$ -10.0° (c=1, MeOH). IR $\nu_{\max}^{\text{Nujol}}$ cm⁻¹: 1740. NMR (D₂O) δ : 1.29 (6H, d, J=6.0 Hz, CH(CH₃)₂), 1.44 (3H, d, J=6.0 Hz, 5-CH₃), 2.80 (2H, d, J=7.5 Hz, 6-H₂), 3.02 (1H, d, J=10.0 Hz, 2-H ax.), 3.09 (1H, d, J=4.0 Hz, 2-H eq.), 3.2—3.8 (1H, m, 5-H), 4.23 (1H, d.d, J=4.0, 10.0 Hz, 3-H), 4.85—5.30 (1H, m, CH(CH₃)₂).

Methyl (3R)-1,4-Thiazane-3-carboxylate (9) and S-(2-Hydroxyethyl)-L-cysteine Methyl Ester (10) ——An aqueous solution (50%) of chloroacetaldehyde (8) $(4.7~{\rm g}, 30~{\rm mmol})$ was added dropwise to a stirred solution of $2a \cdot {\rm HCl}$ (3.4 g, 20 mmol) and KOH (85%, 2.7 g, 41 mmol) in MeOH (50 ml) at 0—5° and stirring was continued for 1 hr at the same temperature. Subsequently, the reaction mixture was acidified with 30% methanolic HCl ($ca.4~{\rm ml}$) at 0—5°. The mixture was stirred for 1 hr at the same temperature, then sodium borohydride (1.5 g, 40 mmol) was added portionwise to the reaction mixture while the temperature was kept below 5°. The reaction mixture was worked up in a manner similar to that described above and then chromatographed on silica gel with CHCl₃-MeOH (20: 1) as an eluent. From the first fraction, 9 was obtained as a colorless syrup (0.5 g) which crystallized as a hydrochloride (0.48 g, 12%). Colorless needles (from MeOH-Et₂O), mp 172—173° (dec.). Anal. Calcd for $C_6H_{11}NO_2S \cdot HCl$: C, 36.45; H, 6.12; N, 7.09; S, 16.22; Cl, 17.94. Found: C, 36.50; H, 6.12; N, 7.10; S, 16.30; Cl, 17.88. $[\alpha]_5^2 - 20.4^\circ$ (c=1, MeOH). IR $r_{\rm max}^{\rm Nuiol}$ cm⁻¹: 2800—2400, 1740. NMR (D₂O) δ : 2.7—3.8 (6H, m, 2-H₂, 5-H₂, 6-H₂), 3.89 (3H, s, CO₂CH₃), 4.42 (1H, d.d, J=5.5, 8.5 Hz, 3-H).

From the second fraction, 10 (0.54 g, 15%) was obtained as a colorless syrup. IR $v_{\text{max}}^{\text{film}}$ cm⁻¹: 1730. NMR (CDCl₃) δ : 2.56 (3H, s, OH, NH₂), 2.6—3.1 (4H, m, -CH₂-S-CH₂-), 3.5—4.1 (3H, m, HOCH₂-, -S-

 CH_2CH_1 , 3.75 (3H, s, CO_2CH_3).

Saponification of 10 with NaOH gave S-(2-hydroxyethyl)-L-cysteine, mp 189—190°, $[\alpha]_D^{25}$ -53.3° (c=1, H_2O), which was identical to the reported compound¹⁶ in terms of physicochemical properties.

General Procedure for the Preparation of (3R,5S)-5-Substituted-1,4-thiazane-3-carboxylic Acids (11a-c)—A solution of NaOH (0.4 g, 10 mmol) in H_2O (5 ml) was added to a stirred solution of a methyl (3R)-1,4-thiazane-3-carboxylate (6a-c) (5 mmol) in MeOH (20 ml) at 10° . After being stirred for 5 hr at room temperature, the reaction mixture was neutralized with 10% HCl. The resulting solution was passed through a column of cation exchanger [Dowex-50 $(H^+-\text{form})$]. The ion exchanger was washed thoroughly with H_2O , and the amino acid was eluted with 2 N ammonium hydroxide. The eluate was concentrated in vacuo to yield a crystalline residue of (3R)-1,4-thiazane-3-carboxylic acids (11a-c). Yields and pysicochemical data of the products were as follows.

(3R,5S)-5-Methyl-1,4-thiazane-3-carboxylic Acid (11a)—The crude product was converted to the hydrochloride, which was recrystallized from Me₂CO-H₂O. Yield 77%. Colorless needles, mp 244—245° (dec.) [lit.⁵) 244°]. Anal. Calcd for C₆H₁₁NO₂S·HCl: C, 36.45; H, 6.12; N, 7.09; S, 16.22; Cl, 17.94. Found: C, 36.52; H, 6.11; N, 7.13; S, 16.05; Cl, 17.94. [α]_D²⁵ - 30.0° (c=1, 2 N HCl) [lit⁵): -23.9° (c=1, 2 N HCl)]. IR ν ^{Nujol} cm⁻¹: 1750. NMR (D₂O) δ : 1.64 (3H, d, J=6.5 Hz, 5-CH₃), 2.7—4.2 (5H, m, 2-H₂, 5-H, 6-H₂), 4.45 (1H, d.d, J=6.0, 8.5 Hz, 3-H).

(3R,5S)-5-Ethyl-1,4-thiazane-3-carboxylic Acid (11b)—Yield 88%. Colorless needles (from Me₂CO-H₂O), mp 275° (dec.). Anal. Calcd for C₇H₁₃NO₂S·H₂O: C, 43.50; H, 7.83; N, 7.25; S, 16.59. Found: C, 43.43; H, 7.69; N, 7.17; S, 16.60. [α]²⁵_D -40.8° (c=1, 1 N HCl) [lit.⁷): -33.8° (c=2.5, 1 N HCl)]. IR ν ^{Nujol} cm⁻¹: 1580. NMR (D₂O) δ : 1.01 (3H, t, J=7.0 Hz, CH₃CH₂-), 1.4—2.2 (2H, m, CH₃CH₂-), 2.6—3.8 (5H, m, 2-H₂, 5-H, 6-H₂), 4.23 (1H, d.d, J=5.0, 11.0 Hz, 3-H).

(3R,5S)-5-Phenyl-1,4-thiazane-3-carboxylic Acid (11c)—Yield 87%. Colorless needles (from H_2O), mp 269—271° (dec.). Anal. Calcd for $C_{11}H_{13}NO_2S$: C, 59.17; H, 5.87; N, 6.27; S, 14.36. Found: C, 59.09; H, 5.88; N, 6.33; S, 14.30. $[\alpha]_D^{25} - 3.9^\circ$ (c = 1, 1 N HCl). IR $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3450, 1585. NMR (CF₃COOD-D₂O) δ : 2.4—3.7 (4H, m, 2-H₂, 6-H₂), 4.3—4.8 (2H, m, 3-H, 5-H), 7.50 (5H, s, arom. H).

Methyl (1S,3R,5S)-5-Methyl-1,4-thiazane-3-carboxylate 1-Oxide (12)—A solution of $6a \cdot \text{HCl}$ (5 g, 23.6 mmol) in H₂O (50 ml) was neutralized with NaHCO₃ (2 g) and then sodium metaperiodate 3 hydrate (10 g, 37.3 mmol) was added portionwise while the temperature was kept below 5°. The mixture was stirred for 2 hr, then the precipitate was filtered off and the filtrate was made alkaline with NaHCO₃ and extracted with CHCl₃. The organic layer was dried over MgSO₄ and concentrated to yield 12 as a syrup, which crystallized as a hydrochloride (3.9 g, 73%). Colorless needles (from MeOH-Et₂O), mp 187° (dec.). Anal. Calcd for C₇H₁₃NO₃S·HCl: C, 36.92; H, 6.20; N, 6.15; S, 14.08; Cl, 15.57. Found: C, 36.88; H, 6.18; N, 6.14; S, 14.12; Cl, 15.57. [α]_D +6.8° (c=1, MeOH). IR ν _{max} cm⁻¹: 1750, 1015. NMR (D₂O) δ : 1.55 (3H, d, J=7.0 Hz, 5-CH₃), 2.6—4.5 (5H, m, 2-H₂, 5-H, 6-H₂), 3.91 (3H, s, CO₂CH₃), 4.88 (1H, d.d, J=2.5, 11.5 Hz, 3-H).

(1S,3R,5S)-5-Methyl-1,4-thiazane-3-carboxylic Acid 1-Oxide: Cycloalliin (1a)——a) From 11a: A solution of 11a·HCl (3.6 g, 18 mmol) in H₂O (50 ml) was oxidized with sodium metaperiodate 3 hydrate (5.9 g, 22 mmol) according to the procedure of Carson et al.¹8) After recrystallization from EtOH-H₂O, 1a hydrochloride hydrate (3.1 g, 74%) was obtained as colorless needles, mp 204—206° (dec.). Anal. Calcd for C₆H₁₁NO₃S·HCl·H₂O: C, 31.10; H, 6.09; N, 6.05; S, 13.84; Cl, 15.30. Found: C, 31.29; H, 6.06; N, 6.01; S, 13.60; Cl, 15.40. [α]²⁵₂₅ -11.4° (c=1, 1 N HCl) [lit.⁵⁾ -11.7° (c=2.5, 1 N HCl)]. IR ν ^{Nujol}_{max} cm⁻¹: 3350, 1730, 1030. NMR (CF₃COOD) δ : 1.74 (3H, d, J=7.0 Hz, 5-CH₃), 2.8—5.0 (5H, m, 2-H₂, 5-H, 6-H₂), 5.22 (1H, d.d, J=2.5, 11.5 Hz, 3-H).

b) From 12: A solution of 12·HCl (2.3 g, 10 mmol) in H_2O (20 ml) was hydrolyzed with NaOH (1.2 g, 30 mmol) by the procedure described for the preparation of 11a-c to yield 1a hydrochloride hydrate (2.2 g, 94%), which was identical with the specimen obtained above.

(1S,3R,5S)-5-Ethyl-1,4-thiazane-3-carboxylic acid 1-oxide (1b) and the (5S)-phenyl analog (1c) were prepared from 11b and 11c, respectively. The yields and physicochemical data were as follows.

1b·HCl: Yield 82%. Colorless needles (from EtOH-H₂O), mp 223—225° [lit.⁷⁾ 230—238° (dec.)]. Anal. Calcd for C₇H₁₃NO₃S·HCl: C, 36.56; H, 6.28; N, 6.15; S, 14.08; Cl, 15.57. Found: C, 36.56; H, 6.28; N, 6.10; S, 13.95; Cl, 15.85. $[\alpha]_D^{25}$ -23.0 (c=1, 3 N HCl) [lit.⁷⁾ -22.7° (c=2.5, 3 N HCl)]. IR $\nu_{\text{max}}^{\text{Nuloi}}$ cm⁻¹: 2800—2400, 1735, 1030, 1020, 1003.

1c: Yield 83%. Colorless needles (from H₂O), mp 200°. Anal. Calcd for C₁₁H₁₃NO₃S: C, 55.21; H, 5.48; N, 5.85; S, 13.40. Found: C, 55.11; H, 5.44; N, 5.90; S, 13.29. $[\alpha]_D^{25}$ -59.5° (c=1, 2 N HCl). IR $\nu_{\max}^{\text{Nujol}}$ cm⁻¹: 1625, 1035, 1015. NMR (CF₃COOD) δ : 3.4—4.2 (4H, m, 2-H₂, 6-H₂), 5.1—5.7 (2H, m, 3-H, 5-H), 7.52 (5H, s, arom. H).

(3R,5R)-5-Methyl-1,4-thiazane-3-carboxylic Acid 1-Oxide (14)—Hydrolysis of 7a (1.0 g, 5.7 mmol) with NaOH (0.3 g, 7.5 mmol) was carried out in the same manner as for 6a to yield (3R,5R)-5-methyl-1,4-thiazane-3-carboxylic acid (13). Recrystallization from Me₂CO-H₂O gave colorless needles (0.75 g, 82%), mp 293—300° (dec.). [lit.⁵) 287—288°]. Anal. Calcd for C₆H₁₁NO₂S: C, 44.70; H, 6.88; N, 8.69; S, 19.89. Found: C, 44.90; H, 6.81; N, 8.55; S, 19.77. [α]²⁵ -84.9° (c=1, 3 n HCl) [lit.⁵) -70.0° (3 n HCl)]. IR ν ^{Nujol} cm⁻¹: 1580. NMR (CF₃COOD-D₂O) δ : 1.45 (3H, d, J=6.0 Hz, 5-CH₃), 2.7—3.0 (2H, m, 6-H₂), 3.28 (2H, d, J=4.0 Hz, 2-H₂), 3.5—4.3 (1H, m, 5-H), 4.61 (1H, t, J=4.0 Hz, 3-H).

Oxidation of 13 (1.5 g, 9.3 mmol) with sodium metaperiodate 3 hydrate (4.2 g, 16 mmol) was carried out in the manner described for 11a—c to give 14 (0.9 g, 55%), mp> 280° , as a mixture of two diastereomers at the S-oxide. Anal. Calcd for $C_6H_{11}NO_3S$: C, 40.66; H, 6.26; N, 7.90; S, 18.09. Found: C, 44.47; H, 6.26; N, 7.85; S, 17.83. IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1580, 1055, 1030.

Acknowledgement The authors thank Dr. I. Chibata and M. Miyoshi for their encouragement throughout the present study.

References and Notes

- 1) A part of this work was presented at the 29th Annual Meeting of the Kinki Branch, Pharmaceutical Society of Japan, Kyoto, November 1979.
- 2) A.I. Virtanen and E.J. Matikkala, J. Suomen Kemistilehti, B29, 134 (1956).
- 3) A.I. Virtanen and E.J. Matikkala, Acta Chem. Scand., 13, 623 (1959).
- 4) K.J. Palmer and K.S. Lee, Acta Cryst., 20, 790 (1966).
- 5) J.F. Carson and L.E. Boggs, J. Org. Chem., 31, 2862 (1966).
- 6) The ring closure of trans-S-(1-propenyl)-L-cysteine S-oxide, which was isolated from Allium cepa (onion), with dilute ammonia provided 1a in reasonable yield; J.F. Carson, R.E. Lundin, and T.M. Lukes, J. Org. Chem., 31, 1634 (1966).
- 7) J.F. Carson, R.E. Lundin, and L.E. Boggs, J. Org. Chem., 34, 1996 (1969).
- 8) J.F. Carson and R.E. Lundin, J. Chem. Soc., Perkin I, 1976, 1195.
- 9) J.F. Carson, L.E. Boggs, and R.E. Lundin, J. Org. Chem., 33, 3739 (1968).
- 10) J.F. Carson and L.E. Boggs, J. Org. Chem., 36, 611 (1971).
- 11) R.K. Agarwal, H.A. Dewer, D.J. Newell, and B. Das, Atherosclerosis, 27, 347 (1977).
- 12) The configuration at the 3-position is R because L-cysteine methyl ester was used as the starting material.
- 13) T. Takemoto, Yahugaku Kenkyu, 32, 645 (1960).
- 14) M. Kuriyama, M. Takagi, and K. Murata, Bull. Fac. Fish Hokkaido Univ., 11, 58 (1960).
- 15) F. Tominaga and K. Oka, J. Biochemistry, 54, 222 (1963).
- 16) J.F. Carson and F. Wong, J. Org. Chem., 29, 2203 (1964).
- 17) E. Dabritz and A.I. Virtanen, Acta Chem. Scand., 18, 837 (1964).
- 18) J.F. Carson, L.E. Boggs, and R.E. Lundin, J. Org. Chem., 35, 1594 (1970).
- 19) M. Bergmann and G. Michalis, Ber., 63, 987 (1930).
- 20) J.R. Catch, D.F. Elliorr, D.H. Hey, and E.R. Jones, J. Chem. Soc., 1948, 278.