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Intramolecular tandem seleno-Michael/aldol reaction: a simple route to hydroxy cyclo-1-ene-1carboxylate esters.

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Supporting Information Placeholder



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ABSTRACT: Intramolecular tandem seleno-Michael/aldol reaction followed by oxidation-elimination process can be an efficient tool for the construction of hydroxy cyclo-1-ene-1-carboxylate esters from oxo- α , β -unsaturated esters. Generation of lithium selenolate from elemental selenium and *n*-BuLi provides a simple and efficient one-pot access to cyclic *endo*-Morita-Baylis–Hillman adducts.

Introduction

The tandem or domino reaction has been of interest for organic synthesis because it offers a convenient and economical way to prepare desired organic molecules.^{1,2} The sequenced Michael/aldol tandem reaction is one of the most important processes for constructing carbon skeletons. Intramolecular Morita-Baylis-Hillman (IMBH) of α,β -unsaturated esters may lead to two types of products: a) "*exo*" when acrylate derivatives are cyclized; b) "*endo*" when the internal double bond is located in the substrate (Scheme 1).

Scheme 1. IMBH of α,β -unsaturated esters



Results

It has been previously reported that simple phosphine and tertiary amines are efficient in promoting the MBH reaction of acrylates but are completely unreactive in the case of linear $0x0-\alpha,\beta$ -unsaturated esters. For this purpose, high nucleophilicity and very low basicity of chalcogenols and metallic chalcogenolates can be employed as initial nucleophiles in the intermolecular reaction leading to Morita-Baylis-Hillman products.^{3–10}

Several examples of endo type products formed in IMBH reaction have been reported^{11–17} and the most common procedures are based on intermolecular MBH followed by ring-closing metathesis (RCM).¹⁸ This strategy fails in the presence of more than two double bonds in the molecule. Elegant work involving use of lithium benzylthiolate was presented by Ono.¹⁹ They presented total synthesis of Neplanocin A and the key intermediate was obtained via thio-Michael/aldol cyclization.

A serious drawback of this methodology is the use of alkylchalcogenols as starting material, since these compound are volatile and have a foul odour. Comasseto *et. al.* recently described a sequence of intermolecular Michael-aldol reaction activated by lithium *n*-butylchalcogenate generated *in situ.*⁶

Table 1. Catalyst screening



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4	1.0 DBU	CHCl ₃ , rt, 7 d	0
5	0.2 PBu ₃	CDCl ₃ , rt, 24 h	up to 70 ¹⁵
6	0.2 PBu ₃	CHCl ₃ , rt, up to 7 d	<5
7	0.2 PMe ₃	CHCl ₃ , rt, up to 7 d	<5
8	1.0 DABCO	THF, rt, 7 d	0
9	1.0 PBu ₃	THF, rt, 7 d	0
10	1.0 PMe ₃	THF, rt, 7 d	0
11	1.0 <i>n</i> -BuSeLi	THF, 0 °C (0.25 h) to rt (1 h)	38

Our initial attempts to induce cyclization simply by exposure of **1** to common catalysts in chloroform or THF at room temperature gave mostly no reaction (Table 1, Entries 1-10). Traces of product have been detected by crude NMR in phosphine catalyzed reactions (Table 1, Entries 6-7). Those results are opposite of previously reported yield.¹⁵ Treatment of **1** with lithium benzenethiolate gave only acyclic sulpha-Michael adduct as previously reported.¹⁷ Detailed NMR studies on the crude mixture containing lithium *n*-butylselenolates and $\infty - \alpha_{,}\beta$ -unsaturated ester, clearly suggested the formation of seleno-Michael adduct and the consumption of aldehyde. We decided to stop the reaction with aqueous ammonium chloride and treated the crude mixture with excess of H₂O₂/pyridine at 50 °C. This protocol gave desired product with 38% isolable yield. The best experimental protocol for lithium *n*-butylselenolates generation was used to investigate the experimental conditions for the chalcogenation/condensation sequence. Several reaction conditions and sequences of addition of the components were tested (Table 2).

Table 2. Optimization of reaction conditions

entry	solvent (v/v)	<i>n</i> -BuSeLi equiv	conditions	yield [%] ^c
1	THF	1.0	A	38
2	THF	1.0	А	49
3	Et ₂ O	1.0	А	0 ^a
4	THF/Et₂O (1:6)	1.0	А	40

5	THF/DMF (1:6)	1.0	А	22
6	THF/MeCN (1:6)	1.0	А	48
7	THF/1,4-dioxane (1:6)	1.0	А	43
8	THF/MeOH (1:6)	1.0	А	0^{b}
9	THF/PhMe (1:6)	1.0	А	53
10	THF/PhMe (1:6)	1.0	А	<10 ^d
11	THF/PhMe (1:6)	1.0	В	65
12	THF/PhMe (1:6)	1.0	С	64
13	THF/PhMe (1:6)	1.0	D	0^{b}
14	THF/PhMe (1:6)	1.0	D (6 h)	0^{b}
15	THF/PhMe (1:6)	1.2	В	72

Reaction conditions: A: 0 °C (0.25 h) to rt (1 h); B: - 20 °C (6 h); C: - 40 °C (6 h); D: - 78 °C (1 h); ^a Nucleophile not soluble in Et₂O. ^b No conversion based on ¹H NMR. ^c The yields refer to the isolate product purified by column chromatography. ^d Opposite sequence of addition (*n*-BuSeLi in THF has been added to aldehyde solution in anhydrous THF).

Omission of the extraction after reaction quenching increased yield to 49% (Table 2, Entry 2). An attempt to improve reaction by employing Et₂O was unsuccessful (Table 2, Entry 3). Reductive cleavage of elemental selenium under these conditions produced an insoluble mixture which when treated with substrate 1 did not provide desire product. Much better results were obtained using mixtures of solvents (Table 2, Entries 4-9). Protic solvents such as methanol stopped the reaction completely (Table 2, Entry 8). The greatest yield was observed in THF/toluene mixture at 0 °C. Alternative sequence of substrate addition gave a complex mixture and produced the desired product with a low yield (Table 2, Entry 10). Optimization of the reaction temperature improved the yield slightly to 65%. A small excess of nucleophile lead to the formation of product with 72% yield. Having identified conditions under which the desired reaction was high yielding, several other potential substrates for this reaction were prepared and tested. As presented in Table 3, our methodology provides a route to both single and fused cyclic systems. Synthesis of the six-membered carbocycles were very high yielding. Furthermore, the syntheses of five and seven-membered rings gave products with moderate (2a and 10a) to very high isolated yields.

Interestingly, formation of bridged bicyclic system (13a) gave a desired product with 68% yield and good diastereomeric ratio (5:1). The simple seven-membered ring (3a) was obtained with 27% yield only. Interestingly, compound 5 did not yield the expected product 5a. Rather, further aromatization was observed to give ethyl 2-naphthoate (5b). Surprisingly, compound 6 also did not give desired product and a complex mixture of degradation products were obtained. For detailed descriptions of substrate syntheses (1-14) please see supporting information.

Table 3. Substrate scope



^a Cyclization in the presence of 0.2 equiv PBu₃, CHCl₃, rt, 3h.

At this point we turned our attention to common IMBH substrates such as $0x0-\alpha,\beta$ -unsaturated ketones. (5*E*)-7-0x0-7-phenylhept-5-enal (14) was chosen as a model substrate. Treatment of 14 with 0.2 equiv of PBu_3 in chloroform for 3 h at room temperature gave 14a in 83% yield in comparison to 96% obtained under our conditions.

In addition, this method has been successfully applied to the synthesis of C-7 aminocyclitol with pharmacological properties. For example, *N*-octyl- β -(+)-valienamine (NOV, **15**) has been found as a potential chemical chaperone for Gaucher disease.²⁰ Fully protected β -(+)-valienamine (**19**) can be accessed from the key intermediate cyclohex-1-ene-1-carboxylate via seleno induced Michael/aldol tandem reaction (Scheme 2). Compound **18** can be obtained by the one-pot intramolecular lithium selenolateinduced Michael-aldol tandem reaction with furthered oxidation-elimination from simple oxo- α , β unsaturated ester **17**.

Scheme 2. Retrosynthetic analysis of β -(+)-valienamine



The synthesis of compound **20** was completed in 5 steps from commercially available D-xylose in 69% overall yield.²¹ Wittig reaction of the partially protected D-xylose **20** and ylide **21** gave a primary alcohol as mixture of isomers (E/Z = 1:0.2) in 65% yield. Oxidation of the remaining primary hydroxyl group gave compound **17** after flash chromatography as a mixture of isomers (E/Z = 90:10) in 90% yield. Both isomers of compound **17** have been isolated and fully characterized (see supporting information).

Scheme 3. Total synthesis of β -(+)-valienamine



With the enoate **17** in hand, the key intramolecular tandem reaction was attempted. Elemental selenium was suspended in dry THF, cooled and equimolar amount of *n*-BuLi in THF was added to generate lithium *n*-butylselenolate. Then, compound **17** was subjected to the Michael reaction with lithium *n*-butylselenolate followed by trapping of the produced enolate containing an aldehyde group to give a cyclic product as an inseparable mixture of isomers. The crude mixture of *n*-butylselenol adducts were treated with H₂O₂/pyridine system in THF at 50 °C to give mixture of cyclic Morita-Baylis-Hillman type products (**18** *anti/syn* = 65:35) in 38% total yield after 3 steps at first attempt. Short optimization gave compound **18** with 66% yield (Table 4). For further investigation we chose THF as the best solvent (Table 4, Entries 7 - 9). Treatment of **17** with excess of *n*-BuSeLi (1.2 equiv) at -20 °C gave **18** as mixture of *anti/syn* diastereomers (3:1) in 66% total. Diastereomeric ratio was determined by ¹H NMR analysis and pure isomers have been isolated by column chromatography. On the basis of above-described results, we decided to examine pure *Z*-**17** and *E*-**17** isomers in cyclization reactions under best reaction conditions. As presented in Table 4 (Entries 8 and 9), both isomers reacted to produce the same diastereomeric ratio, but *E*-isomer gave much better yield than *Z*-isomer. Simple phosphines gave better

selectivity (dr = 8:1) but unfortunately yields were very poor even after 14 or 28 days (Table 4, Entries 10 and 11).

Table 4. Reaction conditions and isolated yields of **18**.

		çO ₂ Et	CO ₂ Et	CO ₂ Et	
	O BnO ^{()''}	$\frac{1. n-BuSeLi}{2. H_2O_2, Py}$	HO BnO'' OBn OBn	HO,, BnO ^{''} OBn OBn	
		17	anti -18	syn- 18	
entry	[equiv] nuc.	solv	rent	anti/syn	yield [%]
1	1.0	THF, 0 °C (0.2	5 h) to rt (1 h)	65:35	38
2	1.0	THF/PhMe, 0 °C ((0.25 h) to rt (1 h)	67:33	39
3	1.0	THF/DMF, 0 °C (0.25 h) to rt (1 h)	nd	traces
4	1.0	THF/MeCN, 0 °C	(0.25 h) to rt (1 h)	54:46	32
5	1.0	THF/1,4-dioxane, 0 °	C (0.25 h) to rt (1 h)	66:34	42
6	1.0	THF/Et ₂ O, 0 °C (0.25 h) to rt (1 h)	67:33	34
7	1.2	THF, - 20	°C (6 h)	74:26	66
8^{a}	1.2	THF, - 20	°C (6 h)	75:25	42
9 ^b	1.2	THF, - 20	°C (6 h)	74:26	69
10 ^c	1.0 PBu3	CHCl ₃ ,	rt, 14 d	87:13	10
11 ^c	1.0 PMe3	CHCl ₃ ,	rt, 28 d	81:19	20

^a Pure Z-isomer of **17**. ^b Pure E-isomer of **17**. ^c PBu₃ or PMe₃ was used instead of *n*-BuSeLi.

With the *anti*-isomer of compound **18** in hand, the reduction of α,β -unsaturated ester reaction was attempted. As presented on Scheme 3, compound **18** was treated with 3.5 equivalents of DIBAL-H in DCM at -10 °C to give corresponding diol and protected with benzyl ethers under standard reaction conditions. Cyclic product (**22**) was treated with 3 equivalents of chlorosulfonyl isocyanate (CSI) under optimal reaction conditions (anhydrous toluene, 0 °C, 15 h) to afford the corresponding 1,2-*anti*-amino alcohol **19** in good yield (74%).²² Only one single diastereomer was isolated. Protection groups were removed using BCl₃ to provide β -(+)-valienamine (**16**).

Conclusions

In conclusion, the tandem Michael/aldol reaction with lithium selenolates and subsequent oxidationelimination of the seleno group is a highly efficient method for the construction of six-membered rings.

 The presented methodology shows some interesting advantages in comparison to the classic IMBH reaction, such as the short reaction times and the substrate scope. We have developed a fast and high yielding route to multi-functionalized IMBH adducts employing in situ generated metallic chalcogenolates. This protocol can be a valuable alternative route to obtain IMBH adducts and derivatives. In this context, the asymmetric version of this reaction is under development in our laboratory.

Experimental section

General information

All starting materials and reagents were purchased from commercial sources and used without purification. Reactions were controlled using TLC on silica [alu-plates (0.2 mm)]. Plates were visualized with UV light (254 nm) and by treatment with: aqueous cerium(IV) sulfate solution with molybdic and sulfuric acid followed by heating. All organic solutions were dried over anhydrous magnesium sulfate. Reaction products were purified by flash chromatography using silica gel 60 (240-400 mesh). Optical rotations were measured at room temperature with a digital polarimeter. CDCl₃, D₂O were used as NMR solvents. ¹H spectra were recorded with 600, 400 and 300 MHz and referenced relative to: CDCl₃ - tetramethylsilane ($\delta = 0$ ppm). Data are reported as follows: chemical shift in parts per million (ppm) , multiplicity (s = singlet, d = doublet, t = triplet, dd = doublet of doublets, ddd = doublet of doublet of doublets, dddd = doublet of doublet of doublet of doublets, m = multiplet), coupling constants (in hertz) and integration. ¹³C NMR spectra were measured at 150, 100 and 75 MHz with complete proton decoupling. Chemical shifts were reported in ppm from the residual solvent as an internal standard: CDCl₃ (δ = 77.16 ppm). High-resolution mass spectra were acquired using ESI-TOF method.

General procedure for ozonolysis/Wittig reaction (Procedure A).¹⁵ Cycloalkene (10 mmol) was dissolved in anhydrous methylene chloride (50 ml) and cooled to -78 °C whereupon ozone was bubbled in the flask until the solution became blue in color and at this point, the reaction was flushed with argon and triphenylphosphine (1 equiv. to the amount of alkene) was added in one portion. Reaction mixture was allowed to slowly warm to 0 °C and phosphorane (0.1-2 equiv., substrate dependent) was added and the reaction mixture was stirred for 12 h in room temperature. Products were purified by column chromatography in Hx/EA gradient.

Ethyl (E)-6-oxohex-2-enoate (2)¹⁵ Procedure A: 1,5-Cyclooktadiene (1.40 g, 12.94 mmol), 21 (9.00 g, 25.88 mmol) gave product as a colorless oil (1.15 g, 7.36 mmol), 28 %. ¹H NMR (600 MHz, CDCl₃) δ 9.80 (s, 1H), 6.94 (dt, *J* = 15.6, 6.7 Hz, 1H), 5.85 (dt, *J* = 15.7, 1.5 Hz, 1H), 4.19 (q, *J* = 7.1 Hz, 2H),

2.64 (t, J = 7.2 Hz, 2H), 2.57 – 2.50 (m, 2H), 1.29 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 200.3, 166.2, 146.2, 122.4, 60.3, 41.8, 24.4, 14.2.

Ethyl (E)-8-oxooct-2-enoate (3)¹⁵ Procedure A: Cyclohexene (2.0 g, 24.35 mmol), 21 (4.24 g, 12.17 mmol) gave product as a colorless oil (1.43 g, 6.53 mmol), 54 %. ¹H NMR (600 MHz, CDCl₃) δ 9.77 (t, J = 1.6 Hz, 1H), 6.94 (dt, J = 15.6, 6.9 Hz, 1H), 5.83 (dt, J = 15.6, 1.6 Hz, 1H), 4.19 (q, J = 7.1 Hz, 2H), 2.46 (td, J = 7.3, 1.6 Hz, 2H), 2.23 (ddd, J = 14.6, 7.3, 1.6 Hz, 2H), 1.71 – 1.62 (m, 2H), 1.51 (dt, J = 15.2, 7.7 Hz, 2H), 1.29 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 202.1, 166.6, 148.2, 121.8, 60.2, 43.6, 31.8, 27.4, 21.5, 14.2.

Ethyl (E)-4-(2-formylphenyl)but-2-enoate (5) Procedure A: Indene (2.00 g, 17.22 mmol), 21 (1.80 g, 5.17 mmol) gave product as a colorless oil (0.42 g, 1.90 mmol), 37%. ¹H NMR (600 MHz, CDCl₃) δ 10.16 (s, 1H), 7.85 (dd, J = 7.6, 1.4 Hz, 1H), 7.56 (td, J = 7.5, 1.5 Hz, 1H), 7.47 (td, J = 7.5, 1.1 Hz, 1H), 7.29 (d, J = 7.6 Hz, 1H), 7.14 (dt, J = 15.6, 6.4 Hz, 1H), 5.72 (dt, J = 15.6, 1.7 Hz, 1H), 4.17 (q, J = 7.1 Hz, 2H), 3.99 (dd, J = 6.4, 1.6 Hz, 2H), 1.27 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 192.5, 166.3, 146.6, 139.7, 134.0, 133.8, 133.6, 131.4, 127.5, 122.6, 60.3, 35.2, 14.2.

Ethyl (E)-3-((1R,3S)-3-formylcyclopentyl)acrylate **(13)** Procedure A: Norbornene (2.16 g, 22.94 mmol), **21** (4.00 g, 11.47 mmol) gave product as a colorless oil (1.61 g, 8.21 mmol), 72%. ¹H NMR (600 MHz, CDCl₃) δ 9.64 (d, *J* = 2.3 Hz, 1H), 6.91 (dd, *J* = 15.6, 7.7 Hz, 1H), 5.83 (dd, *J* = 15.6, 1.2 Hz, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 2.91 – 2.84 (m, 1H), 2.77 – 2.68 (m, 1H), 2.11 – 2.00 (m, 2H), 1.96 – 1.85 (m, 2H), 1.72 (ddd, *J* = 13.1, 9.9, 8.7 Hz, 1H), 1.49 – 1.42 (m, 1H), 1.29 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 202.6, 166.6, 150.8, 120.6, 60.3, 51.2, 42.9, 32.2, 31.8, 25.9, 14.2; HRMS (ESI): calcd. for C₁₁H₁₆O₃Na [M+Na]⁺ 219.0991, found 219.0991.

(*E*)-7-oxo-7-phenylhept-5-enal (14)¹⁵ Procedure A: Cyclopentene (7.16 g, 105.15 mmol), 23 (4.00 g, 10.51 mmol) gave product as colorless oil (1.00 g, 4.94 mmol), 47%. ¹H NMR (600 MHz, CDCl₃) δ 9.81 (t, *J* = 1.4 Hz, 1H), 7.94 (dt, *J* = 8.4, 1.6 Hz, 2H), 7.59 – 7.55 (m, 1H), 7.51 – 7.46 (m, 2H), 7.03 (dt, *J* = 15.4, 6.8 Hz, 1H), 6.92 (dt, *J* = 15.4, 1.4 Hz, 1H), 2.54 (td, *J* = 7.2, 1.4 Hz, 2H), 2.42 – 2.35 (m, 2H), 1.89 (p, *J* = 7.3 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 201.6, 190.5, 148.0, 137.7, 132.7, 128.5, 128.5, 126.6, 43.0, 31.8, 20.5.

General procedure for synthesis salicylic aldehyde derivatives (Procedure B).²³ Different substituent salicylic aldehyde (0.5 g), ethyl 4-bromocrotonate (1.2 equiv.) and K₂CO₃ (2 equiv.) were dissolved in DMF (15 ml) and heated to 40 °C or 70 °C under argon for 4 h. Reaction mixture was allowed to cool to room temperature and filtrated. After water addition (20 ml) the mixture was extracted with ethyl

acetate (3 x 20ml). The combined organic phases were washed with water (2 x 20 ml) and brine (30 ml) and dried over anhydrous MgSO₄. After removal of solvent under reduced pressure, the residue was purified by column chromatography on silica gel in Hx/EA gradient to give the pure products.

Ethyl (E)-4-(2-formylphenoxy)but-2-enoate (7)²³ Procedure B: 70 °C, (0.34 g, 1.43 mmol), 35%, pale yellow solid mp: 68-70 (66-68 °C lit.).²³ ¹**H NMR** (600 MHz, CDCl₃) δ 10.56 (d, J = 0.7 Hz, 1H), 7.86 (dd, J = 7.7, 1.8 Hz, 1H), 7.54 (ddd, J = 8.5, 7.3, 1.8 Hz, 1H), 7.13 – 7.05 (m, 2H), 6.94 (d, J = 8.4 Hz, 1H), 6.22 (dt, J = 15.8, 2.1 Hz, 1H), 4.83 (dd, J = 4.1, 2.1 Hz, 2H), 4.23 (q, J = 7.1 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 189.3, 165.8, 160.1, 141.1, 135.9, 128.8, 125.1, 122.5, 121.4, 112.5, 66.8, 60.7, 14.2.

Ethyl (E)-4-(2-formyl-5-methoxyphenoxy)but-2-enoate (8)²³ Procedure B: 40 °C, (0.72 g, 2.72 mmol) 83%, pale yellow solid, mp: 95-96 °C (91-93 °C lit.).²³ ¹H NMR (600 MHz, CDCl₃) δ 10.38 (d, J = 0.7 Hz, 1H), 7.84 (d, J = 8.7 Hz, 1H), 7.09 (dt, J = 15.8, 4.0 Hz, 1H), 6.59 (ddd, J = 8.7, 2.2, 0.6 Hz, 1H), 6.40 (d, J = 2.2 Hz, 1H), 6.22 (dt, J = 15.8, 2.1 Hz, 1H), 4.80 (dd, J = 4.0, 2.1 Hz, 2H), 4.23 (q, J = 7.1 Hz, 2H), 3.87 (s, 3H), 1.31 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 187.8, 166.0, 165.8, 161.8, 141.0, 130.9, 122.5, 119.2, 106.3, 99.0, 66.8, 60.7, 55.7, 14.2.

Ethyl (E)-4-(2-formyl-5-methylphenoxy)but-2-enoate (9) Procedure B: 40°C, (0.65 g, 2.63 mmol) 72%, pale yellow solid, mp: 63-65 °C. ¹H NMR (600 MHz, CDCl₃) δ 10.47 (d, *J* = 0.6 Hz, 1H), 7.75 (d, *J* = 7.9 Hz, 1H), 7.10 (dt, *J* = 15.8, 4.0 Hz, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 6.73 (s, 1H), 6.21 (dt, *J* = 15.8, 2.1 Hz, 1H), 4.81 (dd, *J* = 4.0, 2.1 Hz, 2H), 4.23 (q, *J* = 7.1 Hz, 2H), 2.39 (s, 3H), 1.31 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 188.9, 165.8, 160.2, 147.4, 141.3, 128.8, 122.9, 122.4, 122.3, 113.0, 66.7, 60.7, 22.3, 14.2. HRMS (ESI): calcd. for C₁₄H₁₆O₄Na [M+Na]⁺ 271.0941, found 271.0941.

Ethyl (E)-4-(2-formyl-4-methoxyphenoxy)but-2-enoate (10)²³ Procedure B: 70 °C, (0.61 g. 2.29 mmol) 70%, pale yellow solid, mp: 83-84 °C (83-85 °C lit.).²³ ¹H NMR (600 MHz, CDCl₃) δ 10.52 (s, 1H), 7.35 (d, *J* = 3.3 Hz, 1H), 7.13 – 7.06 (m, 2H), 6.90 (d, *J* = 9.1 Hz, 1H), 6.20 (dt, *J* = 15.8, 2.1 Hz, 1H), 4.78 (dd, *J* = 4.1, 2.1 Hz, 2H), 4.22 (q, *J* = 7.1 Hz, 2H), 3.81 (s, 3H), 1.31 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 189.0, 165.8, 155.0, 154.1, 141.4, 125.4, 123.4, 122.4, 114.4, 110.6, 67.5, 60.7, 55.8, 14.2.

Ethyl (E)-4-(2-formyl-4-methylphenoxy)but-2-enoate **(11)** Procedure B: 70 °C, (0.53 g, 2.11 mmol) 57%, pale yellow solid, mp: 64-65 °C. ¹H NMR (600 MHz, CDCl₃) δ 10.52 (s, 1H), 7.65 (d, J = 2.1 Hz, 1H), 7.34 (ddd, J = 8.5, 2.4, 0.6 Hz, 1H), 7.09 (dt, J = 15.8, 4.0 Hz, 1H), 6.84 (d, J = 8.5 Hz, 1H), 6.20 (dt, J = 15.8, 2.1 Hz, 1H), 4.80 (dd, J = 4.0, 2.1 Hz, 2H), 4.22 (q, J = 7.1 Hz, 2H), 2.32 (s, 3H), 1.30 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 189.4, 165.8, 158.3, 141.4, 136.5, 130.9, 128.8, 124.8,

122.4, 112.5, 66.9, 60.7, 20.2, 14.2. **HRMS** (ESI): calcd. for $C_{14}H_{16}O_4Na [M+Na]^+$ 271.0941, found 271.0941.

Ethyl (E)-4-(4-chloro-2-formylphenoxy)but-2-enoate $(12)^{23}$ Procedure B: 70°C, (0.57 g, 2.12 mmol), 66%, pale yellow solid, mp: 66-67 (64-66 °C lit.).²³ ¹H NMR (600 MHz, CDCl₃) δ 10.48 (s, 1H), 7.82 (d, J = 2.7 Hz, 1H), 7.49 (dd, J = 8.9, 2.8 Hz, 1H), 7.09 (dt, J = 15.8, 4.1 Hz, 1H), 6.91 (d, J = 8.9 Hz, 1H), 6.19 (dt, J = 15.8, 2.0 Hz, 1H), 4.83 (dd, J = 4.1, 2.0 Hz, 2H), 4.24 (q, J = 7.1 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 187.9, 165.6, 158.6, 140.6, 135.4, 128.3, 127.2, 125.9, 122.8, 114.2, 67.2, 60.8, 14.2.

Preparation of other substrates

Ethvl 7-oxohept-2-enoate (1)¹⁵ To solution of tetrahydro-2*H*-pyran-2-ol (7.00 g, 68.54 mmol) in anhydrous 1.4-dioxane (100 ml) 21 (29.80 g. 85.68 mmol) was added. After 12 h of continuous stirring reaction mixture was heated up to 45 °C for 2 h. After solvent evaporation crude mixture was purified by flash chromatography Hx/EA (95:5 to 3:1) to give ethyl 7-hydroxyhept-2-enoate as a mixture of diastereoisomers E/Z > 95/5. Ethyl-7-hydroxyhept-2-enoate was dissolved in anhydrous methylene chloride (60 ml) and was added to oxidative mixture prepared by addition of anhydrous dimethyl sulfoxide (19.6 ml, 275.93 mmol) to solution of oxalvl chloride (11.8 ml, 137.96 mmol) in anhydrous methylene chloride (100 ml) cooled to -78 °C. After 1 h of stirring in -78 °C triethylamine (58.2 ml, 413.89 mmol) was added and strirring was continued for next 30 min. After that time reaction mixture was allowed to slowly warm to room temperature. The reaction was quenched with addition of sat. NH₄Cl (100 ml), followed by addition of water (100 ml). The aqueous layer was extracted with methylene chloride (3 x 100 ml), and combined organic layers were washed with water (2 x 100 ml) and brine (100 ml). The organic phase was dried over anhydrous MgSO₄ and concentrated under reduced pressure (below 35 °C). Crude product was purified by flash chromatography Hx/EA (95:5) to (4:1) to give 1 as an almost pure E diastereoisomer (traces of Z-product) as colorless oil (6.23 g, 37.01 mmol), 54% after 2 steps. ¹H **NMR** (600 MHz, CDCl₃) δ 9.78 (t, J = 1.4 Hz, 1H), 6.92 (dt, J = 15.6, 6.9 Hz, 1H), 5.84 (dt, J = 15.6, 1.6 Hz, 1H), 4.19 (q, J = 7.1 Hz, 2H), 2.49 (td, J = 7.3, 1.4 Hz, 2H), 2.26 (ddd, J = 14.7, 7.4, 1.6 Hz, 2H), 1.81 (p, J = 7.3 Hz, 2H), 1.29 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 201.6, 166.4, 147.5, 122.3, 60.3, 42.9, 31.2, 20.3, 14.2.

Ethyl (E)-3-(2-formylphenyl)acrylate $(4)^{23}$ To the solution of phthaldialdehyde (1.0 g, 7.45 mmol) in anhydrous toluene **21** (2.6 g, 7.45 mmol) was added in one portion. The reaction mixture was heated to 50 °C under argon for 2 h. After solvent removal, semisolid residue was purified by column chromatography in Hx/EA gradient which results product as a clear, colorless oil (0.74 g, 3.62 mmol), 47%. ¹H

NMR (600 MHz, CDCl₃) δ 10.24 (s, 1H), 8.45 (d, J = 15.9 Hz, 1H), 7.82 (d, J = 7.1 Hz, 1H), 7.59 – 7.52 (m, 2H), 7.49 (td, J = 7.4, 1.8 Hz, 1H), 6.31 (d, J = 15.9 Hz, 1H), 4.23 (q, J = 7.1 Hz, 2H), 1.29 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 191.7, 166.2, 140.9, 136.7, 133.9, 133.8, 132.1, 129.8, 128.0, 123.3, 60.8, 14.3.

Ethyl (E)-3-(2-formylphenoxy)acrylate (6) To the solution of salicylic aldehyde (1.00 g, 8.19 mmol) in anhydrous methylene chloride (10 ml) ethyl propiolate (0.99 ml, 9.83 mmol) was added in one portion. After addition catalytic amount of *N*-methylmorpholine (0.09 ml, 0.82 mmol) reaction mixture was stirred under argon for 12 h in room temperature. After that time solvent was removed under reduced pressure and crude product was purified by column chromatography in Hx/EA gradient (5:1 to 3:1) which gave pure product as a colorless oil (1.51 g, 6.86 mmol), 84 %. ¹H NMR (600 MHz, CDCl₃) δ 10.39 (d, *J* = 0.7 Hz, 1H), 7.95 (dd, *J* = 7.7, 1.7 Hz, 1H), 7.86 (d, *J* = 12.2 Hz, 1H), 7.67 (ddd, *J* = 8.3, 7.4, 1.8 Hz, 1H), 7.34 (t, *J* = 7.6 Hz, 1H), 7.18 (d, *J* = 8.3 Hz, 1H), 5.66 (d, *J* = 12.2 Hz, 1H), 4.23 (q, *J* = 7.1 Hz, 2H), 1.32 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 188.1, 166.5, 158.0, 157.5, 136.0, 128.9, 126.5, 125.4, 118.3, 104.2, 60.4, 14.3.

General procedure for tandem seleno-Michael/aldol-oxidation/elimination reaction (Procedure C). A suspension of elemental selenium (0.50 g, 0.63 mmol) in anhydrous THF (0.8 ml) was cooled in ice bath and *n*-BuLi (1.6 M in hexanes, 0.394 ml, 0.63 mmol) was added dropwise (a clear, almost colorless, solution was produced) and the mixture was stirred 15 min in 0 °C. After that reaction mixture was cooled to -20 °C followed by addition anhydrous toluene (3 ml). Solution of substrate (0.53 mmol in 1.5 ml of anhydrous toluene) was added dropwise and stirring was continued through the next 6 hours. After that time, reaction was quenched by addition of aqueous, saturated solution of NH₄Cl (0.100 ml) following by addition of hydrogen peroxide solution (30 % v/v, 0.325 ml, 3.15 mmol) and pyridine (0.254 ml, 3.15 mmol). Reaction mixture was allowed to slowly back to room temperature and after that was heated to 50 °C for 1 hour (reaction mixture became almost colorless). The solvent was evaporated under reduced pressure and resign of pyridine was co-evaporated with toluene (2 x 15 ml). Crude products were purified by column chromatography Hx/EA gradient.

Ethyl 6-hydroxycyclohex-1-ene-1-carboxylate $(1a)^{15}$ Procedure C: colorless oil (0.065g, 0.38 mmol), 72%. ¹H NMR (600 MHz, CDCl₃) δ 7.10 (t, J = 4.0 Hz, 1H), 4.54 (t, J = 4.4 Hz, 1H), 4.23 (q, J = 7.1 Hz, 2H), 3.15 (s, 1H), 2.33 – 2.23 (m, 1H), 2.18 – 2.10 (m, 1H), 1.87 – 1.72 (m, 3H), 1.65 – 1.55 (m, 1H), 1.31 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 167.3, 142.8, 132.4, 63.5, 60.6, 29.9, 26.1, 17.5, 14.2.

Ethyl 5-hydroxycyclopent-1-ene-1-carboxylate $(2a)^{15}$ Procedure C: colorless oil (0.063 g, 0.40 mmol), 75%. ¹H NMR (600 MHz, CDCl₃) δ 6.91 (t, J = 2.5 Hz, 1H), 5.12 – 5.06 (m, 1H), 4.25 (q, J = 7.2 Hz, 2H), 2.65 (m, 1H), 2.45 – 2.38 (m, 1H), 2.38 – 2.32 (m, 1H), 1.88 (m, 1H), 1.32 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 165.0, 146.2, 138.0, 75.5, 60.4, 31.8, 30.8, 14.2.

Ethyl 7-hydroxycyclohept-1-ene-1-carboxylate $(3a)^{15}$ Procedure C: colorless oil (0.026 g, 0.14 mmol), 27%. ¹H NMR (600 MHz, CDCl₃) δ 7.16 (t, J = 6.5 Hz, 1H), 4.81 (dd, J = 7.6, 2.2 Hz, 1H), 4.21 (q, J = 7.1 Hz, 2H), 3.20 (s, 1H), 2.48 – 2.38 (m, 1H), 2.33 – 2.22 (m, 1H), 2.01 – 1.92 (m, 2H), 1.82 – 1.76 (m, 1H), 1.76 – 1.63 (m, 2H), 1.62 – 1.55 (m, 1H), 1.30 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 168.4, 145.2, 136.8, 68.6, 60.8, 32.3, 27.3, 25.7, 23.5, 14.2.

Ethyl 1-hydroxy-1H-indene-2-carboxylate (4a) Procedure C: colorless oil (0.065 g, 0.32 mmol), 60%. ¹H NMR (600 MHz, CDCl₃) δ 7.61 (d, J = 7.3 Hz, 1H), 7.56 (s, 1H), 7.41 (dd, J = 6.5, 1.0 Hz, 1H), 7.40 – 7.33 (m, 2H), 5.43 (s, 1H), 4.34 (q, J = 7.1 Hz, 2H), 3.19 (s, 1H), 1.38 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 164.8, 145.8, 141.3, 139.7, 139.3, 128.9, 128.9, 124.3, 123.8, 75.6, 60.7, 14.3.

Ethyl 2-naphthoate (5b) Procedure C: colorless oil (0.085 g, 0.42 mmol), 80%. ¹H NMR (600 MHz, CDCl₃) δ 8.63 (d, J = 0.7 Hz, 1H), 8.08 (dd, J = 8.6, 1.7 Hz, 1H), 7.97 (dd, J = 8.1, 0.6 Hz, 1H), 7.89 (d, J = 8.6 Hz, 2H), 7.60 (ddd, J = 8.2, 6.9, 1.3 Hz, 1H), 7.55 (ddd, J = 8.0, 6.9, 1.3 Hz, 1H), 4.46 (q, J = 7.1 Hz, 2H), 1.46 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.8, 135.5, 132.5, 130.9, 129.3, 128.1, 128.1, 127.7 (2C), 126.6, 125.2, 61.1, 14.4.

Ethyl 5-hydroxy-2,5-dihydrobenzo[b] oxepine-4-carboxylate (7a) Procedure C: white, amorphous solid (0.081 g, 0.34 mmol), 65%. ¹H NMR (600 MHz, CDCl₃) δ 7.22 (d, J = 7.4 Hz, 2H), 7.08 – 7.03 (m, 2H), 6.82 (dd, J = 3.8, 2.2 Hz, 1H), 5.49 (d, J = 9.3 Hz, 1H), 4.93 (dd, J = 19.2, 3.9 Hz, 1H), 4.49 (dd, J = 19.2, 2.1 Hz, 1H), 4.24 – 4.17 (m, 2H), 3.14 (d, J = 9.6 Hz, 1H), 1.26 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.2, 156.8, 140.3, 136.7, 133.9, 129.9, 128.6, 125.1, 121.7, 70.8, 69.1, 61.4, 14.2. HRMS (ESI): calcd. for C₁₃H₁₄O₄Na [M+Na]⁺ 257.0784, found 257.0784.

Ethyl 5-hydroxy-8-methoxy-2,5-dihydrobenzo[b] oxepine-4-carboxylate (8a) Procedure C: white, amorphous solid (0.118 g, 0.45 mmol), 85%. ¹H NMR (600 MHz, CDCl₃) δ 7.20 (d, *J* = 8.3 Hz, 1H), 6.90 (dd, *J* = 4.0, 2.2 Hz, 1H), 6.68 (d, *J* = 2.5 Hz, 1H), 6.66 (dd, *J* = 8.3, 2.5 Hz, 1H), 5.51 (d, *J* = 8.5 Hz, 1H), 4.99 (dd, *J* = 19.1, 4.1 Hz, 1H), 4.57 (dd, *J* = 19.1, 2.2 Hz, 1H), 4.34 – 4.22 (m, 2H), 3.80 (s, 3H), 3.00 (d, *J* = 9.0 Hz, 1H), 1.34 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.3, 160.9, 157.9,

140.0, 134.1, 129.6, 128.7, 109.9, 107.8, 70.6, 68.8, 61.4, 55.5, 14.2; **HRMS** (ESI): calcd. for $C_{14}H_{16}O_5Na [M+Na]^+ 287.0890$, found 287.0890.

Ethyl 5-hydroxy-8-methyl-2,5-dihydrobenzo[b] oxepine-4-carboxylate (9a) Procedure C: white, amorphous solid (0.093 g, 0.38 mmol), 71%. ¹H NMR (600 MHz, CDCl₃) δ 7.17 (d, J = 8.0 Hz, 1H), 6.96 – 6.92 (m, 2H), 6.89 (dd, J = 3.9, 2.2 Hz, 1H), 5.53 (d, J = 9.3 Hz, 1H), 4.99 (dd, J = 19.2, 3.9 Hz, 1H), 4.55 (dd, J = 19.2, 2.2 Hz, 1H), 4.32 – 4.25 (m, 2H), 3.11 (d, J = 9.5 Hz, 1H), 2.34 (s, 3H), 1.34 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.3, 156.7, 140.3, 134.0, 133.6, 128.5, 125.6, 122.3, 70.7, 68.9, 61.4, 21.1, 14.2. HRMS (ESI): calcd. for C₁₄H₁₆O₄Na [M+Na]⁺ 271.0941, found 271.0941.

Ethyl 5-hydroxy-7-methoxy-2,5-dihydrobenzo[b] oxepine-4-carboxylate (10a) Procedure C: white, amorphous solid (0.122 g, 0.46 mmol), 87%. ¹H NMR (600 MHz, CDCl₃) δ 7.03 (d, J = 8.7 Hz, 1H), 6.88 (dd, J = 3.7, 2.2 Hz, 1H), 6.83 (d, J = 3.0 Hz, 1H), 6.78 (dd, J = 8.7, 3.0 Hz, 1H), 5.51 (d, J = 9.3 Hz, 1H), 4.94 (dd, J = 19.3, 3.8 Hz, 1H), 4.52 (dd, J = 19.3, 2.1 Hz, 1H), 4.31 – 4.24 (m, 2H), 3.78 (s, 3H), 3.34 (d, J = 9.8 Hz, 1H), 1.33 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.2, 156.5, 150.2, 140.6, 137.7, 133.7, 122.3, 114.4, 113.6, 71.0, 69.1, 61.4, 55.6, 14.2; HRMS (ESI): calcd. for C₁₄H₁₆O₅Na [M+Na]⁺ 287.0890, found 287.0890.

Ethyl 5-hydroxy-7-methyl-2,5-dihydrobenzo[b] oxepine-4-carboxylate (11a) Procedure C: white, amorphous solid (0.111 g, 0.45 mmol), 85%. ¹H NMR (600 MHz, CDCl₃) δ 7.12 – 7.08 (m, 2H), 7.01 (d, J = 7.9 Hz, 1H), 6.89 (dd, J = 3.8, 2.1 Hz, 1H), 5.52 (d, J = 9.1 Hz, 1H), 4.98 (dd, J = 19.3, 3.8 Hz, 1H), 4.54 (dd, J = 19.2, 2.1 Hz, 1H), 4.32 – 4.26 (m, 2H), 3.20 (d, J = 9.6 Hz, 1H), 2.32 (s, 3H), 1.34 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.3, 154.6, 140.5, 136.4, 134.8, 134.0, 130.2, 129.2, 121.4, 70.9, 69.2, 61.4, 20.8, 14.3. HRMS (ESI): calcd. for C₁₄H₁₆O₄Na [M+Na]⁺ 271.0941, found 271.0941.

Ethyl 7-chloro-5-hydroxy-2,5-dihydrobenzo[b] oxepine-4-carboxylate (12a) Procedure C: white, amorphous solid (0.097 g, 0.36 mmol), 68%. ¹H NMR (600 MHz, CDCl₃) δ 7.30 (d, J = 2.6 Hz, 1H), 7.25 (dd, J = 8.5, 2.6 Hz, 1H), 7.05 (d, J = 8.4 Hz, 1H), 6.88 (dd, J = 3.7, 2.3 Hz, 1H), 5.55 (s, 1H), 4.97 (dd, J = 19.2, 3.8 Hz, 1H), 4.56 (dd, J = 19.2, 2.7 Hz, 1H), 4.32 – 4.25 (m, 2H), 3.35 (s, 1H), 1.34 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 166.1, 155.3, 140.1, 138.3, 133.4, 130.1, 129.6, 128.5, 123.1, 70.8, 68.5, 61.6, 14.2. HRMS (ESI): calcd. for C₁₃H₁₃O₄ClNa [M+Na]⁺ 291.0395, found 291.0395.

Ethyl (1R,4R,5S)-4-hydroxybicyclo[3.2.1]oct-2-ene-3-carboxylate (exo-13a) and Ethyl (1R,4S,5S)-4hydroxybicyclo[3.2.1]oct-2-ene-3-carboxylate (endo-13a) Procedure C: colorless oil (0.071 g, 0.36 mmol), 68% (5:1 exo/endo); exo-13a: ¹H NMR (600 MHz, CDCl₃) δ 7.24 (dd, J = 7.0, 1.1 Hz, 1H), 4.23 – 4.17 (m, 2H), 4.11 (d, J = 2.8 Hz, 1H), 2.71 (s, 1H), 2.67 (dd, J = 9.8, 6.3 Hz, 1H), 2.55 – 2.49 (m, 1H), 1.93 – 1.87 (m, 1H), 1.85 (d, J = 11.3 Hz, 1H), 1.71-1.64 (m, 1H), 1.63 – 1.58 (m, 1H), 1.35-1.31 (m, 1H), 1.30 (t, J = 7.1 Hz, 3H), 1.24 – 1.19 (m, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 167.7, 148.6, 129.1, 70.9, 60.5, 38.7, 35.6, 30.3, 30.2, 24.7, 14.2; HRMS (ESI): calcd. for C₁₁H₁₆O₃Na [M+Na]⁺ 219.0991, found 219.0992. *endo*-13a: ¹H NMR (600 MHz, CDCl₃) δ 7.19 (d, J = 6.7 Hz, 1H), 4.77 (d, J = 4.8 Hz, 1H), 4.24 – 4.17 (m, 2H), 3.71 (s, 1H), 2.62 – 2.56 (m, 2H), 2.21 – 2.14 (m, 1H), 1.78 – 1.72 (m, 3H), 1.72 – 1.62 (m, 2H), 1.31 (t, J = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 167.6, 148.2, 129.6, 71.7, 60.6, 39.6, 36.5, 36.3, 32.0, 21.2, 14.2; HRMS (ESI): calcd. for C₁₁H₁₆O₃Na [M+Na]⁺ 219.0991, found 219.0992.

(6-hydroxycyclohex-1-en-1-yl)(phenyl)methanone $(14a)^{15}$ Procedure C: colorless oil (0.103 g, 0.51 mmol), 96%. ¹H NMR (600 MHz, CDCl₃) δ 7.69 – 7.63 (m, 2H), 7.57 – 7.52 (m, 1H), 7.44 (dd, J = 10.7, 4.7 Hz, 2H), 6.77 – 6.71 (m, 1H), 4.74 (dd, J = 6.0, 2.8 Hz, 1H), 3.54 (s, 1H), 2.40 – 2.32 (m, 1H), 2.30 – 2.20 (m, 1H), 1.98 – 1.90 (m, 1H), 1.90 – 1.81 (m, 1H), 1.66 (qd, J = 7.1, 3.8 Hz, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 199.4, 147.2, 139.9, 137.7, 132.0, 129.3, 128.2, 64.0, 29.7, 26.4, 17.4.

Total Synthesis of β -(+)-valienamine.

2,3,4-Tri-O-benzyl-D-xylopyranose (20)^{24,25} To methanolic solution of 1.2 % HCl (100 ml) (prepared by dissolved 2.4 ml SOCl₂ in anhydrous methanol) D-xylose (10 g, 66.6 mmol) was added in one portion and resulting mixture was refluxed for 4.5 h under argon (reaction was monitored by TLC CH₂Cl₂/MeOH 2:1). The reaction mixture was allowed to cool to rt, neutralized by addition of solid NaHCO₃ and concentrated under reduced pressure. The residue was dissolved in EtOH (80 ml) and concentrated to the half of the original volume, toluene (2 x 50 ml) was added and the mixture was two times concentrated to dryness. The residue crude mixture oil was used without further purification in the next step. The viscous oil from the previous step was dissolved in anhydrous DMF (100 ml) and anhydrous THF (100 ml) and in 5 portion was added to 60% NaH suspension in mineral oil (13.6 g, 340 mmol) cooled to 0 °C. After hydrogen evolution was complete the mixture was treated with BnBr (29.6 ml, 244 mmol) and stirred for 30 min in 0 °C and then overnight in rt. The reaction mixture was carefully guenched with 10 % cold, agueous solution of NH₄Cl (60 ml) followed by addition of water (150 ml). The aqueous layer was extracted with ethyl acetate (3 x 100 ml) and combined organic layers were washed with water (2 x 120 ml) and brine (100 ml), then dried over anhydrous MgSO₄ and concentrated under reduced pressure. Flash column chromatography Hx/EA (8:1) to (6:1) allows to remove mayor impurities before next step. The partially purified mixture of xylosides from previous step was heated under reflux for 15 h with 1M H₂SO₄ (90 ml), AcOH (90 ml) and 1,4-dioxane (80 ml). Mixture was

 cooled to rt, hexane (40 ml) and water (200 ml) were added with intense stirring. The precipitate was collected by filtration, washed with hexane (2 x 50 ml) and dried under *vacuo*. Recrystallization from methanol gave product as a white solid (19.48 g, 69 %), mp 137-139 °C (137-138 °C lit.), ¹H NMR and ¹³C NMR spectra were compared with lit.²⁵

Ethvl (4S,5R,6R)-4,5,6-tri-O-benzyl-7-hydroxyhept-2-enoate (20a) To solution of 21 (3.65 g, 10.46 mmol) in anhydrous toluene (10 ml), 20 (2.00 g, 4.76 mmol) was added in one portion and mixture was heated to 95 °C for 8 h (reaction was monitored by TLC CHCl₃/Acetone 20:1). The reaction mixture was concentrated under reduced pressure and pass through thin pad of silica to remove part of triphenylphosphine oxide (product was eluted with ca. 200 ml Et₂O). After concentration under reduced pressure crude product was purified by chromatography CHCl₃/Acetone (1:0) to (93:7). Product as a mixture of diastereoisomers (E/Z, 1:0.25) was obtained as a colorless oil (1.52 g, 65%). E-20a: $[\alpha]_D^{26} = -$ 5.2 (c 1.0, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 7.37 – 7.26 (m, 15H), 6.96 (dd, J = 15.8, 5.9 Hz, 1H), 6.07 (dd, J = 15.8, 1.4 Hz, 1H), 4.72 – 4.67 (m, 2H), 4.65 – 4.56 (m, 3H), 4.40 (d, J = 11.7 Hz, 1H), 4.28 - 4.24 (m, 1H), 4.24 - 4.18 (m, 2H), 3.75 - 3.64 (m, 2H), 3.62 (dd, J = 10.1, 4.5 Hz, 1H), 3.54 $(dt, J = 10.0, 4.8 \text{ Hz}, 1\text{H}), 1.31 (t, J = 7.1 \text{ Hz}, 3\text{H}); {}^{13}C \text{ NMR} (151 \text{ MHz}, \text{CDCl}_3) \delta 165.9, 144.6, 138.1,$ 137.7, 137.3, 128.5, 128.5, 128.4, 128.0, 127.9, 127.8, 123.3, 80.7, 79.3, 78.2, 74.7, 72.9, 71.8, 61.3, 60.5, 14.2; **HRMS** (ESI): calcd. for $C_{30}H_{34}O_6Na [M+Na]^+$ 513.2248, found 513.2238. Z-20a: $[\alpha]_D^{24}=$ +58.5 (c 0.65, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 7.39 – 7.24 (m, 15H), 6.37 (dd, J = 11.7, 8.2 Hz, 1H), 5.80 (dd, J = 11.7, 1.3 Hz, 1H), 5.17 (d, J = 11.6 Hz, 1H), 4.76 (d, J = 11.4 Hz, 2H), 4.65 - 4.57(m, 3H), 4.35 (d, J = 11.8 Hz, 1H), 4.12 (q, J = 7.1 Hz, 2H), 3.85 - 3.80 (m, 2H), 3.69 - 3.64 (m, 1H), 3.50 - 3.45 (m, 1H), 1.25 (t, J = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl3) δ 165.8, 148.9, 138.5, 138.1, 137.5, 128.5, 128.4, 128.2, 127.9, 127.9, 127.7, 127.6, 81.3, 80.3, 75.1, 73.8, 73.2, 71.5, 61.6, 60.4, 14.2; **HRMS** (ESI): calcd. for $C_{30}H_{34}O_6Na [M+Na]^+$ 513.2248, found 513.2248.

Ethyl (4S,5R,6R)-4,5,6-tri-O-benzyl-7-oxohept-2-enoate (17) A solution of oxalyl chloride (0.629 ml, 7.34 mmol) in anhydrous methylene chloride (30 ml) was cooled to -78 °C, after that DMSO (1.042 ml, 14.68 mmol) was dropwise added and stirred for 30 min. The **20a** (1.20 g, 2.45 mmol) was dissolved in dry CH₂Cl₂ (15 ml) and slowly added to oxidative mixture and kept in -78 °C for 30 min. The triethylamine (3.094 ml, 22.01 mmol) was added, and continued stirring for next 30 min, after that reaction mixture was allowed slowly warm to 0 °C. The reaction was quenched with addition of sat. NH₄Cl (20 ml), followed by addition of water (20 ml). The aqueous layer was extracted with CH₂Cl₂ (3 x 20 ml), and combined organic layers were washed with water (2 x 20 ml) and brine (20 ml). The organic phase was dried over anhydrous MgSO₄ and concentrated under reduced pressure (below 35 °C). Crude product was purified by flash chromatography Hx/EA (1:0) to (85:15) to give the title compound as a mix-

ture of diastereoisomers (*E*/*Z* 1:0.1) as colorless oil (1.080 g, 90 %); *E*-**17**: $[\alpha]_D^{26} = -5.4$ (c 1.0, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 9.65 (d, *J* = 0.5 Hz, 1H), 7.38 – 7.27 (m, 13H), 7.22 (dd, *J* = 7.4, 1.8 Hz, 2H), 6.88 (dd, *J* = 15.8, 6.2 Hz, 1H), 6.01 (dd, *J* = 15.8, 1.3 Hz, 1H), 4.72 (d, *J* = 11.8 Hz, 1H), 4.62 (d, *J* = 11.8 Hz, 1H), 4.57 (d, *J* = 11.8 Hz, 1H), 4.50 (d, *J* = 11.7 Hz, 2H), 4.39 (d, *J* = 11.4 Hz, 1H), 4.28 (ddd, *J* = 6.0, 4.6, 1.3 Hz, 1H), 4.23 – 4.17 (m, 2H), 3.88 (dd, *J* = 4.4, 0.4 Hz, 1H), 3.82 (t, *J* = 4.5 Hz, 1H), 1.30 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 201.2, 165.8, 144.2, 137.1, 137.1, 136.9, 128.5, 128.4, 128.4, 128.3, 128.2, 128.2, 128.1, 127.9, 123.6, 81.7, 80.7, 77.4, 74.3, 73.3, 72.0, 60.5, 14.2. HRMS (ESI): calcd. for C₃₀H₃₂O₆Na [M+Na]⁺ 511.2091, found 511.2072. *Z*-**17**: $[\alpha]_D^{24} = +27.1$ (c 0.85, CH₂Cl₂); ¹H NMR (400 MHz, CDCl3) δ 9.74 (d, *J* = 0.7 Hz, 1H), 7.42 – 7.21 (m, 15H), 6.35 (dd, *J* = 11.8, 8.3 Hz, 1H), 5.84 (dd, *J* = 11.8, 1.3 Hz, 1H), 5.37 (ddd, *J* = 8.3, 3.0, 1.3 Hz, 1H), 4.77 (d, *J* = 12.0 Hz, 1H), 4.66 (d, *J* = 12.0 Hz, 1H), 4.64 – 4.55 (m, 2H), 4.51 – 4.43 (m, 2H), 4.14 (q, *J* = 7.1 Hz, 2H), 4.05 (dd, *J* = 5.6, 3.0 Hz, 1H), 3.98 (dd, *J* = 5.6, 0.7 Hz, 1H), 1.25 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 200.3, 165.5, 147.5, 137.4, 137.3, 128.4, 128.4, 128.3, 128.2, 128.0, 128.0, 127.9, 127.7, 122.2, 81.4, 81.1, 74.1, 73.7, 72.9, 72.0, 60.3, 14.1; HRMS (ESI): calcd. for C₃₀H₃₂O₆Na [M+Na]⁺ 511.2091, found 511.2090.

Ethvl (3S,4R,5R,6S)-3,4,5-tri-O-benzvl-6-hydroxvcvclohex-1-enecarboxvlate (anti-18) and Ethvl (3S,4R,5R,6R)-3,4,5-tri-O-benzvl-6-hvdroxvcvclohex-1-enecarboxvlate (svn-18) A suspension of elemental selenium (0.116 g, 1.47 mmol) in anhydrous THF (20 ml) was cooled in ice bath and n-BuLi (1.6 M in hexanes, 0.918 ml, 1.47 mmol) was added dropwise (a clear solution was produced) and the mixture was stirred 15 min in 0 °C. After that reaction mixture was cooled to -20 °C, solution of substrate 17 (0.60 g, 1.23 mmol in 3 ml of anhydrous THF) was added dropwise and stirring was continued through the next 6 hours. After that time, reaction was guenched by addition of aqueous, saturated solution of NH₄Cl (0.500 ml) following by addition of hydrogen peroxide solution (30 % v/v, 0.633 ml, 6.15 mmol) and pyridine (0.495 ml, 6.15 mmol). Reaction mixture was allowed to slowly back to rt and after that was heated to 50 °C for 1 hour (reaction mixture became almost colorless). The solvent was evaporated under reduced pressure and resign of pyridine was co-evaporated with toluene (2 x 30 ml). Crude products (mixture of diastereoisomers anti/syn 74:26) were purified by column chromatography Hx/EA (6:1) to (5:1). Pure syn-18 (0.101 g, 17%) and anti-18 (0.289 g, 48%) diastereoisomers were obtained as a colorless syrups. Syn-18: $[\alpha]_{D}^{26} = +3.4$ (c 1.0, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 7.40 – 7.27 (m, 15H), 6.92 (d, J = 2.4 Hz, 1H), 4.96 (d, J = 10.9 Hz, 1H), 4.85 (d, J = 10.9 Hz, 1H), 4.82 (d, J = 11.6 Hz, 1H), 4.79 (d, J = 11.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.74 (s, 2H), 4.29 – 4.22 (m, 2H), 4.19 (dd, J = 1.6 Hz, 1H), 4.76 (d, J = 3.8 Hz, 7.8, 2.4 Hz, 1H), 4.12 (dd, J = 10.3, 7.8 Hz, 1H), 3.52 (dd, J = 10.3, 3.8 Hz, 1H), 2.75 (s, 1H), 1.31 (t, J = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 165.5, 140.1, 138.6, 137.9, 137.8, 130.6, 128.5, 128.4,

128.3, 128.0, 127.9, 127.8, 127.6, (2C)79.1, 78.9, 75.4, 72.8, 72.4, 64.4, 61.2, 14.2; **HRMS** (ESI): calcd. for $C_{30}H_{32}O_6Na [M+Na]^+$ 511.2091, found 511.2086. *Anti*-**18**: $[\alpha]_D^{26} = +23.5$ (c 1.0, CH₂Cl₂); ¹**H NMR** (600 MHz, CDCl₃) δ 7.39 – 7.25 (m, 15H), 6.78 (dd, J = 2.1, 1.2 Hz, 1H), 4.92 (d, J = 11.2 Hz, 1H), 4.89 (d, J = 11.0 Hz, 1H), 4.86 (d, J = 11.2 Hz, 1H), 4.79 (d, J = 11.0 Hz, 1H), 4.76 – 4.70 (m, 2H), 4.69 – 4.65 (m, J = 5.3 Hz, 1H), 4.28 – 4.25 (m, 1H), 4.24 (q, J = 7.1 Hz, 2H), 3.74 – 3.68 (m, 2H), 3.51 (d, J = 1.6 Hz, 1H), 1.30 (t, J = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 166.0, 138.6, 138.3, 137.8, 131.3, 128.5, 128.4, 128.3, 128.0, 127.9, 127.9, 127.7, 127.6, 82.9, 81.7, 78.4, 75.4, 75.3, 73.1, 71.9, 61.3, 14.2. **HRMS** (ESI): calcd. for C₃₀H₃₂O₆Na [M+Na]⁺ 511.2091, found 511.2096.

(3S,4R,5R,6S)-3,4,5-tri-O-benzyl-6-hydroxy-1-hydroxymethyl-cyclohexene (anti-18a) A solution of compound anti-18 (0.250 g, 0.51 mmol) in anhydrous methylene chloride (5 ml) was cooled to -10 °C and solution of DIBAL-H (1 M in hexane, 1.790 ml, 1.79 mmol) was added dropwise. When the addition was complete, the mixture was stirred at -10 °C for 3 h. After that time, H₂O (0.200 ml) was added to quench the reaction, and mixture was stirred at -10 °C for 30 min. Evaporation of the solvent under reduced pressure gave a crude solid residue. Then, methanol (10 ml) and celite (~1 g) were added and the mixture was vigorously stirred at room temperature for 30 min and then filtered by suction through thin pad of celite. Crude product was eluted with methanol (20-30 ml, TLC-check) and filtrate was concentrated under vacuo to give oily residue. Crude product was purified by column chromatography Hx/EA (2:1) to (1:1). Pure anti-18a was obtained as a colorless syrup (0.139 g, 61%). $[\alpha]_D^{24} = +77.1$ (c 0.8, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.27 (m, 15H), 5.69 (s, 1H), 5.00 (d, J = 11.5 Hz, 1H), 4.88 (s, 2H), 4.72 – 4.66 (m, 3H), 4.38 (d, J = 7.7 Hz, 1H), 4.27 – 4.09 (m, 3H), 3.75 (dd, J = 10.1, 7.4 Hz, 1H), 3.59 (dd, J = 10.1, 7.8 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 138.4, 138.3, 138.2, 138.1, 128.6, 128.4, 127.9, 127.9, 127.8, 127.8, 127.7, 123.7, 83.6, 82.9, 79.7, 75.2, 75.1, 73.0, 72.3, 64.2; HRMS (ESI): caled. for C₂₈H₃₀O₅Na [M+Na]⁺ 469.1985, found 469.1985.

(3*S*, 4*R*, 5*R*, 6*S*)-3, 4, 5, 6-penta-O-benzyl-1-O-benzylmethyl-cyclohexene (22) To stirred suspension of NaH (0.035 g, 0.87 mmol, 60% in mineral oil) in anhydrous DMF (3 ml) was added a solution of *anti*-18a (0.130 g, 0.29 mmol) and BnBr (0.104 ml, 0.87 mmol) at 0 °C under argon. After 30 min stirring, ice bath was removed and stirring was continued overnight in room temperature. The reaction mixture was carefully quenched with a cold, 10% aqueous solution of NH₄Cl (3 ml). The aqueous layer was extracted with ethyl acetate (3 x 5 ml). The organic layer was washed with H₂O (7 ml) and brine (7 ml), dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by column chromatography Hx/EA (9:1) to (8:1). Pure 22 was obtained as an amorphous white solid (0.174 g, 95%). $[\alpha]_D^{24}$ = +59.2 (c 0.36, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.26 (m, 25H), 5.82 (s, 1H), 5.02 (d, J = 10.9 Hz, 1H), 4.94 (s, 2H), 4.89 – 4.81 (m, 2H), 4.73 (d, J = 9.2 Hz, 3H), 4.58 – 4.47 (m,

2H), 4.34 (d, J = 7.0 Hz, 1H), 4.29 – 4.21 (m, 2H), 3.96 (d, J = 12.3 Hz, 1H), 3.90 – 3.77 (m, 2H); ¹³C **NMR** (101 MHz, CDCl₃) δ 138.6, 138.6, 138.4, 138.3, 138.1, 136.4, 128.4, 128.4, 128.3, 128.3, 127.9, 127.9, 127.8, 127.6, 127.6, 127.60, 125.17, 84.4, 83.7, 80.0, 79.8, 75.5, 75.5, 74.9, 72.4, 72.2, 69.8; **HRMS** (ESI): calcd. for C₄₂H₄₂O₅Na [M+Na]⁺ 649.2924, found 649.2924.

Benzyl ((3*S*,4*R*,5*R*,6*S*)-4,5,6-tri-O-benzyl-3-((benzyloxy)methyl)cyclohex-2-en-1-yl)carbamate (**19**) To stirred solution of **22** (0.170 g, 0.27 mmol) in anhydrous toluene (1.8 ml) was added Na₂CO₃ (0.316 g, 2.98 mmol) and chlorosulfonyl isocyanate (0.189 ml, 2.17 mmol) at 0 °C under argon. The reaction mixture was stirred 15 h at 0 °C and quenched with carefully addition H₂O (2 ml). The aqueous layer was extracted with ethyl acetate (3 x 4 ml). The organic layer was added to solution of aqueous 25% Na₂SO₃ (12 ml). The reaction mixture was stirred for 24 h at room temperature. The organic layer was washed with H₂O (~6 ml) and brine (~6 ml), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The solid residue was purified by column chromatography DCM/Acetone (1:0) to (99:1) to afford **19** (0.135 g, 74%) as a white solid, mp. 117-118 °C (116-119 °C for enantiomer lit.).²² $[\alpha]_D^{24} = +83.2$ (c 0.64, CHCl₃); ¹**H NMR** (400 MHz, CDCl₃) δ 7.46 – 7.19 (m, 25H), 5.68 (s, 1H), 5.19 – 5.06 (m, 2H), 4.88 – 4.64 (m, 7H), 4.54 – 4.39 (m, 3H), 4.24 (d, J = 8.6 Hz, 2H), 3.95 – 3.86 (m, 2H), 3.61 – 3.52 (t, J = 7.8 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 155.9, 138.4, 138.2, 138.2, 136.6, 136.4, 128.6, 128.5, 128.5, 128.3, 128.2, 128.0, 128.0, 127.9, 127.9, 127.8, 127.7, 126.1, 82.3, 79.7, 77.9, 74.7, 74.3, 74.2, 72.6, 70.4, 66.8, 52.1; **HRMS** (ESI): calcd. for C₄₃H₄₃NO₆Na [M+Na]⁺ 692.2983, found 692.2983. ¹H NMR and ¹³C NMR spectra were compared with enantiomeric form published in lit.²²

 β -(+)-valienamine (16) To stirred solution of 19 (0.050 g, 0.07 mmol) in anhydrous methylene chloride (1.5 ml) was added BCl₃ (7 ml, 7 mmol, 1M in DCM) at -78 °C. The reaction mixture was stirred for 24 h in -78 °C and after that time quenched with methanol (1.5 ml) was added and kept in -78 °C for next 1 h. The resulting mixture was warmed to room temperature and methylene chloride was distilled. The resulting mixture of hydrogen chloride in methanol was refluxed for 3 h and then evaporated under reduced pressure. The residue was purified by ion-exchange resin DOWEX 50WX8-100 using 0.5 M NH₄OH as eluent to afford free amine 16 (0.009 g, 69%) as a nearly colorless syrup.¹H NMR and ¹³C NMR spectra were compared with lit.²⁶ [α]²⁴_D = +83.5 (c 0.45, D₂O); ¹H NMR (600 MHz, D₂O) δ 5.48 – 5.45 (m, 1H), 4.13 – 4.08 (m, 2H), 3.99 (d, J = 14.1 Hz, 1H), 3.46 (dd, J = 10.4, 8.1 Hz, 1H), 3.41 (d, J = 8.8 Hz, 1H), 3.31 (dd, J = 10.2, 8.9 Hz, 1H); ¹³C NMR (151 MHz, D₂O) δ 139.0, 123.4, 75.9, 74.5, 71.9, 61.0, 53.0; HRMS (ESI): calcd. for C₇H₁₃NO₄Na [M+Na]⁺ 198.0736, found 198.0738, calcd. for C₁₄H₂₆N₂O₈Na [2M+Na]⁺ 373.1581, found 373.1587.

β-(+)-valienamine hydrochloride (16a) 16 (0.009 g, 0,05 mmol) was dissolved in water (1 ml). After addition 3 drops of 1M HCl, water was evaporated under high vacuum. Received syrup was dissolved in D₂O and ¹H and ¹³C NMR spectra were collected. ¹H NMR (300 MHz, D₂O) δ 5.61 – 5.56 (m, 1H), 4.22 – 4.11 (m, 3H), 3.87 (d, J = 7.4 Hz, 1H), 3.67 – 3.52 (m, 2H); ¹³C NMR (75 MHz, D₂O) δ 143.8, 117.2, 76.0, 72.2, 71.9, 61.3, 54.1.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website. ¹H and ¹³C NMR spectra for the synthesized compounds (PDF)

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NOTES

There are no conflicts to declare.

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