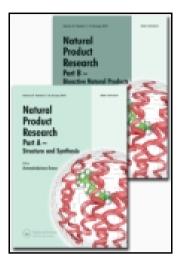
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Leishmanicidal potential of N-substituted morpholine derivatives: Synthesis and structure–activity relationships

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A series of N-substituted morpholines **2–20** was synthesised by reacting various acid chlorides and alkyl halides with morpholine (1). All of the synthesised compounds **2–20** were screened for their leishmanicidal effects using amphotericin B ($IC_{50} = 0.24 \,\mu g \, L^{-1}$) and pentamidine ($IC_{50} = 2.56 \,\mu g \, m L^{-1}$) as standards and a structure-activity relationship (SAR) study was established. The compounds **2** ($IC_{50} = 48 \,\mu g \, m L^{-1}$), **3** ($IC_{50} = 30.0 \,\mu g \, m L^{-1}$), **10** ($IC_{50} = 41.0 \,\mu g \, m L^{-1}$), **15** ($IC_{50} = 33.0 \,\mu g \, m L^{-1}$), **16** ($IC_{50} = 35.0 \,\mu g \, m L^{-1}$) and **20** ($IC_{50} = 47.0 \,\mu g \, m L^{-1}$) showed weak leishmanicidal activities.

Keywords: N-substituted morpholine; synthesis; anti-Leishmanial activity; SAR

1. Introduction

Leishamaniasis is one of the major parasitic diseases, causes enormous suffering in many parts of the tropical and subtropical regions of the world, and contributes to serious health problems (World Health Organisation (WHO), 2001). This disease manifests in different clinical forms, including cutaneous, mucocutaneous and visceral leishmaniasis. The disease is caused by a parasitic protozoa (*Kinetoplastida*: Trypanosomatideae) (Olliaro & Bryceson, 1993; WHO, 2001), which is transmitted as metacyclic flagellated promastigote forms from host to host by the bite of infected sand flies. This species has been isolated from patients with visceral disease or with post-Kala Azar dermal leishmaniasis (Barral et al., 1991; Leon, Machado, Carvalho-Paes, & Grimaldi, 1990).

Therapy of patients with leishmaniasis remains a serious problem. The drugs for the treatment of all clinical forms of leishmaniasis are sodium stibogluconate (pentostam) and meglumine antimonate (glucantime), although they exhibit renal and cardiac toxicity (Raht et al., 2003). Alternative drugs such as pentamidine, amphotericin B and some azo-derivatives are also very toxic, with serious side effects (Mcgregor, 1998). Miltefosine, a phosphocholine analogue, originally developed as an anticancer agent, has been found to be highly effective against leishmaniasis *in vitro* and *in vivo*. Presently, this compound is the

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only oral agent effective against both cutaneous (Soto et al., 2004) and visceral (Prasad, Kumar, Jaiswal, & Singh, 2004) leishmaniasis, although causing severe gastrointestinal problems (Sangraula, Sharma, Rijal, Dwivedi, & Koirala, 2003). Since the chemotherapy against leishmaniasis is still inefficient and with toxic side effects, there is an urgent need for the development of new, efficient and non-toxic drugs for the treatment of this disease (Carvalho, Arribas, & Ferreira, 2000).

In a continuation of research work on bioactive molecules, and as a part of a programme to study potential leishmanicidal agents (Khan et al., 2003, 2008), 19 morpholine derivatives 2–20 (Scheme 1) were synthesised and randomly screened for their leishmanicidal potential. These studies may provide lead compounds for future drug development.

2. Results and discussion

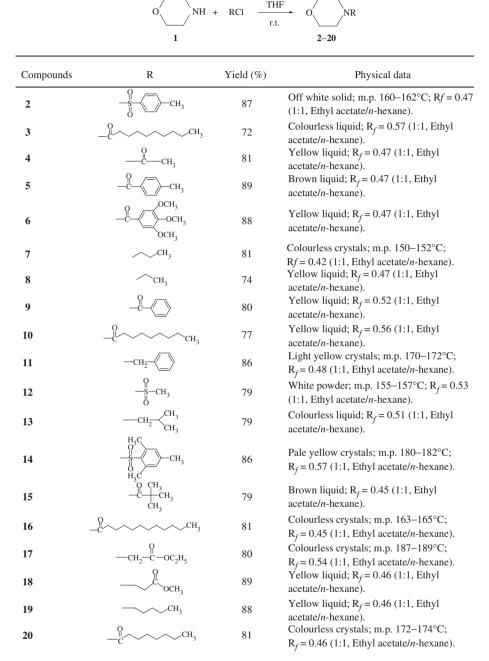
2.1. Chemistry

A series of morpholine derivatives 2–20 (Scheme 1) were synthesised by treating various acid chlorides with morpholine (1) in tetrahydrofuran at room temperature. The structures of the resultant products 2–20 were determined spectroscopically, including UV, IR, NMR, and MS. CHN analyses were found in good agreement with the calculated values.

2.2. Biology

Several reports (Ram & Nath, 1996) describing the leishmanicidal activity of nitrogencontaining compounds suggested random evaluation of the leishmanicidal potential of the synthetic N-substituted morpholines in the present series. The testing was performed at the 100 μ g mL⁻¹ level according to a literature protocol (Atta-ur-Rahman, Choudhary, & Thomsen, 2001) in duplicate. Amphotericin B and pentamidine were used as the positive controls, having an IC₅₀ of 0.24 and 2.56 μ g mL⁻¹, respectively.

All of the synthetic derivatives 2–20 of morpholine (1) were tested for their effects against Lesihmania major and the results of the leishmanicidal activity are collected in Table 1. The compounds 3 and 16 were found to be the most active compounds in the present series, having IC₅₀ values of 30.0 and $33.0 \,\mu g \,m L^{-1}$, respectively. Compounds 3, 10, 16, and 20, which were long, straight chain aliphatic amides, showed IC_{50} values of 30.0, 41.0, 47.0, and $35.0 \,\mu g \,m L^{-1}$, respectively. However, the aliphatic amide 4 exhibited an IC₅₀ value of $80.0 \,\mu g \,m L^{-1}$. The difference in activities of compounds may be explained on the basis of their structures and suggested that the IC_{50} values of these compounds depends on the length of the carbon chain attached to the amide moiety. A nine-carbon chain attached to the amide residue, as in compound 3, was found to be the most suitable for leishmanicidal activity. The slight decline in activity of compound 20, having an IC_{50} value of $47.0 \,\mu g \,m L^{-1}$, may be due to the reduction of one or two carbons in the carbon chain. Unfortunately, a compound with 10 carbons could not be prepared, but compound 16 (IC₅₀ = 35.0 μ g mL⁻¹), with an 11-carbon chain, showed a slight decline in activity, demonstrating that either an increase or a decrease in carbon chain affects the leishmanicidal activity. This hypothesis also establishes that the relative IC_{50} value of compound 4 (IC₅₀ = $80.0 \,\mu g \,m L^{-1}$), which has only one carbon attached to the amide residue, with compound 15 ($IC_{50} = 35.0 \,\mu g \,m L^{-1}$) may be explained on the basis of its branched carbon chain. The aromatic amides 5, 6 and 9 showed very weak activity, which



Scheme 1. Synthetic route and physical data of compounds 2-20.

indicate that an aromatic linkage with an amide residue in this type of compound is not desirable for leishmanicidal activity.

The sulphonamides 2, 12 and 14 were also evaluated for leishmanicidal activity, having IC_{50} values 48.0, 51.0 and $100.0 \,\mu g \,m L^{-1}$, respectively. The difference in activities

Compounds	$IC_{50} \ (\mu g m L^{-1})$
2	48.0
2 3 4 5 6	30.0
4	80.0
5	83.0
	100.0
7	100.0
8	80.0
9	100.0
10	41.0
11	100.0
12	51.0
13	100.0
14	100.0
15	35.0
16	35.0
17	100.0
18	100.0
19	100.0
20	47.0
Amphotericin B	0.24
Pentamidine	2.56

Table 1. Leishmanicidalactivityofcompounds 2–20.

may again be explained on the basis of their structures. Compound 2, which was a 4-methyl phenyl sulphonamide, was found to be the most active sulphonamide, whereas methane sulphonamide showed a slight decline in activity, suggesting that aromatic sulphonamides with suitable substitution may have better potential for leishmanicidal activity as compared to aliphatic sulphonamides. The marked reduction in activity of 2,4,6-trimethylphenyl sulphonamide (14) may be explained on the basis of the steric hindrances of the sulphonamide.

The compounds 7, 8, 11, 13 and 17–19, which were tertiary amine derivatives of morpholine, showed only weak potential for leishmanicidal activity. Only compound 8 showed activity of $<80.0 \,\mu\text{g}\,\text{m}\text{L}^{-1}$, while all other compounds showed $100.0 \,\mu\text{g}\,\text{m}\text{L}^{-1}$. The enhanced activity of compound 8 may be due to the presence of a small carbon chain (ethyl) on the nitrogen atom of morpholine (1), while compounds 11, 13 and 17–19 showed a decrease in activity due to an unsuitable carbon chain attached to the nitrogen atom.

In conclusion, aliphatic, straight carbon chain amides enhance the leishmanicidal activity of morpholine (1), while an aliphatic, branched chain amide linkage at the nitrogen atom of morpholine (1) also increased the leishmanicidal potential of morpholine (1). For the sulphonamides, it was concluded that a suitable substitution on an aromatic sulphonamide may enhance the activity of morpholine (1) and that a small carbon chain sulphonamide of morpholine (1) might play an important role in increasing leishmanicidal activity. Further research on the tertiary amides of compound 1 may also produce good results, either using aliphatic, straight carbon chains with some functional groups like amidine, or hydrazino residues, which may enhance the activity,

as in the case of pentamidine and acyl hydrazides. This research suggested that morpholine may act as possible lead compound for further research for potential leishmanicidal agents.

3. Experimental

3.1. General

Melting points were determined using a Büchi 434 melting point apparatus and are uncorrected. NMR spectroscopy was performed on a Bruker AVANCE 400 MHz. Elemental analyses were carried out on a Carlo Erba Strumentazion-Mod-1106, Italy. UV and IR spectra were recorded on a Perkin-Elmer Lambda-5 UV/VIS and a JASCO IR-A-302 spectrophotometer, respectively. A Finnigan MAT-311A (Germany) spectrometer was used to record mass spectra. Thin layer chromatography (TLC) was performed on pre-coated silica gel glass plates (Kieselgel 60, 254, E. Merck, Germany) and chromatograms were visualised by UV light at 254 and 365 nm or iodine vapours.

3.2. General procedure for the preparation of compounds 2-20

To a mixture of morpholine (1, 1 mmol) dissolved THF (10 mL) was added a mixture of potassium carbonate (2 mmol), and the resultant mixture was stirred at room temperature for 15 min and then various acid chlorides (1 mmol) were added. The mixture was stirred for 48 h and the reaction was monitored by TLC analysis. On completion, the solid was filtered with a sintered glass funnel. The filtrate was evaporated on a rotary evaporator, the residue was dried on a two-stage pump (high vacuum) and the resultant product, if solid, was recrystalised and dried in a desiccator, while the liquid products were purified by vacuum distillation. The compounds were characterised using different spectroscopic techniques, including UV, IR, NMR and MS, elemental analyses, to determine the purity.

3.3. Leishmanicidal bioassay (in vitro)

Leishmania major were grown in bulk in modified NNN biphasic medium by using normal physiological saline. *Leishmania* promastigotes were cultured with RPMI 1640 medium, supplemented with 10% heat-inactivated foetal bovine serum (FBS). Parasites at log phase were centrifuged at 2000 rpm for 10 min, and washed three times with saline at the same speed and time. Parasites were diluted with fresh culture medium to a final density of 10⁶ cells per millilitre.

In a 96-well microtiter plate, $180 \,\mu\text{L}$ of medium was added in the first row and $100 \,\mu\text{L}$ of medium was added in others wells. A total of $20 \,\mu\text{L}$ of the experimental compound was added in medium and serially diluted. A total of $100 \,\mu\text{L}$ of the parasite culture was added in all wells. Two rows were left for negative and positive controls. Negative controls received only medium while the positive control contained varying concentrations of standard anti-leishmanial compounds amphotericin B and petamidine. The plate was incubated $21-22^{\circ}\text{C}$ for 72 h. The culture was examined microscopically on an improved Neubauer counting chamber and IC₅₀ values of compounds were calculated by software EZfit 5.03 (Perella Scientific). All assays were performed in duplicate (Atta-ur-Rahman, Choudhary, & Thomsen, 2001).

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References

- Atta-ur-Rahman, Choudhary, M.I., & Thomsen, W.J. (2001). Bioassay techniques for drug development (pp. 60–64). The Netherlands: Harwood Academic Publisher.
- Barral, A., Pedral-Sampaio, D., Grimaldi, G. Jr, Momen, H., McMahon-Pratt, D., Ribeiro de Jesus, A., et al. (1991). Leishmaniasis in Bahia, Brazil: Evidence that *Leishmania amazonensis* produces a wide spectrum of clinical disease. *The American Journal* of Tropical Medicine and Hygiene, 44, 536–546.
- Carvalho, P.B., Arribas, M.A.G., & Ferreira, E.I. (2000). Leishmaniasis: What do we know about its chemotherapy? *Brazilian Journal of Pharmaceutical Science*, 36, 69–96.
- Khan, K.M., Ahmed, S., Khan, Z.A., Zia-Ullah, Rani, M., Choudhary, M.I., et al. (2008). *In vitro* leishmanicidal activity of 3-substituted isocoumarins: Synthesis and structure-activity relationship. *Medicinal Chemistry (Shāriqah, United Arab Emirates)*, 4, 163–169.
- Khan, K.M., Rasheed, M., Zia-Ullah, Hayat, S., Kaukab, F., Choudhary, M.I., et al. (2003). Synthesis and *in vitro* leishmanicidal activity of some hydrazides and their analogues. *Bioorganic & Medicinal Chemistry*, 11, 1381–1387.
- Leon, L.L., Machado, G.M.C., Carvalho-Paes, L.E., & Grimaldi, G. Jr (1990). Antigenic differences of *Leishmania amazonensis* isolates causing diffuse cutaneous leishmaniasis. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 84, 678–681.
- Mcgregor, A. (1998). WHO warns of epidemic leishmania. Lancet, 351, 575.
- Olliaro, P.L., & Bryceson, A.D.M. (1993). Practical progress and new drugs for changing patterns of leishmaniasis. *Parasitol Today*, 9, 323–328.
- Prasad, R., Kumar, R., Jaiswal, B.P., & Singh, U.K. (2004). Miltefosine: An oral drug for visceral leishmaniasis. *Indian Journal of Pediatrics*, 71, 143–144.
- Raht, S., Trivellin, A., Imbrunito, T.R., Tomazela, D.M., Jesus, M.N., Marzal, P.C., et al. (2003). Antimoniais empregados no tratamento da Leishmaniose: Estado de arte. *Química Nova*, 26, 550–557.
- Ram, V.J., & Nath, M. (1996). For a review see: Progress in chemotherapy of leishmaniasis. Current Medicinal Chemistry, 3, 303–316.
- Sangraula, H., Sharma, K.K., Rijal, S., Dwivedi, S., & Koirala, S. (2003). Orally effective drugs for kala-azar (visceral leishmaniasis): Focus on miltefosine and sitamaquine. *The Journal of the Association of Physicians of India*, 51, 686–690.
- Soto, J., Arana, B.A., Toledo, J., Rizzo, N., Vega, J.C., Diaz, A., et al. (2004). Miltefosine for new world cutaneous leishmaniasis. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*, 38, 1266–1272.
- World Health Organisation. (2001). Tropical Disease Research: Progress 1999–2000; Fifteenth Programme Report of the UNDP/World Bank/WHO Special Programme for Research & Training in Tropical Diseases. (TDR/GEN/01.5). Geneva: World Health Organization.