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Synthesis of various secosteroidal macrocycles by ring-closing metathesis

Malika Ibrahim-Ouali*, Eugénie Romero

Aix Marseille Université, CNRS Institut des Sciences Moléculaires de Marseille UMR 7313, 13397 Marseille, France

ABSTRACT

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1. Introduction

Steroids represent an important class of natural products due to their high ability to penetrate cells and bind to nuclear and membrane receptors. The steroid system, selected by the evolutionary process to perform some of the most fundamental biological functions, has not only inspired biochemists and endocrinologists, but has also become the basis of many important discoveries in organic chemistry.

Secosteroids have attracted considerable interest because of the broad range of biological activities of many naturally occurring representatives, such as Vitamins D [1], anolides [2], and marine steroids [3,4]. Apart from Vitamins D with their innumerable biological effects [5], secosteroids with cytotoxic [6–8], antihistamine [9], and anticancer [10] activity should be mentioned as compounds with great potential for drug development. The activity of seco analogs of normal steroidal hormones in humans and higher animals is a matter of scientific interest as well. Some of these compounds were prepared synthetically and showed hormonal or antihormonal activity [11–16]. It is evident that the higher conformational flexibility of seco steroids in comparison with normal steroids may result in novel, pharmaceutically useful compounds.

Otherwise, macrocyclic compounds have unique physicochemical and topological properties that allow them to exhibit unusual biological properties [17,18]. Macrocycles have the ability to exhibit high target binding affinity, selectivity, and improved oral bioavailability. Additionally, macrocyclization is an efficient way of increasing cellular penetration via the decrease in polarity of peptidic drug leads [19]. Macrocycles have been proven to be efficient as protease inhibitors [20,21], G protein- coupled receptors (GPCRs) [22–24], and protein–protein interaction inhibitors [25,26]. Natural products are one of the sources of bioactive macrocycles such as erythromycin, rapamycin, vancomycin, cyclosporine, and epothilone [27]. Natural products exhibit enormous structural diversity [28]. However, there are several problems associated with their use in screening experiments including difficulties with purification, bioactive component identification, structural assignment, chemical modification, and analog synthesis [29]. These difficulties have motivated medicinal chemistry researchers to develop strategies such as diversity oriented synthesis (DOS) of macrocyclic compounds with known bioactive domains such as peptide motifs [30–32].

We set out to describe an efficient and versatile method for preparing secosteroidal macrocycles from

cholic acid, via an oxidative ring-expansion/ring-opening sequence and a ring-closing metathesis reaction

as the key steps. The characteristic ¹H and ¹³C NMR spectroscopic features of the synthesized compounds

Consequently, many bioactive macrocycles with diversity and complexity are now readily available in chemist's showcases [33,34].

So, it would be interesting to combine secosteroidal skeleton with varied types of macrocycles in order to obtain a new class of biological molecules. We report herein an efficient synthesis of secosteroidal macrocycles from cholic acid **1** (Scheme 1). We show that we can modify the size of the macrocycle and then lead to novel interesting structures. We report here the full details of these syntheses.

2. Experimental section

All reactions were run under argon in oven-dried glassware. ¹H and ¹³C NMR spectra are recorded at 200 or 400 and 50 and 100 MHz respectively, in CDCl₃ solutions. Chemical shift (δ) are reported in ppm with tetramethylsilane as internal standard. NMR signals assignments were made with the aid of a combination of 2D homonuclear (¹H–¹H) and heteronuclear (¹H–¹³C) correlation





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^{*} Corresponding author. Tel.: +33 491288416; fax: +33 491983865. *E-mail address:* malika.ibrahim@univ-amu.fr (M. Ibrahim-Ouali).

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Scheme 1. Retrosynthetic pathway.

techniques, which included ¹H–¹H COSY, ¹H–¹H Nuclear Overhauser Effect Spectroscopy (NOESY), Heteronuclear Single Quantum Correlation (HSQC) and Heteronuclear Multiple Bond Correlation (HMBC). IR spectra were recorded on a Perkin-Elmer 1600 spectrophotometer. Flash chromatography was performed on silica gel (Merk 60 F254) and TLC on silica gel. Dichloromethane was distilled from P2O5 and tetrahydrofuran (THF) over sodium/ benzophenone.

Compounds **14** and **15** were prepared according to the previously described procedure [35]. The nomenclature used for the steroids is not the nomenclature used by Chemical Abstracts [36,37].

2.1. Methyl 3α , 7α -dimethoxy- 12α -hydroxy- 5β -cholan-24-oate (**3**)

To a stirred suspension of NaH (0.62 g, 26 mmol) in THF (10 mL) at 0 °C under argon was added a solution of triol 2 (5 g, 11.8 mmol) in 5 mL of THF. The reaction mixture was stirred for 15 min, and then iodomethane (367 µL, 5.9 mmol) was added dropwise. After 24 h at room temperature, the reaction was diluted with 10 mL of Et₂O and quenched by the slow addition of 10 mL of H₂O. The combined organic extracts $(3 \times 10 \text{ mL})$ were washed with 30 mL of brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel (Et₂O: 100%) to give **7** (5 g, 94%) as a white solid. mp = 80 °C; ¹H NMR (300 MHz, CDCl₃): 0.59 (s, 3H, H-18), 0.83 (d, J = 6.5, 3H, H-21), 0.85 (s, 3H, H-19), 2.94 (m, 1H, H-3β), 3.14 (s, 3H, OCH₃), 3.18 (s, 3H, OCH₃), 3.21 (m, 1H, H-7β), 3.26 (s, 3H, OCH₃), 3.29 (m, 1H, H-12β); ¹³C NMR (75 MHz, CDCl₃): 12.6, 17.5, 22.1, 23.0, 23.3, 26.4, 26.6, 27.5, 27.9, 28.1, 30.9, 31.2, 34.5, 35.2, 35.6, 39.8, 42.1, 42.8, 46.3, 46.4, 55.5, 55.8, 56.0, 77.3, 81.0, 82.1, 180.3. HRMS (EI) for C₂₇H₄₆O₅ [M⁺] calcd 450.3345 found 450.3348.

2.2. Methyl 3α , 7α -dimethoxy-12-oxo-5 β -cholan-24-oate (**4**)

Alcohol **3** (1 g, 2.22 mmol) was mixed in a mortar with pyridinium chlorochromate (PCC) (0.57 g, 2.66 mmol). The mixture was transferred to a pressure-resistant tube (pyrex) and irradiated with MW at 170 °C for 5 min. After cooling to room temperature, the reaction mixture was filtered through a Celite pad and the filtrate and washings (CH₂Cl₂, 3 * 10 mL) were combined and evaporated under reduced pressure. The residue was chromatographed on silica gel (diethyl ether/petroleum ether: 7/3), to afford 0.66 g (70% yield) of 12-oxo steroid **4** as colorless needles. mp = 71–72 °C; IR (neat) 3236, 1736, 1511 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 0.85 (d, *J* = 6.5, 3H, H-21), 1.02 (s, 3H, H-18), 1.03 (s, 3H, H-19), 3.26 (m, 1H, H-3 β), 3.28 (s, 3H, OCH₃), 3.32 (s, 3H, OCH₃), 3.38 (m, 1H, H-7 β), 3.66 (s, 3H, OCH₃); ¹³C NMR (75 MHz, CDCl₃): 18.4, 21.3, 21.4, 22.0, 23.7, 26.4, 27.3, 30.3, 30.9, 31.1, 31.2, 31.3, 34.4, 35.4, 35.6, 37.8, 40.4, 46.2, 51.4, 53.0, 57.1, 57.5, 58.4, 73.4, 78.4, 174.7, 214.0. HRMS (EI) for C₂₇H₄₄O₅ [M⁺] calcd 448.3189 found 448.3192.

2.3. 3α , 7α -Dimethoxy-13-oxa-C-homo-cholan-12-one (**5**)

To a solution of ketone 4 (0.5 g, 1.11 mmol) in dry dichloromethane (30 mL) containing *p*-toluenesulfonic acid (167 mg, 1.11 mmol) *m*-chloroperbenzoic acid (12 mg) was added. The solution was stirred for 24 h at room temperature. The solution was then diluted with water and extracted with dichloromethane (3 * 15 mL). The solution was washed successively with a 5% Na₂S₂₋ O₃ solution (10 mL), saturated brine (10 mL), and water (20 mL) and was dried over anhydrous magnesium sulfate. The oily product, obtained by evaporation of the solvent, was purified by flash chromatography on silica gel (CH₂Cl₂/MeOH: 95/5) to afford 0.51 g of pure lactone **5** (98%) as an oil. ¹H NMR (300 MHz, $CDCl_3$): 0.86 (s, 3H, H-19), 1.05 (d, J = 6.4, 3H, H-21), 1.34 (s, 3H, H-18), 3.25 (m, 1H, H-3β), 3.28 (s, 3H, OCH₃), 3.31 (s, 3H, OCH₃), 3.32 (s, 3H, OCH₃), 3.35 (m, 1H, H-7β); ¹³C NMR (75 MHz, CDCl₃): 14.5, 15.3, 17.7, 22.3, 24.2, 25.2, 26.1, 26.5, 27.5, 31.6, 32.8, 34.6, 35.2, 35.9, 36.3, 41.2, 42.5, 50.1, 55.7, 57.4, 58.5, 73.3, 76.9, 80.2, 86.9, 174.8, 176.5. HRMS (EI) for C₂₇H₄₄O₆ [M⁺] calcd 464.3138 found 464.3143.

2.4. 3α,7α-Dimethoxy-11,12-seco-5β-cholan-12,13α,24-triol (**6**)

A solution of methyl ester **5** (1 g, 2.15 mmol) in dry ether (10 mL) was added in one portion to a suspension of LiAlH₄ (0.25 g, 6.46 mmol) in dry ether (20 mL) at room temperature. After 1 h the reaction was quenched with H₂O (15 mL) and EtOAc (15 mL). The aqueous layer was acidified to pH 2 with diluted HCl, and layers were separated. The aqueous layer was further extracted with EtOAc (3 \times 50 mL), and the combined organic layers

were dried over anhydrous MgSO₄ and evaporated to dryness. The crude product was purified by flash chromatography on silica gel (CH₂Cl₂/MeOH: 9/1) to afford 0.83 g of pure triol **6** (88%) as an oil. IR (neat) 3280 (OH), 1220, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 1.06 (d, *J* = 6.5, 3H, H-21), 1.16 (s, 3H, H-19), 1.50 (s, 3H, H-18), 2.78 (m, 1H, H-3 β), 2.79 (m, 1H, H-7 β), 3.24 (s, 3H, OCH₃), 3.25 (s, 3H, OCH₃), 3.37 (m, 2H, H-24), 3.53 (m, 2H, H-12 β); ¹³C NMR (75 MHz, CDCl₃): 19.1, 19.5, 19.7, 23.5, 24.4, 24.6, 29.9, 30.8, 31.6, 32.0, 33.8, 34.1, 35.7, 38.4, 40.8, 46.2, 46.3, 51.0, 57.1, 57.4, 59.3, 61.6, 74.9, 82.4, 82.9, 89.2. HRMS (EI) for C₂₆H₄₈O₅ [M⁺] calcd 440.3502 found 440.35.09.

2.5. 3α , 7α -Dimethoxy-12,24-diallyloxy-11,12-seco-5 β -cholan-13 α -ol (**7**)

To a stirred suspension of KH (0.32 g, 6.81 mmol) in DMF (10 mL) at 0 °C under argon was added a solution of triol 6 (1 g, 2.27 mmol) in 5 mL of DMF. The reaction mixture was stirred for 15 min, and then allyl bromide (6.43 mL, 4.99 mmol) was added dropwise. After 48 h at room temperature, the reaction was diluted with 10 mL of Et₂O and quenched by the slow addition of 10 mL of H₂O. The solution was extracted with diethyl ether (3 * 10 mL) and the combined organic extracts were washed with 30 mL of brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel (Et₂O: 100%) to give 7 (0.68 g, 58%) as an oil. IR (neat) 2951, 1175, 1096 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 1.05 (d, *J* = 6.4, 3H, H-21), 1.16 (s, 3H, H-19), 1.43 (s, 3H, H-18), 2.79 (m, 1H, H-3β), 2.82 (m, 1H, H-7β), 3.24 (s, 3H, OCH₃), 3.32 (m, 2H, H-24), 3.25 (s, 3H, OCH₃), 3.38 (m, 1H, H-12β), 4.16 (m, 2H, OCH₂-CH=), 4.48 (m, 2H, OCH₂-CH=), 5.23 (m, 4H, CH₂=), 5.78 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): 17.9, 18.8, 24.2, 24.9, 26.1, 26.4, 28.1, 28.2, 29.3, 29.9, 31.9, 32.1, 32.1, 32.8, 33.2, 33.8, 35.7, 41.1, 45.6, 51.2, 57.0, 57.1, 71.2, 72.8, 74.5, 74.9, 82.2, 82.9, 116.3, 116.7, 131.9, 132.8. HRMS (EI) for C₃₂H₅₆O₅ [M⁺] calcd 520.4128 found 520.4133.

2.6. General procedure for ring-closing metathesis reaction

A solution of the secosteroidal ester **7** in dry toluene (c = 0.15– 0.20 mM) was stirred for 5 min in 80 °C and then Grubbs II catalyst (5 mol%) was added to the mixture. Reaction was stirred for 15 min under 80 °C and its progress was monitored by TLC. The total amount of catalyst (20 mol%) was added portionwise to the reaction mixture with stirring within 12 of 24 h. After cooling to room temperature, the reaction was quenched with ethyl vinyl ether and the solvent was evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (CH₂Cl₂/ MeOH: 9/1), to give macrocycle **8** as an oil.

The above method was also used for the RCM reactions of secosteroids **10a-b**, **19** and **22a-b**.

2.7. Macrocycle (8)

Yield 50 mg (65%). Oil. IR (neat) 2981, 1370, 1048 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 1.04 (d, *J* = 6.4, 3H, H-21), 1.18 (s, 3H, H-19), 1.38 (s, 3H, H-18), 2.79 (m, 1H, H-3 β), 2.80 (m, 1H, H-7 β), 3.24 (s, 3H, OCH₃), 3.26 (s, 3H, OCH₃), 3.62 (m, 4H, H-24 and H-12), 4.16 (m, 4H, OCH₂-CH=), 5.92 (td, 1H, *J* = 6.4 and 15.2, CH=), 6.04 (td, 1H, *J* = 6.4 and 15.2, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.2, 18.7, 19.4, 23.8, 24.6, 24.9, 25.0, 25.4, 25.7, 29.8, 29.9, 32.8, 35.7, 35.9, 36.4, 37.8, 42.0, 45.6, 56.8, 57.1, 60.6, 70.8, 71.1, 71.4, 73.6, 74.2, 82.5, 82.7, 122.1, 126.8. HRMS (EI) for C₃₀H₅₂O₅ [M⁺] calcd 492.3815 found 492.3821.

2.8. General procedure for deprotection with trimethylsilyl iodide

To a solution of macrocycle **8** (50 mg, 0.10 mmol) in chloroform (20 mL) was added trimethylsilyl iodide (0.1 mL). The solution was left overnight at room temperature. Methanol (5 mL) was then added to decompose any excess trimethylsilyl iodide. The solution was extracted with diethyl ether (3×10 mL), washed with water (10 mL) and saturated brine (10 mL), dried over anhydrous magnesium sulfate. Evaporation of the solvent gave a crude product, which was purified by chromatography on silica gel (CH₂Cl₂/MeOH: 95/5), to give triol **9**.

The above method was also used for deprotection of macrocycles **13a-b**.

2.9. Macrocycle (9)

Yield 47 mg (92%). Oil. IR (neat) 3429, 2950, 1606, 1089 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.88 (d, *J* = 6.4 Hz, 3H, H-21), 0.89 (s, 3H, H-19), 1.12 (s, 3H, H-18), 3.38 (m, 2H, H-12 β and H-24), 3.66 (m, 1H, H-7), 3.88 (m, 1H, H-3 β), 4.16 (m, 4H, OCH₂-CH=), 5.82 (td, *J* = 6.3 and 15.6 Hz, 1H, CH=), 5.98 (td, *J* = 6.2 Hz and 15.6 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.8, 19.7, 23.3, 23.4, 24.2, 25.6, 29.5, 29.9, 31.5, 32.2, 33.1, 35.7, 36.4, 38.2, 41.4, 45.6, 50.8, 53.8, 59.0, 63.4, 67.2, 69.0, 70.6, 71.7, 72.2, 74.6, 119.4, 125.8. HRMS (EI) for C₂₈H₄₈O₅ [M⁺] calcd 464.3502 found 464.3507.

We give the characterization data for compounds **10-15** later in the Experimental Section.

2.10. Methyl 3α , 12α -dihydroxy-7-oxo-8-oxa-B-homo-cholan-24-oate (16)

To a solution of ketone 15 (3 g, 7.14 mmol) in dry dichloromethane (30 mL) containing *p*-toluenesulfonic acid (1.11 g, 6.43 mmol) *m*-chloroperbenzoic acid (1.84 g, 10.7 mmol) was added. The solution was stirred for 24 h at room temperature. The solution was then diluted with water (20 mL) and extracted with dichloromethane (3 * 15 mL). The solution was washed successively with a 5% Na₂S₂O₃ solution, saturated brine, and water and was dried over anhydrous magnesium sulfate. The oily product, obtained by evaporation of the solvent, was purified by flash chromatography on silica gel (CH₂Cl₂/MeOH: 95/5) to afford 2.55 g of pure lactone 5 (82%) as an oil. ¹H NMR (300 MHz, CDCl₃): 0.66 (s, 3H, H-18), 0.89 (d, *I* = 6.4, 3H, H-21), 0.96 (s, 3H, H-19), 3.58 (s, 3H, OCH₃), 3.89 (m, 1H, H-3β), 4.18 (m, 1H, H-12β); ¹³C NMR (75 MHz, CDCl₃): 12.5, 14.1, 17.3, 20.7, 23.1, 24.9, 27.3, 28.9, 30.8, 31.1, 34.7, 34.9, 35.8, 36.7, 41.6, 45.0, 47.1, 51.5, 53.6, 60.4, 70.4, 71.2, 80.4, 174.6, 175.1. HRMS (EI) for C₂₅H₄₀O₆ [M⁺] calcd 436.2825 found 436.2829.

2.11. Methyl 3α , 12α -di(tert-butyldimethylsilyloxy)-7-oxo-8-oxa-B-homo-cholan-24-oate (**17**)

To a solution of lactone **16** (2 g, 4.59 mmol) in dry DMF (30 mL) containing imidazole (1.40 g, 20.6 mmol) *tert*-butyldimethylsilyl chloride (3.11 g, 20.6 mmol) was added at 0 °C. The solution was stirred for 12 h at room temperature. The solution was then diluted with water (10 mL) and extracted with dichloromethane (3 * 15 mL). The combined organic extracts were washed with 30 mL of brine, dried over anhydrous MgSO₄, and concentrated *in vacuo*. The crude product was purified by flash chromatography on silica gel (Et₂O: 100%) to give **17** (2.77 g, 91%) as an oil. ¹H NMR (300 MHz, CDCl₃): 0.59 (12H, s, Si(CH₃)₂), 0.77 (9H, s, C(CH₃)₃), 0.82 (9H, s, C(CH₃)₃), 0.89 (3H, d, *J* = 6.4 Hz, H-21), 0.94 (3H, s, H-19), 2.92 (1H, m, H-8β), 3.57 (3H, s, OCH₃); 3.85 (1H, m, H-3β), 4.12 (1H, m, H-12β); ¹³C NMR (75 MHz, CDCl₃): -4.6 (2C),

-3.4 (2C), 12.7, 15.4, 17.5, 18.1, 18.2, 23.5, 24.6 (2C), 25.8, 26.0, 27.3, 28.7, 30.6, 30.8, 31.2, 34.8, 34.9, 35.2, 35.8, 36.1, 37.3, 42.0, 42.6, 45.5, 47.1, 47.2, 51.7, 65.9, 71.4, 71.7, 79.9, 174.5, 174.8. HRMS (EI) for $C_{37}H_{68}O_6Si_2$ [M*] calcd 664.4554 found 664.4559.

2.12. 3α , 12α -di(tert-butyldimethylsilyloxy)-7,8-seco-5 β -cholan-7, 8α , 24-triol (**18**)

A solution of diprotected lactone 17 (2 g, 3.01 mmol) in dry ether (30 mL) was added in one portion to a suspension of LiAlH₄ (0.46 g, 12.0 mmol) in dry ether (20 mL) at 0 °C. After 12 h at room temperature, the reaction was guenched with H₂O (10 mL) and EtOAc (10 mL). The aqueous layer was acidified to pH 2 with diluted HCl, and layers were separated. The aqueous layer was further extracted with EtOAc (3×50 mL), and the combined organic layers were dried over anhydrous MgSO₄ and evaporated to dryness. The crude product was purified by flash chromatography on silica gel (CH₂Cl₂/MeOH: 9/1) to afford 1.74 g of pure triol 18 (90%) as an oil. ¹H NMR (300 MHz, CDCl₃): 0.00 (12H, s, Si(CH₃)₂), 0.62 (s, 3H, H-18), 0.82 (18H, s, $C(CH_3)_3$), 0.88 (3H, d, J = 6.4 Hz, H-21), 0.94 (3H, s, H-19), 3.50 (m, 1H, H-8β), 3.56 (m, 2H, H-24), 3.72 (m, 2H, H-12 β and H-3 β), 3.82 (m, 4H, H-24 and H-7); ¹³C NMR (75 MHz, CDCl₃): -4.5 (2C), -3.4 (2C), 12.9, 15.3, 18.4, 19.4 (2C), 21.0, 23.9, 25.8 (2C), 26.4, 27.2, 28.9, 29.3, 30.1, 31.7, 31.8, 32.8, 33.4, 35.9, 36.1, 37.3, 37.6, 41.4, 44.3, 47.4, 47.9, 52.1, 63.2, 65.9, 72.5, 72.9, 73.6. HRMS (EI) for $C_{36}H_{72}O_5Si_2$ [M⁺] calcd 640.4918 found 640.4923.

2.13. 3α , 12α -di(tert-butyldimethylsilyloxy)-7,24-diallyloxy-7,8-seco-5 β -cholan- 8α -ol (**19**)

To a stirred suspension of KH (112 mg, 2.34 mmol) in DMF (10 mL) at 0 °C under argon was added a solution of triol 18 (0.5 g, 0.78 mmol) in 5 mL of DMF. The reaction mixture was stirred for 15 min, and then allyl bromide (152 µL, 1.72 mmol) was added dropwise. After 48 h at room temperature, the reaction was diluted with 10 mL of Et₂O and quenched by the slow addition of 10 mL of H₂O. The combined organic extracts (3 * 10 mL) were washed with 30 mL of brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel (Et₂O: 100%) to give 18 (315 mg, 56%) as an oil. ¹H NMR (300 MHz, CDCl₃): 0.02 (12H, s, Si(CH₃)₂), 0.65 (s, 3H, H-18), 0.83 (18H, s, C(CH₃)₃), 0.89 (3H, d, J = 6.4 Hz, H-21), 0.94 (3H, s, H-19), 3.40 (m, 1H, H-3β), 3.46 (m, 1H, H-8β), 3.56 (m, 2H, H-24), 3.88 (m, 3H, H-12 β and H-7), 4.20 (m, 2H, OCH2-CH=), 4.62 (m, 2H, OCH2-CH=), 5.20 (m, 4H, CH2=), 5.87 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.4 (2C), -3.5 (2C), 13.4, 17.9, 18.3 (2C), 24.6, 24.9, 26.1 (2C), 27.5, 29.6, 30.4, 31.0, 31.8, 31.9, 32.1, 33.5, 35.2, 36.0, 36.2, 37.4, 37.5, 38.2, 41.5, 42.6, 47.3, 47.6, 47.7, 63.5, 68.4, 70.9, 72.1, 72.7, 74.5, 79.1, 116.5, 116.8, 134.7, 134.9. HRMS (EI) for $C_{42}H_{80}O_5Si_2$ [M⁺] calcd 720.5544 found 720.5549.

2.14. Macrocycle (20)

Yield 95 mg (90%). Oil. IR (neat) 3455, 2976, 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.00 (12H, s, Si(CH₃)₂), 0.64 (s, 3H, H-18), 0.83 (18H, s, C(CH₃)₃), 0.83 (3H, s, H-19), 0.88 (3H, d, J = 6.4 Hz, H-21), 3.57 (m, 1H, H-8 β), 3.72 (m, 1H, H-3 β), 3.90 (m, 1H, H-12 β), 4.36 (m, 4H, OCH₂-CH=), 4.88 (m, 4H, H-24 and H-7), 6.12 (td, J = 6.4 Hz, J = 15.3 Hz, 1H, CH=), 6.17 (td, J = 6.2 Hz, J = 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.4 (2C), -3.6 (2C), 12.7, 13.1, 15.4, 17.9, 18.3 (2C), 24.6, 24.7 (2C), 25.3, 26.0, 27.6, 28.3, 29.7, 30.4, 31.5, 31.9, 32.4, 33.0, 35.2, 36.0, 36.2, 37.5, 41.9, 44.6, 47.2, 47.6, 63.7, 66.0, 71.2, 72.3, 72.2, 72.3,

72.7, 129.2, 142.9. HRMS (EI) for $C_{40}H_{76}O_5Si_2$ [M⁺] calcd 692.5231 found 692.5235.

2.15. General procedure for deprotection with TBAF

To a solution of macrocycle **20** (100 mg, 0.14 mmol) in anhydrous THF (20 mL) was added tetra-*n*-butylammonium fluoride (0.35 mL, 1 M, 2.4 equiv). The solution was left overnight at room temperature. The solution was extracted with diethyl ether, washed with water and saturated brine, dried over anhydrous magnesium sulfate. Evaporation of the solvent gave a crude product, which was purified by chromatography on silica gel (CH₂Cl₂/ MeOH: 9/1), to give triol **21** (60 mg, 92%) as an oil.

The above method was also used for deprotection of macrocycles **24a-b**.

2.16. Macrocycle 21

IR (neat) 2965, 1340, 1059 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.66 (s, 3H, H-18), 0.86 (d, J = 6.4 Hz, 3H, H-21), 0.88 (s, 3H, H-19), 3.52 (m, 1H, H-8 β), 3.66 (m, 1H, H-3 β), 3.88 (m, 1H, H-12 β), 4.28 (m, 4H, OCH₂-CH=), 4.82 (m, 4H, H-7 and H-24), 5.95 (td, J = 6.3 and 15.4 Hz, 1H, CH=), 6.12 (td, J = 6.3 Hz and 15.4 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 13.8, 18.1, 23.7, 24.2, 24.6, 25.7, 29.5, 29.9, 31.9, 35.4, 35.7, 36.0, 36.1, 37.3, 41.4, 47.5, 51.2, 53.8, 59.0, 63.4, 67.2, 68.0, 71.7, 71.9, 72.9, 74.0, 119.8, 126.4. HRMS (EI) for C₂₈H₄₈O₅ [M⁺] calcd 464.3502 found 464.3507.

2.17. General procedure for esterification

A solution of diprotected pentahydroxysecocholane **6** or **18** (0.78 mmol), 3-butenoic acid (0.14 g, 1.63 mmol), *N*,*N*-dicyclohexylcarbodiimide (0.14 g, 0.70 mmol) and 4-dimethyl-aminopyridine (86 mg, 0.70 mmol) in dichloromethane (5 mL) was stirred at room temperature until the reaction was completed (about 12 h). The *N*,*N*-dicyclohexyl urea was filtered off and the filtrate was washed with water, 5% acetic acid solution and again water, dried over magnesium sulfate and the solvent was evaporated to afford 3-butenoate **10a** or **22a**.

The above method was also used for preparation of esters **10b-c** from **6** and **22b-c** from **18**.

2.18. Secosteroid 10a

Yield 350 mg (78%). Oil. IR (neat) 3280, 1220, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 0.96 (s, 3H, H-18), 1.02 (d, *J* = 6.4 Hz, 3H, H-21), 1.06 (s, 3H, H-19), 2.88 (m, 1H, H-7 β), 2.92 (m, 1H, H-3 β), 2.96 (m, 4H, CH₂–C=O), 3.22 (s, 3H, OCH₃), 3.24 (s, 3H, OCH₃), 4.16 (m, 4H, H-12 and H-24), 5.22 (m, 4H, CH₂=), 6.01 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.6, 20.8, 23.3, 24.3, 24.7, 26.8, 31.9, 32.2, 32.5, 32.8, 33.6, 35.6, 35.7, 37.9, 38.4, 41.0, 41.2, 45.3, 47.5, 51.4, 57.2, 57.6, 64.2, 64.6, 72.5, 75.4, 81.9, 82.6, 116.9, 117.6, 128.6, 129.1, 168.4, 169.0. HRMS (EI) for C₃₄H₅₆O₇ [M⁺] calcd 576.4026 found 576.4031.

2.19. Secosteroid 10b

Yield 360 mg (76%). Oil. IR (neat) 3418, 2961, 1145 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 0.98 (d, *J* = 6.4 Hz, 3H, H-21), 1.02 (s, 3H, H-18), 1.07 (s, 3H, H-19), 2.28 (m, 4H, CH₂–C=O), 2.34 (m, 4H, CH₂–C=), 2.81 (m, 1H, H-7 β), 2.88 (m, 1H, H-3 β), 3.23 (s, 3H, OCH₃), 3.24 (s, 3H, OCH₃), 4.13 (m, 4H, H-12 and H-24), 5.17 (m, 4H, CH₂=), 5.98 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.9, 20.3, 23.7, 23.9, 24.4, 28.8, 31.1, 32.6, 32.7, 33.2, 34.0, 34.3, 34.7, 35.5, 35.9, 37.6, 38.1, 41.2, 41.7, 44.9, 47.2, 51.3, 57.2, 57.4, 64.8,

65.1, 72.3, 75.7, 81.8, 82.3, 117.4, 118.1, 129.6, 129.9, 172.1, 172.9. HRMS (EI) for $C_{36}H_{60}O_7$ [M⁺] calcd 604.4339 found 604.4344.

2.20. Secosteroid 10c

Yield 360 mg (72%). Oil. IR (neat) 3280, 1220, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 1.00 (d, *J* = 6.4 Hz, 3H, H-21), 1.02 (s, 3H, H-18), 1.06 (s, 3H, H-19), 2.16 (m, 4H, CH₂–C=), 2.28 (m, 4H, CH₂–C=0), 2.82 (m, 1H, H-7 β), 2.92 (m, 1H, H-3 β), 3.23 (s, 3H, OCH₃), 3.25 (s, 3H, OCH₃), 4.15 (m, 4H, H-12 and H-24), 5.21 (m, 4H, CH₂=), 5.92 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.7, 20.6, 23.3, 24.1, 24.4, 24.7, 25.6, 28.3, 31.3, 32.7, 33.0, 33.4, 34.6, 34.1, 34.9, 35.7, 36.0, 37.3, 38.4, 40.9, 41.6, 44.3, 47.8, 51.6, 57.4, 57.7, 65.1, 65.6, 71.9, 75.3, 81.6, 82.9, 118.3, 118.7, 129.1, 129.8, 172.7, 173.0. HRMS (EI) for C₃₈H₆₄O₇ [M⁺] calcd 632.4652 found 632.4657.

2.21. Macrocycle 11a

Yield 60 mg (56%). Oil. IR (neat) 2951, 1732, 1370, 1150 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.98 (s, 3H, H-18), 1.04 (d, *J* = 6.4 Hz, 3H, H-21), 1.07 (s, 3H, H-19), 2.89 (m, 1H, H-7 β), 2.92 (m, 4H, CH₂–C=O), 2.94 (m, 1H, H-3 β), 3.23 (s, 3H, OCH₃), 3.24 (s, 3H, OCH₃), 4.18 (m, 4H, H-12 and H-24), 5.92 (td, *J* = 6.3 and 15.1 Hz, 1H, CH=), 6.09 (td, *J* = 6.3 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.4, 20.6, 23.1, 24.6, 24.9, 27.1, 31.6, 32.4, 32.7, 33.1, 33.8, 34.9, 36.3, 37.1, 38.3, 41.6, 42.0, 45.4, 47.1, 52.3, 56.9, 57.2, 61.3, 67.2, 72.8, 75.7, 82.4, 82.9, 119.9, 126.7, 171.9, 172.6. HRMS (EI) for C₃₂H₅₂O₇ [M⁺] calcd 548.3713 found 548.3716.

2.22. Macrocycle 11b

Yield 55 mg (48%). Oil. IR (neat) 2965, 1728, 1609, 1502 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 1.01 (d, *J* = 6.4 Hz, 3H, H-21), 1.04 (s, 3H, H-18), 1.12 (s, 3H, H-19), 2.35 (m, 4H, CH₂– C=O), 2.44 (m, 4H, CH₂–C=), 2.83 (m, 1H, H-7 β), 2.90 (m, 1H, H-3 β), 3.24 (s, 3H, OCH₃), 3.25 (s, 3H, OCH₃), 4.07 (m, 4H, H-12 and H-24), 5.86 (td, *J* = 6.4 and 15.3 Hz, 1H, CH=), 6.01 (td, *J* = 6.3 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.9, 21.1, 23.4, 24.9, 25.1, 27.8, 31.4, 32.1, 32.9, 33.6, 34.2, 34.9, 35.4, 36.1, 36.8, 37.4, 38.0, 42.2, 42.6, 45.9, 47.4, 52.7, 57.0, 57.7, 61.8, 68.2, 72.8, 74.9, 82.1, 82.7, 119.4, 126.1, 172.8, 173.0. HRMS (EI) for C₃₄H₅₆O₇ [M⁺] calcd 576.4026 found 576.4031.

2.23. Secosteroid **22a**

Yield 500 mg (82%). Oil. IR (neat) 3424, 1338, 1130 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 0.00 (12H, s, Si(CH₃)₂), 0.66 (s, 3H, H-18), 0.85 (18H, s, C(CH₃)₃), 0.97 (3H, d, J = 6.4 Hz, H-21), 1.02 (3H, s, H-19), 3.08 (m, 1H, H-8 β), 3.44 (m, 4H, CH₂–C=O), 3.55 (m, 2H, H-12 and H-3 β), 3.90 (m, 4H, H-24 and H-7 β), 5.18 (m, 4H, CH₂=), 5.83 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): –4.9 (2C), -3.6 (2C), 12.8, 17.5, 18.0, 20.9 (2C), 23.5, 25.8 (2C), 26.0, 27.6, 29.2, 30.3, 31.2, 31.5, 35.1, 37.5, 37.6, 39.0, 41.4, 41.6, 42.5, 42.9, 44.1, 46.3, 47.5, 47.8, 47.9, 49.6, 50.5, 62.2, 63.2, 72.6, 72.7, 73.4, 118.3 (2C), 130.5 (2C), 174.2, 174.4. HRMS (EI) for C₄₄H₈₀O₇Si₂ [M⁺] calcd 776.5443 found 776.5449.

2.24. Secosteroid 22b

Yield 470 mg (75%). Oil. IR (neat) 3280, 1356, 1048 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 0.00 (12H, s, Si(CH₃)₂), 0.64 (s, 3H, H-18), 0.83 (18H, s, C(CH₃)₃), 0.89 (3H, s, H-19), 0.94 (3H, d, J = 6.4 Hz, H-21), 2.40 (m, 8H, CH₂-C=O and CH₂-C=), 3.42 (m, 1H, H-8 β), 3.66 (m, 1H, H-12 β), 3.89 (m, 1H, H-3 β), 4.08 (m, 4H, H-24 and

H-7), 5.01 (m, 4H, CH₂=), 5.80 (m, 2H, CH=); 13 C NMR (75 MHz, CDCl₃): -4.8 (2C), -3.6 (2C), 12.8, 14.2, 17.5, 18.2, 20.2 (2C), 21.0, 22.7, 24.3 (2C), 25.5, 25.9, 26.3, 28.5, 28.9, 32.6, 33.9, 35.0, 35.3, 36.0, 37.2, 44.6, 47.2, 47.5, 51.6, 53.4, 53.8, 60.5, 68.3, 69.3, 72.2, 72.5, 73.8, 79.4, 115.4, 115.5, 136.3, 136.4, 1724.2, 173.4. HRMS (EI) for C₄₄H₈₄O₇Si₂ [M⁺] calcd 804.5756 found 804.5761.

2.25. Secosteroid 22c

Yield 480 mg (74%). Oil. IR (neat) 3429, 1330, 1042 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): 0.00 (12H, s, Si(CH₃)₂), 0.65 (s, 3H, H-18), 0.83 (18H, s, C(CH₃)₃), 0.88 (3H, s, H-19), 0.93 (3H, d, *J* = 6.4 Hz, H-21), 2.04 (4H, m, CH₂–C=), 2.27 (m, 4H, CH₂–C=0), 3.53 (m, 1H, H-8β), 3.77 (m, 1H, H-12β), 3.87 (m, 1H, H-3β), 4.03 (m, 4H, H-24 and H-7), 5.01 (m, 4H, CH₂=), 5.74 (m, 2H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.5 (2C), -3.6 (2C), 12.9, 17.7, 18.2 (2C), 24.2, 24.5 (2C), 24.6, 25.4, 25.9, 27.2, 27.7, 29.5, 29.7, 30.3, 31.4, 31.8, 33.1, 33.7, 34.0, 34.4, 35.0, 35.9, 36.1, 37.4, 37.5, 38.4, 39.2, 41.7, 42.4, 42.9, 44.8, 47.5, 47.7, 64.8, 64.9, 70.3, 72.2, 72.6, 115.3, 115.4, 137.8, 173.7, 173.8. HRMS (EI) for C₄₈H₈₈O₇Si₂ [M⁺] calcd 832.6069 found 832.6074.

2.26. General procedure for the reduction of 11a-b and 23a-b

A flask equipped with a magnetic stirring bar, an argon outlet and a condenser was charged with NaBH4 (90 mg, 0.40 mmol) and anhydr. THF (7 mL) – diglyme (3 mL) under argon. The solution was cooled at 0 °C and then a solution composed of boron trifluoride etherate (0.42 g, 3 mmol), macrocycle **11a-b** or **23a-b** (0.18 mmol) and anhydr. THF (5 mL) was added. After completion of the reaction (TLC), it was quenched by addition of 2 N hydrochloric acid (1 mL) and water (10 mL), the product was extracted with ether (3 × 20 mL). The extracts were dried over MgSO4, filtered and then concentrated under vacuum. The residue was chromatographed on silica gel (Et2O-petroleum ether 1:1).

2.27. Macrocycle 12a

Yield 52 mg (56%). Oil. IR (neat) 3280, 1220, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 1.02 (s, 3H, H-18), 1.06 (d, *J* = 6.4 Hz, 3H, H-21), 1.14 (s, 3H, H-19), 2.26 (m, 4H, CH₂–C=), 2.87 (m, 1H, H-7 β), 2.92 (m, 1H, H-3 β), 3.24 (s, 3H, OCH₃), 3.25 (s, 3H, OCH₃), 3.42 (m, 4H, H-12 and H-24), 5.88 (td, *J* = 6.4 and 15.3 Hz, 1H, CH=), 6.02 (td, *J* = 6.3 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.9, 20.4, 23.6, 24.8, 25.1, 28.4, 31.8, 32.4, 32.6, 32.9, 33.4, 34.6, 36.5, 37.3, 39.1, 41.9, 42.6, 46.2, 47.3, 52.1, 56.4, 57.5, 66.8, 67.4, 70.2, 71.1, 72.4, 75.2, 82.1, 82.6, 119.8, 126.3. HRMS (EI) for C₃₂H₅₆O₅ [M⁺] calcd 520.4128 found 520.4133.

2.28. Macrocycle 12b

Yield 48 mg (48%). Oil. IR (neat) 2981, 1370, 1048 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 1.01 (d, *J* = 6.4 Hz, 3H, H-21), 1.04 (s, 3H, H-18), 1.12 (s, 3H, H-19), 2.35 (m, 4H, CH₂-C=O), 2.44 (m, 4H, CH₂-C=), 2.83 (m, 1H, H-7β), 2.90 (m, 1H, H-3β), 3.24 (s, 3H, OCH₃), 3.25 (s, 3H, OCH₃), 4.07 (m, 4H, H-12 and H-24), 5.86 (td, *J* = 6.4 and 15.3 Hz, 1H, CH=), 6.01 (td, *J* = 6.3 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.7, 21.3, 23.6, 24.5, 24.9, 27.6, 29.6, 31.7, 32.4, 33.1, 34.3, 34.7, 35.1, 36.7, 36.9, 37.1, 37.8, 41.9, 42.3, 44.7, 47.2, 51.9, 57.3, 57.9, 62.3, 69.0, 70.4, 71.6, 73.0, 74.3, 81.9, 82.5, 121.1, 126.3. HRMS (EI) for C₃₄H₆₀O₅ [M⁺] calcd 548.4441 found 548.4446.

2.29. Macrocycle 13a

Yield 45 mg (92%). Oil. IR (neat) 2984, 1374, 1038 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 1.01 (s, 3H, H-18), 1.06 (d, *J* = 6.4 Hz, 3H, H-21), 1.16 (s, 3H, H-19), 2.24 (m, 4H, CH₂–C=), 3.12 (m, 1H, H-7 β), 3.16 (m, 1H, H-3 β), 3.52 (m, 4H, H-12 and H-24), 5.89 (td, *J* = 6.1 and 15.4 Hz, 1H, CH=), 6.04 (td, *J* = 6.3 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.7, 20.6, 23.3, 24.1, 25.3, 28.6, 31.7, 32.6, 32.9, 33.1, 33.7, 34.2, 36.7, 37.8, 39.2, 41.6, 42.1, 44.9, 46.9, 52.4, 62.3, 65.1, 66.5, 68.9, 71.3, 77.8, 72.3, 73.9, 119.6, 126.1. HRMS (EI) for C₃₀H₅₂O₅ [M⁺] calcd 492.3815 found 492.3819.

2.30. Macrocycle 13b

Yield 40 mg (90%). Oil. IR (neat) 3429, 2950, 1606, 1089 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.99 (s, 3H, H-18), 1.02 (d, J = 6.4 Hz, 3H, H-21), 1.10 (s, 3H, H-19), 2.14 (m, 4H, CH₂-C=), 3.22 (m, 1H, H-7 β), 3.31 (m, 1H, H-3 β), 4.02 (m, 8H, CH₂-O, H-12 and H-24), 5.72 (td, J = 6.1 and 15.4 Hz, 1H, CH=), 5.97 (td, J = 6.2 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 18.4, 21.6, 23.9, 24.1, 24.7, 27.3, 29.2, 31.4, 32.0, 32.7, 33.9, 34.6, 35.2, 36.9, 37.1, 37.4, 37.9, 41.6, 43.0, 44.2, 47.1, 52.1, 63.4, 68.8, 70.6, 71.9, 73.1, 73.9, 80.8, 81.7, 119.6, 125.9. HRMS (EI) for C₃₂H₅₆O₅ [M⁺] calcd 520.4128 found 520.4134.

2.31. Macrocycle 23a

Yield 80 mg (54%). Oil. IR (neat) 3280, 1730, 1220, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.00 (12H, s, Si(CH₃)₂), 0.65 (s, 3H, H-18), 0.83 (18H, s, C(CH₃)₃), 0.94 (3H, d, *J* = 6.4 Hz, H-21), 1.02 (3H, s, H-19), 3.38 (m, 1H, H-8β), 3.44 (m, 4H, CH₂– C=O), 3.55 (m, 2H, H-12β and H-3β), 3.90 (m, 4H, H-24 and H-7), 5.76 (td, *J* = 6.4 and 15.2 Hz, 1H, CH=), 6.02 (td, *J* = 6.1 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.4 (2C), -3.5 (2C), 12.6, 15.3, 17.9, 18.3 (2C), 24.3, 24.7 (2C), 25.1, 26.2, 27.4, 29.8, 30.6, 31.2, 31.6, 31.9, 33.2, 35.4, 36.1, 36.3, 36.6, 37.8, 41.3, 42.1, 46.2, 47.0, 47.3, 50.7, 52.6, 65.4, 66.2, 72.5, 72.9, 76.5, 127.6, 133.9, 174.4, 174.6. HRMS (EI) for C₄₂H₇₆O₇Si₂ [M⁺] calcd 748.5130 found 748.5135.

2.32. Macrocycle 23b

Yield 78 mg (50%). Oil. IR (neat) 3448, 1732, 1607, 1041 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.00 (12H, s, Si(CH₃)₂), 0.64 (s, 3H, H-18), 0.83 (18H, s, C(CH₃)₃), 0.92 (3H, d, *J* = 6.4 Hz, H-21), 1.22 (3H, s, H-19), 2.31 (m, 8H, (CH₂)₂–C=), 3.36 (m, 1H, H-8 β), 3.42 (m, 4H, CH₂–C=O), 3.52 (m, 2H, H-12 β and H-3 β), 3.86 (m, 4H, H-24 and H-7), 5.48 (td, *J* = 6.4 and 15.4 Hz, 1H, CH=), 5.93 (td, *J* = 6.2 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.6 (2C), -3.6 (2C), 12.8, 18.1, 18.6 (2C), 22.6, 24.3, 25.8 (2C), 26.0, 26.2, 26.7, 26.9, 29.6, 31.3, 31.9, 34.1, 34.7, 34.9, 35.5, 35.6, 35.9, 37.2, 37.4, 38.0, 40.9, 41.6, 44.5, 46.3, 47.2, 47.5, 53.5, 64.8, 68.0, 70.5, 72.5, 72.7, 128.4, 129.4, 172.1, 173.2. HRMS (EI) for C₄₄H₈₀O₇Si₂ [M⁺] calcd 776.5443 found 776.5448.

2.33. Macrocycle 24a

Yield 40 mg (52%). Oil. IR (neat) 3420, 1264, 905 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.00 (12H, s, Si(CH₃)₂), 0.66 (s, 3H, H-18), 0.85 (18H, s, C(CH₃)₃), 1.02 (3H, d, J = 6.4 Hz, H-21), 1.08 (3H, s, H-19), 3.34 (m, 1H, H-8 β), 3.43 (m, 2H, H-12 β and H-3 β), 3.66 (m, 4H, CH₂–O), 3.90 (m, 4H, H-24 and H-7), 5.72 (td, J = 6.3 and 15.2 Hz, 1H, CH=), 5.96 (td, J = 6.3 Hz and 15.4 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.6 (2C), -3.5 (2C), 12.2, 14.9, 18.1, 18.8 (2C), 24.1, 24.9 (2C), 25.4, 26.0, 27.5, 28.9, 30.1, 31.4,

31.8, 32.0, 33.3, 35.6, 36.3, 36.7, 37.2, 38.4, 39.6, 41.3, 41.7, 46.6, 47.1, 51.2, 52.3, 66.2, 68.1, 70.6, 70.9, 72.4, 73.1, 76.4, 127.4, 129.9. HRMS (EI) for $C_{42}H_{80}O_5Si_2$ [M⁺] calcd 720.5544 found 720.5549.

2.34. Macrocycle 24b

Yield 35 mg (46%). Oil. IR (neat) 3435, 2990, 1058 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.00 (12H, s, Si(CH₃)₂), 0.66 (s, 3H, H-18), 0.89 (18H, s, C(CH₃)₃), 0.98 (3H, d, J = 6.4 Hz, H-21), 1.17 (3H, s, H-19), 2.31 (m, 8H, (CH₂)₂–C=), 3.41 (m, 1H, H-8β), 3.48 (m, 2H, H-12β and H-3β), 3.67 (m, 4H, CH₂–O), 3.89 (m, 4H, H-24 and H-7), 5.42 (td, J = 6.1 and 15.4 Hz, 1H, CH=), 5.87 (td, J = 6.2 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): -4.9 (2C), -3.6 (2C), 12.4, 18.7, 19.1 (2C), 23.1, 24.5 (2C), 25.6, 25.9, 26.4, 26.8, 27.1, 29.2, 31.4, 32.0, 34.1, 34.3, 34.9, 35.1, 35.8, 36.6, 37.4, 37.5, 38.6, 39.8, 41.3, 43.2, 45.9, 46.2, 47.4, 54.1, 64.2, 67.9, 69.9, 70.5, 71.1, 72.8, 73.2, 128.1, 129.6. HRMS (EI) for C₄₄H₈₄O₅Si₂ [M⁺] calcd 748.5857 found 748.5861.

2.35. Macrocycle 25a

Yield 28 mg (95%). Oil. IR (neat) 3410, 2940, 1608 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.64 (s, 3H, H-18), 1.04 (3H, d, *J* = 6.4 Hz, H-21), 1.12 (3H, s, H-19), 3.32 (m, 2H, H-12 β and H-3 β), 3.42 (m, 1H, H-8 β), 3.48 (m, 4H, CH₂–O), 3.72 (m, 4H, H-24 and H-7), 5.67 (td, *J* = 6.4 and 15.3 Hz, 1H, CH=), 5.89 (td, *J* = 6.3 Hz and 15.4 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 14.1, 17.9, 23.6, 24.4, 27.1, 27.8, 29.1, 31.0, 31.6, 31.7, 32.4, 33.7, 34.8, 36.2, 36.9, 37.6, 38.9, 45.7, 47.8, 51.6, 52.1, 65.8, 67.9, 70.4, 71.2, 72.6, 73.4, 75.9, 126.9, 129.1. HRMS (EI) for C₃₀H₅₂O₅ [M⁺] calcd 492.3815 found 492.3821.

2.36. Macrocycle 25b

Yield 25 mg (96%). Oil. IR (neat) 3386, 1070, 820 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (major isomer): 0.67 (s, 3H, H-18), 0.99 (3H, d, J = 6.4 Hz, H-21), 1.11 (3H, s, H-19), 2.16 (m, 8H, (CH₂)₂-C=), 3.21 (m, 2H, H-12β and H-3β), 3.39 (m, 4H, CH₂-O), 3.45 (m, 1H, H-8β), 3.67 (m, 4H, H-24 and H-7), 5.48 (td, J = 6.2 and 15.4 Hz, 1H, CH=), 5.89 (td, J = 6.2 Hz and 15.3 Hz, 1H, CH=); ¹³C NMR (75 MHz, CDCl₃): 12.9, 18.4, 23.3, 24.8, 25.9, 27.1, 29.3, 30.7, 32.4, 33.7, 34.1, 34.9, 35.5, 35.9, 36.2, 37.1, 38.0, 38.9, 42.2, 45.6, 46.3, 47.1, 53.9, 64.6, 68.5, 69.1, 70.7, 71.4, 72.6, 73.7, 127.2, 129.8. HRMS (EI) for C₃₂H₅₆O₅ [M⁺] calcd 520.4128 found 520.4132.

3. Results and discussion

Olefin metathesis has become one of the most powerful and attractive tools for the formation of carbon-carbon double bonds and is widely used in organic synthesis. The spectacular improvements in this reaction achieved over the last two decades are well known to most chemists. An increasing number of papers devoted to applications of olefin metathesis in the synthesis of natural products are observed [38–44]. Although ring-closing metathesis (RCM) is known to be a powerful tool for the preparation of macrocycles [45–49], in the field of steroid chemistry, only few syntheses have been reported based on metathesis reactions [50].

For some years, we have been interested to develop new synthetic approaches to prepare secosteroidal molecules. We recently described a simple preparation of 12,13- (Scheme 2) and 7,8-secosteroids (Scheme 4) possessing a macrocycle using a ring-closing metathesis reaction as the key step [51]. Indeed, this combination



Reaction conditions : (a) MeOH, PTSA, Δ, 2 h, 99%; (b) CH₃I, NaH, THF, r.t., 94%; (c) PCC, MW,5 min, 70%; (d) *m*-CPBA, PTSA, CH₂Cl₂, r.t., 24 h, 98%; (e) LiAlH₄, THF, 0 °C to r.t., 12 h, 88%; (f) allyl bromide, KH, DMF, 0 °C to r.t., 12 h, 58%; (g) Grubbs-II catalyst (20 mol%), CH₂Cl₂, r.t., 12 h, 65%; (h) ISi(CH₃)₃, CHCl₃, r.t., 24 h, 92%.

Scheme 2. Synthesis of a 12,13-secosteroidal macrocycle 9 from cholic acid through 8 steps sequence.

of secocholanic skeleton with varied types of macrocycles, produces high levels of skeletal diversity and complexity.

Our strategy is based on a sequential ring-expansion/ring-opening and on a ring closing metathesis reaction [51]. In the two previously described syntheses (Schemes 2 and 4), we prepared the RCM substrates from cholic acid, a commercial bile acid both inexpensive and readily available. In the case of 12,13-secosteroidal macrocycles matching a cis A/B ring junction, the key reactions are depicted in Scheme 2. Simple esterification of cholic acid 1 led to methyl cholate [52], which was methylated with methyl iodide-sodium hydride in THF affording methyl 3α , 7α -dimethoxy cholate 3 in a yield of 94%. Simultaneous protection of the secondary hydroxyl groups at C-3 and C-7 was needed prior to the reductive opening of the lactone ring. Microwave (MW) [53] irradiation of 3 with pyridinium chlorochromate furnished ketone 4 very quickly, in a few minutes, and in 70% yield. Baeyer-Villiger oxidation of ketocholane 4 led to lactone 5 as the single regioisomer, as a result of a higher migration aptitude of the guaternary C-13 compared to the secondary C-11. Next, the simultaneous reduction of both the lactone moiety on ring C and the ester function at C-24 afforded the diprotected pentahydroxysecocholane 6 in 88% yield. Diallylation of primary alcohols at C-24 and C-12 produced the corresponding diallyl adduct **7** in moderate yield (58%).

The key step of the synthesis, the RCM reaction of **7** was accomplished efficiently with the use of a catalytic amount of Grubbs-II catalyst (20 mol%) in toluene at 80 °C providing secosteroid **8** in good yield (65%). However, the reaction was not stereoselective, a mixture of geometric isomers **8** was obtained with the E isomer. The structure of this macrocycle was completely characterized by NMR (400 MHz) and mass spectroscopy methods (see experimental part). A 8:2 ratio of isomers was obtained with the *trans* (*J* = 15.2 Hz) isomer prevailing over the *cis* (*J* = 10.6 Hz) isomer.

Finally, removal of methoxy groups of **8** was carried out with trimethylsilyl iodide [33] to afford the desired compound **9** in 92% yield.

Encouraged by this result and to show that this ring enlargement is generally applicable, different types of olefins were subjected to metathesis reaction (Scheme 3 and Table 1). We present now results of our further study on metathesis of secosteroidal esters with various sized acid chain (C_4 , C_5 and C_6). Dienes **10a-c** were prepared in high yields by reaction of steroid **6** with alkenoic acids in the presence of DCC and DMAP. All esters



 $\begin{array}{l} \mbox{Reaction conditions : (a) $CH_2=CH(CH_2)_n$COOH, DCC, DMAP, 0 °C to r.t., 12 h; $(b) Grubbs-II catalyst (20 mol%), see table 1; (c) $NaBH_4$, $BF_3.Et_2O$, THF-diglyme, 0 °C, 4 h; (d) ISi(CH_3)_3$, CHCl_3, r.t., 24 h. $ \end{array}$

Scheme 3. Synthesis of 12,13-secosteroidal macrocycles 13 from diprotected secosteroid 6.

Table 1Metathesis reactions via Scheme 3.

Entry	Olefin	п	Cat ^a	Condition	11: yield (%)
1	10a	1	А	CH ₂ Cl ₂ , rt, 24 h	18
2	10a	1	В	CH2Cl2, rt, 24 h	29
3	10a	1	В	MePh, Δ , 4 h	56
4	10b	2	Α	CH ₂ Cl ₂ , rt, 24 h	11
5	10b	2	В	CH ₂ Cl ₂ , rt, 24 h	23
6	10b	2	В	MePh, Δ , 4 h	48
7	10c	3	Α	CH ₂ Cl ₂ , rt, 24 h	-
8	10c	3	В	CH ₂ Cl ₂ , rt, 24 h	-
9	10c	3	В	MePh, Δ , 4 h	-

^a A = $RuCl_2(PCy_3)_2$ = CHCHCMe₂; B = (IMES)(PCy)₃Cl₂Ru = CHPh; IMES = 1,3-dimesityl-4,5-dihydroimidazol-2-ylidene.

obtained were found to be stable and were subjected to RCM reactions. According to the literature, synthesis of unsaturated lactones through RCM has been accomplished using either Grubbs, first or second generation, or Hoveyda catalyst.

We decided to test two commercially available metathesis promoters: Grubbs first-generation (GI) and Grubbs second-generation GII (Table 1). In all cases, the Grubbs catalyst of second generation was more efficient than the first generation. We carried out the first experiments under relatively mild conditions: 20 mol% of catalyst in dry and degassed dichloromethane at room temperature. Using these experimental conditions, we observed fair yields for **11a** and **11b**. The yields of the cyclization products of **10a** and **10b** were improved by increasing the temperature (Table 1, entries 3 and 6). Although many successful applications of RCM to the synthesis of macrocycles with n = 1 and n = 2, in the case of n = 3, the RCM did not occur and the remaining material was mainly the unreacted starting diene **10c**.

Otherwise, the reaction was not stereoselective. All metathesis products were obtained as mixtures of geometric E/Z-isomers with the E isomer predominating. The structure of these new secosteroidal macrocycles **11a-b** was completely characterized by NMR (400 MHz) and mass spectroscopic methods (see experimental part).

In the following step, in order to obtain ether-linked macrocycles as for compound **9** (n = 0, see Scheme 2), we reduced esters **11a** and **11b** to the corresponding ethers **12a** and **12b**, was carried out using sodium borohydride and boron trifluoride, in diglymetetrahydrofuran at 0 °C during 4 h. Thus, in the two cases, reduction according to the Pettit and Piatak procedure [54], afforded the expected macrocycle derivatives **12a** and **12b** in satisfactory yields : 56% for **12a** and 48% for **12b**.

Finally, removal of methoxy groups of macrocycles **12a** and **12b** was carried out with trimethylsilyl iodide [55,56] to afford the desired compounds **13a** and **13b** in good yields. The problem of formation of isomers can be, of course, overcome by hydrogenation of the newly formed double bond.

As a continuation of our synthetic and stereochemical studies on secosteroidal systems containing a macrocycle, a similar strategy was used to obtain 7,8-secosteroidal macrocycles (Scheme 4). The regioselective oxidation of the hydroxyl group at C-7 was performed with NBS [57]. The resulting 7-keto derivative **14** was then converted into cholate **15** in good yield (95% yield over two steps).

As expected, the Baeyer–Villiger oxidation of ketocholane **15** furnished exclusively regioisomer **16**, as a result of the favored

Reaction conditions : (a) NBS, H₂O, NaHCO₃, 12 h at r.t. then 2h at 80-85 °C, 95%; (b) MeOH, PTSA, Δ , 2h, 100%; (c) *m*-CPBA, PTSA, CH₂Cl₂, r.t., 24 h, 82%; (d) TBDMSCl, imidazole, DMF, r.t., 12 h, 91%; (e) LiAlH₄,THF, 0 °C to r.t., 12 h, 90%; (f) allyl bromide, KH, DMF, 0 °C to r.t., 12 h, 56%; (g) Grubbs-II catalyst (20 mol%), CH₂Cl₂, r.t., 12 h, 90%; (h) TBAF, THF, r.t., 12 h, 92%.

Scheme 4. Synthesis of a 7,8-secosteroidal macrocycle 21 from cholic acid through 8 steps sequence.

migration of the tertiary C-8 compared to the secondary C-6. Consecutively, a reaction sequence consisted in TBDMS protection of OH functionalities, followed by the reductive opening of the lactone ring with simultaneous reduction of the ester function at C-24 and finally regioselective diallylation reaction of primary alcohols at C-24 and C-7 led to the diallyl secocholane **19** in 38% overall yield (over four steps). The RCM was promoted by the second generation Grubbs' catalyst and the reaction was very efficient. Using the same conditions as described above for **9**, we obtained the macrocyclic product **20** in 90% yield. Here too, this latter **20** was completely characterized by NMR and mass spectroscopic methods (see experimental part). The E/Z 9:1 ratio observed in the ¹H NMR spectrum of the crude substance was strongly in favor of the *E*-isomer (*J* = 15.6 Hz) with small amounts of sterically less favored *Z*-double bond (*J* = 10.4 Hz).

The 3- and 12-TBS ethers on the A and C rings of **20** were deprotected in a standard manner to afford the corresponding secosteroidal alcohol **21** in good yield.

We have then turned our attention toward the synthesis of new macrocycles by introducing ester functionalities on the secosteroidal alcohol **18** (Scheme 5). An interesting feature of this methodology is the possibility for the expansion of the ring size by adjusting the length of the acid source used for the synthesis of RCM precursors **22**.

Thus, three alkenoates (compounds **22a-c**) were prepared in good yields by reaction of diprotected pentadihydroxysecocholane **18** with alkenoic acids in presence of DCC and DMAP. Here too, esters **22a-c** were found to be stable and were subjected to RCM reactions using Grubbs first-generation (GI) or second-generation (GII) as catalyst (see Table 2).

It is worth pointing out similar results with the work we described above for the synthesis of 12,13-secosteroidal macrocycles. Indeed, the cyclization of **22** was observed for n = 1 and n = 2 but without any success for n = 3. Here too, the remaining material was mainly the unreacted starting diene **22c**. It is interesting to note that the yields and the isomer ratio observed for **23a** and **23b** are exactly or quite the same as those reported for the 12,13-secosteroidal series. Otherwise, in some experiments, the reaction temperature was raised to 40 °C without significant change of reaction course. Only the use of the catalyst of second generation (Grubbs II) at 80 °C in dry and degassed toluene proved effective in the RCM products formation **23a** or **23b** (Table 2,

Reaction conditions : (a) $CH_2=CH(CH_2)_nCOOH$, DCC, DMAP, 0 °C to r.t., 12 h; (b) Grubbs-II catalyst (20 mol%), see table 2; (c) $NaBH_4$, BF_3 . Et_2O , THF-diglyme, 0 °C, 4 h; (d) TBAF, THF, r.t., 12 h.

Scheme 5. Synthesis of 7,8-secosteroidal macrocycles 25 from diprotected secosteroid 18.

Table 2Metathesis reactions via Scheme 5.

Entry	Olefin	п	Cat ^a	Condition	23: yield (%)
1	22a	1	А	CH ₂ Cl ₂ , rt, 24 h	12
2	22a	1	В	CH ₂ Cl ₂ , rt, 24 h	26
3	22a	1	В	MePh, Δ, 4 h	54
4	22b	2	А	CH ₂ Cl ₂ , rt, 24 h	9
5	22b	2	В	CH ₂ Cl ₂ , rt, 24 h	24
6	22b	2	В	MePh, Δ, 4 h	50
7	22c	3	А	CH ₂ Cl ₂ , rt, 24 h	-
8	22c	3	В	CH ₂ Cl ₂ , rt, 24 h	-
9	22c	3	В	MePh, Δ , 4 h	-

^a A = RuCl₂(PCy₃)₂==CHCHCMe₂; B = (IMES)(PCy)₃Cl₂Ru==CHPh; IMES = 1,3-dimesityl-4,5-dihydroimidazol-2-ylidene.

entries 3 and 6). The influence of other reaction conditions on the reaction course, e.g. dilution (tested concentration: 0.5–1.5 mM), mode of reagent addition, appeared to be much less important.

Further synthesis of macrocycles consisted of reduction of the obtained lactones (**23a**, **23b**) with a metal hydride. As described above for the 12,13-secosteroidal series, reduction of **23a** and **23b** using sodium borohydride and boron trifluoride in diglymetetrahydrofuran [54] afforded the corresponding ethers **24a** and **24b** in 52% and 46% yield respectively.

In the final step, the 3α - and 12α -hydroxyl groups in compounds **24a** and **24b** were deprotected with TBAF in a standart manner to afford the corresponding alcohols **25a** and **25b** in excellent yield (see experimental part).

In conclusion, in this paper, it was proven that the RCM strategy can be successfully applied to the synthesis of diverse secosteroids from commercially available cholic acid. The key steps of this synthesis are an oxidative ring-expansion/ring-opening sequence and a ring-closing metathesis. The subsequent reduction of the obtained RCM products afforded new macrocycle derivatives. Tests to check the biological activity of these steroids and application of this strategy to obtain new structures are being conducted by our group.

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References

- [1] Feldman D, Glorieux FH, Pike JW. Vitamin D. New York: Academic Press; 1997.
- [2] Kirson I, Zaretskii Z, Glotter E, Withaphysalin C. A naturally occurring 13,14seco-steroid. J Chem Soc Perkin Trans 1976;11:1244–7.
- [3] Aiello A, Fattorusso E, Menna M. Steroids from sponges: recent reports. Steroids 1999;64(10):687–714.
- [4] Stonick VA. Marine polar steroids. Russ Chem Rev 2001;70(8):673-715.
- [5] Norman AW, Bouillon R, Thomasset M, editors. Vitamin D, a pluripotent steroid hormone: structural studies, molecular endocrinology and clinical applications. Berlin: Walter de Gruyter; 1994.
- [6] Pika J, Andersen RJ. Blancasterol, a cytotoxic 9,11-secosteroid isolated from the Northeastern Pacific marine sponge of the species Pleraplysilla. Tetrahedron 1993;49(39):8757–60.
- [7] Seo Y, Cho KW, Chung H, Lee HS, Shin J. New secosteroids from a gorgonian of the genus *Muricella*. J Nat Prod 1998;61(11):1441–3.
- [8] Rueda A, Zubia E, Ortega MJ, Carballo JL, Salva J. New metabolites from the sponge Spongia agaricina. J Nat Prod 1998;61(2):258–61.
- [9] Dopeso J, Quinoa E, Riguera R, Debitus C, Bergquist PR. Euryspongiols: ten new highly hydroxylated 9,11-secosteroids with antihistaminic activity from the sponge *Euryspongia* sp. Stereochemistry and reduction. Tetrahedron 1994;50(12):3813–28.
- [10] Morris LA, Christie EM, Jaspars M, Van Ofwegen LP. A bioactive secosterol with an unusual A- and B-ring oxygenation pattern isolated from an Indonesian soft coral *Lobophytum* sp. J Nat Prod 1998;61(4):538–41.
- [11] Zerhouni NA, Maes M, Sultan C, Rothwell S, Migeon CJ. Selective inhibition by secosteroids of 5α-reductase activity in human sex skin fibroblasts. Steroids 1979;33(3):277–85.
- [12] Penning TM, Covey DF, Talalay P. Irreversible inactivation of Δ5-3-ketosteroid isomerase of *Pseudomonas testosteroni* by acetylenic suicide substrates. Mechanism of formation and properties of the steroid-enzyme adduct. J Biol Chem 1981;256(13):6842–50.

- [13] Penning TM. Inactivation of Δ5-3-ketosteroid isomerase(s) from beef adrenal cortex by β, γ-acetylenic ketosteroids. Steroids 1982;39(3):301–11.
- [14] Vazquez MH, Tezon JG, Blaquier JA. Studies on the mechanism of the antiandrogenic effect of a putative 5α -reductase inhibitor. J Steroid Biochem 1987;28(2):227–31.
- [15] Hu Y, Covey DF. Synthesis of 1,10-seco-5α-estr-1-ynes: potential mechanismbased inhibitors of 3α- and 3β-hydroxysteroid dehydrogenases. J Chem Soc Perkin Trans 1993;1(4):417-22.
- [16] Reich IL, Lardy H, Wei Y, Marwah P, Kneer N, Powell DR. Ergosteroids III. Syntheses and biological activity of seco-steroids related to dehydroepiandrosterone. Steroids 1998;63(10):542–53.
- [17] Marsault E, Peterson ML. Macrocycles are great cycles: applications, opportunities, and challenges of synthetic macrocycles in drug discovery. J Med Chem 2011;54(7):1961–2004.
- [18] Madsen CM, Clausen MH. Biologically active macrocyclic compounds from natural products to diversity-oriented synthesis. Eur J Org Chem 2011:3107–15.
- [19] Adessi C, Soto C. Converting a peptide into a drug: strategies to improve stability and bioavailability. Curr Med Chem 2002;9(9):963–78.
- [20] Madala PK, Tyndall JDA, Nall T, Fairlie DP. Update 1 of: proteases universally recognize beta strands in their active sites. Chem Rev 2010;110(6):1–31.
- [21] Loughlin WA, Tyndall JDA, Glenn MP, Hill TA, Fairlie DP. Update 1 of: betastrand mimetics. Chem Rev 2010;110(6):32–69.
- [22] Jones RM, Boatman PD, Semple G, Shin YJ, Tamura SY. Clinically validated peptides as templates for de novo peptidomimetic drug design at G-proteincoupled receptors. Curr Opin Pharmacol 2003;3(5):530–43.
- [23] Klabunde T, Hessler G. Drug design strategies for targeting G-protein-coupled receptors. ChemBioChem 2002;3(10):928–44.
- [24] Horton DA, Bourne GT, Smythe ML. Exploring privileged structures: the combinatorial synthesis of cyclic peptides. Mol Diversity 2002;5(4):289–304.
 [25] Ma B, Nussinov R. Trp/Met/Phe hot spots in protein-protein interactions:
- [25] Ma B, Russhov K, Th/Met/He flot spots in protent-protein interactions.
 potential targets in drug design. Curr Top Med Chem 2007;7(10):999–1005.
 [26] Johnson VA, Singh EK, Nazarova LA, Alexander LD, McAlpine SR. Macrocyclic
- inhibitors of Hsp90. Curr Top Med Chem 2010;10(14):1380–402.
- [27] Driggers EM, Hale SP, Lee J, Terrett NK. The exploration of macrocycles for drug discovery – an underexploited structural class. Nat Rev Drug Discovery 2008;7(7):608–24.
- [28] Grabowski K, Schneider G. Properties and architecture of drugs and natural products revisited. Curr Chem Biol 2007;1(1):115–27.
- [29] Galloway WRJD, Bender A, Welch M, Spring DR. The discovery of antibacterial agents using diversity-oriented synthesis. Chem Commun 2009:2446–62.
- [30] Jackson S, DeGrado W, Dwivedi A, Parthasarathy A, Higley A, Krywko J, et al. Template-constrained cyclic peptides: design of high-affinity ligands for GPIIb/ IIIa. | Am Chem Soc 1994;116(8):3220-30.
- [31] Gottschling D, Boer J, Schuster A, Holzmann B, Kessler H. Combinatorial and rational strategies to develop non-peptidic α4β7-integrin antagonists from cyclic peptides. Angew Chem Int Ed 2002;41(16):3007–11.
- [32] Schreiber SL. Target-oriented and diversity-oriented organic synthesis in drug discovery. Science 2000;287:1964–9.
- [33] Agalave SG, Maujan SR, Pore VS. Click chemistry: 1,2,3-triazoles as pharmacophores. Chem Asian J 2011;6(10):2696–718.
- [34] Zhang J, Kemmink J, Rijkers DTS, Liskamp RMJ. Cu(I)- and Ru(II)-mediated "Click" cyclization of tripeptides toward vancomycin-inspired mimics. Org Lett 2011;13(13):3438–41.
- [35] Iuliano A, Pieraccini I, Félix G, Salvadori P. Synthesis of four cholic acid-based CSPs containing 2-naphthoyl carbamate and 3,5-dinitrophenylcarbamate

moieties and their evaluation in the HPLC resolution of racemic compounds. Tetrahedron asymmetry 2002;13:1265–75.

- [36] Differences between the IUPAC names of the hydrocarbons and the corresponding Chemical Abstracts names are noted in the review of Loening KL. Hydrocarbons, nomenclature, Kirk-Othmer encycl. chem. technol, 3rd ed., vol. 12; 1980, p.892–900.
- [37] Moss GP. Nomenclature of steroids. Pure Appl Chem 1989;61:1783-822.
- [38] Copéret C. Stereoselectivity of supported alkene metathesis catalysts: a goal and a tool to characterize active sites. J Org Chem 2011;7:13–24.
- [39] Hoveyda AH, Malcolmson SJ, Meek SJ, Zhugralin AR. Catalytic enantioselective olefin metathesis in natural product synthesis. Chiral metal-based complexes that deliver high enantioselectivity and more. Angew Chem Int Ed 2010;49(1):34–44.
- [40] Aljarilla A, Lopez JC, Plumet J. Metathesis reactions of carbohydrates: recent highlights in cross-metathesis. Eur J Org Chem 2010:6123–43.
- [41] Nolan SP, Clavier H. Chemoselective olefin metathesis transformations mediated by ruthenium complexes. Chem Soc Rev 2010;39(8):3305–16.
- [42] Van Otterlo WAL, De Koning CB. Metathesis in the synthesis of aromatic compounds. Chem Rev 2009;109(8):3743–82.
- [43] Tori M, Mizutani R. Construction of eight-membered carbocycles with trisubstituted double bonds using the ring closing metathesis reaction. Molecules 2010;15:4242–60.
- [44] Nicolaou KC, Bulger PG, Sarlah D. Palladium-catalyzed cross-coupling reactions in total synthesis. Angew Chem Int Ed 2005;44(29):4442–89.
- [45] Smith AB, Adams CM, Kozmin SA, Paone DV. Total synthesis of (-)cylindrocyclophanes A and F exploiting the reversible nature of the olefin cross metathesis reaction. J Am Chem Soc 2001;123(25):5925–37.
- [46] Hu X, Nguyen KT, Verlinde CLMJ, Hol WGJ, Pei D. Structure-based design of a macrocyclic inhibitor for peptide deformylase. J Med Chem 2003;46(18):3771-4.
- [47] Vassilikogiannaki G, Margaros I, Tofi M. Olefin metathesis: remote substituents governing the stereoselectivity of 11-membered-ring formation. Org Lett 2004;6(2):205-8.
- [48] Shi ZD, Lee K, Wei CQ, Roberts LR, Worthy KM, Fisher RJ, et al. Synthesis of a 5methylindolyl-containing macrocycle that displays ultrapotent grb2 sh2 domain-binding affinity. J Med Chem 2004;47(4):788–91.
- [49] Boger DL, Hong J. Asymmetric total synthesis of ent-(-)-roseophilin: assignment of absolute configuration. J Am Chem Soc 2001;123(35):8515–9.
- [50] Morzycki JW. Application of olefin metathesis in the synthesis of steroids. Steroids 2011;76(10–11):949–66.
- [51] Ibrahim-Ouali M, Zoubir J, Romero E. A ring-closing metathesis approach to secosteroidal macrocycles. Tetrahedron Lett 2011;52:7128–31.
- [52] Fieser LF, Rajagopalan S. Oxidation of steroids. Selective oxidations and acylations in the bile acid series. J Am Chem Soc 1950;72:5530-6.
- [53] Lidstrom P, Tierney J, Wathey B, Westman J. Microwave assisted organic synthesis: a review. Tetrahedron 2001;57(45):9225-83.
- [54] Pettit GR, Piatak DM. Steroids and related products. Reduction of ester to ethers. | Org Chem 1962;27:2127–30.
- [55] Jung MF, Lyster MA. Quantitative dealkylation of alkyl esters via treatment with trimethylsilyl iodide. A new method for ester hydrolysis. J Am Chem Soc 1977;99(3):968–9.
- [56] Olah GA, Narang SC, Gupta BGB, Malhotra R. Synthetic methods and reactions. Transformations with chlorotrimethylsilane/sodium iodide, a convenient in situ iodotrimethylsilane reagent. J Org Chem 1979;44(8):1247–51.
- [57] Fieser TL, Rajagopalan S. Selective oxidation with N-bromosuccinimide. Cholic acid. J Am Chem Soc 1949;71:3935–8.