

Article



Subscriber access provided by Gothenburg University Library

Stereospecific Synthesis of 3,4-Dihydro-2H-naphtho-1,4-oxazin-2-ones by Unification of Benzoxepine-4-carboxylates with Chiral Amino Acid Ethyl Esters

Veera Prasad Kasagani, Siva Hariprasad Kurma, and China Raju Bhimapaka

J. Org. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.joc.9b02624 • Publication Date (Web): 29 Jan 2020

Downloaded from pubs.acs.org on January 30, 2020

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.

is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

Stereospecific Synthesis of 3,4-Dihydro-2*H*-naphtho-1,4-oxazin-2-ones by Unification of Benzoxepine-4-carboxylates with Chiral Amino Acid Ethyl Esters

Veera Prasad Kasagani, Siva Hariprasad Kurma and China Raju Bhimapaka*

Department of Organic Synthesis and Process Chemistry, CSIR-Indian Institute of Chemical Technology, Hyderabad-500007, India



ABSTRACT: A novel and efficient stereocontrolled method has been developed for the preparation of chiral 3,4-dihydro-2*H*-naphtho[1,2-*b*][1,4]oxazin-2-ones by the reaction of benzoxepine-4-carboxylates with chiral amino acid ethyl esters for the first time. The chiral 3,4-dihydro-2*H*-naphtho-1,4-oxazinones have been achieved in one step by the formation of C-N, C-C and C-O bonds.

INTRODUCTION

Naphthoxazinones and benzoxazinones constitute an important class of bio-active compounds and useful intermediates for the preparation of pharmaceuticals.¹ These heterocyclic compounds particularly benzoxazinones are synthetic building blocks and present in a variety of natural products.² Further, these compounds are photo active molecules which possess fluorescent, photophysical and photochemical properties.³ Similarly, chiral 3,4-dihydro-1,4naphthoxazin-2-ones and 3,4-dihydro-1,4-benzoxazin-2-ones represent the structural motif of several natural products with interesting biological properties. Hence, the preparations of these heterocyclic compounds have attracted much attention to synthetic organic and medicinal chemists. To the best of our knowledge only two approaches are available for the preparation of chiral dihydrobenzoxazinones (Scheme 1). The enantioselective cycloaddition of ortho-benzoquinone imides with chiral ketene enolates furnished the chiral dihydrobenzoxazinones.4 Asymmetric reduction of benzoxazinones is the another approach to access the dihydrobenzoxazinones.⁵ Interestingly, there is only one method reported by Gorohovsky et al⁶ for the preparation of chiral dihydronaphthoxazinones by the reaction of 2,3dichloro-1,4-naphthoguinones with natural amino acids as chiral source (Scheme 1). Therefore, the development of new

general strategies for the synthesis of enantiopure dihydronaphthoxazinone framework has become attractive and challenging.



Scheme 1. Previous approaches for the preparation of chiral dihydro[1,4]benzoxazinones and naphthoxazinone

ACS Paragon Plus Environment

Our research group is interested in the development of novel methods for the preparation of heterocyclic compounds.⁷ The research work on benzoxepine-4-carboxylates have produced novel heterocyclic compounds.⁸ In continuation of our work on benzoxepine-4-carboxylates, recently we have prepared useful oxygenated heterocycles.⁹ The present work envision to develop a stereospecific method for the construction of 3,4-dihydro-2*H*-1,4-naphtho-1,4-oxazin-2-ones by the reaction of benzoxepine-4-carboxylates with chiral amino acid ethyl esters.

RESULTS AND DISCUSSION

In an initial experiment, we have carried out the reaction of benzoxepine-4-carboxylate (1a, 1 equiv. Prepared as per our with (S)-2-amino-3reported method) previous phenylpropanoic acid (2, 1 equiv.) in the presence of p-TsOH (30 mol %) with ethanol under reflux conditions (Scheme 2). solid was obtained after column А pale yellow chromatography (20% yield). The compound was identified as (S)-3-benzyl-2-oxo-3,4-dihydro-2H-naphtho[1,2ethyl b][1,4]oxazine-5-carboxylate 3a based on spectral data. The reaction is stereocontrolled and specifically obtained compound 3a. This work represents the first example for the stereospecific construction of 3,4-dihydro-2H-naphtho[1,2b][1,4]oxazin-2-one by the formation of C-N, C-C and C-O bonds in one-pot starting from benzoxepine-4-carboxylate.

Scheme 2. Preparation of compound 3a



We observed that the above reaction is very slow and produced compound 3a in low quantity. The unreacted starting materials were recovered even after prolonged reaction time. This may be due to the amino acid is not reacted well under these conditions. We thought the amino acid ethyl ester is

better species when compared to its acid. Therefore, the ethyl (S)-2-amino-3-phenylpropanoate 2a was prepared from 2 by standard esterification protocol.¹⁰ Thus obtained ester 2a (1 equiv.) was reacted with 1a (1 equiv.) in the presence of p-TsOH (30 mol %) with ethanol under reflux conditions (Scheme 2). This furnished compound 3a with improved yield (45%, Table 1, entry 2). Other solvents such as MeOH, toluene, xylene, diethyl ether, DCM, DCE, MeCN and THF (Table 1, entry 3-10) were also studied to see the effect of solvents. We observed that DCM and DCE solvents are superior when compared to other tested solvents and produced compound 3a in good yields (69%, 58%, Table 1, entry 7-8). The reaction at room temperature with DCM is inferior when compared to reflux condition (3a, 36% yield, Table 1, entry 11). A similar result was obtained with both low (20 mol %) and high (50 mol %) equivalents of p-TsOH (Table 1, entry 12-13) when we tested the reaction of 1a with 2a in DCM under reflux conditions.

Table 1. Optimization Study

			U _S UC ₂ H ₅	
	CO ₂ C ₂ H ₅	CO ₂ C ₂ H ₅ NH ₂		H O O O
1a	2a			3a
Entry	Catalyst (equiv)	Solvent	Time[h]	Yield ^[d]
01 ^{<i>a,b</i>}	<i>p</i> -TsOH (0.3)	EtOH	16	20
02 ^b	<i>p</i> -TsOH (0.3)	EtOH	16	45
03 ^b	<i>p</i> -TsOH (0.3)	MeOH	18	31
04 ^b	<i>p</i> -TsOH (0.3)	toluene	18	20
05 ^b	<i>p</i> -TsOH (0.3)	xylene	18	38
06 ^b	<i>p</i> -TsOH (0.3)	ether	32	42
07 ^b	<i>p</i> -TsOH (0.3)	DCM	24	69
08 ^b	<i>p</i> -TsOH (0.3)	1,2-DCE	18	58
09 ^b	<i>p</i> -TsOH (0.3)	MeCN	18	47
10 ^b	<i>p</i> -TsOH (0.3)	THF	18	43
11 ^c	<i>p</i> -TsOH (0.3)	DCM	48	36
12 ^b	<i>p</i> -TsOH (0.2)	DCM	32	62
13 ^b	<i>p</i> -TsOH (0.5)	DCM	22	68
14 ^b	HCI (1.0)	DCM	15	31
15 ^b	AcOH (0.3)	DCM	18	37
16 ^b	TFA (0.3)	DCM	16	30
17 ^b	AICI ₃ (0.3)	DCM	07	34
18 ^b	FeCl ₃ (0.3)	DCM	16	33
19 ^b	Cu(OAc) ₂ (0.3)	DCM	16	Trace
20 ^b	Cul (0.3)	DCM	18	Trace
21 ^b	Bi(OTf) ₃ (0.3)	DCM	10	49
22 ^b	La(OTf) ₃ (0.3)	DCM	10	46

- - . .

^a(S)-2-Amino-3-phenylpropanoic acid used as reactant, ^b reflux,

^c room temperature, ^disolated yield.

Having optimized the reaction conditions, next, the reaction of **1a** with **2a** was tested with various catalysts such as protic acids (Table 1, entry 14-16), Lewis acids (Table 1, entry

2

3 4

5

6 7

8

9

10 11

12

13

14 15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44 45

46

47

48

49

50

51

52

53

54

55 56

57 58 59

60

17-20) and triflates (Table 1, entry 21-22) in DCM under reflux conditions. All these reactions have produced compound **3a** in moderate yields. No product formation was observed when the reaction was carried out in absence of catalyst. The above results revealed that 30 mol % *p*-TsOH is sufficient to promote the reaction in DCM under reflux conditions to produce the compound **3a** (69% yield, Table 1, entry 7). The compound **3a** was obtained stereospecifically and this was confirmed by chiral HPLC analysis (Table 2, enantiomeric purity = 99.89%, see SI). Then we have carried out the reaction of **1a** with ethyl (*R*)-2-amino-3-

Table 2. Preparation of 3,4-dihydro-2H-naphtho[1,2-b][1,4]oxazine-5-carboxylates 3a-m



phenylpropanoate **2b** under our optimized reaction conditions to obtain **3b** (Table 2, enantiomeric purity = 99.95%, see SI). Further, we also carried out the reaction of **1a** with ethyl 2amino-3-phenylpropanoate **2c** (Table 2) to furnish racemic (\pm) naphthoxazinone compound **3c** (*S* = 50.61%; *R* = 49.39%, see SI). The HPLC analysis of the chiral compounds **3a-b** was compared with the racemic compound **3c**. The present protocol was found to be stereocontrolled and specifically formed the chiral compounds.

Further to explore the present protocol, we have investigated the reaction with substrates present on benzoxepine-4-carboxylates. Accordingly, the electron withdrawing (bromo 1b, chloro 1c) and electron donating (methoxy 1d, methyl 1e-f) groups present on benzoxepine-4carboxylates were prepared as per our earlier reported method.^{8a} The benzoxepines **1b-f** were reacted with ethyl (S)-2-amino-3-phenylpropanoate 2a and ethyl (R)-2-amino-3phenylpropanoate 2b under our optimized conditions. All these reactions proceeded smoothly and produced the corresponding chiral naphthoxazinones 3b-m (Table 2). The electron donating groups produced the compounds with better yields when compared to electron withdrawing groups. All the compounds were characterized by spectral data and the enantiomeric purity of the compounds were analyzed by chiral HPLC (>97-99%, see SI).

To broaden the scope of the present stereospecific reaction, the following amino acid ethyl esters 2d-v have been prepared from their corresponding amino acids (see SI).10 Accordingly, 1a was reacted with 2d-t under our optimized reaction conditions (Table 3).11 The reactions underwent smoothly with amino acid ethyl esters such as 2d-l, 2n-p and 2r provided the corresponding chiral naphthoxazinones 3n-v, 3x-z and 3zb with >92-99% enantiomeric purity (Table 3). Whereas, the amino acid ethyl esters such as 2m, 2q and 2s-t could not provide the target compounds under these reaction conditions and the starting materials were recovered. The reaction of 1a with ethyl (R)-2-amino-2-phenylacetate 2u and ethyl (S)-2-amino-2-phenylacetate 2v provided the compounds 3ze and 3zf (Table 4). The compound 3ze afforded with low enantiomeric purity (*R* isomer = 16.25%, *S* isomer = 83.75%see SI) and compound 3zf obtained in racemic (±) mixture (R isomer = 46.46%, S isomer = 53.54%, see SI). As the α - hydrogen is more acidic in case of 2-amino-2-phenylacetic acid this type of recemization is possible. 2D NMR spectra were recorded for the compounds **3zb**, **3ze** and **3zg** and incorporated in the SI.

Table 3. Preparation of 3,4-dihydro-2H-naphtho[1,2-b][1,4]oxazine-5-carboxylates 3n-zd



Since we obtained racemic target compound $3\mathbf{z}\mathbf{f}$ from ethyl (*S*)-2-amino-2-phenylacetate $2\mathbf{v}$, then we sought to conduct the reaction with electron withdrawing and electron donating group present on benzoxepine-4-carboxyltes in order to check the racemization. Accordingly, we have carried out the

reaction of benzoxepine-4-carboxylates 1c and 1f with 2v under our optimized conditions (Table 4). Interestingly, the electron donating group present on benzoxepine produced chiral compound 3zg (enantiomeric purity for *R* isomer = > 99%, see SI) and electron withdrawing group present on benzoxepine could not provided the compound 3zh (Table 4, see SI). This result indicated that the racemization has not been observed when electron donating group present on benzoxepine. All the obtained compounds were characterized by spectroscopy and enantiomeric purity analyzed by chiral HPLC (see SI).

Table 4. Preparation of compounds 3ze-zh



To further demonstrate the present protocol, chiral 3,4dihydro-2*H*-naphtho[1,2-*b*][1,4]oxazin-2-ones **3a**, **3c** and **3f** have been prepared in gram scale by the reaction of benzoxepine compounds **1a** (2.5 g) with **2a**, **1a** (2.2 g) with **2c**, and **1c** (1.2 g) with **2a** under our optimized conditions and the results were depicted in Scheme 3.

Scheme 3. Gram scale preparation of compounds 3a, 3c & 3f



A plausible reaction pathway for the preparation of 3a was depicted in Scheme 4. The benzoxepine-4-carboxylate 1a with ethyl (S)-2-amino-3-phenylpropanoate 2a under acidic medium forms the Schiff base A. The imine (Schiff base) upon rearrangement provides enamine intermediate B. The rearrangement of intermediate **B** and subsequent intramolecular cyclization through **C-E** furnish the compound **3a** with the elimination of ethanol.

Scheme 4. Plausible Reaction Mechanism



CONCLUSIONS

In conclusion, a facile stereospecific method has been developed for the preparation of chiral 3,4-dihydro-1,4naphthoxazin-2-ones by the reaction of benzoxepine-4carboxylates with chiral amino acid ethyl esters for the first time. The present protocol is efficient and practical one-step process to construct the chiral naphthoxazinones. The synthesized compounds can be utilized as lead compounds in the area of pharmaceuticals, agrochemicals and material science.

EXPERIMENTAL SECTION

General Information. Salicylaldehydes, ethyl 4-chloro-3oxobutanoate and amino acids such as glycine, phenylglycine, alanine, phenylalanine, valine, serine, tyrosine, methionine, leucine, isoleucine and cystein were procured from Sigma-Aldrich. Triphenylphosphine, *p*-TsOH, thionyl chloride and solvents were obtained from local suppliers. All the reactions were carried out under reflux conditions using oil bath. The reactions were monitored by thin layer chromatography (TLC) on pre-coated silica gel 60 F_{254} (mesh) and spots were visualized under UV light. Merck silica gel (60-120 mesh) was used for chromatography. Melting points were determined in open glass capillary tubes on a Stuart melting point apparatus and are uncorrected. ¹H NMR and ¹³C NMR spectra were recorded on Avance 400 MHz and 500 MHz spectrometers in CDCl₃ using TMS as internal standard. FT-IR spectra were recorded on a Thermo Nicollet Nexus 670 spectrometer. Mass spectra were obtained on Agilent LCMS instrument. HRMS were measured on Agilent Technologies 6510, O-TOFLC/MS ESI-Technique. Optical rotations were recorded on Anton Paar, MCP-200 polarimeter with 1 mL cell. Chiral HPLC analysis was carried out by Shimadzu SPD-M20A, PDA detector using Chiral column: CHIRALPAK IC-3, 250 mm. Eluent: n-hexane/isopropyl alcohol = 85:15; flow rate: 0.6 mL/min; 200-500 nm. The Chiral HPLC analyses for the amino acid ethyl esters 2e-f and 2p-q have been analyzed by CHIRALPAK IC-G, 250 mm. Eluent: nhexane/isopropyl alcohol = 85:15; flow rate: 0.5 mL/min using RID detector.

General procedure for the preparation of ethyl 3-oxo-2,3dihydrobenzo[b]oxepine-4-carboxylates 1a-f: The benzoxepine-4-carboxylates 1a-f have been prepared from ethyl 2-(chloromethyl)-2-hydroxy-2*H*-chromene-3-carboxylates by Wittig homologation as per our previous reported method and the compounds were compared with our reported compounds.^{8a,c,9}

Typical procedure for the preparation of 3,4-dihydro-2*H***-naphtho**[1,2-*b*][1,4]oxazin-2-ones: *p*-TsOH (0.057 g, 0.3 mmol) was added to the stirred solution of ethyl (*S*)-2-amino-3phenylpropanoate (**2a**, 0.229 g, 1.0 mmol) and ethyl 3-oxo-2,3dihydrobenzo[*b*]oxepine-4-carboxylate (**1a**, 0.232 g, 1.0 mmol) in DCM at room temperature. The contents were refluxed. After completion of the reaction (TLC, 24h), the solvent was removed under reduced pressure. The crude product was purified by column chromatography using silica gel (60:120, hexane: ethyl acetate, 98:2) as eluent afforded ethyl (*S*)-3-benzyl-2-oxo-3,4dihydro-2*H*-naphtho[1,2-*b*][1,4]oxazine-5-carboxylate **3a** as pale yellow solid in 69% yield. The target compounds **3b-v**, **3x-z**, **3zb** and **3ze-zg** were prepared by the reaction of ethyl 3-oxo-2,3dihydrobenzo[*b*]oxepine-4-carboxylates **1a-f** with amino acid ethyl esters **2b-l**, **2n-p**, **2r** and **2u-v** under similar conditions.

Ethyl (*S*)-3-benzyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (**3a**). Pale yellow solid (108 mg, 69% Yield); $[\alpha]_D^{20} = +20.12$ (c = 0.33, CHCl₃); mp: 168-170 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.34 (s, 1H), 8.06 (dd, J = 8.5, 0.7 Hz, 1H), 7.78 (d, J = 8.3 Hz, 1H), 7.58-7.54 (m, 1H), 7.36-7.31 (m, 3H), 7.27 (dd, J = 9.5, 3.4 Hz, 1H), 7.21-7.18 (m, 2H), 7.11 (s, 1H), 4.38-4.25 (m, 3H), 3.35 (dd, J = 13.8, 3.8 Hz, 1H), 3.02 (dd, J = 13.8, 10.0 Hz, 1H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.8, 165.5, 135.9, 134.1, 129.6, 129.4, 129.1, 128.9, 128.8, 127.3, 126.3, 126.0, 124.2, 119.2, 115.1, 61.3, 55.6, 37.3, 14.3. FT-IR (KBr): 3332, 2990, 1768, 1688, 1466, 1298, 1211, 1154, 1047, 740 cm⁻¹. MS-ESI: *m/z* 362 [M+H]⁺; HRMS-ESI: calcd for $C_{22}H_{20}NO_4$ [M+H]⁺ 362.1392; found 362.1386. HPLC analysis: enantiomeric purity = 99.89%.

1

2

3 4

5

6

7

8

9

10

11 12

13

14

15

16 17

18

19

20 21

22

23

24

25 26

27

28

29 30

31

32

33 34

35

36

37 38

39

40

41

42 43

44

45

46 47

48

49

50 51

52

53

54

55 56

57 58 59

60

Ethyl (*R*--3-*benzyl*-2-*oxo*-3, 4-*dihydro*-2*H*-*naphtho*[1,2*b*][1,4]*oxazine*-5-*carboxylate* (**3b**). Pale yellow solid (106 mg, 68% Yield); [α]_D²⁰ = -19.52 (c = 0.33, CHCl₃); mp: 167-169 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.34 (s, 1H), 8.06 (d, J = 8.5 Hz, 1H), 7.79 (d, J = 8.3 Hz, 1H), 7.58-7.54 (m, 1H), 7.37-7.31 (m, 3H), 7.27 (dd, J = 9.5, 3.4 Hz, 1H), 7.20 (d, J = 7.0 Hz, 2H), 7.11 (s, 1H), 4.37-4.26 (m, 3H), 3.35 (dd, J = 13.8, 3.8 Hz, 1H), 3.02 (dd, J = 13.8, 10.1 Hz, 1H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.8, 165.2, 135.9, 134.1, 129.6, 129.4, 129.1, 128.9, 128.8, 127.3, 126.3, 126.0, 124.2, 119.2, 115.1, 61.3, 55.6, 37.3, 14.3. FT-IR (KBr): 3331, 2989, 1768, 1688, 1465, 1298, 1210, 1154, 1047, 740 cm⁻¹. MS-ESI: *m/z* 362 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₁₉NNaO₄ [M+Na]⁺ 384.1206; found 384.1229. HPLC analysis: enantiomeric purity = 99.95%.

Ethyl 3-benzyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (3c). Pale yellow solid (93 mg, 59% Yield); mp: 152-154 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.35 (s, 1H), 8.06 (dd, J = 8.6, 0.8 Hz, 1H), 7.79 (dd, J = 8.0, 4.5 Hz, 1H), 7.60-7.52 (m, 1H), 7.37-7.26 (m, 4H), 7.22-7.17 (m, 2H), 7.11 (s, 1H), 4.38-4.25 (m, 3H), 3.35 (dd, J = 13.7, 3.8 Hz, 1H), 3.02 (dd, J = 13.7, 10.1 Hz, 1H), 1.40-1.35 (m, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.7, 165.5, 135.8, 134.0, 129.6, 129.3, 129.1, 128.6, 128.8, 127.3, 126.3, 125.7, 124.1, 119.1, 115.1, 61.3, 55.5, 37.2, 14.3. FT-IR (KBr): 3563, 3371, 3328, 1766, 1702, 1641, 1520, 1475, 1382, 1304, 1212 cm⁻¹. MS-ESI: *m/z* 362 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₂₀NO4 [M+H]⁺ 362.1392; found 362.1392. HPLC analysis: enantiomeric purity = 50.61% for (*S*) and 49.39% for (*R*).

Ethyl (*S*)-3-*benzyl-8-bromo-2-oxo-3,4-dihydro-2Hnaphtho*[*1,2-b*][*1,4*]*oxazine-5-carboxylate* (*3d*). Pale yellow solid (40 mg, 28% Yield) ; $[\alpha]_D^{20} = +6.91$ (*c* = 0.33, CHCl₃); mp: 170-172 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.23 (s, 1H), 7.93 (dd, *J* = 7.9, 5.3 Hz, 2H), 7.60 (dd, *J* = 9.0, 1.5 Hz, 1H), 7.32 (t, *J* = 7.4 Hz, 2H), 7.27 (d, *J* = 7.6 Hz, 1H), 7.19 (d, *J* = 7.3 Hz, 2H), 7.16 (s, 1H), 4.38-4.27 (m, 3H), 3.36 (dd, *J* = 13.8, 3.8 Hz, 1H), 3.02 (dd, *J* = 13.8, 9.9 Hz, 1H), 1.37 (t, *J* = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.5, 165.1, 135.7, 134.1, 132.5, 130.8, 130.1, 129.3, 128.9, 127.6, 127.4, 127.2, 124.3, 121.0, 117.7, 115.9, 61.5, 55.4, 37.5, 14.3. FT-IR (KBr): 3336, 2992, 1772, 1690, 1465, 1330, 1277, 1159, 1048, 810 cm⁻¹. MS-ESI: m/z 440 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₁₉BrNO₄ [M+H]⁺ 440.0497; found 440.0500. HPLC analysis: enantiomeric purity = 99.76%.

Ethyl (*R*)-3-benzyl-8-bromo-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3e). Pale yellow solid (38 mg, 27% Yield); $[\alpha]_D^{20} = -8.60$ (c = 0.2, CHCl₃); mp: 168-170 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.23 (s, 1H), 7.92 (d, J = 9.7 Hz, 2H), 7.60 (dd, J = 9.0, 1.8 Hz, 1H), 7.32 (t, J = 7.2 Hz, 2H), 7.27 (d, J = 7.4 Hz, 1H), 7.19 (d, J = 6.9 Hz, 2H), 7.16 (s, 1H), 4.38-4.26 (m, 3H), 3.35 (dd, J = 13.8, 3.8 Hz, 1H), 3.02 (dd, J = 13.7, 9.9 Hz, 1H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.5, 165.1, 135.7, 134.1, 132.5, 130.8, 130.1, 129.4, 128.9, 127.6, 127.4, 127.2, 124.3, 121.0, 117.7, 115.9, 61.5, 55.4, 37.5, 14.3. FT-IR (KBr): 3335, 2990, 1772, 1690, 1464, 1330, 1276, 1159, 1048, 810 cm⁻¹. MS-ESI: m/z 440 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₁₉BrNO₄ [M+H]⁺ 440.0497; found 440.0479. HPLC analysis: enantiomeric purity = 98.63%.

Ethvl (S)-3-benzyl-8-chloro-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3f). Pale yellow solid (36 mg, 25% Yield); $[\alpha]_D^{20} = +7.55$ (*c* = 0.05, CHCl₃); mp: 168-170 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.23 (s, 1H), 7.99 (d, J = 9.1 Hz, 1H), 7.76 (d, J = 1.9 Hz, 1H), 7.48 (dd, J = 9.1, 2.0 Hz, 1H), 7.32 (t, J = 7.3 Hz, 2H), 7.27 (d, J = 8.4 Hz, 1H), 7.19 (d, J = 7.0 Hz, 2H), 7.15 (s, 1H), 4.38-4.27 (m, 3H), 3.36 (dd, J =13.8, 3.8 Hz, 1H), 3.02 (dd, J = 13.8, 9.9 Hz, 1H), 1.37 (t, J = 7.1Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.5, 165.1, 135.7, 134.1, 130.1, 129.9, 129.8, 129.4, 128.9, 127.7, 127.5, 127.4, 126.7, 124.2, 121.0, 116.1, 61.5, 55.4, 37.5, 14.3. FT-IR (KBr): 3337, 2925, 1770, 1690, 1465, 1333, 1279, 1153, 1046, 809 cm⁻¹. MS-ESI: m/z 396 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₁₉ClNO₄ [M+H]⁺ 396.1003; found 396.0965. HPLC analysis: enantiomeric purity = 99.99%.

Ethyl (*R*)-3-benzyl-8-chloro-2-oxo-3,4-dihydro-2*H*naphtho[1,2-b][1,4]oxazine-5-carboxylate (**3g**). Pale yellow solid (33 mg, 24% Yield); $[\alpha]_D{}^{20} = -7.0$ (c = 0.2, CHCl₃); mp: 167-169 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.23 (s, 1H), 7.99 (d, J = 9.1 Hz, 1H), 7.76 (d, J = 1.9 Hz, 1H), 7.48 (dd, J = 9.1, 2.0 Hz, 1H), 7.32 (t, J = 7.3 Hz, 2H), 7.27 (d, J = 8.4 Hz, 1H), 7.19 (d, J = 7.0 Hz, 2H), 7.15 (s, 1H), 4.38-4.27 (m, 3H), 3.36 (dd, J =13.8, 3.8 Hz, 1H), 3.02 (dd, J = 13.8, 9.9 Hz, 1H), 1.37 (t, J = 7.1Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.5, 165.1, 135.7, 134.1, 130.1, 129.9, 129.8, 129.3, 128.9, 127.7, 127.5,

2

3 4

5

6

7

8

9

10

11 12

13

14

15

16 17

18

19

20 21

22

23

24

25 26

27

28

29 30

31

32

33 34

35

36

37

38 39

40

41

42 43

44

45

46 47

48

49

50 51

52

53

54

55 56

57 58 59

60

127.4, 126.7, 124.1, 120.9, 116.0, 61.5, 55.4, 37.5, 14.3. FT-IR (KBr): 3336, 2924, 1770, 1690, 1464, 1333, 1279, 1152, 1046, 809 cm⁻¹. MS-ESI: m/z 396 [M+H]⁺; HRMS-ESI: calcd for $C_{22}H_{19}CINO_4$ [M+H]⁺ 396.1003; found 396.0971. HPLC analysis: enantiomeric purity = >99%.

Ethvl (S)-3-benzyl-8-methoxy-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3h). Pale yellow solid (99 mg, 59% Yield); $[\alpha]_D^{20} = +26.23$ (*c* = 0.33, CHCl₃); mp: 166-168 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.22 (s, 1H), 7.98 (d, J = 9.3 Hz, 1H), 7.33 (t, J = 7.2 Hz, 2H), 7.29-7.22 (m, 2H), 7.19 (d, J = 6.9 Hz, 2H), 7.08 (d, J = 2.2 Hz, 1H), 6.95 (s, 1H), 4.36-4.27 (m, 2H), 4.26-4.21 (m, 1H), 3.90 (s, 3H), 3.34 (dd, *J* = 13.8, 3.7 Hz, 1H), 3.00 (dd, J = 13.7, 10.1 Hz, 1H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.8, 165.7, 156.4, 136.0, 134.8, 129.4, 128.9, 128.2, 127.5, 127.3, 127.0, 122.6, 121.7, 120.9, 115.6, 106.5, 61.3, 55.6, 55.4, 37.2, 14.3. FT-IR (KBr): 3320, 2969, 2935, 1757, 1686, 1517, 1339, 1289, 1160, 1024, 826, 698 cm⁻¹. MS-ESI: *m/z* 392 [M+H]⁺; HRMS-ESI: calcd for C₂₃H₂₂NO₅ [M+H]⁺ 392.1498; found 392.1495. HPLC analysis: enantiomeric purity = >99%.

Ethvl (R)-3-benzyl-8-methoxy-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3i). Pale yellow solid (99 mg, 59% Yield); $[\alpha]_D^{20} = -24.92$ (*c* = 0.33, CHCl₃); mp: 165-167 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.24 (s, 1H), 7.99 (d, J = 9.2 Hz, 1H), 7.33 (t, J = 7.3 Hz, 2H), 7.30-7.23 (m, 2H), 7.19 (d, J = 7.1 Hz, 2H), 7.09 (d, J = 2.4 Hz, 1H), 6.95 (s, 1H), 4.36-4.28 (m, 2H), 4.24 (ddt, J = 10.0, 3.6, 1.8 Hz, 1H), 3.90 (s, 3H), 3.34 (dd, J = 13.8, 3.8 Hz, 1H), 3.01 (dd, J = 13.8, 10.1 Hz, 1H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.8, 165.7, 156.5, 135.9, 134.8, 129.4, 128.9, 128.2, 127.5, 127.3, 127.0, 122.6, 121.7, 120.9, 115.6, 106.5, 61.3, 55.7, 55.4, 37.2, 14.3. FT-IR (KBr): 3319, 2967, 2934, 1757, 1686, 1517, 1338, 1287, 1160, 1024, 824, 698 cm⁻¹. MS-ESI: m/z 392 $[M+H]^+$; HRMS-ESI: calcd for C₂₃H₂₂NO₅ $[M+H]^+$ 392.1492; found 392.1500. HPLC analysis: enantiomeric purity = 99.98%.

Ethyl (*S*)-3-benzyl-8-methyl-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (**3***j*). Pale yellow solid (87 mg, 57% Yield); $[\alpha]_D{}^{20} = +18.32$ (c = 0.33, CHCl₃); mp: 186-188 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.25 (s, 1H), 7.96 (d, J = 8.7 Hz, 1H), 7.55 (s, 1H), 7.40 (d, J = 8.7 Hz, 1H), 7.33 (t, J =7.3 Hz, 2H), 7.29 -7.24 (m, 1H), 7.19 (d, J = 7.3 Hz, 2H), 7.02 (s, 1H), 4.35-4.27 (m, 2H), 4.27-4.22 (m, 1H), 3.34 (dd, J = 13.8, 3.7 Hz, 1H), 3.00 (dd, J = 13.7, 10.2 Hz, 1H), 2.47 (s, 3H), 1.36 (t, J =7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.9, 165. 7, 135.9, 134.3, 133.8, 131.8, 129.4, 128.9, 128.8, 128.1, 127.8, 127.3, 126.7, 124.3, 119.1, 115.1, 61.3, 55.6, 37.2, 21.4, 14.3. FT-IR (KBr): 3331, 2973, 2927, 1757, 1689, 1488, 1337, 1292, 1158, 1049, 809 cm⁻¹. MS-ESI: m/z 376 [M+H]⁺; HRMS-ESI: calcd for C₂₃H₂₂NO₄ [M+H]⁺ 376.1543; found 376.1554. HPLC analysis: enantiomeric purity = 99.56%.

Ethyl (R)-3-benzyl-8-methyl-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3k). Pale yellow solid (85 mg, 58% Yield); $[\alpha]_D^{20} = -18.12$ (*c* = 0.33, CHCl₃); mp: 183-185 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.26 (s, 1H), 7.97 (d, J = 8.7 Hz, 1H), 7.56 (s, 1H), 7.43-7.39 (m, 1H), 7.33 (t, J = 7.2Hz, 1H), 7.30-7.24 (m, 2H), 7.19 (d, *J* = 6.9 Hz, 2H), 7.03 (s, 1H), 4.32 (qd, J = 7.1, 3.6 Hz, 2H), 4.25 (dd, J = 10.8, 3.2 Hz, 1H), 3.34 (dd, J = 13.7, 3.7 Hz, 1H), 3.01 (dd, J = 13.7, 10.1 Hz, 1H),2.47 (s, 3H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.9, 165.7, 135.9, 134.3, 133.8, 131.7, 129.4, 128.9, 128.8, 128.1, 127.8, 127.3, 126.7, 124.3, 119.1, 115.1, 61.3, 55.6, 37.2, 21.4, 14.3. FT-IR (KBr): 3330, 2972, 2927, 1757, 1689, 1488, 1336, 1292, 1158, 1049, 808 cm⁻¹. MS-ESI: m/z 376 [M+H]⁺; HRMS-ESI: calcd for C₂₃H₂₁NNaO₄ [M+Na]⁺ 398.1363; found 398.1386. HPLC analysis: enantiomeric purity = 99.59%.

Ethyl (S)-3-benzyl-10-methyl-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (31). Pale yellow solid (79 mg, 55% Yield); $[\alpha]_D^{20} = +19.02$ (*c* = 0.33, CHCl₃); mp: 189-191 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.30 (s, 1H), 7.61 (d, J = 8.1 Hz, 1H), 7.32 (dd, J = 13.2, 5.9 Hz, 3H), 7.27 (d, J = 8.1Hz, 1H), 7.18 (dd, J = 6.9, 5.0 Hz, 4H), 4.37-4.28 (m, 2H), 4.19 (ddd, J = 9.8, 3.7, 1.5 Hz, 1H), 3.34 (dd, J = 13.8, 3.8 Hz, 1H),3.02 (dd, J = 13.7, 9.9 Hz, 1H), 2.93 (s, 3H), 1.37 (t, J = 7.1 Hz)3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.7, 165.6, 136.2, 135.9, 132.2, 131.9, 130.8, 129.8, 129.4, 128.9, 127.8, 127.7, 127.3, 125.8, 123.8, 114.7, 61.3, 54.9, 36.9, 24.4, 14.3. FT-IR (KBr): 3333, 2989, 2930, 1759, 1688, 1448, 1325, 1292, 1163, 1047, 785 cm⁻¹. MS-ESI: *m/z* 376 [M+H]⁺; HRMS-ESI: calcd for C₂₃H₂₂NO₄ [M+H]⁺ 376.1549; found 376.1545. HPLC analysis: enantiomeric purity = 99.30%.

Ethyl (*R*)-3-benzyl-10-methyl-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (**3m**). Pale yellow solid (76 mg, 54% Yield); $[\alpha]_D^{20} = -18.92$ (c = 0.33, CHCl₃); mp: 187-189 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.30 (s, 1H), 7.61 (d, J = 8.1 Hz, 1H), 7.32 (dd, J = 13.9, 6.4 Hz, 3H), 7.27 (d, J = 8.6Hz, 1H), 7.22-7.16 (m, 4H), 4.37-4.28 (m, 2H), 4.19 (ddd, J = 9.9, 3.9, 1.6 Hz, 1H), 3.34 (dd, J = 13.8, 3.9 Hz, 1H), 3.03 (dd, J =13.8, 9.9 Hz, 1H), 2.93 (s, 3H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 166.7, 165.6, 136.2, 135.9, 132.2, 131.9, 130.8, 129.7, 129.4, 128.9, 127.8, 127.7, 127.3, 125.8, 123.8, 114.7, 61.3, 54.9, 36.9, 24.4, 14.3. FT-IR (KBr): 3332, 2987, 2929, 1759, 1688, 1448, 1324, 1291, 1163, 1047, 784 cm⁻¹. MS-ESI: *m/z* 376 [M+H]⁺; HRMS-ESI: calcd for C₂₃H₂₁NNaO₄ [M+Na]⁺ 398.1363; found 398.1389. HPLC analysis: enantiomeric purity = 99.83%.

Ethyl 2-oxo-3,4-dihydro-2H-naphtho[1,2-b][1,4]oxazine-5carboxylate (**3n**). Pale yellow solid (21 mg, 18% Yield); mp: 127-129 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.37 (s, 1H), 8.06 (d, J = 8.5 Hz, 1H), 7.80 (d, J = 8.3 Hz, 1H), 7.57 (t, J = 7.7 Hz, 1H), 7.35 (t, J = 7.5 Hz, 1H), 7.21 (s, 1H), 4.44 (q, J = 7.1 Hz, 2H), 4.17 (d, J = 1.5 Hz, 2H), 1.47 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 167.4, 163.9, 134.2, 130.9, 129.5, 129.1, 128.8, 126.3, 126.3, 124.2, 119.1, 114.7, 61.4, 44.5, 14.3. FT-IR (KBr): 3364, 2924, 2854, 1777, 1695, 1461, 1376, 1300, 1042, 786 cm⁻¹. MS-ESI: *m*/*z* 272 [M+H]⁺. HRMS-ESI: calcd for C₁₅H₁₄NO₄ [M+H]⁺ 272.0923; found 272.0920.

Ethyl (*S*)-3-methyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (**3o**). Pale yellow solid (38 mg, 31% Yield); [α]_D²⁰ = +9.01 (c = 0.2, CHCl₃); mp: 102-104 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.36 (s, 1H), 8.09-8.03 (m, 1H), 7.80 (d, J = 8.3 Hz, 1H), 7.56 (m, 1H), 7.38-7.32 (m, 1H), 7.17 (s, 1H), 4.44 (q, J = 7.1 Hz, 2H), 4.09 (m, 1H), 1.63 (d, J = 6.7 Hz, 3H), 1.47 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.4, 166.5, 134.4, 130.8, 129.4, 129.1, 128.6, 126.3, 126.2, 124.2, 119.2, 114.7, 61.4, 49.8, 17.3, 14.3. FT-IR (KBr): 3330, 2980, 2925, 1772, 1692, 1466, 1377, 1298, 1212, 1042, 784 cm⁻¹. MS-ESI: m/z 286 [M+H]⁺; HRMS-ESI: calcd for C₁₆H₁₆NO₄ [M+H]⁺ 286.1074; found 286.1090. HPLC analysis: enantiomeric purity = 97.84%.

Ethyl (*R*)-3-methyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (**3***p*). Pale yellow solid (68 mg, 55% Yield); $[\alpha]_D^{20} = -8.81$ (c = 0.3818, CHCl₃); mp: 124-126 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.36 (s, 1H), 8.06 (dd, J = 8.6, 0.9Hz, 1H), 7.83-7.75 (m, 1H), 7.61-7.51 (m, 1H), 7.35 (m, 1H), 7.19 (s, 1H), 4.48-4.40 (m, 2H), 4.13-4.06 (m, 1H), 1.63 (d, J =6.7 Hz, 3H), 1.50-1.44 (m, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.4, 166.5, 134.3, 130.8, 129.3, 129.1, 128.5, 126.3, 126.1, 124.1, 119.2, 114.7, 61.4, 49.7, 17.2, 14.2. FT-IR (KBr): 3363, 3023, 1174, 1696, 1471, 1381, 1304, 1215 cm⁻¹. MS-ESI: m/z 286 [M+H]⁺; HRMS-ESI: calcd for C₁₆H₁₆NO₄ [M+H]⁺ 286.1079; found 286.1094. HPLC analysis: enantiomeric purity = 99.63%. *Ethyl* (*S*)-*3*-*isopropyl*-2-*oxo*-*3*,*4*-*dihydro*-2*H*-*naphtho*[*1*,2*b*][*1*,*4*]*oxazine*-*5*-*carboxylate* (*3q*). Pale yellow solid (60 mg, 44% Yield); $[\alpha]_D^{20} = +65.16$ (*c* = 0.33, CHCl₃); mp: 72-74 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.35 (s, 1H), 8.04 (d, *J* = 8.5 Hz, 1H), 7.77 (d, *J* = 8.2 Hz, 1H), 7.57-7.51 (m, 1H), 7.45 (s, 1H), 7.32 (t, *J* = 7.5 Hz, 1H), 4.45 (tt, *J* = 7.2, 3.6 Hz, 2H), 3.94 (dd, *J* = 6.1, 1.8 Hz, 1H), 2.28 (m, 1H), 1.47 (t, *J* = 7.1 Hz, 3H), 1.12 (d, *J* = 6.9 Hz, 3H), 1.03 (d, *J* = 6.7 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.5, 165.0, 133.5, 130.2, 129.3, 129.1, 128.6, 125.9, 123.8, 119.1, 114.3, 61.4, 59.9, 30.3, 19.0, 17.8, 14.3. FT-IR (KBr): 3327, 2963, 2928, 1764, 1692, 1475, 1342, 1296, 1156, 1046, 783 cm⁻¹. MS-ESI: *m/z* 314 [M+H]⁺; HRMS-ESI: calcd for C₁₈H₂₀NO₄ [M+H]⁺ 314.1392; found 314.1391. HPLC analysis: enantiomeric purity = >99%.

Ethyl (*R*)-3-isopropyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (**3***r*). Pale yellow solid (60 mg, 44% Yield); $[\alpha]_D^{20} = -60.66$ (c = 0.33, CHCl₃); mp: 70-72 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.35 (s, 1H), 8.04 (d, J = 8.5 Hz, 1H), 7.77 (d, J = 8.2 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.45 (s, 1H), 7.31 (t, J = 7.5 Hz, 1H), 4.45 (dd, J = 13.2, 6.7 Hz, 2H), 3.94 (d, J = 5.0 Hz, 1H), 2.28 (m, 1H), 1.47 (t, J = 7.1 Hz, 3H), 1.12 (d, J = 6.8 Hz, 3H), 1.03 (d, J = 6.7 Hz, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 167.5, 165.0, 133.5, 130.2, 129.3, 129.1, 128.6, 125.9, 123.8, 119.1, 114.4, 61.4, 59.9, 30.3, 19.0, 17.8, 14.4. FT-IR (KBr): 3326, 2962, 2928, 1764, 1692, 1474, 1342, 1295, 1156, 1045, 783 cm⁻¹. MS-ESI: *m*/z 314 [M+H]⁺; HRMS-ESI: calcd for C₁₈H₂₀NO₄ [M+H]⁺ 314.1387; found 314.1376. HPLC analysis: enantiomeric purity = 99.88%.

Ethyl (*S*)-*3*-*isobutyl*-2-*oxo*-*3*,*4*-*dihydro*-2*H*-*naphtho*[*1*,2*b*][*1*,*4*]*oxazine*-*5*-*carboxylate* (*3s*). Pale yellow solid (58 mg, 41% Yield); $[a]_D^{20} = +4.76$ (c = 0.4181, CHCl₃); mp: 76-78 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.36 (s, 1H), 8.08-8.02 (m, 1H), 7.82-7.76 (m, 1H), 7.58-7.52 (m, 1H), 7.33 (dd, *J* = 9.2, 5.1, 1.7 Hz, 2H), 4.48-4.40 (m, 2H), 4.11 (dd, *J* = 8.8, 4.9, 1.6 Hz, 1H), 1.91-1.73 (m, 3H), 1.49-1.45 (m, 3H), 1.00 (dd, *J* = 6.2, 5.3 Hz, 6H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.4, 166.2, 133.9, 130.2, 129.3, 129.1, 128.6, 126.2, 126.0, 124.0, 119.1, 114.7, 61.4, 52.2, 39.6, 24.3, 23.1, 21.3, 14.3. FT-IR (KBr): 3324, 3053, 2978, 2928, 1728, 1652, 1490, 1316, 1232, 1101, 1047 cm⁻¹. MS-ESI: *m/z* 328 [M+H]⁺; HRMS-ESI: calcd for C₁₉H₂₂NO₄ [M+H]⁺ 328.1549; found 328.1557. HPLC analysis: enantiomeric purity = >99%.

Ethyl (*R*)-3-isobutyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (3t). Pale yellow solid (66 mg,

1

2

3 4

5

6

7

2

3 4

5

6

7

8 9

10

11

12 13

14

15

16 17

18

19

20

21 22

23

24

25 26

27

28

29 30

31

32

33 34

35

36

37

38 39

40

41

42 43

44

45

46 47

48

49

50

51 52

53

54

55 56

57 58 59

60

47% Yield); [α]_D²⁰ = -2.24 (c = 0.6454, CHCl₃); mp: 90-92 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.28 (d, J = 6.9 Hz, 1H), 7.98 (dd, J = 8.5, 0.7 Hz, 1H), 7.71 (t, J = 6.3 Hz, 1H), 7.52-7.45 (m, 1H), 7.30-7.23 (m, 2H), 4.40-4.33 (m, 2H), 4.06-4.00 (m, 1H), 1.84-1.67 (m, 3H), 1.42-1.38 (m, 3H), 0.92 (dd, J = 6.2, 5.2 Hz, 6H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.4, 165.2, 133.0, 129.2, 128.3, 128.1, 127.6, 125.2, 125.0, 123.0, 118.1, 113.7, 60.4, 51.2, 38.6, 23.4, 22.1, 20.3, 13.3. FT-IR (KBr): 3355, 2964, 1776, 1697, 1640, 1525, 1476, 1382, 1306, 1210 cm⁻¹. MS-ESI: m/z 328 [M+H]⁺; HRMS-ESI: calcd for C₁₉H₂₂NO₄ [M+H]⁺ 328.1531; found 328.1549. HPLC analysis: enantiomeric purity = >99%.

Ethyl (*S*)-3-(2-(methylthio)ethyl)-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (**3u**). Pale yellow solid (60 mg, 40% Yield); $[\alpha]_D^{20} = +9.18$ (c = 0.4636, CHCl₃); mp: 111-113 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.37 (s, 1H), 8.05 (dd, *J* = 8.6, 0.9 Hz, 1H), 7.84-7.75 (m, 1H), 7.56 (m, 1H), 7.40-7.32 (m, 2H), 4.49-4.40 (m, 2H), 4.29 (m, 1H), 2.80-2.65 (m, 2H), 2.35-2.23 (m, 1H), 2.20-2.15 (m, 4H), 1.50-1.44 (m, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.3, 165.7, 133.9, 129.9, 129.4, 129.1, 128.7, 126.3, 126.0, 124.1, 119.1, 114.7, 61.9, 61.4, 52.7, 30.2, 29.7, 15.4, 14.3. FT-IR (KBr): 3601, 3358, 3012, 2555, 1777, 1699, 1641, 1526, 1480, 1382, 1301, 1215 cm⁻¹. MS-ESI: *m/z* 346 [M+H]⁺; HRMS-ESI: calcd for C₁₈H₂₀NO₄S [M+H]⁺ 346.1113; found 346.1104. HPLC analysis: enantiomeric purity = 96.11%.

Ethyl (*R*)-3-(2-(methylthio)ethyl)-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (**3**ν). Pale yellow solid (56 mg, 44% Yield); $[\alpha]_D^{20} = -9.70$ (c = 0.2909, CHCl₃); mp: 102-104 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.37 (s, 1H), 8.05 (dd, *J* = 8.6, 0.8 Hz, 1H), 7.80 (d, *J* = 8.3 Hz, 1H), 7.56 (dd, *J* = 8.4, 6.8, 1.2 Hz, 1H), 7.37 (d, *J* = 1.1 Hz, 1H), 7.36-7.32 (m, 1H), 4.48-4.41 (m, 2H), 4.29 (dd, *J* = 7.4, 5.0, 1.4 Hz, 1H), 2.80-2.65 (m, 2H), 2.34-2.23 (m, 1H), 2.20-2.10 (m, 4H), 1.50-1.44 (m, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.4, 165.7, 133.9, 129.9, 129.4, 129.1, 128.7, 126.3, 126.0, 124.1, 119.1, 114.7, 61.4, 52.7, 30.2, 29.8, 15.4, 14.3. FT-IR (KBr): 3684, 3022, 2405, 1773, 1695, 1215 cm⁻¹. MS-ESI: *m/z* 346 [M+H]⁺; HRMS-ESI: calcd for C₁₈H₂₀NO₄S [M+H]⁺ 346.1113; found 346.1102. HPLC analysis: enantiomeric purity = 97.28%.

Ethyl (*S*)-3-(4-hydroxybenzyl)-2-oxo-3, 4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (**3x**). Pale yellow solid (33 mg, 19% Yield); $[\alpha]_D^{20} = +2.0$ (*c* = 0.3, CHCl₃); mp: 158-160 °C; ¹H NMR (300 MHz, CDCl₃): δ 8.34 (s, 1H), 8.05 (d, J = 8.5 Hz, 1H), 7.78 (d, J = 8.2 Hz, 1H), 7.56 (t, J = 7.7 Hz, 1H), 7.34 (t, J = 7.5 Hz, 1H), 7.12 (s, 1H), 7.05 (d, J = 8.4 Hz, 2H), 6.79 (d, J = 8.4 Hz, 2H), 4.34 (qd, J = 7.1, 3.6 Hz, 2H), 4.26-4.19 (m, 1H), 3.27 (dd, J = 13.9, 3.8 Hz, 1H), 2.95 (dd, J = 13.9, 10.0 Hz, 1H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (75 MHz, CDCl₃): δ 166.9, 165.7, 155.0, 134.0, 130.6, 129.7, 129.4, 129.13, 128. 9, 127.8, 126.3, 126.0, 124.2, 119.2, 115.8, 114.9, 61.4, 55.6, 36.5, 14.3. FT-IR (KBr): 3446, 3332, 2985, 1765, 1687, 1511, 1471, 1332, 1298, 1210, 1158, 1046, 783 cm⁻¹. MS-ESI: *m/z* 378 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₂₀NO₅ [M+H]⁺ 378.1341; found 378.1346. HPLC analysis: enantiomeric purity = 99.79%.

Ethyl (R)-3-(4-hydroxybenzyl)-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3y). Pale yellow solid (31 mg, 18% Yield); $[\alpha]_D^{20} = -1.67$ (c = 0.3, CHCl₃); mp: 160-162 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.35 (s, 1H), 8.05 (d, J = 8.5 Hz, 1H), 7.79 (d, J = 8.2 Hz, 1H), 7.56 (t, J = 7.6 Hz, 1H), 7.34 (t, J = 7.5 Hz, 1H), 7.13 (s, 1H), 7.06 (d, J = 8.2 Hz, 2H), 6.79 (d, J = 8.2 Hz, 2H), 4.40-4.30 (m, 2H), 4.23 (d, J = 7.4 Hz, 1H), 3.27 (dd, J = 13.9, 3.6 Hz, 1H), 2.96 (dd, J = 13.8, 10.0 Hz, 1H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.9, 165.7, 154.9, 134.0, 130.6, 129.7, 129.4, 129.13, 128.9, 127.9, 126.3, 126.0, 124.2, 119.2, 115.8, 115.0, 61.4, 55.6, 36.5, 14.3. FT-IR (KBr): 3445, 3330, 2986, 1766, 1687, 1510, 1471, 1330, 1298, 1211, 1155, 1044, 820 cm⁻¹. MS-ESI: m/z 378 [M+H]⁺; HRMS-ESI: calcd for C₂₂H₁₉NNaO₅ [M+Na]⁺ 400.1155; found 400.1149. HPLC analysis: enantiomeric purity = 99.79%.

Ethyl (*S*)-3-(*hydroxymethyl*)-2-oxo-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (3z). Pale yellow solid (19 mg, 14% Yield); $[\alpha]_D{}^{20} = +3.86$ (*c* = 0.23, CHCl₃); mp: 152-154 °C: ¹H NMR (400 MHz, CDCl₃): δ 8.37 (s, 1H), 8.04 (d, J = 8.5 Hz, 1H), 7.79 (d, J = 8.3 Hz, 1H), 7.56 (t, J = 7.4 Hz, 1H), 7.47 (s, 1H), 7.35 (t, J = 7.4 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 4.20 (dd, J = 13.0, 8.4 Hz, 2H), 4.08 (d, J = 5.7 Hz, 1H), 2.38 (s, 1H), 1.46 (t, J = 7.1 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.4, 165.1, 133.8, 130.2, 129.5, 129.2, 128.9, 126.3, 126.1, 124.2, 119.1, 114.7, 62.6, 61.5, 55.3, 14.3. FT-IR (KBr): 3335, 2981, 1769, 1693, 1471, 1377, 1300, 1176, 1041, 786 cm⁻¹. MS-ESI: *m/z* 324 [M+Na]⁺; HRMS-ESI: calcd for C₁₆H₁₅NNaO₅ [M+Na]⁺ 324.0827; found 324.0842. HPLC analysis: enantiomeric purity =>99%.

Ethyl (*S*)-3-sec-butyl-2-oxo-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (**3zb**). Pale yellow liquid (47 mg, 33% Yield); $[\alpha]_D^{20} = +2.63$ (c = 0.5727, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.26 (s, 1H), 7.95 (d, *J* = 8.5 Hz, 1H), 7.69 (d, *J* = 8.3 Hz, 1H), 7.49-7.43 (m, 1H), 7.34 (s, 1H), 7.22 (dd, *J* = 6.3, 4.6, 0.7 Hz, 1H), 4.40-4.33 (m, 2H), 3.95 (dd, J = 6.0, 1.9 Hz, 1H), 1.99-1.90 (m, 1H), 1.62-1.45 (m, 2H), 1.41-1.37 (m, 3H), 1.07-0.97 (m, 3H), 0.82 (t, J = 7.4 Hz, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 166.4, 163.9, 132.3, 129.2, 128.2, 128.1, 127.6, 124.9, 124.8, 122.7, 117.9, 113.2, 60.3, 58.0, 35.8, 23.6, 13.3, 10.2. FT-IR (KBr): 3735, 3594, 3374, 2972, 1778, 1698, 1526, 1476, 1383, 1339, 1302, 1207 cm⁻¹. MS-ESI: *m/z* 328 [M+H]⁺; HRMS-ESI: calcd for C₁₉H₂₂NO₄ [M+H]⁺ 328.1549; found 328.1553. HPLC analysis: enantiomeric purity = >99%.

Ethyl (*S*)-2-oxo-3-phenyl-3,4-dihydro-2H-naphtho[1,2b][1,4]oxazine-5-carboxylate (**3**ze). Pale yellow solid (64 mg, 43% Yield); $[\alpha]_D^{20} = -24.26$ (c = 0.1909, CHCl₃); mp: 138-140 °C; ¹H NMR (500 MHz, CDCl₃): δ 8.40 (s, 1H), 8.06 (dd, J = 8.5, 0.7 Hz, 1H), 7.79 (t, J = 8.6 Hz, 1H), 7.72 (s, 1H), 7.56 (dd, J =8.3, 6.8, 1.1 Hz, 1H), 7.47-7.40 (m, 2H), 7.39-7.30 (m, 4H), 5.30 (d, J = 1.5 Hz, 1H), 4.48-4.41 (m, 2H), 1.49-1.44 (m, 3H). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 167.4, 164.2, 136.2, 133.8, 130.0, 129.4, 129.1, 128.9, 128.8, 127.0, 126.3, 126.1, 124.2, 119.2, 114.5, 61.4, 58.2, 14.3. FT-IR (KBr): 3307, 3022, 1779, 1698, 1636, 1525, 1383, 1300, 1214, 1049 cm⁻¹. MS-ESI: *m/z* 348 [M+H]⁺; HRMS-ESI: calcd for C₂₁H₁₈NO₄ [M+H]⁺ 348.1236; found 348.1240. HPLC analysis: enantiomeric purity = 83.75%.

Ethyl-2-oxo-3-phenyl-3,4-dihydro-2H-naphtho[1,2-b][1,4]oxazine-5-carboxylate (*3zf*). Pale yellow solid (62 mg, 42% Yield); mp: 142-144 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.40 (s, 1H), 8.06 (dd, *J* = 8.6, 0.8 Hz, 1H), 7.80 (d, *J* = 8.3 Hz, 1H), 7.72 (s, 1H), 7.60-7.53 (m, 1H), 7.46-7.41 (m, 2H), 7.39-7.31 (m, 4H), 5.30 (d, *J* = 1.7 Hz, 1H), 4.49-4.40 (m, 2H), 1.50-1.44 (m, 3H). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 167.5, 164.3, 136.3, 133.8, 130.1, 129.5, 129.2, 129.0, 128.9, 128.8, 127.1, 126.3, 126.2, 124.2, 119.3, 114.6, 61.5, 58.3, 14.3. FT-IR (KBr): 3364, 3075, 2928, 2341, 1774, 1706, 1387, 1216, 1153 cm⁻¹. MS-ESI: *m/z* 348 [M+H]⁺; HRMS-ESI: calcd for C₂₁H₁₈NO₄ [M+H]⁺ 348.1236; found 348.1240. HPLC analysis: enantiomeric purity of *R* isomer = 46.46%, *S* isomer = 53.54%.

Ethyl (*R*)-10-methyl-2-oxo-3-phenyl-3,4-dihydro-2Hnaphtho[1,2-b][1,4]oxazine-5-carboxylate (**3**zg). Pale yellow solid (45 mg, 31% Yield); $[\alpha]_D^{20} = +0.69$ (c = 0.7272, CHCl₃); mp: 108-110 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.36 (s, 1H), 7.81 (s, 1H), 7.63 (d, J = 8.1 Hz, 1H), 7.43 (d, J = 6.6 Hz, 2H), 7.36-7.29 (m, 4H), 7.19 (t, J = 7.6 Hz, 1H), 5.21 (s, 1H), 4.45 (q, J = 7.0 Hz, 2H), 2.91 (s, 3H), 1.46 (t, J = 7.0 Hz, 3H). ¹³C {1H} NMR (101 MHz, CDCl₃): δ 166.3, 151.3, 147.7, 145.4, 136.3, 134.4, 134.1, 131.9, 131.5, 129.5, 129.0, 128.5, 128.3, 127.9,

ASSOCIATED CONTENT

Supporting Information

The copies of ¹H, ¹³C NMR spectras, structures of the amino acid ethyl esters 2a-v, HPLC chromatograms and HRMS spectra were provided in SI. This material is available free of charge via the internet at <u>http://pubs.acs.org</u>.

found 362.1400. HPLC analysis: enantiomeric purity = >99%.

AUTHOR INFORMATION

Corresponding Author

*E-mail: chinaraju@iict.res.in

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

K.V.P thanks UGC-New Delhi for the fellowship. The authors are thankful to Mr. T. Ramesh Babu, Fluoro-Agrochemicals division for chiral HPLC analysis and Mr. K. Sai Teja, JRF (P), OSPC division for helping in experimental work. B. China Raju acknowledges Science & Engineering Research Board (SERB), Department of Science and Technology (DST), New Delhi for the financial support (EEQ/2017/000314, GAP-0743). IICT Communication No. IICT/Pubs./2018/359.

REFERENCES

(1) (a) Sengupta, S. K.; Trites, D. H.; Madhavarao, M. S.; Beltz, W. R. Actinomycin D Oxazinones as Improved Antitumor Agents. J. Med. Chem. 1979, 22, 797-802. (b) Jarvest, R. L.; Connor, S. C.; Gorniak, J. G. L.; Jennings, J.; Serafinowska, H. T.; West, A. Potent Selective Thienoxazinone Inhibitors of Herpes Proteases. Bioorg. Med. Chem. Lett. 1997, 7, 1733-1738. (c) Bredenberg, J. B. S.; Honkanen, E.; Virtanen, A. I. The Kinetics and Mechanism of the Decomposition of 2,4-Dihydroxy-1,4-benzoxazin-3-one. Acta Chem. Scand. 1962, 16, 135-141. (d) Belattar, A.; Saxton, J. E. Total Synthesis of Heptacyclic Aspidosperma Alkaloids. Part 3 Synthesis of an Advanced Intermediate in the Synthesis of Alalakine. J. Chem. Soc. Perkin

2

3 4

5

6

7

8

9

10

11

12 13

14

15

16 17

18

19

20

21 22

23

24

25 26

27

28

29

30

31

32

33

34 35

36

37

38

39

40

41

42 43

44

45

46

47 48

49

50

51

52

53

54

55

56 57

58 59

60

Trans. 1 1992, 679-683. (e) Krohn, K.; Kirst, H. A.; Maag, H. Antibiotics and Antiviral Compounds VCH, Weinheim, 1993. (f) Kleemann, A.; Engel, J.; Kutscher, B.; Reichert, D Pharmaceutical Substances, 4th ed., Thieme, Stuttgart, New York, 2001, 2521, 599. (g) Achari, B.; Mandal, S. B.; Dutta, P. K.; Chowdhury, C. Perspectives on 1,4-Benzodioxins, 1,4-Benzoxazines and Their 2,3-Dihydro Derivatives. Synlett 2004, 2449-2467. (h) Li, X.; Liu, N.; Zhang, H.; Knudson, S. E.; Slayden, R. A.; Tonge, P. Synthesis and SAR Studies of 1,4-Benzoxazine MenB Inhibitors: Novel Antibacterial Agents Against Mycobacterium Tuberculosis. *Bioorg. Med. Chem. Lett.* 2010, 20, 6306-6309. (i) Bondock, S.; Adel, S.; Etman, H. A.; Badria, F. A. Synthesis and Antitumor Evaluation of Some New 1, 3, 4-Oxadiazole-Based Heterocycles. *Eur. J. Med. Chem.* 2012, 48, 192-199.

(2) (a) Macias, F. A.; Marin, D.; Bastidas, A. O.; Molinillo. J. M.
G. Rediscovering the Bioactivity and Ecological Role of 1,4-Benzoxazinones. *Nat. Prod. Rep.* 2009, *26*, 478-489. (b) Saith, C.; Rodriguez, H.; Marquez, A.; Canete, A.; Jullian, C.; Zanocco, A.
New Synthesis of Naphtho and Benzoxazoles: Decomposition of Naphtho and Benzoxazinones with KOH. *Synth. Commun.* 2001, *31*, 135-140.

(3) (a) Duval, R. A.; Lewin, G.; Peris, E.; Chahboune, N.; Garofano, A.; Drese, S.; Cortes, D.; Brandt, U.; Hocqumiller, R. Heterocyclic Analogues of Squamocin As Inhibitors of Mitochondrial Complex I. On the role of the terminal lactone of annonaceous acetogenins. Biochemistry 2006, 45, 2721-2728. (b) Hu, M.; Fan, J.; Li, H.; Song, K.; wang, S.; Cheng, G.; Peng, X. Fluorescent Chemodosimeter for Cys/Hcy with a Large Absorption Shift and Imaging in Living Cells. Org. Biomol. Chem. 2011, 9, 980-983. (c) Azuma, K.; Suzuki, S.; Uchiyama, S.; Kajiro, T.; Santa, T.; Imal, K. A Study of the Relationship Between the Chemical Structures and the Fluorescence Quantum Yields of Coumarins, Quinoxalinones and Benzoxazinones for the Development of Sensitive Fluorescent Derivatization Reagents. Photochem. Photobiol. Sci. 2003, 2, 443-449. (d) Nishio, T. Photochemical Reactions of Quinoxalin-2-ones and Related Compounds. J. Chem. Soc., Perkin Trans. 1, 1990, 565-570. (e) Nonell, S.; Fenrrares, L. R.; Caete, A.; Lemp, E.; Gunther, G.; Pizarro, N.; Zanocco, A. L. Photophysics and Photochemistry of Naphthoxazinone Derivatives. J. Org. Chem. 2008, 73, 5371-5378.

(4) (a) Wolfer, J.; Bakele, T.; Abraham, C. J.; Isonagie, C. D.;
Lectka, T. Catalytic, Asymmetric Synthesis of 1,4-Benzoxazinones: A Remarkably Enantioselective Route to

α-Amino Acid Derivatives from *o*-Benzoquinone Imides. *Angew. Chem. Ind. Ed.* **2006**, *45*, 7398-7400. (b) Paull, D. H.; Danforth, E. A.; Wolfer, J.; Isonagie, C. D.; Abraham, C. J.; Lectka, T. An Asymmetric, Bifunctional Catalytic Approach to Non-Natural α-Amino Acid Derivatives. *J. Org. Chem.* **2007**, *72*, 5380-5382.

(5) (a) Liang, Q. L.; Yuehui, L.; Kathrin, J.; Matthias, B. Relay Iron/Chiral Brønsted Acid Catalysis: Enantioselective Hydrogenation of Benzoxazinones. J. Am. Chem. Soc. 2015, 137, 2763-2772. (b) Xue, Z. Y.; Jiang, Y.; Peng, X. Z.; Yuan, W. C.; Zhang, X. M. The First General, Highly Enantioselective Lewis Base Organo-Catalyzed Hydrosilylation of Benzoxazinones and Quinoxalinones. Adv. Synth. Catal. 2010, 352, 2132-2136. (c) Rueping, M.; Antonchick, A. P.; Theissmann, T. Remarkably Low Catalyst Loading in Bronsted Acid Catalyzed Transfer Hydrogenations: Enantioselective Reduction of Benzoxazines, Benzothiazines, and Benzoxazinones. Angew. Chem., Int. Ed. 2006, 45, 6751-6755.

(6) Gorohovsky, S.; Bittner, S. Novel N-Quinonyl Amino Acids and their Transformation to 3-Substituted *p*-Isoxazinones. *Amino Acids* **2001**, *20*, 135-144.

(7) (a) Rajkumar, K.; Murthy, T. R.; Zehra, A.; Khursade, P. S.; Kalivendi, S. V.; Tiwari A. K.; Prakasham, R. S.; Raju, B. C. A One-pot Facile Construction of 1*H*-1,2,3-Triazolyl 1,2-Dihydropyridyl Derivatives and Evaluation of Bioactivity Profile. *ChemistrySelect* **2018**, *3*, 13729-13735. (b) Dayakar, CH.; Raju, B. C. A Selective Three-Component, One-Pot Approach for the Synthesis of 1,2-Dihydroquinazolines and Quinazolines. *ChemistrySelect* **2018**, *3*, 9388-9392.

(8) (a) Raju, B. C.; Saidachary, G.; Kumar, J. A. Wittig Homologation of 2-(Chloromethyl)-2H-Chromen-2-ol Derivatives: A New Facile Synthesis of Substituted 2,3-Dihydrobenzoxepine-4-carboxylates. Tetrahedron 2012, 68, 6289-6297. (b) Saidachary, G.; Prasad, K. V.; Divya, D.; Singh, A.; Ramesh, U.; Sridhar, B.; Raju, B. C. Convenient One-Pot Synthesis, Anti-Mycobacterial and Anticancer Activities of Novel Benzoxepinoisoxazolones and Pyrazolones. Eur. J. Med. Chem. 2014, 76, 460-469. (c) Raju, B. C.; Prasad, K. V.; Saidachary, G.; Sridhar, B. A Novel Approach for C-C, C-N, and C-O Bond Formation Reactions: A Facile Synthesis of Benzophenazine, Quinoxaline, and Phenoxazine Derivatives via Ring Opening of Benzoxepines. Org. Lett. 2014, 16, 420-423. (d) Prasad, K. V.; Saidachary, G.; Hariprasad, K. S.; Nagaraju, P.; Rao, V. J.; Raju, B. C. Copper-Catalyzed C-H Oxygenation of Benzoxepine-4-carboxylates: Facile Synthesis and Photophysical Properties of Naphtho[2,1-d]oxazoles and Benzo[c]phenoxazines.

Asian J. Org. Chem. 2016, 5, 819-827.

(9) Prasad, K. V.; Hariprasad, K. S.; Raju, B. C. A Facile Construction of Oxygen Heterocycles by the Reaction of Benzoxepine-4-carboxylates with Dihaloalkanes and Activated Alkynes. *Org. Biomol. Chem.* **2019**, *17*, 6645-6653.

(10) (a) Isamu, A.; Ichiro, M. A Simple and Convenient Method for Esterification of Tryptophan and Other Amino Acids. J. Org. Chem. 1983, 48,121-123. (b) Rivero, I. A.; Heredia, S.; Ochoa, A. Esterification of Amino Acids and Mono Acids Using Triphosgene. Synth. Commun. 2001, 31, 2169-2175. (c) Olga, I. S.; Nikolay, E. S.; Elizabeth, S. B.; Gerd-Volker, R.; Valentine, G. N. Friedel-Crafts Alkylation of Natural Amino Acid-Derived Pyrroles with CF₃-Substituted Cyclic Imines. Mendeleev Commun. 2013, 23, 92-93. (d) Takanori, K.; Tomoyuki, T.; Shuhachi K. Polymerization Degree of Oligomethionine to Determine Its Bioavailability When Added to a Low-protein Diets. Biosci. Biotech. Biochem. 1996, 60, 828-834. (e) Maarten van, D.; Tobias, M. P.; Dirk, T. S. R.; Rob, M. J. L.; Cornelus, F. N.; Wim E. H. Synthesis and Characterization of Tailorable Biodegradable Thermoresponsive Methacryloylamide Polymers based on L-Serine and L-Threonine Alkyl Esters. Polymer 2010, 51, 2479-2485. (f) Hans, J. F.; Erik, K.; Lars, L.; Dichloromethane as Reactant in Synthesis: An Expedient Transformation of Prolinamide to a Novel Pyrrolo[1,2-c Jimidazolone. J. Org. Chem. 1990, 55, 2254-2256.

(11) We thank a reviewer's suggestion to elaborate the scope of the manuscript.

