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Mechanistic Investigation of the Oxidation of the Carbanion of Methyl 2-Methoxy-2-phenylacetate by an Isoalloxazine

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Summary Evidence from product studies and radical trapping experiments indicates that the mechanism of the oxidation of the carbanion of methyl 2 methoxy 2-

phenylacetate by a model flavin compound in basic methanol is free radical in nature

VARIOUS flavoenzymes, including lactic acid oxidase, amino acid oxidases, and succinic acid oxidase, catalyse dehydrogenation reactions which introduce unsaturation $\alpha\beta$ to a carbonyl group.¹ In an effort to gain a better understanding of the mechanism(s) of these reactions we have investigated the dehydrogenation of alcoholic substrates and their corresponding methyl ethers mediated by the model flavin 10-(2,6-dimethylphenyl) isoalloxazine (Fl_{ox}) in basic methanol solution. Our results with the model substrates 9-hydroxy- and 9-methoxy-fluorene have previously appeared.² We now report the results of our studies with methyl mandelate (1) and methyl 2-methoxy-2-phenylacetate (2) which compounds resemble the actual biological substrates of the flavoenzyme oxidases more closely.

PhCH(OH) CO₂Me

(1)

When (1) (0.10 M) and Fl_{0x} (1×10^{-4} M) were combined in the dark in basic methanol (1.0 M NaOMe; 30 °C) in the absence of O₂, the characteristic visible spectrum of Fl_{0x} disappeared in a first-order manner (k_{0bs} 2.0 × 10⁻³ s⁻¹) and was replaced by a spectrum characteristic of a reduced flavin. Admission of O₂ at the end of the reaction resulted in the restoration of the spectrum of the original oxidized flavin (λ_{max} 441 nm; ϵ 8900 l mol⁻¹ cm⁻¹). Product studies carried out using higher Fl_{0x} concentrations showed that the oxidation product of methyl mandelate is methyl benzoylformate (isolated in 80% yield) as expected.²⁻⁴ Kinetic studies carried out by following the disappearance of Fl_{0x} indicated that the rate law is of the form shown in equation (1), which is characteristic of the rate law observed

$$-d[Fl_{ox}]/dt = k[(1)] [NaOMe] [Fl_{ox}]$$
(1)

for other weak carbon acid substrates^{2,3a,c,e} and is consistent with the previously demonstrated mechanism involving interaction of Fl_{0x} with the carbanion of the substrate in the rate determining step.^{2,3}

When (2) (1.0 m) and Fl_{ox} (1 \times 10⁻⁴ m) were mixed under the same conditions as above the visible spectrum of Fl_{ox} disappeared at a much slower rate than with (1) $(k_{obs} 4.5 \times 10^{-5} \text{ s}^{-1})$ to give a spectrum characteristic of a reduced flavin. Interestingly, (2) has previously been reported to be unreactive towards flavins under basic conditions.⁴ At long reaction times (> 24 h) a subsequent reaction began to take place which resulted in an increase in absorbance at longer wavelengths (λ_{max} 620 nm). When O₂ was admitted after 3 days a spectrum due to a new oxidized flavin (λ_{max} 445 nm) appeared in two phases. (i) Approximately 80% of the final absorbance at 445 nm appeared immediately upon addition of O₂. (ii) The remaining absorbance at 445 nm appeared more slowly over a period of about 30 min and was accompanied by the disappearance of the long-wavelength absorbance band. A product study was initiated at higher Flox concentrations $(2 imes 10^{-2} \, {
m M})$. The reaction was quenched before completion (ca. 2 days) to avoid complications owing to the subsequent reaction noted above. The only product that could be detected was a flavin adduct (3) [equation (2)] which could be separated from the changed starting flavin by column chromatography on silica gel using ethyl acetate- CH_2Cl_2 (5:95) as eluant.[†] The adduct was isolated in 90% yield after correction for recovered unchanged Fl_{0x} . The visible spectrum of (3b) was identical to that of the oxidized flavin recorded at the end of the kinetic experiment involving (2). The identity of the material with λ_{max} 620 nm was not determined, but it could be shown that (3a) incubated in 1.0 M methanolic sodium methoxide slowly gave rise to the same long wavelength absorption observed in the kinetic experiment.



The radical trapping agents O_2 and 4-hydroxy-2,2,6,6-tetramethylpiperidine 1-oxyl⁵(>N-O·) were capable of altering the course of the reaction as shown in the Table.

TABLE. % Yields of products obtained upon reaction of (2) (0.5 M) with Fl_{0x} (0.05 M) in methanol containing NaOMe (1.0 M) in the presence of an excess of O_2 or $\sim N-O \cdot (2.0 M)$.

	With O ₂		With >N-O.
Product	2 days ^a	7 daysª	7 days ^a
PhCO ₂ Me	27	56	53
PhCO,H	8	16	0
(2)	64	16	12
PhCOCO ₂ Me	0	0	26
Flox	85	72	47
(3b)	0	18	27

^a Incubated at 25 \pm 2 °C in the dark for the indicated length of time before products were isolated. ^b The only materials detected from product studies in the absence of radical traps were unchanged (2) and Fl_{ox}, and the adduct (3b). These studies were performed under conditions similar or identical to those involving the radical traps, except for the presence of the trapping agent, and incubation periods ranged from 2 to 5 days. The total yield of Fl_{ox} and (3b) recovered accounted for more than 90% of the original flavin in all cases in which radical traps were not present.

[†] Identification of the partial structure of (3) was based on the following spectroscopic data for (3b): n.m.r. (CDCl₃), δ 8.60 (s, 1NH), 8.4-6.9 (m, 11H), 3.70 (s, 3H), 3.15 (s, 3H), and 1.87 (d, J 3.5 Hz, 6H); mass spectrum (EI 70V) m/e 496, 481, and 317; u.v.-vis. (in methanol) λ_{max} 435 nm (ϵ 11,300 l mol⁻¹ cm⁻¹), 340 (8000); u.v.-vis (in 1.0 M sodium methoxide in methanol) λ_{max} 445 nm (ϵ 9,500 l mol⁻¹ cm⁻¹) 342(9700). The u.v.-visible spectrum of (3a) was typical of a reduced flavin (in methanol) λ_{max} 360 nm (ϵ 6200 l mol⁻¹ cm⁻¹) (in 1.0 M sodium methoxide in methanol) λ_{max} 362 nm (ϵ 6700 l mol⁻¹ cm⁻¹). High resolution mass spectrum of (3b): m/e 496; calc. for C₂₈H₂₄N₄O₅, 496·175; obs., 496·180. All data indicate that (3) is not a mixture of adducts, but is a single compound.

The yields reported are based on the original amounts of starting materials in the reaction mixture Control experiments showed that (3b) is stable in basic methanol solution in the presence of O_2 , H_2O_2 , and $>N-O \cdot$ so that the products reported in the Table are not due to decomposition of (3b) under the reaction conditions In all cases it could be shown that no reaction took place in the absence of Fl_{ox} or sodium methoxide

The mechanisms shown in the Scheme are most consistent with the results of the product studies involving (2) and Fl_{ox} As is true in all other cases studied^{2,3} the carbanion, (2⁻), is the active form of the substrate The radical, (2.), formed from the reaction of (2^{-}) with Γl_{ox} cannot be oxidized to ketoester by dissociation of H⁺ followed by oneelectron transfer or by H. transfer as can occur in the oxidation of α -hydroxycarbonyl compounds ⁶ Instead (2.) is trapped by FI^- . to form (3a) as shown The radical traps function by scavenging $(2\cdot)$ before it can react with The fact that some (3b) is formed in the presence of F1-.



Scheme

The mechanism shown in the Scheme accounts for the production of methyl benzoate in the presence of O₂ The small amount of benzoic acid isolated in these cases could be due to hydrolysis of methyl benzoate by water produced from decomposition of H_2O_2 which may result from the reaction of Fl-. or FlH2 with O27 It is unclear whether methyl benzoylformate isolated in the experiments involving >N–O• is produced *via* an electron transfer mechanism or via an hydroxylamine type intermediate 8 Control experiments do show that methyl benzoate is formed from methyl benzoylformate in basic methanol in the presence of a mixture of >N-O and the corresponding hydroxylamine

Under comparable conditions the rate of disappearance of Fl_{0x} in the presence of (2) is ca 450-fold slower than in the presence of (1) It could be argued that this rate difference indicates that the oxidation of (1) by Fl_{ox} does not involve radical intermediates but that (1) is oxidized by an alternative mechanism not available to $(2)^{2,3,7}$ However, we believe that the differences in the rates of oxidation of (1) and (2) are a reflection of the greater stability of the resonance stabilized 1,2-enediol radical (1.) Such a resonancestabilized structure is not possible for the analogous radical derived from (2) In contrast, the carbanions of 9-hydroxyand 9-methoxy-fluorene are oxidized by Flox at comparable rates² The radicals derived from the oxidation of these two carbanions would be of similar stability ² The results of this study provide convincing evidence for the radical nature of the oxidation of (2) by Fl_{ox} This and the similar conclusions of our earlier study² suggest that the oxidations of other carbon acid substrates by flavins proceed via the same mechanism



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¹ Recent reviews include P Hemmerich, G Nagelshneider and C Veeger FEBS Lett 1970 8 69 A H Neims, and L Hellerman, Ann Rev Biochem , 1970, 39, 867 'Flavins and Flavoproteins 'ed H Kamin University Park Press, Baltimore Md , 1971, 'Flavins and Flavoproteins,' ed T P Singer, Elsevier, New York, 1976, 'Mechanisms of Oxidizing Enzymes' eds T P Singer and R N Ondarza, Elsevier, New York, 1978, pp 43-78

² M Novak and T C Bruice, J Am Chem Soc 1977, 99, 8079 ³ (a) L Main, G J Kasperek, and T C Bruice Biochem, 1972, 11, 3991, (b) S Shinkai and T C Bruice, J Am Chem Soc 1973, ³ (a) L. Main, G. J. Kasperek, and T. C. Bruice. Biochem, 1972, 11, 3991, (b) S. Shinkai and T. C. Bruice, J. Am. Chem. Soc. 1973, 95, 7528, (c) S. Shinkai, T. Kunitake, and T. C. Bruice. ibid, 1976, 98, 7759, (e) T. C. Bruice and J. P. Taulane, ibid, 1976, 98, 7769, (f) R. F. William, S. Shinkai, and T. C. Bruice. ibid, 1977, 99, 921
⁴ L. E. Brown and G. A. Hamilton, ibid, 1970, 92, 7225
⁵ T. W. Chan and T. C. Bruice, J. Am. Chem. Soc., 1977, 99, 2387, W. K. Robbins and R. H. Eastman, ibid, 1970, 92, 6077
⁶ T. C. Bruice, Prog. Bioorg. Chem., 1976, 4, 1
⁷ C. Kemal, T. W. Chan, and T. C. Bruice, J. Am. Chem. Soc., 1977, 99, 7272, and references therein
⁸ G. A. Russell, E. G. Janzen, A. G. Bemis, E. J. Geels, A. J. Moye, S. Mak, and E. T. Strom, Adv. Chem. Ser., 1965, 51, 112, G. A. Russell, A. G. Bemis, E. J. Geels, E. G. Janzen, and A. J. Moye, ibid, 1968, 75, 174