

## FLAVONOID AND TERPENOID CONSTITUENTS OF *EUPATORIUM ODORATUM*

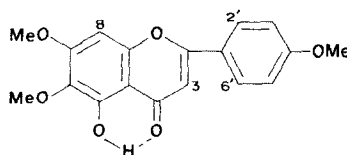
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**Key Word Index**—*Eupatorium odoratum*, Compositae flavone salvigenin triterpenoids lupeol  $\beta$ -amyrin

*Eupatorium odoratum* (Compositae, Subfam. Corymbiferae), distributed in temperate Himalayas, belongs to the tribe Eupatorieae.<sup>1</sup> The plant is used in the country as fish poison.<sup>2</sup> Isolation of a number of flavonoid compounds<sup>3</sup> from several *Eupatorium* species,<sup>4</sup> and a host of sesquiterpenoids from different Compositae plants prompted us to investigate the constituents of *Eupatorium odoratum*. Earlier workers reported a sesquiterpene alcohol eupatol,<sup>5</sup> in its essential oil, and a trihydric alcohol,  $C_{25}H_{34}O_5$  (m.p. 278–280°) along with anisic acid.<sup>6</sup> The present study has revealed the occurrence of two common triterpene alcohols, lupeol and  $\beta$ -amyrin, and a rarely occurring flavone, salvigenin (**1**) (first natural occurrence in Compositae) reported recently from *Salvia triloba*<sup>7</sup> (Labiatae) and *Alnus japonica*<sup>8</sup> (Betulaceae).



(1)

The cold light petrol (b.p. 60–80°) extract of the whole plant (3 kg) upon concentration yielded an oily residue (free from basic component). The latter was chromatographed over silica gel (660 g). Elution with  $CHCl_3$  gave a solid crystallizing from  $CHCl_3$ -light petrol in pale yellow needles (yield 0.01%), m.p. 185–186°,  $R_f$  0.4 (TLC on silica gel,  $CHCl_3$ ), analyzed for  $C_{18}H_{16}O_6$  ( $M^+$  328). A pink coloration in Shinoda test and the UV spectrum [ $\lambda_{max}^{OH}$  274 nm ( $\epsilon$  18 700) and 330 nm ( $\epsilon$  26 500)] are characteristic of a flavonoid. The compound gave an intense green colour with  $FeCl_3$  but no IR band for OH indicating the probable presence of a 5-OH. This was supported by the appropriately deshielded phenolic proton (1H,  $\delta$  12.76 ppm, exchangeable with  $D_2O$ ) and IR bands at 1635

<sup>1</sup> BENTHAM G. (1865) *Hand Book of British Flora* Vol 1 p. 413 Lovell Reeve London

<sup>2</sup> CHOPRA R. N., NAYL S. L. and CHOPRA I. C. (1956) *Glossary of Indian Medicinal Plants* p. 113 Publications and Information Directorate CSIR New Delhi

<sup>3</sup> HARBORNI J. B. (1967) *Comparative Biochemistry of Flavonoids* Academic Press London

<sup>4</sup> KUPCHAN, S. M., SIGEL, C. W., HINGWAY, R. J., KNOX, J. R. and UDAYAMURTHI, M. S. (1969) *Tetrahedron* **25**, 1603; WAGNER, H., JYNGER, M. A., HORHAMMER, L. and HERZ, W. (1972) *Phytochemistry* **10**, 1504

<sup>5</sup> CHOPRA, R. N., CHOPRA, I. C. and VARMA, B. S. (1969) *Supplement to the Glossary of Indian Medicinal Plants* p. 28 Publications and Information Directorate CSIR New Delhi

<sup>6</sup> AHMAD, M. (1969) *Sci. Res. (Dacca Pak)* **6**, 37

<sup>7</sup> ULUBELIN, A., OZTURK, S. and ISILDATICI, S. (1968) *J. Pharm. Sci.* **57**, 1037

<sup>8</sup> WOLLINWFER, E. and WASSER, M. (1972) *Tetrahedron Letters* 797

(chelated CO) and  $830\text{ cm}^{-1}$  (*p*-substituted phenyl ring) The 100 MHz PMR spectrum ( $\text{CDCl}_3$ ,  $\delta$ ) displayed the following signals in addition to the signal at 12.76 ppm which fit in well with structure (1) 3.90, 3.94, 3.98 (3H, *s* each, three OMe's), 6.54 and 6.58 (1H, *s* each, H-3 and H-8), 7.84\* (2H, *d*, H-2' and H-6'), 7.02\* ppm (2H, *d*, H-3' and H-5',  $J_{2',3'}$  or  $J_{5',6'}$  9 Hz) All the above data suggested its identity with salvigenin (1), which was confirmed by the preparation of the corresponding acetate, ( $\text{Ac}_2\text{O}$ -Py,  $R_f$  24 hr), m.p. 166–167° (lit.<sup>8</sup> 169–172°) and the fully methylated product ( $\text{MeI}$ - $\text{K}_2\text{CO}_3$  in refluxing acetone, 6 hr), m.p. 159–160° (lit.<sup>9</sup> 162–163°) The MS measured in a Hitachi Perkin Elmer Spectrometer at 70 eV [ $m/e$  328 ( $\text{M}^+$ ), 313, 299, 285, 282, 269, 253, 214, 181, 167, 153, 135, 133, 132] is in full agreement with the assigned structure and is similar† to that reported earlier.<sup>8</sup>

From the main chromatogram,  $\text{C}_6\text{H}_6$  eluted in poor yield (0.005%) a solid containing mainly lupeol ( $R_f$  0.38) mixed with small amount (estimated to be 3–5% from relative intensities of the spots) of  $\beta$ -amyrin ( $R_f$  0.42) as evident from TLC comparison on silica gel plates impregnated with 12.5%  $\text{AgNO}_3$  ( $\text{C}_6\text{H}_6$ -50%  $\text{CHCl}_3$ , spots developed by spraying with  $\text{EtOH}$ -5%  $\text{Ac}_2\text{O}$ -5% conc  $\text{H}_2\text{SO}_4$  followed by heating at 120° for 15 min) with authentic materials and their respective acetates (lupeol acetate 0.35,  $\beta$ -amyrin acetate 0.54-light petrol-50%  $\text{C}_6\text{H}_6$ ) and benzoates (lupeol benzoate 0.86,  $\beta$ -amyrin benzoate 0.92-light petrol-50%  $\text{C}_6\text{H}_6$ ) Several successive chromatographies of the crude acetate over Brockmann alumina afforded pure lupeol acetate, m.p. 215°, [hydrolysis product, m.p. 213° ( $\text{CHCl}_3$ - $\text{MeOH}$ ),  $[\alpha]_D^{+26}$ , identified (m.m.p., IR) as lupeol] The other fractions contained lupeol acetate enriched with  $\beta$ -amyrin acetate which, however, could not be isolated in pure state due to paucity of material

The cold  $\text{CHCl}_3$  extract of the marc, when processed in a similar way, yielded the same compounds in low yields, (salvigenin 0.006%, lupeol and  $\beta$ -amyrin mixture 0.0014%) A voucher specimen No T/E o/3/72 has been preserved in this laboratory

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\* Ulubelen *et al.*<sup>7</sup> erroneously assigned H-2', H-3', H-5' and H-6' as singlets at 7.93, 7.1, 6.93 and 7.68 ppm respectively

† The intensities of the peaks at  $m/e$  167 (4.4%) and 135 (8.8%) were much less than those reported earlier (51 and 19% respectively)

<sup>9</sup> GEISSMAN, T. A. (1962) *The Chemistry of Flavonoid Compounds*, p. 421, Pergamon Press, Oxford