# SYNTHESIS OF DI- AND TRI-SACCHARIDES CORRESPONDING TO RECEPTOR STRUCTURES RECOGNISED BY PYELONEPHRITOGENIC *E. coli* FIMBRIAE (PILI)

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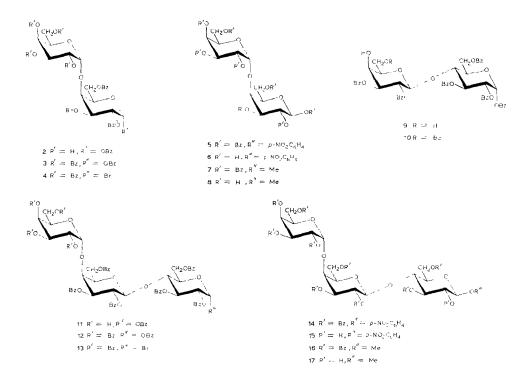
### ABSTRACT

Syntheses are described of di- and tri-saccharides required for studies of the inhibition of adhesion by means of fimbriae of pyelonephritogenic *E. coli* bacteria to epithelium cells in the urinary tract containing suitable receptor sites. The di-saccharides are the *p*-nitrophenyl and methyl glycosides of  $4-O-\alpha$ -D-galactopyranosyl- $\beta$ -D-galactopyranose, and the trisaccharides are the *p*-nitrophenyl and methyl glycosides of  $4-O-(4-O-\alpha-D-galactopyranosyl-<math>\beta$ -D-galactopyranosyl- $\beta$ -D-galactopyranosyl)- $\beta$ -D-glucopyranose. The key aglycons were the 1,2,3,6-tetrabenzoate of  $\alpha$ -D-galactose and the 1,2,3,6,-2',3',6'-heptabenzoate of  $\alpha$ -lactose. Glycosidations were performed with 2,3,4,6-tetra-O-benzyl- $\alpha$ -D-galactopyranosyl chloride as glycosylating agent and silver triflate as promoter.

## INTRODUCTION

Adhesion of *E. coli* to the urinary-tract epithelium is related to the specific binding to receptor structures present on the urinary-tract cells. These receptor structures are present not only on the urinary-tract epithelium, but also on human erythrocytes<sup>1</sup>. The result of haemagglutination studies suggested that the receptor structure relevant for pyelonephritogenic *E. coli* was the trihexosylceramide glycosphingolipid corresponding to the P<sup>k</sup> antigen<sup>2</sup>. The structure of the oligosaccharide moiety of this glycosphingolipid is  $4 - O - (4 - O - \alpha - D - \text{galactopyranosyl} - \beta - D - \text{galactopyranose}$ . In order to study these interactions, the following diand tri-saccharides were synthesised: *p*-nitrophenyl  $4 - O - (4 - O - \alpha - D - \text{galactopyranoside} (8)$ , *p*-nitrophenyl  $4 - O - (4 - O - \alpha - D - \text{galactopyranosyl} - \beta - D - \text{glucopyranoside} (15)$ , and methyl  $4 - O - (4 - O - \alpha - D - \text{galactopyranosyl} - \beta - D - \text{glucopyranosyl} - \beta - D - \text{glucopyranoside} (17). Of these, 17 has previously been synthesised<sup>3</sup> by a different route.*$ 

<sup>\*</sup>After the submission of the present paper, the synthesis of 8 by a different route was reported [M.-L. Milat, P. A. Zollo, and P. Sinaÿ, *Carbohydr. Res.*, 100 (1982) 263-271].



**RESULTS AND DISCUSSION** 

D-Galactose was benzoylated with 4 mol of benzoyl chloride in pyridine at  $0^{\circ}$ , and subsequent chromatography yielded 38% of the 1,2,4,6-tetrabenzoate 1. The constitution of 1 was proved, inter alia, by methylation analysis, including methylation with methyl triflate<sup>4,5</sup>. Glycosidation of 1 with 2,3,4,6-tetra-O-benzyl- $\alpha$ -Dgalactopyranosyl chloride, using silver triflate as promoter<sup>6</sup>, followed by catalytic hydrogenolysis of the product yielded 77% of the  $\alpha$ -galactoside 2. A high yield on glycosidation of a sterically hindered hydroxyl group using this galactosylation procedure was demonstrated earlier<sup>7</sup>. The disaccharide 2 was fully benzoylated to give the octabenzoate 3 (68% from 1). Treatment of 3 with hydrogen bromide in acetic acid gave the  $\alpha$ -bromide 4, which was converted into the protected *p*-nitrophenyl disaccharide 5 by using silver imidazolate and zinc chloride as promoter<sup>8</sup> for the glycosidation reaction. Debenzovlation of 5 afforded the first target substance 6. Silver triflate-promoted treatment of 4 with methanol gave the methyl  $\beta$ -D-glycoside 7, debenzovlation of which afforded the second target substance 8. The anomeric configurations of 6 and 8 were demonstrated by <sup>1</sup>H- and <sup>13</sup>C-n.m.r. spectroscopy, and the  $(1 \rightarrow 4)$  linkages by methylation analysis<sup>9</sup>.

A synthesis strategy analogous to that described above was used in order to obtain the remaining two target substances 15 and 16. Partial benzoylation of  $9^{10}$  with benzoyl chloride in pyridine at 0° afforded 75% of the lactose derivative 10

having the axial HO-4' free; 10 was then galactosylated with 2,3,4,6-tetra-O-benzyl- $\alpha$ -D-galactopyranosyl chloride, and the product was debenzylated by catalytic hydrogenolysis to give the trisaccharide heptabenzoate 11. Benzoylation of 11 yielded 12 (59% from 9) which, with hydrogen bromide and glacial acetic acid in dichloromethane, afforded the  $\alpha$ -bromide 13. The reaction of 13 with *p*-nitrophenol gave the glycoside 14 and the reaction with methanol gave the glycoside 16, as in the corresponding syntheses of glycosides 6 and 8 described above. Debenzoylation of 14 and 16 yielded the target compounds 15 and 17, respectively. The structures of 15 and 17 were confirmed by <sup>1</sup>H- and <sup>13</sup>C-n.m.r. spectroscopy, and by methylation analysis<sup>9</sup>.

The above routes for synthesis demonstrate facile access to galactose and lactose derivatives having free axial HO-4 and HO-4', respectively. These are useful aglycons for the synthesis of  $\alpha$ -D-galactosides using silver triflate as promoter. The synthesis route is flexible, in that various aglycons may be linked to C-1 of the disaccharides obtained.

Immunological studies using the above di- and tri-saccharides have been described elsewhere<sup>11</sup>.

#### EXPERIMENTAL

General methods were the same as those previously reported<sup>7</sup>. Column chromatography on silica gel was performed in the flash mode<sup>12</sup>. Sugar<sup>13,14</sup> and methylation<sup>9</sup> analyses were in agreement with the postulated structures.

1,2,3,6-Tetra-O-benzoyl- $\alpha$ -D-galactopyranose (1). — A solution of benzoyl chloride (16 g) in pyridine (5 mL) was added dropwise to a stirred suspension of D-galactose (5.0 g) in pyridine (100 mL) at 0°. After 1 h, ice was added and the mixture was extracted with dichloromethane. The organic layer was washed with 2M sulfuric acid, saturated aqueous sodium hydrogencarbonate and water, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. Column chromatography (toluene-ethyl acetate, 4:1) then afforded amorphous 1 (6.0 g, 38%),  $[\alpha]_D^{22} + 147°$  (c 0.8, chloroform). N.m.r. data: <sup>1</sup>H (100 MHz, CDCl<sub>3</sub>, internal Me<sub>4</sub>Si),  $\delta$  6.80 (d, 1 H,  $J_{1,2}$  4 Hz, H-1); <sup>13</sup>C (25 MHz, CDCl<sub>3</sub>, internal Me<sub>4</sub>Si),  $\delta$  90.9 (C-1). Treatment of 1 in sequence with methyl triflate<sup>4</sup>, sodium methoxide, sodium borohydride, and acetic anhydride-pyridine gave 1,2,3,5,6-penta-O-acetyl-4-O-methylgalactitol as the only product, as shown by g.l.c.-m.s.

1,2,3,6-Tetra-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- $\alpha$ -D-galactopyranosyl)- $\alpha$ -D-galactopyranose (3). — A solution of 2,3,4,6-tetra-O-benzyl- $\alpha$ -D-galactopyranosyl chloride<sup>7</sup> (10.0 g) in dry toluene (20 mL) was added dropwise to a stirred mixture of 1 (4.0 g), silver trifluoromethanesulfonate (5.5 g), 2,4,6-trimethylpyridine (2.1 g), and dry toluene (20 mL) at  $-10^{\circ}$ . The mixture was allowed to attain room temperature, and then stirred overnight, filtered, washed with dilute hydrochloric acid, water, saturated aqueous sodium hydrogencarbonate, and water, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The syrupy residue was hydrogenolysed overnight in glacial acetic

acid (20 mL) over 10% Pd/C (0.2 g). The mixture was filtered and concentrated, and the residue was subjected to column chromatography (chloroform-methanol, 9:1), to yield syrupy **2** (4.9 g, 77%),  $[\alpha]_{D}^{22}$  +166° (c 1.5, chloroform). N.m.r. data: <sup>1</sup>H,  $\delta$  6.80 (d, 1 H,  $J_{1,2}$  4 Hz, H-1); <sup>13</sup>C,  $\delta$  90.9 (C-1) and 101.1 (C-1').

Benzoyl chloride (5.8 g) was added dropwise with stirring to a solution of **2** (4.0 g) in pyridine (100 mL) at 0°. After 2 h at room temperature and work-up as described above in the preparation of **1**, the product was purified by column chromatography (toluene–ethyl acetate, 12:1), to yield syrupy **3** (5.5 g, 89°,  $_{0}$ ),  $[\alpha]_{D}^{22}$  +182° (c 1.3, chloroform). N.m.r. data: <sup>1</sup>H,  $\delta$  6.89 (d, 1 H,  $J_{1,2}$  4 Hz, H-1);<sup>13</sup>C,  $\delta$  90.8 (C-1) and 98.7 (C-1').

p-Nitrophenyl 2,3,6-tri-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- $\alpha$ -D-galactopyranosyl)- $\beta$ -D-galactopyranoside (5). — Compound 3 was treated with acetic acid saturated with hydrogen bromide, and the resulting glycosyl bromide 4, obtained after the usual work-up<sup>15</sup>, was used without purification. A mixture of 4 (0.90 g), p-nitrophenol (0.16 g), silver imidazolate (0.10 g), and dry zinc chloride (0.60 g) in dichloromethane (25 mL) containing 4Å molecular sieves was stirred in the dark overnight at 40° and then filtered, washed with saturated aqueous sodium hydrogencarbonate and water, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The product was purified by column chromatography [light petroleum (b.p. 40-60°)-chloroformethyl acetate, 3:1:1], to yield amorphous 5 (0.80 g, 87%),  $[\alpha]_D^{22} + 124°$  (c 1.1, chloroform). <sup>13</sup>C-N.m.r. data:  $\delta$  98.3 and 98.7 (C-1 and C-1').

Anal. Calc. for  $C_{67}H_{53}NO_{20}$ : C, 67.5; H, 4.48; N, 1.17. Found: C, 67.7; H, 4.59; N, 1.19.

p-Nitrophenyl 4-O- $\alpha$ -D-galactopyranosyl- $\beta$ -D-galactopyranoside (6). — Debenzoylation of 5 (0.12 g) with a catalytic amount of sodium methoxide in methanol followed by elution of the product from a column of Bio-gel P-2 afforded amorphous 6 (50 mg, 91 %),  $[\alpha]_D$  +53° (c 1, chloroform). N.m.r. data (D<sub>2</sub>O): <sup>1</sup>H,  $\delta$  5.02 (d, 1 H,  $J_{1',2'}$  1 Hz, H-1') and 5.23 (d, 1 H,  $J_{1,2}$  8 Hz, H-1); <sup>13</sup>C,  $\delta$  102.0 and 102.6 (C-1, C-1').

Methyl 2,3,6-tri-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- $\alpha$ -D-galactopyranosyl)- $\beta$ -D-galactopyranoside (7). — A solution of 4 (0.60 g) in toluene (2 mL) was added with stirring to a mixture of silver trifluoromethanesulfonate (0.154 g), 2,4,6-trimethylpyridine (30 mg), methanol (60 mg), and toluene (5 mL) at  $-0^{\circ}$ . The mixture was allowed to attain room temperature, and was then stirred overnight, washed successively with aqueous sodium thiosulfate, water, 2M sulfuric acid, and saturated aqueous sodium hydrogencarbonate, dried (MgSO<sub>4</sub>), filtered, and concentrated. The product was purified by column chromatography (toluene-ethyl acetate, 2:1), to yield 7 (0.50 g, 90%), m.p. 222–223°,  $[\alpha]_D$  +96° (c 1.3, chloroform). <sup>13</sup>C-N.m.r. data:  $\delta$  56.4 (OMe), 98.4 (C-1'), and 101.8 (C-1).

Anal. Calc. for C<sub>62</sub>H<sub>52</sub>O<sub>18</sub>: C, 68.6; H, 4.83. Found: C, 68.6; H, 4.94.

Methyl 4-O- $\alpha$ -D-galactopyranosyl- $\beta$ -D-galactopyranoside (8). — Debenzoylation of 7 (0.10 g) with a catalytic amount of sodium methoxide in methanol followed by elution of the product from a column of Bio-gel P-2 afforded amorphous 8

(28 mg, 85%),  $[\alpha]_D^{22}$  +88° (c 1.2, water). N.m.r. data (D<sub>2</sub>O): <sup>13</sup>C,  $\delta$  58.3 (OMe), 101.3 (C-1'), and 104.8 (C-1); <sup>1</sup>H,  $\delta$  4.32 (d, 1 H,  $J_{1,2}$  7.1 Hz, H-1) and 4.96 (d, 1 H,  $J_{1',2'}$  2.2 Hz, H-1').

1,2,3,6-Tetra-O-benzoyl-4-O-(2,3,6-tri-O-benzoyl-β-D-galactopyranosyl)-α-Dglucopyranose (10). — A solution of benzoyl chloride (10.7 g) in pyridine (5 mL) was added dropwise to a solution of 1,2,3,6,2',3'-hexa-O-benzoyl-α-D-lactose<sup>10</sup> (9, 4.0 g) in pyridine (50 mL) at 0°. After 2 h at room temperature, the mixture was poured onto ice and stirred for 30 min. The crystals were collected, washed with cold water, and recrystallised from ethanol, to yield 10 (3.5 g, 75%). Conventional work-up and chromatography of the mother liquor yielded more (0.60 g, 15%) 10, m.p. 196–197°,  $\lceil \alpha \rceil_{D}^{22} +98°$  (c 1, water). <sup>13</sup>C-N.m.r. data: δ 89.8 (C-1) and 101.3 (C-1').

Anal. Calc. for C<sub>61</sub>H<sub>50</sub>O<sub>18</sub>: C, 68.4; H, 4.70. Found: C, 68.1; H, 4.69.

1,2,3,6-Tetra-O-benzoyl-4-O-[2,3,6-tri-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl-α-D-galactopyranosyl)-β-D-galactopyranosyl]-α-D-glucopyranose (12). — Compound 10 (2.50 g) was treated with 2,3,4,6-tetra-O-benzyl-α-D-galactopyranosyl chloride<sup>7</sup> (2.5 g), silver trifluoromethanesulfonate (1.20 g), and 2,4,6-trimethyl-pyridine (0.50 g) in toluene (20 mL), the mixture was worked-up, and the product hydrogenolysed, as described above, to yield amorphous 11 (1.80 g, 64%),  $[\alpha]_D^{22}$  +124° (c 1, chloroform). <sup>13</sup>C-N.m.r. data: δ 89.7 (C-1), 100.2 (C-1″), and 101.6 (C-1′).

Benzoylation of 11 (1.5 g), as described above, yielded amorphous 12 (1.84 g, 92%),  $[\alpha]_D^{22} + 72^\circ$  (c 1, chloroform). <sup>13</sup>C-N.m.r. data:  $\delta$  89.8 (C-1), 98.9 (C-1"), and 101.8 (C-1').

p-Nitrophenyl 2,3,6-tri-O-benzoyl-4-O-[2,3,6-tri-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- $\alpha$ -D-galactopyranosyl)- $\beta$ -D-galactopyranosyl]- $\beta$ -D-galactopyranoside (14). — A solution of 12 in dry dichloromethane was treated with glacial acetic acid saturated with hydrogen bromide. After 1 h at room temperature, the solution was washed with water, saturated aqueous sodium hydrogencarbonate, and water, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated, to give 13 which was used directly.

Compound 13 (0.95 g) was treated with *p*-nitrophenol (0.13 g), silver imidazolate (0.084 g), and zinc chloride (0.80 g) in dry dichloromethane (30 mL) containing powdered 4Å molecular sieves. Work-up, as described above, gave amorphous 14 (0.80 g, 80%),  $[\alpha]_D^{22} + 51^\circ$  (*c* 1, chloroform). <sup>13</sup>C-N.m.r. data:  $\delta$  97.4 (C-1), 98.8 (C-1"), and 101.4 (C-1').

p-Nitrophenyl 4-O-(4-O- $\alpha$ -D-galactopyranosyl- $\beta$ -D-galactopyranosyl)- $\beta$ -D-glucopyranoside (15). — Debenzoylation of 14 (0.50 g) and purification of the product, using the procedures described above, yielded 15 (0.17 g, 91%),  $[\alpha]_D^{22} + 12^\circ$  (c 0.76, water). N.m.r. data (D<sub>2</sub>O): <sup>1</sup>H,  $\delta$  4.51 (d, 1 H,  $J_{1',2'}$  7 Hz, H-1'), 4.96 (d, 1 H,  $J_{1'',2''}$ 2 Hz, H-1"), and 5.21 (d, 1 H,  $J_{1,2}$  7.3 Hz, H-1); <sup>13</sup>C,  $\delta$  100.4 (C-1"), 101.5 (C-1), and 104.5 (C-1').

Methyl 2,3,6-tri-O-benzoyl-4-O-[2,3,6-tri-O-benzoyl-4-O-(2,3,4,6-tetra-O-benzoyl- $\alpha$ -D-galactopyranosyl)- $\beta$ -D-galactopyranosyl]- $\beta$ -D-galactopyranoside (16). — Compound 13 (1.5 g) was treated with methanol (0.50 mL), silver trifluoromethanesulfo-

nate (0.20 g), and 2,4,6-trimethylpyridine (0.09 g) in dry toluene (30 mL) as described above. Column chromatography (toluene-ethyl acetate, 15:1) of the product afforded **16** (1.2 g, 76%), m.p. 223–224° (from ethanol),  $[\alpha]_D$  +70° (c 1, chloroform). <sup>13</sup>C-n.m.r. data:  $\delta$  98.8 (C-1″), 101.3 (C-1), and 101.6 (C-1′).

Anal. Calc. for C<sub>89</sub>H<sub>74</sub>O<sub>26</sub>: C, 68.5; H, 4.78. Found: C, 68.5; H, 4.85.

*Methyl* 4-O-(4-O- $\alpha$ -D-galactopyranosyl- $\beta$ -D-galactopyranosyl)- $\beta$ -D-glucopyranoside (17). — Treatment of 16 (0.70 g) with a catalytic amount of sodium methoxide in methanol afforded, after chromatography on Bio-gel P-2, amorphous 17 (0.20 g, 86%),  $[\alpha]_{\rm D}$  +65° (c 1, water); lit.<sup>3</sup>  $[\alpha]_{\rm D}$  +63° (water). N.m.r. data (D<sub>2</sub>O): <sup>1</sup>H,  $\delta$  4.94 (d, 1 H,  $J_{1",2"} \sim 1.7$  Hz, H-1"), 4.47 (d, 1 H,  $J_{1',2'} \sim 7.6$  Hz, H-1'), and 4.34 (d, 1 H,  $J_{1,2} \sim 7.8$  Hz, H-1); <sup>13</sup>C,  $\delta$  58.4 (OMe), 61.3 (C-6), 61.6 (C-6'), 61.7 (C-6"), 101.5 (C-1"), 104.2 (C-1'), and 104.5 (C-1).

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#### REFERENCES

- 1 R. RACE AND R. SANGER, Blood Groups in Man, 6th edn., Blackwell, Oxford, 1975.
- 2 G. Källenius, R. Möllby, S. B. Svenson, J. Winberg, A. Lundblad, S. Svensson, and B. Cedergren, *FEMS Microbiol. Lett.*, 7 (1980) 297–302.
- 3 D. D. Cox, E. K. METZNER, AND E. J. REIST, Carbohydr. Res., 63 (1978) 139-147.
- 4 J. ARNARP, L. KENNE, B. LINDBERG, AND J. LÖNNGREN, Carbohydr. Res., 44 (1975) C5-C7.
- 5 J. M. BERRY AND L. D. HALL, Carbohydr. Res., 47 (1976) 307-310.
- 6 R. U. LEMIEUX, R. M. RATCLIFFE, B. ARREGUIN, A. R. DE VIVAR, AND M. J. CASTILLO, *Carbohydr. Res.*, 55 (1977) 113–120.
- 7 P. J. GAREGG, H. HULTBERG, AND C. LINDBERG, Carbohydr. Res., 83 (1980) 157-162.
- 8 P. J. GAREGG, H. HULTBERG, C. ORTEGA, AND B. SAMUELSSON, Acta Chem. Scand., Ser. B, 36 (1982) 513-514.
- 9 H. BJÖRNDAL, C. G. HELLERQVIST, B. LINDBERG, AND S. SVENSSON, Angew. Chem. Int. Ed. Engl., 9 (1970) 610-619.
- 10 H. H. BAER AND S. ABBAS, Carbohydr. Res., 77 (1979) 117-129.
- 11 G. KÄLLENIUS, R. MÖLLBY, S. B. SVENSON, J. WINBERG, AND H. HULTBERG, *Infection*, 8 (1980) 288–293.
- 12 W. C. STILL, M. KAHN, AND A. MITRA, J. Org. Chem., 43 (1978) 2923-2925.
- 13 J. S. SAWARDEKER, J. H. SLONEKER, AND A. JEANES, Anal. Chem., 37 (1965) 1602-1604.
- 14 O. S. CHIZHOV, L. S. GOLOVKINA, AND N. S. WULFSON, Izv. Akad. Nauk SSSR, Ser. Khim., (1966) 1915.
- 15 P. J. GAREGG, B. LINDBERG, AND T. NORBERG, Acta Chem. Scand., Ser. B, 33 (1979) 449-452.