THE BITTER IRIDOIDS FROM VIBURNUM URCEOLATUM

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Abstract—Four new bitter iridoid glucosides, including three bis-iridoids, were isolated from *Viburnum urceolatum* and their structures elucidated. They are composed of loganin or deoxyloganin as the aglucone esterified with either aromatic glucoside or with glucose in the 4-position.

INTRODUCTION

In the course of an investigation on the bitter principles of *Viburnum* species, we investigated a monoterpenoid glycoside, urceolide, isolated from *Viburnum urceolatum* [1]. As a result of our continued work on the same plant, the elucidation of the structures of three new bis-iridoids and an iridoid is reported now.

All of the new iridoids (named urceolatoside A-D) were recognized as the sources of bitterness from V. *urceolatum*.

RESULTS AND DISCUSSION

The isolation and purification of the compounds are described in detail in the Experimental.

Compound 1, urceolatoside A, $C_{33}H_{42}O_{14}$, amorphous; mp 136–140°, $[\alpha]_D - 56^\circ$, showed a typical iridoid colour reaction by hydrochloric acid, and the IR spectrum showed strong absorption of a hydroxyl group (3400 cm⁻¹), benzene ring (1630, 1528), unsaturated ester (1720) and enol ether (1635). The ¹H NMR spectrum confirmed the existence of these functional groups $[\delta 1.08]$



(6H, d, J = 7 Hz), 4.93 (2H, m), 5.02 (2H, s), 6.99, 7.30 (4H, A_2B_2 , J = 10 Hz) and 7.38 (2H, br s)]. Acetylation of 1 gave a hexa-acetate (2), $C_{45}H_{54}O_{20}$, amorphous; $[\alpha]_D - 55^\circ$, whose IR spectrum showed no hydroxyl absorption, and in the ¹H NMR spectrum, the signals from six acetoxyl groups were seen. Furthermore, doublets at $\delta 5.93$ and 6.00 showed the presence of two acetylated acetalic centers in 2.

Upon methanolysis under mild conditions, 1 gave an anomeric mixture of dimethoxy compound 3 as seen from the ¹H NMR spectrum. Acetylation gave the corresponding mixture of tetra-acetates (4) which was separated into the diastereoisomeric mixtures of 4a $[\alpha]_D + 61^\circ$ (major product) and 4b $[\alpha]_D - 8^\circ$. Compound 4a was hydrolysed with 2 N sodium hydroxide to afford two anomeric mixtures of carboxylic acids (5 and 6), and a phenolic glycoside (7). Compound 5, $C_{11}H_{16}O_4$, was identical (IR, ¹H NMR) with authentic 1-0-methyl deoxyloganic acid aglycone (anomeric mixture) and 6 was further purified to give 6a and 6b, both having molecular formula of $C_{11}H_{16}O_5$, and their methyl esters ($[\alpha]_D - 23.5^\circ$ and $+ 190^\circ$) were identical (IR, ¹H NMR, $[\alpha]_D$) with those reported for 1-0-methyl loganin aglycone and its anomer, respectively [2–4]. Compound 7 was hydrolysed with hydrochloric acid to give *p*-hydroxybenzyl alcohol and pglucose. In the ¹H NMR spectrum, the signal due to the benzylic protons of 7 originally at $\delta 4.80$ (2H, *s*, pyridine d_5) was shifted to 5.00 (CDCl₃)* by acetylation, and the chemical shift of the acetoxyl groups of the penta-acetate (7a) were at 2.07 and 2.08 (total 3H × 5). These data indicated that 7 had a benzylic hydroxyl group. The $W_{1/2}$ value of the anomeric proton of 7a (at $\delta 5.68$, J = 10 Hz) suggested that the glycosidic linkage must be β in 7. Thus, the structure of 7 was determined to be *p*-(hydroxymethyl)phenyl- β -p-glucoside.

Treatment of **4a** with dilute sodium hydroxide in methanol gave an ester (**8**) as a mixture of diastereoisomers, $C_{23}H_{32}O_8$. The IR spectrum (1720, 1636, 960 and 930 cm⁻¹) and ¹H NMR spectrum [δ 1.05 (3H, d, J = 6 Hz), 1.07 (3H, d, J = 6 Hz), 3.48. 3.60, 3.72 (total 9H, s) and 7.50 (2H, s)] suggested that **8** had a bis-iridoid structure. The chemical shift (δ 5.02) of the signal of the benzylic protons of **1** showed the existence of acylated benzyl methylene in **1**. From the above data, the structure of **1** was determined to be as shown.

Compound 9, urceolatoside B, was isolated as white crystals, mp 148–152°, $[\alpha]_D = -15^\circ$, with a molecular formula $C_{26}H_{36}O_{13}\cdot 1/2$ H₂O. Upon hydrolysis with acidic resin, 9 yielded D-glucose. The UV absorption at 246 nm (ε 11 000) was characteristic of the conjugated enol ether system which was confirmed by IR absorptions at 1720–1680 and 1620 cm⁻¹. The ¹H NMR spectrum exhi-



^{*}The chemical shift of benzylic protons in the spectrum (in $CDCl_3$) of *p*-hydroxy benzylalcohol was $\delta 4.54$.

bited two singlets at δ 7.54 (1H) and 7.58 (1H) due to the olefinic protons at C-3 and C-3' of the iridoid, suggesting that 9 was likely to have a bis-iridoid structure. Acetylation of 9 afforded an amorphous hexa-acetate (10), C₃₈H₄₈O₁₉, whose ¹H NMR spectrum showed the signals due to six acetoxyl groups at $\delta 2.00$, 2.08 and 2.14. The three broad doublets which appeared at $\delta 5.97$ (1H, J = 3 Hz), 6.08 (1H, J = 2 Hz) and 6.30 (1H, J = 3 Hz) were assigned to acetylated hemiacetal protons. Methanolysis of 9 with methanol containing a catalytic amount of conc. hydrochloric acid vielded a diastereoisomeric mixture of methyl ethers (11), whose ¹H NMR spectrum showed signals at $\delta 3.40-3.52$ (total 9H, s). On acetylation with acetic anhydride-pyridine compound 11 afforded an amorphous triacetate (12) and its ¹H NMR spectrum indicated the presence of three acetoxyl groups at $\delta 2.00-2.10$ (3H × 3, s). Upon hydrolysis with 2 N sodium hydroxide, compound 12 afforded three acids which were identical with 5, 6a and 6b, respectively. On mild alkaline hydrolysis, 12 gave the same kind of diastereoisomeric mixture as 8a, C₂₂H₃₀O₅, and its methyl ester.

These facts confirmed that 9 had a bis-iridoid skeleton and C-1 of D-glucose in 9 was not acylated. In order to determine the position of the linkage between D-glucose and aglycone 8, compound 11 was permethylated by the Purdie method, followed by alkaline hydrolysis. This procedure gave a mixture of methyl-2,3,6-tri-O-methyl-Dglucoside, 5, 6a and 6b, showing that the ester linkage to the sugar was located at C-4 of D-glucose. The structure of 9 was, thus, shown to be as indicated.

Compound 14, (urceolatoside C), was an amorphous powder, $C_{33}H_{42}O_{15}$, $[\alpha]_D - 42.1^\circ$. The IR, UV and ¹H NMR spectra showed that 14 had an iridoid skeleton with an aromatic ring. [IR v_{max} cm⁻¹: 3400, 1695, 1633 and 1518; UV λ_{max} nm (ϵ): 228 (11 000) and 238 (sh 10 600); ¹H NMR: $\delta 1.07$ (6H, br d, J = 6Hz), 5.07 (2H, s), 7.04, 7.41 $(4H, A_2B_2, J = 9 Hz)$ and 7.44 (2H, br s)]. Acetylation of 14 afforded a hepta-acetate (15), amorphous, mp 88-89°, $C_{47}H_{56}O_{22}$, $[\alpha]_{D} - 22.5^{\circ}$ and its ¹H NMR spectrum confirmed the presence of seven aliphatic acetoxyl groups $[2.04 (3H \times 5, s) \text{ and } 2.09 (3H \times 2, s)]$. Upon mild acidic methanolysis, 14 gave a mixture of diastereoisomers (16) and the ¹H NMR spectrum revealed the presence of two methoxyl groups at δ 3.39 (s), 3.41 (s) and 3.44 (s) (total 6H). Without separation, the dimethyl ether, 16, was hydrolysed with 2 N sodium hydroxide to give an epimeric mixture of acids 6 (2 mol) which was separated into two pure acids, 6a and 6b, and a phenolic glucoside (1 mol) which was identical with 7. On acetylation, 16 gave a penta-acetate (17) whose IR spectrum showed no hydroxyl absorption and, in the ¹H NMR spectrum the signals due to the protons of five acetoxyl groups appeared at $\delta 2.05$ (3H × 5, s). These facts suggested that, like 1 and 9, compound 14 has a bis-iridoid structure in which two loganic acid aglycones are linked to each other through an ester linkage.

The Purdie methylation of 16 afforded a hepta-methyl ether (18) and a hexa-methyl ether (19). The minor product 18 was shown to be the permethyl ether of 14 by its IR (no hydroxyl absorption) and ¹H NMR spectra $[\delta 3.32, 3.39, 3.44, 3.49$ (3H each, s) and 3.59 (3H × 3, s)]. Upon alkaline hydrolysis, compound 18 gave two anomeric mixtures of carboxylic acids which were identified as 1,7-di-O-methyl loganic acid aglycone and 6, and a phenolic glycoside (20) which was hydrolysed with hydrochloric acid to give *p*-hydroxybenzyl alcohol and a methylated p-glucose identical with an authentic sample of 2,3,4,6-tetra-O-methyl-p-glucose. The IR spectrum of 19 showed the absorption of a hydroxyl group, and the ¹H NMR spectrum revealed the presence of six methoxyl groups [δ 3.36, 3.43, 3.49, 3.51 (3H each, *s*) and 3.60 (3H \times 2, *s*)]. Upon alkaline hydrolysis, compound 19 gave a single epimeric mixture of 6 (2 mol) and a phenolic methylated glycoside (1 mol) which was identical with 20. On the basis of the above data, compound 14 has the structure shown.

Compound 21 (urceolatoside D), was obtained as a hygroscopic powder, mp 153–161°, $C_{23}H_{30}O_{11}$, $[\alpha]_D$ -42.5°, whose UV (236 nm, ε 6500), IR (3400, 1700, 1632 and 1518 cm⁻¹) and ¹H NMR spectra [δ 1.10 (3H, d, J = 6 Hz), 1.40–2.50 (4H), 5.15 (2H, s), 7.19, 7.47 (4H, A₂B₂, J = 9 Hz) and 7.57 (1H, s)] indicated the presence of an aromatic ring and an iridoid skeleton besides a sugar moiety. Acetylation of 21 afforded a hexa-acetate (22) whose ¹H NMR spectrum showed the signals due to six acetoxyl groups $[\delta 2.05 (3H \times 5, s) \text{ and } 2.09 (3H, s)]$ and an acetylated hemiacetal proton [6.05 (1H, d, J = 3 Hz)]. Upon treatment with hydrochloric acid, 21 gave D-glucose and a black polymeric product. On mild methanolysis, 21 afforded an epimeric mixture (2:1) of the monomethyl ether (23). This mixture was acetylated without separation, to give a penta-acetate (24). Upon hydrolysis with alkali, 24 gave an epimeric mixture of carboxylic acids which was identical with 6 and which was purified further to give 6a and 6b, and a phenolic glycoside which was identical with 7. The chemical shift of the benzylic protons $(\delta 5.15)$ indicated the presence of an acylated benzylmethylene in 21. The structure of 21 was, therefore, revealed to be as shown.



In the ¹H NMR spectra of 1, 9, 14 and 21, the signals due to the hemiacetal protons of the iridoids and Dglucose were overlapped and the chemical shifts and coupling constants of the protons were not so clear as to permit the determination of the configurations of the anomeric carbons of the iridoid moieties. Also, the possibility that each of these compounds was a mixture of diastereoisomers, could not be excluded. However, all of these compounds may have the β -configuration at C-1 (s) of the loganic and deoxyloganic acid moieties, because each of them gave a single acetate and the iridoids, which have so far been reported as agluconic compounds, were indicated to be β -anomers [5]. The bis-iridoids, in which secoiridoids are included, have so far been isolated from *Cantleya* (Icacinaceae) [6], *Fraxinus* (Oleaceae) [7], *Dipsacus* (Dipsacaceae) [5] and *Erythraea* (Gentianaceae) [8] species and, recently, Uesato *et al.* [9] reported a dimeric iridoid in which the two iridoid moieties are linked through an acetal linkage. Compounds 1, 9, and 14 are the first examples of bisiridoids which consist of two intact iridoid aglucones joined through an ester linkage.

EXPERIMENTAL

Isolation. Fresh leaves of V. urceolatum (2 kg) were extracted with MeOH (2 × 10 l.). The combined MeOH solns were concd to dryness to afford a dark green residue (210 g). The residue was diluted with H₂O and extracted with Et₂O and then EtOAc. The EtOAc extract was evaporated to dryness (92 g), and the residue was extracted with CHCl₃–MeOH (93:7) to give a dark green syrup (37 g). The syrup was chromatographed on a column of Si gel, eluting with CHCl₃–MeOH with increasing MeOH content. From the fractions eluted with CHCl₃–MeOH (93:7), a monoterpenoid glycoside (130 mg) and 1 (500 mg) were afforded, successively.

The fractions eluted with $CHCl_3$ -MeOH (9:1) gave 9 (640 mg), and those eluted with $CHCl_3$ -MeOH (8:2) afforded 14 (420 mg) and 21 (400 mg) in succession.

Compound 1. Amorphous powder from MeOH and Et₂O, mp 136–140°, $\lceil \alpha \rceil_D^{25} = 56$ (MeOH; c 0.36), UV $\lambda \frac{\text{MeOH}}{\text{max}}$ nm (e): 204 sh (6600), 228 (18 000) and 240 (16 300); IR $\nu \frac{\text{Nijol}}{\text{max}}$ cm⁻¹: 3400, 1720, 1635 and 1520; ¹H NMR (60 MHz; CD₃COCD₃): δ1.08 (6H, d, J = 7 Hz, H-10 and H-10'), 4.93 (3H, m, H-1, H-1' and H-1"), 5.02 $(2H, s, benzyl H), 6.99, 7.30 (4H, A_2B_2, J = 10 Hz, Ar-H) and 7.38$ (2H, br s, H-3 and H-3'). In the ¹H NMR spectra of 1, 9, 14 and 21, the signals due to the protons of sugar moieties were seen in the range δ 3.40–5.20. (Found: C, 59.58; H, 6.01 % C₃₃H₄₂O₁₄ requires: C, 59.84; H, 6.39 %) Acetylation of 1 (80 mg) with Ac_2O -pyridine gave a hexa-acetate (2) (90 mg), amorphous, $[\alpha]_D^{25} = 55^{\circ}$ (MeOH; c 0.32), UV λ_{max}^{MeOH} nm (ϵ): 228 (27 700); IR ν_{max}^{Nujol} cm⁻¹: 1770, 1720, 1640 and 1520; ¹H NMR (100 MHz; CDCl₃): δ 1.11 (6H, br d, H-10 and H-10'), 2.03, 2.05, 2.12 (6H each, s, COMe), 5.93, 6.00 (1H each, d, J = 3 Hz, H-1 and H-1'), 6.96, 7.34 (4H, A_2B_2 , J = 9 Hz, Ar-H) and 7.37 (2H, br s, H-3 and H-3'). (Found: C, 58.73; H, 5.94 %. C45H54O20 requires: C, 59.08; H, 5.91 %.)

Methanolysis of 1. Compound 1 (200 mg) was dissolved in 10 ml dry MeOH containing one drop conc. HCl. The soln was stirred at room temp. under N2 stream; after 12 hr the soln was diluted with H₂O and then extracted with Et₂O. The Et₂O extract was chromatographed over a column of Si gel (CHCl₃-MeOH, 97:3) to give an anomeric mixture of the dimethyl ether (3), amorphous powder (200 mg), $[\alpha]_D^{25} - 25^{\circ}$ (MeOH; c 0.40), UV λ_{max}^{MeOH} nm (ϵ): 218 (10000); IR ν_{mx}^{fl} $n cm^{-1}$: 3400, 1710, 1690, 1630 and 1515; ¹H NMR (100 MHz; CD_3COCD_3): $\delta 1.12$ (6H, d, J = 7 Hz, H-10 and H-10'), 3.57–3.60 (total 6H, s, OMe-1 and OMe-1'), 5.28 (2H, s, benzyl H), 7.35 and 7.65 (4H, A_2B_2 , J = 10 Hz, Ar-H), and 7.38 (2H, br s, H-3 and H-3'). (Found: C, 60.66; H, 6.66 %. C₃₅H₄₆O₁₄ requires: C, 60.87; H, 6.67 %). Compound 3 (140 mg) was acetylated with Ac₂O-pyridine (each 0.5 ml), worked-up as usual, and the crude product (4) was purified by Si gel chromatography (CHCl₃-MeOH, 95:5) to give tetra-acetates 4a and 4b. Compound 4a (major product 64 mg), amorphous, $[\alpha]_D^{25} + 61^\circ$ (MeOH; c 0.18); IR v_{max}^{film} cm⁻¹: 1770, 1710, 1690, 1630 and 1518; ¹H NMR (100 MHz; CDCl₃): δ 1.04 (6H, d, J = 8 Hz, H-10 and H-10'), 2.20 (3H × 4, s, COMe), 3.38-3.53 (total 6H, s, OMe-1 and OMe-1'), 7.00 and 7.40 (4H, A_2B_2 , J = 10 Hz, Ar-H) and 7.51

(2H, br s, H-3 and H-3'). Compound **4b**, amorphous, $[\alpha]_{D}^{25} - 4^{\circ}$ (MeOH; c 0.26); IR v_{max}^{film} cm⁻¹: 1750, 1695, 1620 and 1500; ¹H NMR (100 MHz; CDCl₃): δ 1.05 (6H, m, H-10 and H-10'), 2.05 (3H × 4, s, COMe), 3.42–3.48 (total 6H, s, OMe-1 and OMe-1'), 6.98 and 7.33 (4H, A₂B₂, J = 10 Hz, Ar-H) and 7.45 (2H, br s, H-3 and H-3').

Alkaline hydrolysis of 4a. Compound 4a (200 mg) was refluxed with 2 N NaOH (20 ml) and MeOH (20 ml) for 30 min. The soln was acidified with HCl, extracted with Et₂O; the Et₂O extract was washed with H₂O, dried and evaporated to dryness. The residue was chromatographed on a column of Si gel to give oily 5 (55 mg) and 6 (48 mg). The H₂O layer was evaporated to dryness in vacuo, and the residue was dissolved in CHCl3-MeOH (4:1) and subjected to CC on Si gel. The fractions eluted with CHCl₃-MeOH (4:1) gave a crystalline phenolic glucoside, 7 (35 mg). Compound 5, IR v_{max}^{film} cm⁻¹: 2650–2350, 1680 and 1630; ¹H NMR (100 MHz; CDCl₃): δ 1.07 (3H, d, J = 7 Hz, H-10), 3.42-3.49 (total 3H, s, OMe-1), 4.52 and 4.86 (total 1H, d each, J = 7 and 4 Hz, H-1) and 7.47 (1H, s, H-3); MS m/z 212.1032 [M]⁺; C11H16O4 requires, 212.1047. Compound 5 was identical to an authentic sample of an epimeric mixture of 1-Omethyldeoxyloganic acid aglycone. Compound 6, $[\alpha]_D^{25} + 27.5^\circ$ (CHCl₃; c 0.40), was purified further by Si gel chromatography (CHCl₃-MeOH, 97:3) to give epimers 6a and 6b. Compound 6a, major epimer, IR v film cm⁻¹: 3400, 2650–2400, 1695, 1635 and 1190; ¹H NMR (100 MHz; CDCl₃): δ 1.10 (3H, d, J = 7 Hz, H-10), 1.60-2.40 (4H, m, H-6, H-8 and H-9), 3.10 (1H, m, H-5), 3.48 (3H, s, OMe-1), 4.12 (1H, m, H-7), 4.60 (1H, d, J = 4 Hz, H-1),5.20 (OH), 7.51 (1H, br s, H-3); MS m/z 228.1032 [M]⁺; C11H16O5 requires, 228.1010. Compound 6a was methylated with CH_2N_2 in Et_2O to give a methyl ester (10 mg) $[\alpha]_D^{25} - 23.8^\circ$ (CHCl₃; c 0.20); IR v_{max}^{film} cm⁻¹: 3450, 1705, 1633 and 1180; ¹H NMR (60 MHz; CDCl₃): δ 1.10 (3H, d, J = 6.5 Hz, H-10), 1.40-2.50 (4H, m, H-6, H-8 and H-9), 3.10 (1H, m, H-5), 3.45 (3H, s, OMe-1), 3.66 (3H, s, COMe), 4.08 (1H, m, H-7), 4.59 (1H, d, J = 4 Hz, H-1) and 7.40 (1H, d, J = 1.5 Hz, H-3); EIMS (70 eV) m/z (rel. int.): 242 [M]⁺ (6), 224 (5), 210 (7), 192 (11), 178 (8), 139 (9), 117 (14) and 85 (100).

These data were coincident with those of authentic (1*R*) 1-*O*methyl loganin aglycone [2-4]. Compound **6b** (8 mg) IR spectrum differed from that of **6a** only in the range 1100–900 cm⁻¹, ¹H NMR (60 MHz; CDCl₃): δ 1.10 (3H, *d*, *J* = 6.5 Hz, H-10), 3.43 (3H, *s*, OMe-1), 4.94 (1H, *d*, *J* = 3 Hz, H-1) and 7.56 (1H, *d*, *J* = 1 Hz, H-3); methyl ester of **6a** had an optical rotation of + 190° (CHCl₃; *c* 0.10) and the spectra of the ester were coincident with those of (1S) 1-*O*-methyl loganin aglycone in the lit. [2, 3].

1-O-methyl deoxyloganic acid aglycone (5) from deoxyloganin. Deoxyloganin (226 mg) was dissolved in a buffer soln (NaOAc-HOAc, pH 4.9, 9 ml) and 40 mg β -glucosidase was added; the soln was allowed to stand for 2 days at 38°, then extracted with Et₂O. The Et₂O extract was evaporated to dryness. Deoxyloganin aglycone (98 mg), oil, was afforded after purification by Si gel chromatography, IR v_{max}^{film} cm⁻¹: 3400, 1720, 1690 and 1630. The aglycone was dissolved in MeOH (2 ml) containing a drop of conc. HCl, stirred at room temp. overnight, then H₂O (20 ml) added. Extraction with Et₂O, and chromatography on a column of Si gel (CHCl₃-MeOH, 97:3) gave an ether, oil, (38 mg) IR v_{max}^{film} cm⁻¹: 1750, 1720 and 1630; ¹H NMR (100 MHz; CDCl₃): δ 1.07 (3H, d, J = 6 Hz, H-10), 3.48, 3.51 (total 3H, s, OMe-1), 3.70 (3H, s, COOMe). 4.54, 4.89 (total 1H, d each, J = 6 and 3 Hz, H-1) and 7.44 (1H, s, H-3). This was dissolved in 2 N NaOH (0.5 ml) and MeOH (1.5 ml), refluxed for 45 min and 10 ml H_2O added followed by extraction with Et_2O . The aq. layer was acidified with dil, HCl and extracted with Et₂O. The Et₂O extract was evaporated to dryness and the residue was chromatographed on a column of Si gel (CHCl3-MeOH, 99:1) to

give 5 (8 mg) whose IR and ${}^{1}H$ NMR spectra were superimposable with those of the acid (5) derived from 3.

Hydrolysis of 7. Compound 7, needles from MeOH-CHCl₃, mp 155–156°, IR v_{max}^{Nujol} cm⁻¹: 3350, 1615 and 1510; ¹H NMR (100 MHz; pyridine-d₅): δ3.90-4.60 (6H, m, sugar protons), 4.80 (2H, s, benzyl H), 5.54 (1H, $W_{1/2} = 10$ Hz, glucose H-1), 7.31 and 7.41 (4H, A_2B_2 , J = 8 Hz, Ar-H). Acetylation of 7 in Ac₂O-pyridine gave a penta-acetate, IR v_{max}^{film} cm⁻¹: 1740, 1610, 1590 and 1510; ¹H NMR (100 MHz; CDCl₃): δ 2.08 (3H × 5, s, COMe), 5.00 (2H, s, benzyl H), 6.95 and 7.21 (4H, A₂B₂, J = 8 Hz, Ar-H). Compound 7 (30 mg) was dissolved in 5 ml 2 N HCl, refluxed for 1 hr, and the soln was extracted with Et₂O. The extract was evaporated to dryness and the residue was crystallized from MeOH to give needles (11 mg), mp 125-126° which were identical with authentic p-hydroxybenzyl alcohol. The aq. layer was neutralized with Amberlite IR-400, evaporated to dryness and the residue was identified as D-glucose by PC (n-BuOH-HOAc- H_2O , 4:1:2, R_f 0.34).

Mild alkaline hydrolysis of 4a. Compound 4a (140 mg) in 0.45 N NaOH (5 ml) and MeOH (9 ml) was stirred at 60° for 1 hr. The soln was extracted with Et₂O; the extract was washed with H_2O , dried and evaporated to dryness, to give 8 as an oil (27 mg). The aq. layer was acidified with dil. HCl and extracted with Et₂O; evaporation of the solvent gave an oily acid (8a) (26 mg). Compound 8, IR v^{film}_{max} cm⁻¹: 1720, 1636, 960 and 860; ¹H NMR (60 MHz; CDCl₃): δ 1.05 (3H, d, J = 6 Hz, H-10 or H-10'), 1.07 (3H, d, J = 6 Hz, H-10 or H-10'), 3.48, 3.60 [total 6H, two s, intensity (4:3), OMe-1 and OMe-1'], 3.72 (3H, s, COOMe) and 7.50 (2H, br s, H-3 and H-3'). MS m/z 436.2118 [M]⁺; C₂₃H₃₂O₈ requires, 436.2097. Compound 8a, IR v film cm⁻¹: 1720, 1700, 1680, 1640 and 960; ¹H NMR (100 MHz; CDCl₃): δ 1.06 (6H, d, H-10 and H-10'), 3.48 (6H, s, OMe-1 and OMe-1'), 7.38 and 7.50 (1H each, s, H-3 and H-3'). Upon methylation with CH_2N_2 in Et₂O, 8a (10 mg) gave an ester (10 mg) whose IR and ¹H NMR spectra were superimposable with those of 8.

Compound 9. Prisms from Me₂CO-H₂O, mp 148-152°, $[\alpha]_D^{25}$ -15° (MeOH; c 0.30), UV λ_{\max}^{MeOH} nm (ϵ): 246 (11000); IR v_{max}^{Nujol} cm⁻¹: 3400, 1720, 1700, 1680 and 1620; ¹H NMR (100 MHz; pyridine- d_5): $\delta 0.84$ (3H, d, J = 7 Hz, H-10 or H-10'), 0.94 (3H, d, J = 7 Hz, H-10 or H-10'), 7.54 and 7.58 (each 1H, s, s)H-3 and H-3') besides signals for sugar protons. (Found: C, 55.00; H, 6.53 %. C₂₆H₃₆O₁₃·1/2 H₂O requires: C, 55.21; H, 6.59 %.) Compound 9 (100 mg) was acetylated with Ac₂O-pyridine (0.5 ml each) at room temp. The crude product was chromatographed on a column of Si gel. Elution with CHCl₃-MeOH (99:1) gave a hexa-acetate (10) (54 mg) as an amorphous powder mp 85–87°, $[\alpha]_D^{25}$ + 71.4° (MeOH; c 0.06), IR ν_{max}^{Nujol} cm⁻¹: 1770, 1720, 1640 and 1220; ¹H NMR (100 MHz; CDCl₃): δ1.10 (6H, br d, H-10 and H-10'), 2.00, 2.08, 2.14 (total 18H, s, COMe), 5.97, 6.08, 6.30 (1H each, br d, J = 3, 2 and 3 Hz, H-1, H-1', and sugar H-1), 7.31 and 7.37 (1H each, s, H-3 and H-3'). (Found: C, 56.37; H, 6.11%. C₃₈H₄₈O₁₉ requires: C, 56.43; H, 5.98%.)

Methanolysis of 9. Compound 9 (100 mg) was dissolved in dry MeOH (3 ml) containing one drop of conc. HCl. The soln was stirred at 60° under a N₂ atmosphere for 1 hr. The soln was diluted with H₂O and extracted with Et₂O. The Et₂O extract was chromatographed on a column of Si gel to give an amorphous powder of 11 (75 mg), IR v_{max}^{Nujol} cm⁻¹: 3400, 1700 and 1630; ¹H NMR (100 MHz; CDCl₃): δ 1.07 (6H, d, J = 6 Hz, H-10 and H-10'), 3.40, 3.48, 3.52 [total 9H, three s, (5:3:2), C-1, C-1' and sugar OMe-1), 7.49 (1H, s, H-3 or H-3') and 7.54 (1H, br s, H-3 or H-3'). Compound 11 (62 mg) was acetylated with Ac₂O-pyridine (0.5 ml each) at room temp. to give a tri-acetate (12) (53 mg), IR v_{max}^{Nujol} cm⁻¹: 1760, 1710, 1635 and 1230; ¹H NMR (100 MHz; CDCl₃): δ 1.03 (3H, d, J = 7 Hz, H-10 or H-10'), 1.08 (3H, d, J = 8 Hz, H-10 or H-10'), 2.00, 2.05, 2.10 (total 9H, s, COMe), 3.40, 3.42, 3.46, 3.52 [total 9H, four s, (1:2:2:1), C-1, C-1' and sugar OMe-1] and 7.46 (2H, s, H-3 and H-3').

Alkaline hydrolysis of 12. Compound 12 (51 mg) was refluxed with 2 N NaOH (2 ml) and MeOH (1.2 ml) for 30 min. The soln was acidified with dil. HCl and extracted with Et_2O . The Et_2O extract was washed with H_2O , dried and concd to dryness. The residue was chromatographed on a column of Si gel to give three oily acids which were identical with 5 (6 mg), 6a (3 mg) and 6b (2.5 mg).

Mild alkaline hydrolysis of 12. Compound 12 (150 mg) was dissolved in MeOH (9 ml) and 0.45 N NaOH (5 ml). The soln was stirred at 62° for 1 hr, extracted with Et_2O ; the Et_2O extract was washed with H_2O , dried and evaporated to dryness. The IR and ¹H NMR spectra of the purified product (21 mg) were superimposable with those of 8. From the acidified H_2O layer, 20 mg of the corresponding acid was recovered.

Purdie methylation followed by hydrolysis of 11. Compound 11 (75 mg) in DMF (1 ml) was stirred with MeI (1 ml) and Ag₂O (200 mg) at 5° for 48 hr in the dark. The soln was diluted with CHCl₃ and filtered. The filtrate was evaporated and the residue was chromatographed on a column of Si gel to afford 13 as an oil (72 mg), IR $v_{\text{max}}^{\text{ilm}}$ cm⁻¹: no OH absorption, 1700 and 1625; ¹H NMR (60 MHz; CDCl₃): δ 1.08 (6H, br d, H-10 and H-10'), 3.40 (3H × 6, s, OMe). Compound 13 (40 mg) in MeOH (1 ml) and 2 N NaOH (1 ml) was refluxed for 1 hr. The soln was acidified and extracted with CHCl₃, dried and evaporated to dryness. The residue was chromatographed on a Si gel column to give a mixture of acids and a methylated D-glucose (4 mg) which was identical with an authentic sample of methyl-2,3,6-tri-O-methyl D-glucoside by comparison on TLC (CHCl₃-MeOH, 97:3, R_f 0.32) and IR and ¹H NMR spectra.

Compound 14. Amorphous powder, mp 134–138°, $[\alpha]_{D5}^{25}$ – 42.1° (MeOH; *c* 0.19), UV λ_{max}^{MeOH} nm (ϵ): 228 (11000) and 238 sh (10350); IR v_{max}^{Nujol} cm⁻¹: 3400, 1695, 1633 and 1518; ¹H NMR (60 MHz; CD₃COCD₃): δ 1.07 (6H, br d, J = 6 Hz, H-10 and H-10'), 5.07 (2H, *s*, benzyl H), 7.00, 7.36 (4H, A₂B₂, J = 9 Hz, Ar-H) and 7.44 (2H, br s, H-3 and H-3'). (Found: C, 58.48; H, 6.46 %, C₃₃H₄₂O₁₅ requires: C, 58.40; H, 6.24 %) Acetylation of 14 (100 mg) in Ac₂O–pyridine (1 ml each) afforded a hepta-acetate (15) (95 mg), amorphous, mp 88–89°, $[\alpha]_{D5}^{25}$ – 22.5° (MeOH; *c* 0.20); IR v_{max}^{Nujol} cm⁻¹: 1765, 1720, 1638 and 1515; ¹H NMR (60 MHz; CDCl₃): δ 1.05 (6H, br d, J = 6 Hz, H-10 and H-10'), 2.04 (3H × 5, s, COMe), 2.09 (3H × 2, s, COMe), 6.07 (2H, d, J = 3 Hz, H-1 and H-1'), 7.00, 7.33 (4H, A₂B₂, J = 9.5 Hz, Ar-H) and 7.38 (2H, br d, J = 1.5 Hz, H-3 and H-3'). (Found: C, 57.65; H, 6.08 %, C₄₇H₅₆O₂₂ requires: C, 58.02; H, 5.76 %.)

Methanolysis of 14. Compound 14 (85 mg) was dissolved in dry MeOH (6 ml), two drops of conc. HCl added and the mixture allowed to stand for 20 hr. The soln was diluted with H_2O and extracted with EtOAc, washed with H2O, dried and evaporated to dryness in vacuo. The residue was chromatographed on a column of Si gel, to give a mixture of diastereoisomers of dimethyl ether (16), $[\alpha]_{D}^{25} - 62.5^{\circ}$ (MeOH; c 0.40), IR v^{Nujol} cm⁻¹: 3450, 1718, 1635 and 1517; ¹H NMR (60 MHz; CD_3COCD_3): $\delta 1.07$ (6H, br d, J = 6 Hz, H-10 and H-10'), 3.39, 3.41, 3.44 (total 6H, s, OMe-1 and OMe-1'), 5.08 (2H, s, benzyl H), 7.14, 7.33 (4H, A_2B_2 , J = 9.5 Hz, Ar-H) and 7.50 (2H, d, J = 1.5 Hz, H-3 and H-3'). (Found: C, 59.45; H, 6.55 %. C₃₅H₄₆O₁₅ requires: C, 59.49; H, 6.52%) Acetylation of 16 (38 mg) in Ac₂O-pyridine (0.5 ml each) gave a penta-acetate (17) (mixture, 40 mg), amorphous, IR v_{max}^{Nujol} cm⁻¹: 1763, 1715, 1638 and 1517; ¹H NMR (60 MHz; CDCl₃): δ 1.07 (6H, d, J = 6.5 Hz, H-10 and H-10'), 2.05 (3H × 5, s, COMe), 3.40, 3.46, 3.48 (total 6H, s, OMe-1 and OMe-1'), 4.63, 4.90 (total 2H, d, J = 4.5 and 3 Hz, H-1 and H-1'), 7.14, 7.33 (4H, A_2B_2 , J = 9.5 Hz, Ar-H) and 7.48 (2H, d, J= 1.5 Hz, H-3 and H-3').

Hydrolysis of 16. Compound 16 (60 mg) in MeOH (2 ml) and 2 N NaOH (8 ml) was refluxed for 1 hr, diluted with H₂O and acidified with $10\frac{9}{6}$ HCl (congo red), extracted with EtOAc, washed with H₂O (twice), dried and evaporated to dryness. The residue (33 mg) was chromatographed on a column of Si gel, to give an acid (24 mg) which was identical with 6a together with a small amount of an acid identical with 6b. The H₂O layer was worked-up as described above to give a glucoside (15 mg) identical with 7.

Purdie methylation of 16. Compound 16 (100 mg) was dissolved in DMF (4 ml) and 1 g Ag₂O and 0.8 ml MeI added. The mixture was stirred for 2 days in the dark, a small amount of CHCl₃ added and the mixture filtered. The filtrate was workedup as described for the methylation of 3. On Si gel chromatography, fractions eluted with CHCl₃ gave a hepta-methyl ether (18) (25 mg), and fractions eluted with CHCl₃-MeOH (97:3) afforded a hexa-methyl ether (19) (60 mg). Compound 18, amorphous, IR v film cm⁻¹: no OH absorption, 1720, 1635, and 1513; ¹H NMR (60 MHz; CDCl₃): δ 1.03 (6H, br d, J = 5.5 Hz, H-10 and H-10'), 3.32, 3.39, 3.44, 3.49 (3H each, s, OMe), 3.59 (9H, s, OMe), 5.02 (2H, s, benzyl H), 7.01, 7.33 (4H, A_2B_2 , J = 9 Hz, Ar-H), and 7.41 (2H, d, J = 1.5 Hz, H-3 and H-3'). Compound 19, IR v_{max}^{neat} cm⁻¹: 3450, 1710, 1622 and 1510; ¹H NMR (60 MHz; $CDCl_3$): $\delta 1.02$, 1.10 (3H each, d, J = 6 Hz, H-10 and H-10'), 3.36, 3.43, 3.46, 3.51 (3H each, s, OMe), 3.60 (6H, s, OMe), 5.05 (2H, s, benzyl H), 7.04, 7.32 (4H, A_2B_2 , J = 9.5 Hz, Ar-H) and 7.39 (2H, br d, H-3 and H-3').

Alkaline hydrolysis of 18. Upon hydrolysis in 2 N NaOH (1 ml) and MeOH (1 ml), compound 18 (25 mg) gave a mixture of two acids (13 mg) and a phenolic glycoside (20) (9 mg). The mixture of the acids was separated further by chromatography on a column of Si gel to give 6 and an acid (6') which was identified as 1,7-di-Omethyl loganic acid aglycone (anomeric mixture). Compound 6', IR v^{film}_{max} cm⁻¹: 3200–2400, 1680, 1630 and 1190; ¹H NMR (60 MHz; CDCl₃): δ 1.02 (3H, br d, J = 6 Hz, H-10), 3.21, 3.43 (total 6H, s, OMe), 4.5-5.1 (total 2H, m, H-1 and H-7), and 7.53 (1H, br s, H-3); EIMS (70 eV) m/z (rel. int.): 242 $[M]^+$ (3), 224 (1), 211 (3), 210 (5), 139 (16) and 85 (100). Compound 20, amorphous, IR v_{max}^{Nujol} cm⁻¹: 3460, 1613, 1510 and 1090; ¹H NMR (60 MHz; CDCl₃): δ 3.19, 3.50, 3.62 (total 12H, s, OMe), 3.0-3.8 (6H, glucose protons), 4.59 (2H, s, benzyl H), 4.90 (1H, m, glucose H-1), 7.02 and 7.28 (4H, A_2B_2 , J = 10 Hz, Ar-H). Compound **20** (8 mg) was refluxed in 2 N NaOH (1 ml) and MeOH (0.5 ml) for 1 hr. Workup as usual gave p-hydroxybenzyl alcohol (mp 125-126°) and a methylated D-glucose which was identical with an authentic sample of 2,3,4,6-tetra-O-methyl-D-glucose by comparison of IR and ¹H NMR spectra and TLC (Si gel, CHCl₃–MeOH 90:10; R_{f} 0.70)

Alkaline hydrolysis of 19. Compound 19 (50 mg) was dissolved in 2 N NaOH (1 ml) and MeOH (1 ml), refluxed for 1 hr and worked-up as above. The product was subjected to chromatography on Si gel (CHCl₃-MeOH, 97:3) to give a phenolic glucoside (16 mg) which was identical with 20. The H₂O layer was acidified with $10\frac{0}{10}$ HCl and extracted with Et₂O. The Et₂O extract was evaporated to give an acid (32 mg) which was identical with 6, and afforded 6a and 6b by further purification. *Compound* (21). Hygroscopic powder, mp 153-161², $[\alpha]_{D}^{25}$

 -42.5° (MeOH; c 0.2), UV λ_{max}^{MeOH} nm (ϵ): 236 (6550); IR ν_{max}^{Nujol} cm⁻¹: 3400, 1700, 1632 and 1517; ¹H NMR (60 MHz; CD₃COCD₃): δ 1.10 (3H, d, J = 6 Hz, H-10), 1.40–2.50 (4H), 5.15 (2H, *s*, benzyl H), 7.19, 7.47 (4H, A₂B₂, J = 9 Hz, Ar-H) and 7.57 (1H, *s*, H-3). (Found: C, 57.53; H, 6.57 $^{\circ}_{0.6}$ C₂₃H₃₀O₁₁ requires: C, 57.25; H, 6.26 $^{\circ}_{0.6}$). Acetylation of **21** with Ac₂O–pyridine gave a hexa-acetate (**22**), amorphous, mp 67–72°, $[x]_{D}^{25}$ = 26.9° (MeOH; *c* 0.27), IR v_{max}^{nuyol} cm⁻¹: 1765, 1720, 1640 and 1518; ¹H NMR (60 MHz; CDCl₃): δ 1.05 (3H, d, J = 6 Hz, H-10), 2.05 (15H, *s*, COMe) 2.09 (3H, *s*, COMe), 3.10 (1H, *m*, H-5), 5.10 (2H, *s*, benzyl H), 6.05 (1H, d, J = 3 Hz, H-1), 6.99, 7.27 (4H, A₂B₂, J = 9 Hz, Ar-H) and 7.35 (1H, d, J = 1.5 Hz, H-3). (Found: C, 57.27; H, 5.92 $^{\circ}_{0.6}$ C₃₅H₄₂O₁₇ requires: C, 57.22; H, 5.72 $^{\circ}_{0.6}$)

Methanolysis of **21**. Compound **21** (100 mg) in MeOH (10 ml) containing one drop of conc. HCl was allowed to stand for 20 hr. The soln was worked-up as described above to yield 80 mg methyl ether (**23**), $[\alpha]_{D}^{25} + 45^{\circ}$ (MeOH; *c* 0.30), IR v_{max}^{Nujol} cm⁻¹: 3400, 1708, 1680, 1635, 1518 and 1080; ¹H NMR (60 MHz; CD₃COCD₃): δ 1.09 (3H, *d*, *J* = 6 Hz, H-10), 3.41, 3.46 (total 3H, *s*, OMe-1), 4.78, 5.08 (total 1H, *d*, *J* = 4 and 2.5 Hz, H-1), 5.20 (2H, *s*, benzyl H), 7.13 and 7.51 (4H, A₂B₂, *J* = 9 Hz Ar-H). Compound **23** (80 mg) was acetylated in Ac₂O--pyridine (0.5 ml each) to give a penta-acetate mono-methyl ether (**24**) (75 mg), amorphous, IR v_{max}^{Nujol} cm⁻¹: 1770, 1718, 1637 and 1516; ¹H NMR (60 MHz; CDCl₃): δ 1.01 (3H, *d*, *J* = 6.5 Hz, H-10), 2.02 (15H, *s*, COMe), 3.36, 3.44 (total 3H, *s*, OMe-1), 4.57, 4.86 (total 1H, *d*, *J* = 4 and 3 Hz, H-1) 6.96, 7.29 (4H, A₂B₂, *J* = 10 Hz, Ar-H) and 7.41 (1H, br s, H-3). (Found: C, 58.15; H, 6.14. C₃₄H₄₂O₁₆ requires: C, 57.79; H, 5.95 °_o.)

Alkaline hydrolysis of 24. Compound 24 (60 mg) was hydrolysed in 2 N NaOH (2 ml) and MeOH (1.5 ml) and worked-up as described for the hydrolysis of 3 to yield a single acidic product (22 mg) which was identical with 6 and a glucoside (26 mg) which was identical with 7.

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