Practical Syntheses of Immunologically Relevant β -Glycosides of 2-Acetamido-2-deoxy-p-mannopyranose. Methyl N-Acetyl- β -p-mannosaminide, N-Acetyl- β -p-mannosaminyl- $(1\rightarrow 6)$ -p-galactose, and Methyl N-Acetyl- β -D-mannosaminyl- $(1\rightarrow 4)$ - α -D-glucopyranoside

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Highly stereorelective, expedient syntheses are described for three immunologically relevant β -glycosides of N-acetyl-p-mannosamine, i.e. β -p-ManpNAc-1-OMe (8), β -p-ManpNAc-(1 \rightarrow 6)-p-Gal (15), and β -p-ManpNAc-(1→4)-α-p-Glcp-1-OMe (23). Basic mannosamine progenitor in each case is 3,4,6-tri-O-benzoyl-2-(benzoyloxyimino)-2-deoxy- α -D-arabino-hexopyranosyl bromide (1a), which via the three step-sequence β -selective glycosidation \rightarrow hydroboration \rightarrow deblocking is converted to 8, 15, and 23 in overall yields of 61, 56, and 39% for the glycosyl acceptors methanol, 1,2:3,4-di-O-isopropylidene-α-p-galactose, and methyl 2,3,6-tri-O-benzyl-α-pglucopyranoside, respectively.

B-Glycosidically linked 2-acetamido-2-deoxy-Dmannopyranose (N-acetyl- β -D-mannosamine) carries a type-specific immunogeneity in various bacteria associated with invasive diseases. 1,2) Even simple alkyl glycosides such as methyl N-acetyl-β-D-mannosaminide acts as a specific hapten against the immunoglobulin of IgA class originated from a mouse myeloma MOPC 406;3) more complex N-acetyl- β -D-mannosaminides constitute a number of bacterial K antigens of capsular polysaccharides²⁾ as well as O antigens of lipopolysaccharides.4)

Investigations directed towards the molecular design of artificial antigens require as a principal prerequisite preparatively useful, chemical procedures for the practical construction of oligosaccharides containing N: acetyl-\beta-p-mannosamine. Such procedures are still lacking despite of the possibility of using 2-azido-2deoxy-D-mannopyranosyl halides,5,6) which are accessible in syrupy form via low yield, multistep reaction sequences, and furthermore, require careful optimiza-

tion of glycosidation conditions to reach useful β selectivities.⁵⁾ A similarly laborious approach⁷⁾ comprises the a priori construction of the respective disaccharide for example, and the a posteriori introduction of the 2-amino function via oxidation of a 2-hydroxyl group, oximation and subsequent reduction.7)

Our approach to the problem of a practical synthesis for β -D-mannosaminides, that has been elaborated on lactose-derived derivatives, 8,9) comprises 2-(benzoyloxyimino)glycosyl bromides of type 1 as the key compounds readily accessible from glucose^{10,11)} or disaccharides8) in a few high-yielding steps and in crystalline form. These key building blocks I are ideal β -D-mannosamine progenitors, since β -selective glycosidation, reduction of the benzoyloxyimino function to amino group, and deprotection yielding β -Dmannosamine-containing oligosaccharides, a protocol, which combines high stereoselectivity with efficiency of operation and large-scale adaptability

Scheme 1.

(Scheme 1).

In complementation of previous work concentrating on trisaccharides with central β -D-mannosamine units, 9) we here wish to describe 12) the utilization of the D-glucose-derived monosaccharide building block 1a for the preparation of disaccharides with a nonreducing N-acetyl- β -D-mannosamine residue, of which one, i.e. 23, constitutes a disaccharide unit of the capsular polysaccharide from *Streptococcus pneumoniae* type $9N^2$ and $19F.^{13}$

Methyl N-Acetyl- β -D-mannosaminide (8) A preparatively reliable synthesis of methyl N-acetyl- β -D-mannosaminide (8), as of now, is not available, since direct glycosidation of 2-acetamido-2-deoxy-D-mannose with methanol in the presence of acid (Fischer method) results in a mixture of the four possible methyl N-acetyl-D-mannosaminides from which the β -D-

pyranoside (8) is isolable in yields of 1—2% only. ¹⁴⁾ The major isomers were α -D-pyranoside (9%), α -D-furanoside (8%), and β -D-furanoside (2—3%).

Our synthesis of **8** involves methanolysis of the building block **1a**, reduction of the resultant β -glycoside **6** to mannosaminide, N-acetylation, and O-debenzoylation, so that the synthetic efficiency depends on the β -selective methanolysis and mannoselective reduction (Scheme 2).

The glycosidation of 1a with excess methanol (tenfold molar amount) in the presence of several kinds of condensation promotors are summarized in Table 1. Of a variety of reaction conditions examined, only silver carbonate in dichloromethane resulted in good yield (93%) and high stereoselectivity ($\alpha: \beta \approx 5:95$). Other conditions employed, such as silver triflate-tetramethylurea or 2,4,6-collidine-1a (cf. Runs 2—5,

BzO OBz
$$Ag_2CO_3/I_2$$
 OBz $I)B_2H_6 \cdot THF$ BzO NHAC

BzO OME $2)Ac_2O$ BzO OME

BzO OME CH_2CI_2 NOBz $I)B_2H_6 \cdot THF$ BzO OME

BzO OME I NOBz I NOB

Table 1. Glycosidation of Tri-O-benzoyl-2-benzoyloxyimino-2-deoxy-α-p-arabino-hexopyranosyl Bromide (1a) with Methanol and Partially Protected Sugars

Run	Ålcohol	Molar Ratio (Alcohol/ 1a)	Promotor	Solvent	Time	Yield	$lpha$: $oldsymbol{eta}^{\mathbf{a})}$
Run					h	%	
1	MeOH	10.0	Ag ₂ CO ₃ /I ₂	CH ₂ Cl ₂	72	93	5:95
2	MeOH	10.0	AgOTf/TMU ^{b)}	CH_2Cl_2	24	87	20:80
3	MeOH	10.0	AgOTf/TMU	Dioxane	24	83	40:60
4	MeOH	10.0	$2,4,6$ -Collidine/ I_2	Dioxane	72	7 5	35:65
5	MeOH	10.0	$Hg(CN)_2$	Dioxane	72	17	35:65
6	6-OH Gal iP ₂ c)	1.1	Ag_2CO_3/I_2	CH_2Cl_2	24	86	5:95
7	6-OH Gal iP ₂	1.1	Ag_2CO_3/I_2	CH_2Cl_2	72	94	5:95
8	6-OH Gal iP ₂	1.1	AgOTf/TMU	Dioxane	16	89	70:30
9	4-OH Glc Bz ₃ d)	1.1	Ag_2CO_3/I_2	CH_2Cl_2	72	<24	e)
10	4-OH Glc Bz ₃	1.1	AgClO ₄ /TMU	CH_2Cl_2	24	< 50	e)
11	4-OH Glc Bz ₃	1.1	AgOTf/TMU	Dioxane	16	<16	e)
12	4-OH Glc Bn ₃ f)	1.1	Ag_2CO_3/I_2	CH_2Cl_2	48	52	5:95
13	4-OH Glc Bn ₃	2.0	Ag_2CO_3/I_2	CH_2Cl_2	48	55	5:95
14	4-OH Glc Bn ₃	1.1	AgClO ₄ /TMU	CH_2Cl_2	48	<24	e)
15	4-OH Glc Bn ₃	1.1	Ag_2O/I_2	CH_2Cl_2	48	<44	e)
16	4-OH Glc Bn ₃	1.5	Ag-silicate	CH_2Cl_2	48	<28	e)

a) Estimated on the basis of ¹H NMR signals for the anomeric protons. b) Silver trifluoromethanesulfonate/1,1,3,3-tetramethylurea. c) 1,2:3,4-Di-O-isopropylidene- α -D-galacto-pyranose (11). d) Methyl 2,3,6-tri-O-benzoyl- α -D-glucopyranoside (17). e) Contaminated with byproducts that impeded the estimation of α : β ratio. f) Methyl 2,3,6-tri-O-benzyl- α -D-glucopyranoside (16).

Table 1) gave anomeric mixtures with only partial predominance of β -anomers. β -Glycoside **6** was then subjected to hydroboration with twelve-fold molar excess diborane in tetrahydrofuran to afford on Nacetylation methyl N-acetyl-tri-O-benzoyl-β-D-mannosaminide (7) in 88% yield. The corresponding β -pgluco isomer 10 was isolated in only 1.3% yield, so that the preference for the β -D-manno form is in the remarkable, practically very useful range of around 50:1. The subsequent O-debenzoylation of 7 was cleanly effected with 0.05 M sodium methoxide (1 M=1 mol dm⁻³) in methanol (Zemplén method) to furnish methyl N-acetyl- β -D-mannosaminide (8) in 87% yield. Furthermore, 8 was hydrolyzed with 1 M hydrochloric acid giving p-mannosamine hydrochloride 9 in quantitative yield, which was unequivocally identified by comparison with an authentic sample.

The following stereochemical aspects are worthy of note: β -oriented methyl 2-(benzoyloxyimino)glycoside **6** seems to be distorted towards the twist-boat form as indicated in the formula considering from relatively small coupling constant of $J_{3,4}=J_{4,5}=5.7$ Hz. In contrast, **1a** and **7** prefer 4C_1 coformation, as evidenced by $J_{3,4}$ and $J_{4,5}$ values of 8.4—9.6 Hz. This may be attributed to substantial steric congestion caused by the 2-benzoyloxyimino and anomeric methoxyl groups in **6**. Similar conformational propensity for 2-(benzoyloxyimino)- β -glycosides were observed previously.^{8,9)}

¹H and ¹³C NMR spectra of methyl β-D-mannosaminide **8** were fully elucidated with the aid of ¹H-¹H and ¹H-¹³C shift correlated two-dimentional (2D) NMR spectra. ¹H-¹H coupling of $J_{1,2}$ =1.5 and $J_{2,3}$ =4.5 Hz cogently reflect β-D-manno configuration, and a $J_{Cl,Hl}$ coupling of 163.6 Hz agrees well with that (ca. 160 Hz) of methyl β-glycopyranosides with axial H-1.¹⁵⁾ A calculation of differences of ¹³C-chemical shifts ($\Delta\delta_c$) by δ_{gluco} - δ_{manno} between methyl N-acetyl- β - D-mannosaminide and the corresponding glucosaminide¹⁶⁾ showed that major differences appeared at C-2 ($\Delta\delta_c$ =-4.1 ppm) and C-4 (-3.2), whilst they are less pronounced at C-3 (-2.0), C-1 (-1.5), C-5 (+0.5), and C-6 (-0.4). These findings were found to be helpful in the structural elucidation of β-D-mannosamine-containing oligosaccharides.

N-Acetyl- β -D-mannosaminyl- $(1\rightarrow 6)$ -D-galactose (15) and Methyl N-Acetyl- β -D-mannosaminyl- $(1\rightarrow 4)$ - α -Dglucopyranoside (23). Replacement of the glycosyl acceptor in the above glycosidation $1a \rightarrow 6$ by suitably blocked sugar alcohols similarly gave the desired disaccharide derivatives in preparatively highly useful fashion. Thus, glycosylation of 1,2:3,4-di-O-isopropylidene-D-galactose (11)17) with (benzoyloxyimino)glycosyl bromide la in the presence of silver carbonateiodine in dichloromethane (Run 7 in Table 1) cleanly gave the β -glycoside 12 in the satisfactory yield of 94% (Scheme 3). In contrast, silver triflate in dioxane (Run 8) gave rise to α -predominance (α : $\beta \approx 70:30$), a distinct promotor-dependent inversion, that was also observed in the glycosylation of 11 with the disaccharide donor 1b.9) The β -glycoside 12 was then subjected to hydroboration and subsequent N-acetylation as described for 7 to give 13 in 74% yield, whereby the presence of the β -D-gluco epimer could not be detected. *O*-Debenzoylation of 13 followed by removal of isopropylidene groups with 95% aqueous trifluoroacetic acid gave Nacetyl- β -D-mannosaminyl- $(1\rightarrow 6)$ -D-galactose (15) in 81% yield. The structure of 15 was confirmed by ¹³C NMR spectroscopy.

Scheme 4 depicts the synthetic pathway to methyl N-acetyl- β -D-mannosaminyl- $(1\rightarrow 4)$ - α -D-glucopyranoside (23). Glycosidations of 1a with partially protected methyl glucopyranosides with a free C-4 hydroxyl group were somewhat troublesome because of the low reactivity of the secondary hydroxyl function. After

Scheme 3.

$$1a + HO OR RO OMe RO$$

much attempts shown in Table 1, the $\beta(1\rightarrow 4)$ -glycoside (18) could be secured in 55% yield (Run 13 in Table 1) by the use of methyl 2,3,6-tri-O-benzyl- α -D-glucopyranoside (16)¹⁸⁾ as an acceptor and silver carbonate-iodine as the promotor, whilst methyl 2,3,6-tri-O-benzoyl- α -D-glucopyranoside (17)¹⁹⁾ afforded multi-component mixtures in all the cases tested (Runs 9—11). Other glycosylation catalysts, such as silver oxide, silver perchlorate, or the allegedly highly superior²⁰⁾ silver silicate were noneffective (Runs 14—16).

Under the conditions of Run 13, i.e. glycosylation of 16 with two molar equivalents of 1a and Ag₂CO₃ in dichloromethane, substantial amounts of 1-hydroxy derivative 19 was generated, its identity being established by its conversion into the 1-acetate 20α , that in turn was unequivocally identified by comparison with an authentic sample prepared by reaction of la with sodium acetate in dioxane. Hydroboration of 18 was carried out as for $6\rightarrow7$, yet proceeded in an essentially stereospecific manner, to provide the protected Nacetyl- β -D-mannosaminide (21) in a yield of 81%. Subsequent debenzoylation readily gave 22 (96% yield), which was catalytically hydrogenated over palladium/ carbon to quantitatively afford desired methyl Nacetyl- β -D-mannosaminyl- $(1\rightarrow 4)$ - α -D-glucopyranoside (23).

The concise, practical route reported here for the chemical synthesis of methyl or glycosyl N-acetyl- β -D-mannosaminides, as of now, appears to be the most straightforward methodology for the construction of heterooligosaccharides with a terminal β -D-mannosamine unit; it can be expected to find application for the immunologically relevant repeating units of Strep-tococcus pneumoniae-derived capsular polysaccharide

and a series of others, some of which we hope to report in due course.

Experimental

General. Melting points were determined on a Yamato MP-1 apparatus and are uncorrected. Spectral data were recorded on the following instruments; IR; Jasco IR-810 spectrophotometer; $\lceil \alpha \rceil_D$: Jasco DIP-180 digital polarimeter; MS: JMS D-100 Spectrometer; ¹H NMR: Varian EM-390 (90 MHz), VXR-300 (300 MHz), or XL-400 (400 MHz) spectrometers in chloroform-d solution unless otherwise noted. ¹H-¹H and ¹H-¹³C shift correlated 2D NMR and ¹³C NMR: Varian VXR-300 or XL-400 spectrometer. TLC was carried out on silica gel 60 F₂₅₄ (Merck Art. 5735) developed with the same solvent systems as described for column chromatography in the individual experimental section. The spots were detected by UV light (254 nm) or charring with 10% aq. sulfuric acid. Column chromatography was achieved on a silica gel 60 (Merck Art. 7734). Paper chromatography (PC) was carried out on Toyo filter paper No. 525 and detected with 0.2% ninhydrin in pyridine solution at 100 °C for 10

Methyl 3,4,6-Tri-*O*-benzoyl-2-(benzoyloxyimino)-2-deoxy- β -D-arabino-hexopyranoside (6). (Benzoyloxyimino)glycosyl bromide $1a^{10}$ (135 mg, 0.2 mmol) was added to a mixture of methanol (0.08 ml, 2 mmol), silver carbonate (279 mg, 1 mmol), and iodine (51 mg, 0.2 mmol) in dry dichloromethane (2 ml) containing Molecular Sieves 3A (100 mg, powder). The mixture was stirred in the dark at room temperature for 3 days, whereafter all the educt had been consumed (TLC). Dilution with dichloromethane (10 ml), filtration through Celite, consecutive washing of the filtrate with saturated aq. NaHCO₃ (10 ml) and water (3×10 ml), drying (Na₂SO₄) and evaporation to dryness in vacuo gave a residue which was eluted from a silica-gel column with toluene-ethyl acetate (8:1). The major fraction was concentrated and crystallized from diethyl ether-pentane: 116 mg (93%) of

6; mp 57—65 °C (decomp); $[\alpha]_{0}^{20}$ —42.8° (*c* 1.0, CHCl₃); ¹H NMR (400 MHz) δ=3.70 (3H, s, CH₃), 4.51 (1H, m, H-5), 4.77 (1H, dd, H-6a), 4.84 (1H, dd, H-6b), 5.92 (1H, dd, H-4), 5.93 (1H, s, H-1), 6.25 (1H, d, H-3), 7.3—8.1 (aromatic H); $J_{3,4}=J_{4,5}=J_{5,6a}=5.5$, $J_{5,6b}=7.0$, $J_{6a,6b}=11.5$ Hz; ¹³C NMR (100 MHz) δ=57.02 (OMe), 64.64 (C-6), 68.42 (C-3), 68.61 (C-4), 73.24 (C-5), 93.98 (C-1), 128.2—133.7 (phenyl C), 156.56 (C-2), 162.29, 164.52, 164.74, and 165.82 (benzoyl C=O).

Found: C, 67.09; H, 4.61; N, 2.36%. Calcd for $C_{35}H_{29}NO_{10}$: C, 67.41; H, 4.69; N, 2.25%.

Methyl 2-Acetamido-3,4,6-tri-O-benzoyl-β-D-mannopyranoside (7). A 1 M solution of diborane in tetrahydrofuran (15.5 ml) was added to a solution of 6 (807 mg, 1.3 mmol) in tetrahydrofuran (15 ml) at -10 °C under atmosphere of nitrogen. The mixture was stirred at -10 °C for 0.5 h and at ambient temperature for 2 h. Excess reductant was quenched with methanol (15 ml) followed by addition of acetic anhydride (8 ml) for N-acetylation. After stirring for 1 h at ambient temperature, the mixture was passed through a basic resin (Amberlite IR-45), and washed with methanol. The eluate was concentrated in vacuo and the residue was purified by elution from a silica-gel column with chloroform-ethyl acetate (1:1). The major fraction was concentrated and the residue crystallized from ethyl acetate-diethyl ether (1:2)-pentane to give 622 mg (88%) of 7: mp 95—105 °C; $[\alpha]_D^{22}$ -55.8 ° (c 0.5, CHCl₃); IR (KBr) 3450, 3380 cm⁻¹ (NH); ¹H NMR (400 MHz) δ =2.03 (3H, s, COCH₃), 3.55 (3H, s, OCH₃), 4.11 (1H, td, H-5), 4.57 (1H, dd, H-6a), 4.64 (1H, dd, H-6b), 4.79 (1H, d, H-1), 4.98 (1H, td, H-2), 5.47 (1H, dd, H-4), 5.87 (1H, d, NH), 7.3-8.1 (aromatic H); $J_{1,2}=2.0$, $J_{2,3}$ =4.0, $J_{2,NH}$ =8.5, $J_{3,4}$ =9.5, $J_{4,5}$ =9.0, $J_{5,6a}$ =6.0, $J_{5,6b}$ =3.5, $J_{6a,6b}$ =12.0 Hz; ¹³C NMR (100 MHz) δ =23.38 (COCH₃), 50.19 (C-2), 57.15 (OCH₃), 63.46 (C-6), 67.45 (C-4), 71.92 (C-3), 72.60 (C-5), 100.12 (C-1), 128.2—133.5 (phenyl C), 165.32, 165.53, and 165.86 (3×C=O), 170.28 (NHCO).

Found: C, 65.70; H, 5.35; N, 2.84%. Calcd for $C_{30}H_{29}NO_{9}$. C, 65.80; H, 5.34; N, 2.56%.

On concentration of the minor fraction, dissolution of the residue in diethyl ether, and addition of pentane, β -D-gluco isomer **10** was obtained in a yield of 9.4 mg (1.3%): mp 80—88 °C; [α] $_{\rm D}^{22}$ +8.8° (c 0.25, CHCl $_{\rm 3}$); $^{\rm 1}$ H NMR (90 MHz) δ =1.85 (3H, s, COCH $_{\rm 3}$).

Methyl 2-Acetamido-2-deoxy-β-D-mannopyranoside (8). A solution of 7 (212 mg, 0.39 mmol) in 0.05 M methanolic sodium methoxide (20 ml) was stirred at ambient temperature for 20 h and was subsequently neutralized with a dry acidic resin (Dowex 50WX8) and filtered. The filtrate was evaporated to dryness and the residue was partitioned between dichloromethane (30 ml) and water (30 ml). The aqueous layer was separated, washed with dichloromethane (2×40 ml), and concentrated to dryness. The residue was purified by elution from a silica-gel column with chloroform-methanol (1:1), followed by concentration and crystallization from methanol-diethyl ether to give 71 mg (74%) of 8 as 1/2 hydrate: mp 123—127 °C; $[\alpha]_D^{23}$ —70° (c 0.25, H₂O; $lit,^{14)} [\alpha]_0^{24} -68 \ ^{\circ}(c \ 1.5, \ H_2O)); \ ^{1}H \ NMR \ (D_2O, \ 400 \ MHz)$ δ =1.95 (3H, s, COCH₃), 3.29 (1H, td, H-5), 3.29 (3H, s, OCH₃), 3.40 (1H, dd, H-4), 3.69 (1H, dd, H-3), 3.70 (1H, dd, H-6a), 3.80 (1H, dd, H-6b), 4.37 (1H, dd, H-2), 4.58 (1H, d, H-1); $J_{1,2}=1.5$, $J_{2,3}=4.5$, $J_{3,4}=J_{4,5}=9.5$, $J_{5,6a}=5.0$, $J_{5,6b}=2.5$, $J_{6a,6b}=12.0 \text{ Hz}$; ¹³C NMR (D₂O, 100 MHz), $\delta=22.82 \text{ (COCH}_3)$, 53.83 (C-2), 57.81 (OCH₃), 61.24 (C-6), 67.60 (C-4), 72.83 (C-3), 77.22 (C-5), 101.24 (C-1), 176.24 (NHCO); $J_{\text{Cl.H1}}=163.6$ Hz

Found: C, 44.23; H, 7.22; N, 5.62%. Calcd for $C_9H_{17}NO_6$ $1/2H_2O$: C, 44.26; H, 7.43; N, 5.73%.

The monohydrate of **8** was also isolated in a yield of 13 mg (14%) along with the 1/2 hydrate; Found: C, 42.84; H, 7.61; N, 5.40%. Calcd for $C_9H_{17}NO_6$ H₂O: C, 42.68; H, 7.56; N, 5.53%.

2-Amino-2-deoxy-p-mannopyranose Hydrochloride (9). Methyl *N*-acetyl-β-p-mannosaminide (8) 1/2 hydrate (30 mg, 0.124 mmol) was dissolved in 1 M HCl (10 ml) and heated at 95 °C for 2 h. After concentration of the solution, the residue crystallized from methanol-diethyl ether to afford 27 mg (quantitative yield) of **9** as colorless crystals: mp 173—175 °C (decomp); $[\alpha]_{\rm E}^{24}$ –4.2° (c 0.5, H₂O), (lit,²¹⁾ mp 178—180 °C, $[\alpha]_{\rm E}^{23}$ –3.0° (c 2.0, H₂O)); PC (n-BuOH-pyridine-H₂O, 6:4:3 v/v): $R_{\rm Glc}$ =0.73 (cf., authentic p-ManpN HCl: $R_{\rm Glc}$ =0.73, p-GlcpN HCl: $R_{\rm Glc}$ =0.68).

6-O-[3,4,6-Tri-O-benzoyl-2-(benzoyloxyimino)-2-deoxy-β-Darabino-hexopyranosyl]-1,2:3,4-di-O-isopropyridene- α -Dgalactopyranose (12). A mixture of diacetonegalactose (11)¹⁷⁾ (573 mg, 2.2 mmol), silver carbonate (2.76 g, 10 mmol), iodine (508 mg, 2.0 mmol), and (benzoyloxyimino)glycosyl bromide 1a (1.35 g, 2.0 mmol) in dry dichloromethane (20 ml) with Molecular Sieves 3A (2.0 g, powder) was stirred in the dark at ambient temperature for 3 days. Subsequent dilution with dichloromethane (50 ml), filtration through celite, washing of the filtrate with 1 M aqueous Na₂S₂O₃ (50 ml), water (50 ml), saturated aqueous NaHCO₃ (50 ml), and water (3×100 ml) was followed by drying (Na₂SO₄) and evaporation to dryness. The residue was eluted through a silica-gel column with toluene-ethyl acetate (6:1). Concentration of the major fraction, followed by crystallization from diethyl ether-pentane gave 1.61 g (94%) of 12: mp 83-86 °C; $[\alpha]_D^{22}$ -95.6° (c 0.5, CHCl₃); ¹H NMR (400 MHz) δ =1.27, 1.28, 1.35, and 1.40 (each 3H, s, 2×Me₂C), 3.90 (1H, dd, H-6a), 4.04 (1H, td, H-5), 4.12 (1H, dd, H-4), 4.24 (1H, dd, H-6b), 4.28 (1H, dd, H-2), 4.47 (1H, m, H-5'), 4.54 (1H, dd, H-3), 4.84 (2H, m, H-6'a,b), 5.46 (1H, d, H-1), 5.92 (1H, t, H-4'), 6.09 (1H, s, H-1'), 6.25 (1H, d, H-3'), 7.3-8.2 (aromatic H); $J_{1,2}=5.0$, $J_{2,3}=2.5$, $J_{3,4}=8.0$, $J_{4,5}=2.0$, $J_{5,6a}=6.8$, $J_{5,6b}$ =5.5, $J_{6a,b}$ =9.7, $J_{3',4'}$ = $J_{4',5'}$ =5.3 Hz; ¹³C NMR (100 MHz) δ =24.32, 24.89, 25.88, and 25.96 (2×Me₂C), 64.55 (C-6'), 66.74 (C-5), 68.65 (C-3'), 68.86 (C-4' and 6), 70.45 (C-2), 70.60 (C-3), 70.89 (C-4), 73.03 (C-5'), 93.19 (C-1'), 96.22 (C-1), 108.65 and 109.45 (2×Me₂C), 128.0—134.0)phenyl C), 156.21 (C-2'), 162.68, 164.70, 164.92, and 165.91 (4×C=O).

Found: C, 64.60; H, 5.29; N, 1.59%. Calcd for $C_{46}H_{45}NO_{15}$: C, 64.86; H, 5.32; N, 1.64%.

6-*O*-(2-Acetamido-3,4,6-tri-*O*-benzoyl-2-deoxy-β-D-mannopyranosyl)-1,2:3,4-di-*O*-isopropylidene-α-D-galactopyranose (13). A solution of the disaccharide 12 (426 mg, 0.5 mmol) in dry tetrahydrofuran (6 ml) was treated with a 1 M solution of diborane in tetrahydrofuran (6 ml) as described for $6\rightarrow 7$. Termination with methanol (6 ml) and subsequent *N*-acetylation with acetic anhydride (3 ml) followed by processing of the mixture as described for **7** gave a syrup, which crystallized from ethyl acetate-diethyl ether (1:2)-pentane providing 288 mg (74%) of 13: mp 171–173 °C; $[\alpha]_{L}^{22}$ =66.1° (*c* 0.5, CHCl₃); ¹H NMR (90 MHz) δ=1.29 and 1.40 (each 6H, s, 2×Me₂C), 1.98 (3H, s, COCH₃), 4.27 (1H, dd, H-2), 4.53 (1H, dd, H-3), 4.55—4.70 (2H, m, H-6a,b), 4.90 (1H, td, collapsed to dd on deuteration), 7.2—8.1 (aromatic H), 3.7—4.2 (other protons); $J_{1,2}$ =5.1, $J_{2,3}$ =2.4, $J_{3,4}$ =7.8, $J_{1',2'}$ =3.0, $J_{2',3'}$ =3.6,

 $J_{2',NH}$ =8.4, $J_{3',4'}$ =7.2, $J_{4',5'}$ =7.8 Hz.

Found: C, 63.23; H, 5.81; N, 1.64%. Calcd for $C_{41}H_{45}NO_{14}$: C, 63.48; H, 5.85; N, 1.81%.

6-O-(2-Acetamido-2-deoxy-β-p-mannopyranosyl)-p-galactopyranose (15). A solution of **13** (343 mg, 0.44 mmol) in 0.05 M methanolic sodium methoxide (20 ml) was stirred at ambient temperature for 20 h. Subsequent neutralization (Dowex 50 W×8), filtration through Celite, and evaporation to dryness gave a residue, which was eluted through a silicagel column with chloroform-methanol (4:1). Concentration of the major fraction gave debenzoylated product **14** as a colorless syrup (200 mg, quantitative), which was subjected to further deblocking of the isopropylidene groups without further purification.

The syrup was dissolved in 95% aqueous trifluoroacetic acid (3 ml) and stirred at ambient temperature for 20 min. Evaporation of the mixture was followed by co-evaporation with methanol to give a residue, which crystallized from ethanol-diethyl ether affording 140 mg (81%) of **15** as a monohydrate: mp 170 °C (decomp, with sintering at 115 °C): $[\alpha]_{\rm b}^{2l}-14^{\circ}$ (c 0.25, H₂O): ${}^{1}{\rm H}$ NMR (D₂O, 90 MHz) δ =1.90 (3H, s, COCH₃), 7.73 (1H, d, NH); ${}^{13}{\rm C}$ NMR (D₂O, 25.4 MHz) δ =22.8 (COCH₃), 53.9 (C-2′), 61.3 (C-6′), 67.6 (C-4′), 69.1 and 69.6 (C-6 for respective anomers), 77.3 (C-5′), 93.2 (C-1 for α -anomer), 97.3 (C-1 for β -anomer), 101.6 (C-1′), 176.5 (NHCO). The anomeric carbon signals of the reducing end suggested that the ratio of the anomers was estimated as α : β =ca. 2:3.

Found: C, 41.68; H, 6.50; N, 3.38%. Calcd for $C_{14}H_{25}NO_{11}H_2O$: C, 41.89; H, 6.78; N, 3.49%.

Methyl 4-O-[3,4,6-Tri-O-benzoyl-2-(benzoyloxyimino)-2deoxy-β-D-arabino-hexopyranosyl]-2,3,6-tri-O-benzyl-α-Dglucopyranoside (18). Silver carbonate (138 mg, 0.5 mmol) and iodine (25 mg, 0.1 mmol) were added to a stirred solution of methyl 2,3,6-tri-O-benzyl- α -p-glucopyranoside (16)¹⁸⁾ (93 mg, 0.2 mmol) in dry dichloromethane (1 ml) containing Molecular Sieves 3A (100 mg, powder) and then stirred in the dark for 1 h under atmosphere of nitrogen. A solution of (benzoyloxyimino)glycosyl bromide la (67 mg, 0.1 mmol) in dry dichloromethane (1 ml) was added and the mixture was further stirred at ambient temperature for 2 days. The mixture was diluted with dichloromethane (10 ml) and filtered through Celite. The filtrate was washed with saturated aqueous NaHCO₃ (10 ml) and water (3×15 ml). Drying (Na₂SO₄) and evaporation in vacuo gave a syrup, which was eluted through a silica-gel column with toluene-ethyl acetate (6:1). The major fraction eluted first was concentrated and the residue crystallized from diethyl ether-pentane to give 58 mg (55%) of **18**: mp 57—59 °C; $[\alpha]_D^{20}+21.8^{\circ}$ (c 0.5, CHCl₃); 1 H NMR (90 MHz), δ =3.29 (3H, s, OCH₃), 4.98 (1H, s, H-1), 5.92 (1H, t, H-4'), 6.22 (1H, d, H-3'), 6.74 (1H, s, H-1'), 3.4-4.9 (15H, other protons), 6.9-8.2 (aromatic H); $J_{3',4'}=J_{4',5'}=5.4$ Hz.

Found: C, 70.51; H, 5.44; N, 1.33%. Calcd for $C_{62}H_{57}NO_{15}$: C, 70.69; H, 5.20; N, 1.30%.

The minor fraction eluted next was treated as above to give 10.5 mg (18%) of **19** as 1/2 hydrate: mp 177—179 °C (decomp); $[\alpha]_D^{20}$ +32.4° (c 0.25, CHCl₃); IR (KBr) 3400 cm⁻¹ (OH); ¹H NMR (DMSO- d_6 , 90 MHz), δ =4.5—4.7 (2H, m, H-6a,b), 4.75 (1H, m, H-5), 5.87 (1H, t, H-4), 6.36 (1H, d, H-3), 6.50 (1H, t, H-1), 8.27 (1H, d, OH, disappeared on deuteration), 7.3—8.2 (aromatic H); $J_{1, OH}$ =6.0, $J_{3,4}$ = $J_{4,5}$ =10.5 Hz.

Found: C, 66.29; H, 4.42; N, 2.19%. Calcd for C₃₉H₂₇NO₁₀ 1/2H₂O: C, 66.02; H, 4.56; N, 2.26%.

Acetylation of 19 with acetic anhydride-pyridine in a usual manner gave the corresponding 1-acetate 20α which proved to be identical with the product obtained by acetylation of 1a as described below.

1-O-Acetyl-3,4,6-tri-O-benzoyl-2-(benzoyloxyimino)-2-deoxy- α -D-arabino-hexopyranose (20 α) and Its β -Anomer (20 β). A mixture of (benzoyloxyimino)glycosyl bromide Ia (1.0 g, 1.5 mmol), sodium acetate (369 mg, 4.5 mmol), and Molecular Sieves 3A (1 g, powder) in dry dioxane (20 ml) was stirred at ambient temperature for 3 days. The resulting mixture was diluted with dichloromethane (50 ml) and filtered through Celite. The filtrate was washed with water (3×50 ml), dried (Na₂SO₄), and evaporated to dryness. Elution of the residue through a silica-gel column with toluene-ethyl acetate (6:1) provided both anomeric isomers, which crystallized from diethyl ether-pentane to afford 365 mg (37%) of 20 α and 200 mg (20%) of the corresponding β -anomer 20 β .

20 α : Mp 75—77 °C; $[\alpha]_{0}^{22}$ +53.4° (c 0.3, CHCl₃); ¹H NMR (90 MHz) δ =2.25 (3H, s, COCH₃), 4.4—4.7 (2H, m, H-6a,b), 4.68 (1H, m, H-5), 5.98 (1H, dd, H-4), 6.44 (1H, d, H-3), 7.2—8.2 (H-1 and aromatic H); $J_{3,4}$ =9.6, $J_{4,5}$ =9.0 Hz.

Found: C, 66.46; H, 4.49; N, 2.18%. Calcd for C₃₆H₂₉NO₁₁: C, 66.36; H, 4.49; N, 2.15%.

20 β : Mp 64—66 °C (with sintering at 60 °C); [α]_D²³ -43.2° (c 0.5, CHCl₃); ¹H NMR (90 MHz) δ =4.5—5.0 (3H, m, H-5 and 6a,b), 5.88 (1H, dd, H-4), 6.28 (1H, d, H-3), 7.2—8.2 (H-1 and aromatic H); $J_{3,4}$ =5.1, $J_{4,5}$ =4.8 Hz.

Found: C, 66.74; H, 4.46; N, 2.21%. Calcd for $C_{36}H_{29}NO_{11}$: C, 66.36; H, 4.49; N, 2.15%.

Methyl 4-O-(2-Acetamido-3,4,6-tri-O-benzoyl-2-deoxy-β-D-mannopyranosyl)-2,3,6-tri-O-benzyl-α-D-glucopyranoside (21). A solution of 18 (618 mg, 0.59 mmol) in dry tetrahydrofuran (8 ml) was treated with a 1 M solution of diborane in tetrahydrofuran (7.1 ml) as described for $6\rightarrow7$. Subsequent acetylation with acetic anhydride (4 ml) followed by processing of the mixture as described for 7 gave 461 mg (81%) of syrupy 21. Crystallization from diethyl ether afforded 411 mg of 21 as colorless prisms: mp 163-164°C (decomp); $[\alpha]_D^{22} - 21.2^{\circ}$ (c 0.25, CHCl₃); IR (KBr) 3430 (NH), 1720 (ester C=O), 1685 cm⁻¹ (amide C=O); ¹H NMR (300 MHz) δ =1.83 (3H, s, COCH₃), 3.37 (3H, s, OCH₃), 3.48 (1H, m, H-5'), 3.49 (1H, dd, H-2), 3.65-3.75 (3H, m, H-5 and H-6a,b), 3.88 (1H, t, H-3), 4.01 (1H, t, H-4), 4.24 (1H, dd, H-6'a), 4.39 (1H, dd, H-6'b), 4.55, 4.72, and 4.91 (each 2H, dd, 3×CH₂Ph), 4.61 (1H, d, H-1), 4.81 (1H, td, H-2'), 4.86 $(1H,\,d,\,H\text{-}1'),\,5.12\,(1H,\,dd,\,H\text{-}3'),\,5.49\,(1H,\,t,\,H\text{-}4'),\,5.70\,(1H,\,t)$ d, NH), 7.2-8.0 (aromatic H); $J_{1,2}=3.5$, $J_{2,3}=J_{3,4}=J_{4,5}=9.3$, $J_{1',2'}=1.5$, $J_{2',NH}=8.7$, $J_{2',3'}=4.0$, $J_{3',4'}=J_{4',5'}=10.0$, $J_{5',6'a}=5.0$, $J_{5',6'b}$ =3.5, $J_{6'a,b}$ =12.0 Hz; ¹³CNMR (75 MHz) δ =23.05 (COCH₃), 51.09 (C-2'), 55.29 (OCH₃), 63.25 (C-6'), 67.26 (C-4'), 68.45 (C-6), 69.35 (C-5), 72.34 (C-5') 72.43 (C-3'), 73.44, 73.56, and 75.00 (3×CH₂Ph), 76.51 (C-4), 79.38 (C-2), 80.41 (C-3), 98.27 (C-1), 98.87 (C-1'), 126.6—139.2 (aromatic C), 165.64, 165.64, and 165.90 (3×C=O), 170.34 (NHCO).

Found: C, 70.00; H, 5.87; N, 1.42%. Calcd for C₅₇H₅₇NO₁₄: C, 69.85; H, 5.86; N, 1.43%.

Methyl 4-O-(2-Acetamido-2-deoxy- β -p-mannopyranosyl)-2,3,6-tri-O-benzyl- α -p-glucopyranoside (22). A 0.05 M solution of sodium methoxide in dry methanol (18 ml) was added to 21 (352 mg, 0.36 mmol). The solution was stirred at

ambient temperature for 20 h, neutralized with acidic resin (Dowex 50W×8), and filtered through Celite. The filtrate was concentrated in vacuo to give a residue, which was eluted through a silica-gel column with chloroform-methanol (8:1). Concentration of the major fraction, followed by recrystallization from methanol-pentane gave 210 mg (87%) of **22** as colorless needles: mp 170—172 °C; $[\alpha]_{b}^{22}$ +14.2° (c 0.5, CH₃OH); IR (KBr) 3400—3300 (NH and OH), 1640 cm⁻¹ (amide C=O); ¹H NMR (DMSO- d_{6} , 90 MHz) δ =1.83 (3H, s, COCH₃), 3.29 (3H, s, OCH₃), 7.04 (1H, s, NH), 7.1—7.5 (aromatic H), 2.8—5.1 (other protons).

Found: C, 64.98; H, 6.89; N, 2.04%. Calcd for $C_{36}H_{45}NO_{12}$: C, 64.75; H, 6.79; N, 2.10%.

Methyl 4-O-(2-Acetamido-2-deoxy-β-D-mannopyranosyl)α-D-glucopyranoside (23). A solution of 22 (172 mg, 0.26 mmol) in acetic acid (10 ml) was hydrogenated in the presence of 10% palladium on carbon (85 mg) under atmosphere of hydrogen (3.45×10⁵ Pa) for 24 h. The mixture was filtered through Celite and the filtrate was concentrated in vacuo to give a syrup, which was purified by elution from a silica-gel column with chloroform-methanol (1:2). The major fraction was concentrated and the residual solid was washed with diethyl ether to give 110 mg (quantitative) of 23 as 1/2 hydrate: mp 190 °C (decomp, with sintering at 110 °C); $[\alpha]_D^{23}$ $+45.2^{\circ}$ (c 0.25, H₂O); ¹³C NMR (D₂O, 25.4 MHz) δ =22.8 (COCH₃), 54.1 (C-2'), 55.8 (OCH₃), 60.9 and 61.3 (C-6 and 6'), $\overline{67}$.5 (C-4'), 70.8 (C-2), 71.9, 72.5, and 72.8 (C-3, C-3', and C-5), 77.4 (C-5'), 79.6 (C-4), 99.2 (C-1), 100.2 (C-1'), 176.4 (CONH).

Found: C, 44.62; H, 7.13; N, 3.16%. Calcd for C₁₅H₂₇NO₁₁ 1/2H₂O: C, 44.33; H, 6.94; N, 3.45%.

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