Letter

Synthesis of Two Stereoisomers of Potentially Bioactive 13,19,20-Trihydroxy Derivative of Docosahexaenoic Acid

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Abstract The C16–C22 fragment with the acetylene terminus was constructed through the asymmetric dihydroxylation of the corresponding olefin, while the 15-iodo-olefin corresponding to the C11–C15 part was prepared via the asymmetric transfer hydrogenation of the corresponding acetylene ketone followed by hydrozirconation/io-dination. Both pieces were joined by a Sonogashira coupling, and the product was further converted into the title compound via a Wittig reaction with the remaining C1–C10 segment and Boland reduction using Zn with TMSCI.

Key words DHA metabolite, trihydroxylated DHA, organic synthesis, enyne, Boland reduction, TMSCI, Hansen protocol

Discovery of new metabolites of docosahexaenoic acid (DHA) formed by lipoxygenases and/or cytochrome P450 enzyme pathways is an active area of study because most of the metabolites isolated to date show properties of resolving and preventing inflammation.¹ Recently, we isolated compound **1**, a dihydroxylated metabolite of DHA² (Figure 1). Interestingly, **1** displayed wound-healing activity, a unique property for a DHA metabolite.³ Subsequently, the compound was synthesized, and the structure was determined.⁴ Similar properties were then found in new metabolite 2 ('maresin like'),⁵ and the structure of 2 was also determined after organic synthesis as well.⁶ We then envisaged that hydroxylated compound 3 would possess similar activities because of the structural similarity in the positions of the hydroxylated carbon atoms relative to those in 1 and 2. Based on the fact that the ω -epoxide⁷ (19,20-epoxide) of DHA is converted into 19,20-dihydroxy-DHA by soluble epoxide hydrolases^{7b,8} and that 13-hydroxy-19,20-epoxy derivative of DHA is isolated,⁹ compound **3** is likely produced by 13-lipoxygenation^{8e,10,11} of the former and/or by epoxide hydrolysis of the latter. It was envisaged that, in a similar way, epoxide hydrolysis of 11-hydroxy-17,18-epoxy-EPA¹² and 11-lipoxygenation^{10,13} of 17,18-dihydroxy-EPA would produce **4**, which have high potential to be bioactive.¹⁴





Among **3** and **4**, we chose **3** as a synthetic target because it has the same origin (DHA) as **1** and **2**, and we hoped that the method would be applicable to the synthesis of **4**. We established the 19,20-dihydroxy moiety as *syn*, based on Downloaded by: Western University. Copyrighted material.

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the same stereochemistry seen in leukotoxin diol and isoleukotoxin diol,¹⁵ in which the diol unit is nonconjugated, as that seen in 3. Consequently, diastereoisomers 3aa and 3ab, shown in Figure 2, represent different relative stereochemistry and would be useful stereoisomers for biological study. We also hoped that, once the method was established, enantiomers of 3aa and 3ab could also be synthesized as well.



Retrosynthetic analysis of 3aa indicated acetylene 5aa to be a key precursor, which could be disconnected to diol derivative **6a**, iodo-olefin **7a**, and Wittig reagent **8** (Scheme 1). We expected the asymmetric dihydroxylation $(AD)^{16}$ and the asymmetric transfer hydrogenation¹⁷ to produce the stereogenic centers in **6a** and **7a**. respectively. In practice, the syntheses were successful with high enantioselectivity, as summarized in Schemes 2 and 3. Wittig reagent 8 was prepared by a straightforward method, as delineated in Scheme 4.



Synthesis of **6a** was started with mesylation of alcohol **9**, which was followed by the CuI-assisted coupling¹⁸ with TMS-acetylene to afford 11 and the S_N2' regioisomer (structure not shown) in a 72:28 ratio, as determined with ¹H NMR spectroscopy (Scheme 2). Although the mixture was difficult to separate by chromatography on silica gel, the diols after the AD reaction were separated by chromatograDownloaded by: Western University. Copyrighted material.

phy on silica gel to afford 12 in 35% yield with 86% ee, as determined by ¹H NMR spectroscopy of the derived MTPA ester. Diol **12**¹⁹ was protected as TBS ether **13**, which produced 6a in 93% yield after TMS-desilylation with K₂CO₃.



Synthesis of 7a was carried out according to our previous method.²⁰ In brief, 1,3-propanediol was converted into alcohol 15 with 96% ee, as determined by HPLC analysis, via the asymmetric transfer hydrogenation¹⁷ of ketone 14²¹ (Scheme 3). TBDPS protection of the hydroxy group in 15



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and subsequent deprotection of the TMS group afforded **16**, which was subjected to hydrozirconation with in situ generated $Cp_2Zr(H)Cl^{22}$ iodination, and TBS desilylation to give *trans*-iodide **17** stereo- and regioselectively in 80% yield. In a similar manner, ketone **14** was converted into *ent*-**15** with 98% ee, and the subsequent transformations furnished *ent*-**7a**.

Wittig reagent **8** was synthesized from alcohol **18**²³ by the method delineated in Scheme 4. Thus, mesylation of **18** followed by Cul-assisted coupling¹⁸ with acetylene **19** af-

forded **20**, which gave olefin **21** in 81% yield upon reduction with P-2 Ni²⁴ under hydrogen. Desilylation, iodination, and the reaction of the resulting iodide with PPh₃ proceeded cleanly to furnish Wittig reagent **8** in a good yield.

Construction of the target compound **3aa** began with the Sonogashira coupling of 7a with 6a (1.1 equiv) to afford 22 in 83% yield (Scheme 5). Oxidation with SO₃·pyridine(Py)/Et₃N afforded the somewhat unstable aldehyde 23, which was then subjected to a Wittig reaction with the vlide generated from 8 and NaHMDS in the presence of HMPA (11 equiv, THF/HMPA = 5:1).²⁵ Olefin **24** was produced stereoselectively and cleanly, and subsequently, treated with TBAF to furnish the key intermediate 5aa in 63% yield from alcohol 22. Next, 5aa was subjected to the Boland reduction of the triple bond under the modified conditions reported by Hansen,²⁶ who added TMSCl to Zn(Cu/Ag) in aqueous MeOH. In practice, the reduction was successful in cleanly affording 25 in 81% yield. By contrast. an attempted reduction under the standard conditions²⁷ (without TMSCI) caused elimination of the hydroxy group at C13 and afforded a mixture of 25 and the elimination byproducts in a 73:27 weight ratio.²⁸ A similar elimination reaction has been reported previously.²⁹ Finally, hydrolysis of 25 produced 3aa in 84% vield.



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In a similar way, the coupling reaction of *ent*-**7a** with **6a** gave **26**, which was converted into **5ab** in 66% yield. Reduction of **5ab** with Zn/TMSCl proceeded cleanly, and the resulting ester **27** was hydrolyzed to acid **3ab**. The ¹H NMR spectra of **3aa** and **3ab** were completely superimposed, whereas some of the olefinic carbons in the ¹³C NMR spectrum were observed at different positions (see the Supporting Information).

In summary, we developed a method to produce the title triol and successfully performed the syntheses of **3aa** and **3ab** through the Boland reduction of enyne **5aa** and **5ab** with Zn(Cu/Ag), which proceeded cleanly in the presence of TMSCL³⁰ The ¹³C NMR spectra of these products were found to be useful for determining the relative stereochemistry between the carbon atoms at C13 and *syn* C19,C20. Synthesis of the enantiomers of **6a** and **7a** is undoubtedly possible by changing the chirality of the catalysts and would furnish the enantiomers of **3aa** and **3ab**. The asymmetric dihydroxylation reaction of the *cis* olefin corresponding to **11** produces the *anti* diol, which is the key intermediate for the synthesis of another set of targets with *anti* stereochemistry at C19 and C20.

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Supporting Information

Supporting information for this article is available online at https://doi.org/10.1055/s-0040-1706415.

References and Notes

(1) (a) Pham, T. L.; Kakazu, A. H.; He, J.; Jun, B.; Bazan, N. G.; Bazan, H. E. P. Sci. Rep. 2020, 10, 4582. (b) Saini, R. K.; Keum, Y.-S. Life Sci. 2018, 203, 255. (c) Basil, M. C.; Levy, B. D. Nat. Rev. Immunol. 2016, 16, 51. (d) Serhan, C. N.; Dalli, J.; Colas, R. A.; Winkler, J. W.; Chiang, N. Biochim. Biophys. Acta 2015, 1851, 397. (e) Kuhn, H.; Banthiya, S.; van Leyen, K. Biochim. Biophys. Acta 2015, 1851, 308. (f) Isobe, Y.; Arita, M. J. Clin. Biochem. Nutr. 2014, 55, 79. (g) Serhan, C. N.; Nicos, A.; Petasis, N. A. Chem. Rev. 2011, 111, 5922. (h) Lukiw, W. J.; Cui, J.-G.; Marcheselli, V. L.; Bodker, M.; Botkjaer, A.; Gotlinger, K.; Serhan, C. N.; Bazan, N. G. J. Clin. Invest. 2005, 115, 2774. (i) Arita, M.; Bianchini, F.; Aliberti, J.; Sher, A.; Chiang, N.; Hong, S.; Yang, R.; Petasis, N. A.; Serhan, C. N. J. Exp. Med. 2005, 201, 713. (j) Bazan, N. G. Mol. Neurobiol. 2005, 31, 219. (k) Bazan, N. G. Brain Pathol. 2005, 15, 159. (1) Marcheselli, V. L.; Hong, S.; Lukiw, W. J.; Tian, X. H.; Gronert, K.; Musto, A.; Hardy, M.; Gimenez, J. M.; Chiang, N.; Serhan, C. N.; Bazan, N. G. J. Biol. Chem. 2003, 278, 43807.

- (2) (a) Lu, Y.; Tian, H.; Hong, S. J. Lipid Res. 2010, 51, 923. (b) Tian,
 H.; Lu, Y.; Shah, S. P.; Hong, S. J. Cell. Biochem. 2010, 111, 266.
- (3) (a) Tian, H.; Lu, Y.; Shah, S. P.; Wang, Q.; Hong, S. Stem Cells Dev. **2012**, 21, 1187. (b) Tian, H.; Lu, Y.; Shah, S. P.; Hong, S. Am. J. Pathol. **2011**, 179, 1780. (c) Tian, H.; Lu, Y.; Shah, S. P.; Hong, S. J. Biol. Chem. **2011**, 286, 4443.
- (4) Nishimura, K.; Sakaguchi, T.; Nanba, Y.; Suganuma, Y.; Morita, M.; Hong, S.; Lu, Y.; Jun, B.; Bazan, N. G.; Arita, M.; Kobayashi, Y. J. Org. Chem. 2018, 83, 154.
- (5) Hong, S.; Lu, Y.; Tian, H.; Alapure, B. V.; Wang, Q.; Bunnell, B. A.; Laborde, J. M. *Chem. Biol.* **2014**, *21*, 1318.
- (6) Hong, S.; Lu, Y.; Morita, M.; Saito, S.; Kobayashi, Y.; Jun, B.; Bazan, N. G.; Xu, X.; Wang, Y. Synlett **2019**, *30*, 343.
- (7) (a) Roy, J.; Watson, J. E.; Hong, I. S.; Fan, T. M.; Das, A. J. Med. Chem. 2018, 61, 5569. (b) McDougle, D. R.; Watson, J. E.; Abdeen, A. A.; Adili, R.; Caputo, M. P.; Krapf, J. E.; Johnson, R. W.; Kilian, K. A.; Holinstat, M.; Das, A. Proc. Natl. Acad. Sci. U.S.A. 2017, 114, E6034. (c) Shearer, G. C.; Harris, W. S.; Pederson, T. L.; Newman, J. W. J. Lipid Res. 2010, 51, 2074.
- (8) (a) Pickens, C. A.; Sordillo, L. M.; Zhang, C.; Fenton, J. I. *Metab. Clin. Exp.* **2017**, 70, 177. (b) Tajima, Y.; Ishikawa, M.; Maekawa, K.; Murayama, M.; Senoo, Y.; Nishimaki-Mogami, T.; Nakanishi, H.; Ikeda, K.; Arita, M.; Taguchi, R.; Okuno, A.; Mikawa, R.; Niida, S.; Takikawa, O.; Saito, Y. *Lipids Health Dis.* **2013**, *12*, 68. (c) Morisseau, C.; Inceoglu, B.; Schmelzer, K.; Tsai, H.-J.; Jinks, S. L.; Hegedus, C. M.; Hammock, B. D. *J. Lipid Res.* **2010**, *51*, 3481. (d) Oliw, E. H.; Sprecher, H. W. *Biochim. Biophys. Acta* **1991**, *1086*, 287. (e) VanRollins, M.; Baker, R. C.; Sprecher, H. W.; Murphy, R. C. *J. Biol. Chem.* **1984**, *259*, 5776.
- (9) Arita, M.; Arai, H.; Isobe, Y.; Kubota, T. WO2012023254 A1, **2012**.
- (10) Mortimer, M.; Järving, R.; Brash, A. R.; Samel, N.; Järving, I. Arch. Biochem. Biophys. **2006**, 445, 147.
- (11) (a) Fredman, G.; Hellmann, J.; Proto, J. D.; Kuriakose, G.; Colas, R. A.; Dorweiler, B.; Connolly, E. S.; Solomon, R.; Jones, D. M.; Heyer, E. J.; Spite, M.; Tabas, I. *Nat. Commun.* **2016**, *7*, 12859. (b) Hong, S.; Gronert, K.; Devchand, P. R.; Moussignac, R.-L.; Serhan, C. N. J. Biol. Chem. **2003**, *278*, 14677. (c) Serhan, C. N.; Hong, S.; Gronert, K.; Colgan, S. P.; Devchand, P. R.; Mirick, G.; Moussignac, R.-L. J. Exp. Med. **2002**, *196*, 1025.
- (12) Kubota, T.; Arita, M.; Isobe, Y.; Iwamoto, R.; Goto, T.; Yoshioka, T.; Urabe, D.; Inoue, M.; Arai, H. *FASEB J.* **2014**, *28*, 586.
- (13) (a) Wang, C.; Liu, W.; Yao, L.; Zhang, X.; Zhang, X.; Ye, C.; Jiang, H.; He, J.; Zhu, Y.; Ai, D. Br. J. Pharmacol. 2017, 174, 2358.
 (b) Coffa, G.; Hill, E. M. Lipids 2000, 35, 1195. (c) Hawkins, D. J.; Brash, A. R. J. Biol. Chem. 1987, 262, 7629.
- (14) A similar compound would be derived from n-3 docosapentaenoic acid, see: Vik, A.; Dalli, J.; Hansen, T. V. *Bioorg. Med. Chem. Lett.* **2017**, *27*, 2259.
- (15) (a) Young, U. J.; Miklossy, G.; Chai, X.; Wongwiwatthananukit, S.; Toyama, O.; Songsak, T.; Turkson, J.; Chang, L. C. *Fitoterapia* **2014**, *93*, 194. (b) Blée, E.; Schuber, F. *Eur. J. Biochem.* **1995**, *230*, 229. (c) Morris, L. J.; Crouchman, M. L. *Lipids* **1972**, *7*, 372. (d) Morris, L. J.; Crouchman, M. L. Lipids **1969**, *4*, 50.
- (16) Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. Chem. Rev. 1994, 94, 2483.
- (17) Matsumura, K.; Hashiguchi, S.; Ikariya, T.; Noyori, R. J. Am. Chem. Soc. **1997**, 119, 8738.
- (18) Spinella, A.; Caruso, T.; Martino, M.; Sessa, C. Synlett 2001, 1971.
- (19) Caruso, T.; Spinella, A. Tetrahedron: Asymmetry 2002, 13, 2071.
- (20) Ogawa, N.; Tojo, T.; Kobayashi, Y. Tetrahedron Lett. **2014**, 55, 2738.

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- (21) (a) Saito, S.; Nanba, Y.; Morita, M.; Kobayashi, Y. Synlett 2019, 30, 1085. (b) Hartmann, O.; Kalesse, M. Org. Lett. 2012, 14, 3064.
- (22) (a) Kiyotsuka, Y.; Igarashi, J.; Kobayashi, Y. Tetrahedron Lett.
 2002, 43, 2725. (b) Huang, Z.; Negishi, E. Org. Lett. 2006, 8, 3675.
- (23) (a) Akoto, C. O.; Rainier, J. D. Synthesis 2019, 51, 3529.
 (b) Chakraborty, T. K.; Purkait, S.; Das, S. Tetrahedron 2003, 59, 9127.
- (24) Brown, C. A.; Ahuja, V. K. J. Chem. Soc., Chem. Commun. **1973**, 553.
- (25) The use of 1 equiv of HMPA caused the elimination of the TBDPS-oxy group.
- (26) Mohamed, Y. M. A.; Hansen, T. V. Tetrahedron 2013, 69, 3872.
- (27) Boland, W.; Schroer, N.; Sieler, C. Helv. Chim. Acta 1987, 70, 1025.
- (28) Determined by ¹H NMR spectroscopy and R_f values (0.60 and 0.33 for the byproduct and **25** (hexane/EtOAc = 1:2).
- (29) Dayaker, G.; Durand, T.; Balas, L. *Chem. Eur. J.* **2014**, *20*, 2879; see the Supporting Information on page S35.
- (30) To an argon-bubbled suspension of Zn powder (4.30 g, 65.8 mmol) in H₂O (20 mL) were added Cu(OAc)₂ (436 mg, 2.40 mmol) and, after 20 min, AgNO₃ (442 mg, 2.60 mmol). The suspension was stirred for 45 min and filtered by suction, and the

remaining solids were washed with H₂O, MeOH, acetone, and Et₂O, sequentially. A solution of triol **5aa** (129 mg, 0.330 mmol) in H₂O/MeOH (1:1, 20 mL) was added to the above solids. Subsequently, TMSCl (0.410 mL, 3.28 mmol) was added. The mixture was stirred at rt for 18 h and filtered through a pad of Celite. The filtrate was concentrated, and the residue was purified first by chromatography on silica gel (hexane/EtOAc) and second by recycling HPLC (LC-Forte/R equipped with YMC-Pack SIL-60, ø 20 × 250 mm, hexane/EtOAc 30:70, 25 mL/min) to afford methyl ester 25 (104 mg, 81%) as a colorless liquid: $R_f = 0.22$ (hexane/EtOAc = 1:2); $[\alpha]_D^{21} - 2.3$ (c 1.0, CHCl₃). IR (neat): 3480, 1733, 1652, 759 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 0.99 (t, J = 7.4 Hz, 3 H), 1.44-1.66 (m, 2 H), 1.77 (br s, 1 H), 2.22-2.52 (m, 10 H), 2.77-2.90 (m, 4 H), 3.40 (dt, J = 8.4, 4.2 Hz, 1 H), 3.53 (q, J = 5.8 Hz, 1 H), 3.68 (s, 3 H), 4.25 (q, J = 6.0 Hz, 1 H), 5.31–5.60 (m, 7 H), 5.77 (dd, J = 15.2, 6.0 Hz, 1 H), 6.16 (t, J = 11.2 Hz, 1 H), 6.55 (dd, J = 15.2, 11.2 Hz, 1 H). ¹³C NMR (100 MHz, CDCl₃): δ = 10.1, 22.9, 25.7, 25.9, 26.6, 32.3, 34.1, 35.4, 51.7, 71.9, 73.5, 75.1, 125.0, 125.3, 127.5, 127.9, 128.0, 128.3, 129.4, 130.8, 131.2, 136.5, 173.8. HRMS (FD): m/z calcd for C₂₃H₃₆O₅ [M]⁺: 392.25627; found: 392.25781.

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