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N-Bromosuccinimide-Mediated Conversion of Allyl Glycosides to Glycosyl Hemiacetals

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A novel reaction condition has been developed for the hydrolysis of differentially functionalized allyl glycosides to their corresponding glycosyl hemiacetal derivatives in the presence of N-bromosuccinimide (NBS). The reaction condition is exceptionally fast, mild, and compatible with most of the functional groups used in the oligosaccharide synthesis, and yields were excellent.

Keywords Allyl glycoside; N-Bromosuccinimide; Glycosyl hemiacetal; Hydrolysis

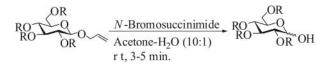
INTRODUCTION

1-Hydroxy sugars or glycosyl hemiacetal derivatives are useful intermediates for the synthesis of complex oligosaccharides.^[1,2] They can be used directly in the dehydrative glycosylation reactions^[3] or can be converted to reactive glycosyl donors^[4–6] to be used in the oligosaccharides or natural product synthesis.^[7,8] Because of their usefulness, a number of reports appeared in the literature dealing with the preparation of glycosyl hemiacetal derivatives, which include (a) removal of the anomeric acetyl group using hazardous hydrazine salts^[9] or organic bases^[10] or acidic conditions^[11]; (b) hydrolysis of alkyl glycosides^[12] and oxidative removal of 4-methoxyphenyl glycosides^[13]; and (c) hydrolysis of thioglycoside derivatives using a variety of thiophilic reagents.^[14–20] Hydrolysis of thioglycoside derivatives has been found to be beneficial over other methods due to the less toxic and mild reaction conditions as well as compatibility of other functional groups present in the sugar skeleton.

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Suitably functionalized allyl glycosides have been used as glycosyl acceptors in the synthesis of several complex oligosaccharides.^[21] In a multistep oligosaccharide synthetic strategy the anomeric allyl group has been used as a temporary protecting group that can be removed after glycosylation to generate hemiacetal derivatives for their use in the next step. Conventionally, removal of the allyl group is carried out using expensive palladium or rhodium salts^[22]; in a two-step reaction sequence involving the isomerization of the O-allyl group to the more labile O-propenyl group in the presence of expensive iridium salt^[23]; or using irradiation^[24] followed by hydrolysis of the resulting O-propenyl group using N-bromosuccinimide (NBS). In some cases the removal of the anomeric allyl group became exceptionally problematic because of the functional groups present in the substrates and thereby required special reaction conditions.^[25,26] However, many of these methods for the removal of the allyl group suffer from limitations such as use of expensive reagents, incompatibility with acidic functional groups, relatively low yield, and sometimes harsh reaction conditions. In this context, it would be useful to develop an economically convenient mild reaction protocol for the preparation of glycosyl hemiacetal derivatives having sensitive protecting groups. Although NBS has been used to remove allyl ether under the special reaction conditions mentioned earlier, we disclose herein a novel metal-free reaction condition for the preparation of functionalized glycosyl hemiacetal derivatives by direct NBSmediated hydrolysis of allyl glycosides, avoiding the use of expensive reagents or stringent reaction conditions (Sch. 1).



Scheme 1: *N*-Bromosuccinimide (NBS)-mediated hydrolysis of allyl glycoside for the preparation of glycosyl hemiacetal derivatives.

RESULTS AND DISCUSSION

During the synthesis of oligosaccharides^[27] using suitably protected allyl glycoside as the glycosyl acceptor under the thioglycoside activation condition in the presence of *N*-iodosuccinimide (NIS) and trifluoromethane sulfonic acid (TfOH) combination, we isolated a considerable amount of unwanted product, which was characterized as the hemiacetal derivative generated from the allyl glycoside. Taking this clue from the earlier experiments, allyl glycoside (1) was allowed to react separately with NBS and NIS in the presence and absence of an acid in different solvents, for example, CH_2Cl_2 , CH_3CN-H_2O (9:1), acetone- H_2O (10:1), THF. It was observed that the use of 1.1 equiv. of NBS in acetone- H_2O can form hemiacetal derivative in excellent yield from the corresponding

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allyl glycosides without requiring any acid as activator at room temperature in 3 minutes. Although NIS was almost equally effective for this transformation, we opted to use economically cheaper NBS. However, NBS has some limitation because of its tendency for radical reactions. To expand the scope of the reaction, a series of differentially protected allyl glycoside was transformed into corresponding hemiacetal derivative in excellent yield (Sch. 1, Table 1) following similar reaction conditions. Use of CH₃CN-H₂O (9:1) as solvent can furnish similar yield of the product, but acetone- H_2O has been used because of its cheap availability. Most of the functional groups used in the protection of hydroxy groups of the carbohydrate backbone (i.e., benzylidene, isopropylidene acetal, benzyl, 4-methoxybenzyl, benzoyl, acetyl, tert-butyldiphenylsilyl, etc.) remain unaffected under the reaction condition. Although NBS has been used in the oxidative opening of benzylidene acetal under the Hanessian-Hullar reaction condition, it has no effect on the benzylidene acetal using the present reaction condition. A comparative study on the effect of solvent and halogenating agents on the formation of hemiacetal derivatives is presented in Table 2. Using α - and β -allyl glycosides, similar results were obtained.

We presumed that bromonium ion (Br^+) generated from NBS and the allyl group are in a close proximity to form an addition product, which after hydrolysis resulted in the glycosyl hemiacetal derivatives from the allyl glycoside.

In summary, a novel, metal-free mild reaction condition has been developed for the direct hydrolysis of allyl glycosides in the presence of NBS. This finding can also explain the low-yielding glycosylation reactions using functionalized allyl glycoside as glycosyl acceptor under iodonium ion-mediated thioglycoside activation conditions. Use of readily available reagents, without requirement of heavy metallic salts, high boiling solvents, or expensive Lewis acid additives, makes this exceptionally fast reaction protocol for the preparation of glycosyl hemiacetal derivatives an attractive alternative to the existing methods.

EXPERIMENTAL

General Procedure

All reactions were monitored by thin layer chromatography over silica gel-coated TLC plates. The spots on TLC were visualized by warming ceric sulphate (2% Ce(SO₄)₂ in 2N H₂SO₄)-sprayed plates in hot plate. Silica gel 230–400 mesh was used for column chromatography. ¹H and ¹³C NMR spectra were recorded on Bruker Avance DRX 500 MHz using CDCl₃ as solvents and TMS as internal reference unless stated otherwise. Chemical shift value is expressed in δ ppm. ESI-MS were recorded on a Micromass Quattro II triple

Thioglycosides	Products	Time (min)	Yield (%) (α/β)	Ref
1 Aco Aco	15 Aco Aco OH	3	90 (2:1)	(14)
Aco OAc 2 Aco Aco	AcO OAc 16 AcO ACO OH	<2	92 (3:1)	(14)
3 ACO OAC ACO O	17 AcO OAc AcO OAc	3	90 (5:1)	(14)
BzO OBz 4 BzO BzO O BzO	BzO OBz 18 BzO BzO OH	5	88 (2:1)	(14)
BnO OBn 5 BnO BnO O	BnO OBn 19 BnO BnO OH	5	82 (2:1)	(14)
6 BzO BzO BzO BzO	20 BzO BzO OH	5	80 (3:1)	(16)
7 Aco Joh Aco OAc	21 Aco OAc	<2	90 (4:1)	(14)
8 DOBn BnO	$22 \qquad \qquad \overbrace{OBn}^{OZ} \xrightarrow{OH}_{OBn}$	<2	85 (3:1)	(14)
9 Aco Aco	23 Aco Aco OH	3	86 (2:1)	_
10 Phto O Aco PhthN	24 Phto Of Control Phth Of Phth Of Of Control Phth Of Control Phth Of Control Phth Of Control Phth Of Control Phthe Of Contro	5	90 (1:3)	(18)
Ph O Bro BzO	25 Bno Bzo OH	5	82 (3:1)	_
$12 \text{ MBnO} \xrightarrow{\text{Ph}}_{\text{AcO}} -\infty$	26 MBnO Aco OH	3	80 (2:1)	—

Table 1: Hydrolysis of allyl glycosides using N-bromosuccinimide (NBS)^a at room temperature

(Continued on next page)

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 Table 1: Hydrolysis of allyl glycosides using N-bromosuccinimide (NBS)^a at room

 temperature (Continued)

Thioglycosides	Products	Time (min)	Yield (%) (α/β)	Ref
13 Aco 200	27 Aco 0,0 OH	5	88 (3:1)	(16)
AcO OAc 14 AcO AcO AcO	Aco OAc 28 Aco Aco Aco OH	5	82 (2:1)	(16)

NPhth, N-phthalimido; MBn, 4-methoxybenzyl. $^{\circ}$ 1.1 equiv. of NBS was used in acetone-H₂O (10:1).

quadrupole mass spectrometer. Commercially available grades of organic solvents of adequate purity are used in many reactions.

Typical Experimental Protocol for The Preparation of Glycosyl Hemiacetal Derivatives

To a solution of functionalized allyl glycoside (1 mmol) in acetone-H₂O (2 mL; 10:1, v/v) was added NBS (1.1 mmol) at 0 to 5°C and the reaction mixture was allowed to stir at the same temperature for the appropriate time (Table 1). The reaction mixture was diluted with CH_2Cl_2 (20 mL) and the organic layer was washed with 5% $Na_2S_2O_3$ and H_2O , dried (Na_2SO_4), and evaporated to dryness. The crude product was purified over SiO₂ using hexane-EtOAc as eluant to give pure glycosyl hemiacetal derivative. An inseparable anomeric mixture of hemiacetal derivative was formed in every case; the ratio was determined from the integration of their ¹H NMR spectra. ¹H NMR and ¹³C NMR spectra of the known glycosyl hemiacetal derivatives matched with the data reported in the cited references. Spectral data for new compounds,

Table 2: Optimization of the reaction condition for the formation of glycosylhemiacetal derivative (15) from allyl glycoside (1)

Entry	Substrate	Catalyst	Solvent	Time (min)	Yield (%)
1 2 3 4 5 6	1 1 1 1 1	NBS NBS NBS NIS NIS I ₂	$\begin{array}{c} CH_2Cl_2\\ CH_3CN-H_2O\\ Acetone-H_2O\\ CH_3CN-H_2O\\ Acetone-H_2O\\ Acetone-H_2O\\ Acetone-H_2O\end{array}$	30 5 3 15 15 45	85 88 90 85 90 82

which were not reported earlier, are presented below. Although both α - and β -anomers formed under the reaction conditions, spectral data for the major isomer are presented for the sake of simplicity.

2,3-Di-O-acetyl-4,6-O-benzylidene- α -D-galactopyranose (23)

¹H NMR (CDCl₃, 300 MHz): δ 7.48–7.33 (m, 5 H, Ar-H), 5.46 (s, 1 H, PhCH), 5.32–5.29 (m, 1 H, H-2), 4.94 (dd, J = 10.4, 3.4 Hz, 1 H, H-3), 4.53 (d, J = 3.4 Hz, 1 H, H-1), 4.32 (br s, 1 H, H-4), 3.85–3.77 (m, 2 H, H-6_{a,b}), 3.50–3.49 (m, 1 H, H-5), 2.09 (s, 6 H, 2 COCH₃); ESI-MS: calcd. for C₁₇H₂₀O₈: m/z 352.12; found: m/z 335.1 [M-H₂O+1].

2-O-benzoyl-3-O-benzyl-4,6-O-benzylidene- α -Dgalactopyranose (25)

¹H NMR (CDCl₃, 300 MHz): δ 8.02–7.17 (m, 15 H, Ar-H), 5.59–5.54 (m, 1 H, H-2), 5.46 (s, 1 H, PhC*H*), 4.67 (d, *J* = 12.4 Hz, 1 H, PhC*H*_{2a}), 4.58 (d, *J* = 9.0 Hz, 1 H, H-1), 4.56 (d, *J* = 12.4 Hz, 1 H, PhC*H*_{2b}), 4.20 (br s, 1 H, H-4), 4.03–4.01 (m, 1 H, H-3), 3.76–3.69 (m, 2 H, H-6_{a,b}), 3.39–3.37 (m, 1 H, H-5); ESI-MS: calcd. for C₂₇H₂₆O₇: *m/z* 462.17; found: *m/z* 445.1 [M-H₂O+1].

2-O-acetyl-4,6-O-benzylidene-3-O-(4-methoxybenzyl)-α-Dgalactopyranose (26)

 $^{1}\mathrm{H}$ NMR (CDCl₃, 300 MHz): δ 7.51–6.80 (m, 9 H, Ar-H), 5.46 (s, 1 H, PhCH), 5.32–5.26 (m, 1 H, H-2), 4.58–4.43 (m, 2 H, MeOPhCH₂), 4.52 (d, J = 3.0 Hz, 1 H, H-1), 4.30–4.27 (m, 1 H, H-3), 4.15 (br s, 1 H, H-4), 3.87–3.84 (m, 2 H, H-6_{a,b}), 3.79 (s, 3 H, OCH₃), 3.57–3.55 (m, 1 H, H-5); ESI-MS: calcd. for C₂₃H₂₆O₈: m/z 430.16; found: m/z 413.1 [M-H₂O+1].

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REFERENCES

1. (a) Boons, G.-J.; Hale, K.J. Organic Synthesis with Carbohydrates. Academic: Sheffield, **2000**, pp. 155–172; (b) Barresi,F.; Hindsgaul, O. Chemically synthesized oligosaccharides, 1994. A searchable table of glycosidic linkages. J. Carbohydr. Chem. **1995**, *14*, 1043–1087; (c) Zhu, X.; Schmidt, R.R. New principles for glycoside-bond formation. Angew. Chem. Int. Ed. Engl. **2009**, *48*, 1900–1934.

2. Hanessian, S. Total Synthesis of Natural Products: The "Chiron" Approach. Pergamon Press, New York, New York, 1983, pp. 1–291.

3. (a) Garcia, B.A.; Poole, J.L.; Gin, D.Y. Direct glycosylations with 1-hydroxy glycosyl donors using trifluoromethanesulfonic anhydride and diphenyl sulfoxide. *J. Am. Chem.*

82 R. Panchadhayee and A.K. Misra

Soc. **1997**, *119*, 7597–7598; (b) Garcia, B.A.; Gin, D.Y. Dehydrative glycosylation with activated diphenyl sulfonium reagents. Scope, mode of C(1)-hemiacetal activation, and detection of reactive glycosyl intermediates. J. Am. Chem. Soc. **2000**, *122*, 4269–4279; (c) Garcia, B.A.; Gin, D.Y. Synthesis of glycosyl-1-phosphates via dehydrative glycosylation. Org. Lett. **2000**, *2*, 2135–2138.

4. Caputo, R.; Kunz, H.; Mastroianni, D.; Palumbo, G.; Pedatella, S.; Solla, F. Mild synthesis of protected α -D-glycosyl iodides. *Eur. J. Org. Chem.* **1999**, 3147–3150.

5. Schmidt, R.R.; Jung, K.-H. *Preparative Carbohydrate Chemistry*; Hanessian, S. Ed.; Marcel Dekker: New York, **1996**, pp. 283–312.

6. Yokoyama, M. Methods of synthesis of glycosyl fluorides. *Carbohydr. Res.* 2000, 327, 5–14.

7. (a) Schmidt, R.R.; Klotz, W. Glycoside bond formation via anomeric O-alkylation: how many protective groups are required? *Synlett* **1991**, 168–170; (b) Smith, A.B. III; Hale, K.J.; Vaccaro, H.A. The total synthesis of (+)-payllanthostatin 2. *Tetrahedron Lett.* **1987**, 28, 5591–5594.

8. Satoh, Y.; Nakahata, T.; Kuwahara, S. Synthesis of the glycosyl lactol moiety of halichoblelide.*Tetrahedron* **2007**, *63*, 11733–11737.

9. Excoffier, G.; Gagnaire, D.; Utille, J.-P. Coupure sélective par l'hydrazine des groupements acétyles anomères de résidus glycosyles acétylés. *Carbohydr. Res.* **1975**, *39*, 368–373.

10. Sambaiah, T.; Fanwick, P.E.; Cushman, M. Regioselective 1-O-acyl hydrolysis of peracylated glycopyranoses by mercuric chloride and mercuric oxide. *Synthesis* **2001**, 1450–1452.

11. Tiwari, P.; Misra, A.K. Selective removal of anomeric *O*-acetate groups in carbo-hydrates using HClO₄-SiO₂. *Tetrahedron Lett.* **2006**, *47*, 3573–3576.

12. Koto, S.; Morishima, N.; Miyata, Y.; Zen, S. Preparation of 2,3,4,6-tetra-O-benzyl-D-mannose. Bull. Chem. Soc. Jpn. 1976, 49, 2639–2640.

13. (a) Mori, M.; Ito, Y.; Ogawa, T. A highly stereoselective and practical synthesis of cyclomannohexaose, $Cyclo\{\rightarrow 4\}$ - $[\alpha$ -D-Manp- $(1\rightarrow 4)$ - $]_5$ - α -D-Manp- $(1\rightarrow \}$, a manno isomer of cyclomaltohexaose. Carbohydr. Res. **1989**, 192, 131–146; (b) Dan, A.; Ito, Y.; Ogawa, T. A convergent and stereocontrolled synthetic route to the core pentasaccharide structure of asparagine-linked glycoproteins. J. Org. Chem. **1995**, 60, 4680–4681.

14. Bujar Barua, P.M.; Sahu, P.R.; Mondal, E.; Bose, G.; Khan, A.T. A mild and environmentally benign synthetic protocol for catalytic hydrolysis of thioglycosides. *Synlett* **2002**, 81–84.

15. Motawia, M.S.; Marcussan, J.; Moeller, B.L. A general method based on the use of *N*-bromosuccinimide for removal of the thiophenyl group at the anomeric position to generate a reducing sugar with the original protecting groups still present. *J. Carbohydr. Chem.* **1995**, *14*, 1279–1294; (b) Uchiro, H.; Wakiyama, Y.; Mukaiyama, T. A new efficient method for catalytic hydrolysis of thioglycoside. *Chem. Lett.* **1998**, 567–568; (c) Nicolaou, K.C.; Seitz, S.P.; Papahatjis, D.P. A mild and general method for the synthesis of *O*-glycosides. *J. Am. Chem. Soc.* **1983**, *105*, 2430–2434.

16. Misra, A.K.; Agnihotri, G. Chloramine-T-mediated chemoselective hydrolysis of thioglycosides into glycosyl hemiacetals under neutral conditions. *Carbohydr. Res.* **2004**, *339*, 885–890.

17. Duynstee, H.I.; de Koning, M.C.; Ovaa, H.; van der Marel, G.A.; van Boom, J.H. Synthesis of verbascoside: a dihydroxyphenylethyl glycoside with diverse bioactivity. *Eur. J. Org. Chem.* **1999**, 2623–2632.

18. Mandal, P.K.; Misra, A.K. Mild and efficient hydrolysis of thioglycosides to glycosyl hemiacetals using *N*-iodosaccharin. *Synlett* **2007**, 1207–1210.

19. Dinkelaar, J.; Witte, M.D.; van denBos, L.J.; Overkleeft, H.S.; van derMarel, G.A. NIS/TFA: a general method for hydrolyzing thioglycosides. *Carbohydr. Res.* **2006**, *341*, 1723–1729.

20. Oshitari, T.; Shibasaki, M.; Yoshizawa, T.; Tomita, M.; Takao, K.; Kobayashi, S. Synthesis of 2-O-(3-O-carbamoyl-α-D-mannopyranosyl)-L-gulopyranose: sugar moiety of antitumor antibiotic bleomycin. *Tetrahedron* **1997**, *53*, 10993–11006.

21. (a) Aspinall, G.O.; Gammon, D.W.; Sood, R.K.; Chatterjee, D.; Rivoire, B.; Brennan, P.J. Structures of the glycopeptidolipid antigens of serovars 25 and 26 of the *My*cobacterium avium serocomplex, synthesis of allyl glycosides of the outer disaccharide units and serology of the derived neoglycoproteins. *Carbohydr. Res.* **1992**, 237, 57–77; (b) Dubber, M.; Lindhorst, T.K. Synthesis of octopus glycosides: core molecules for the construction of glycoclusters and carbohydrate-centered dendrimers. *Carbohydr. Res.* **1998**, 310, 35–41; (c) Lee, R.T.; Lee, Y.C. Synthesis of allyl 3-deoxy- and 4-deoxy- β -Dgalactopyranoside and simultaneous preparation of Gal(1 \rightarrow 2)- and Gal(1 \rightarrow 3)-linked disaccharide glycosides. *Carbohydr. Res.* **1994**, 251, 69–79; (d) Zhu, Y.; Kong, F. A facile and effective synthesis of α -(1 \rightarrow 6)-linked mannose di-, tri-, tetra-, hexa-, octa-, and dodecasaccharides, and β -(1 \rightarrow 6)-linked glucose di-, tri-, tetra-, hexa-, and octasaccharides using sugar trichloroacetimidates as the donors and unprotected or partially protected glycosides as the acceptors. *Carbohydr. Res.* **2001**, 332, 1–21.

22. Greene, T.W.; Wuts, P.G.M. Protective groups in organic synthesis, 3rd Ed., John Wiley & Sons, Inc., New York, New York, **1999**, pp. 67–74.

23. (a) Shiozaki, M.; Doi, H.; Tanaka, D.; Shimozato, T.; Kurakata, S.-i. Syntheses of glucose-containing E5564 analogues and their LPS-antagonistic activities. *Tetrahedron* **2006**, *62*, 205–225; (b) Nakamura, T.; Shiozaki, M. Total synthesis of sphingofungin E from D-glucose derivative. *Tetrahedron* **2002**, *58*, 8779–8791; (c) Nakamura, T.; Shiozaki, M. Total synthesis of sphingofungin E. Tetrahedron Lett. **2001**, *42*, 2701–2704.

24. Diaz, R.R.; Melgarejo, C.R.; Lopez-Espinosa, M.T.P.; Cubero, I.I. A novel, mild, and practical regeneration of alcohols from their allylic ethers by NBS/H₂O. *J. Org. Chem.* **1994**, *59*, 7928–7929.

25. Yu, B.; Zhang, J.; Lu, S.; Hui, Y. A novel and efficient deprotection of the allyl group at the anomeric oxygen of carbohydrates. *Synlett* **1998**, 29–30.

26. (a) Nakayama, K.; Uoto, K.; Higashi, K.; Soga, T.; Kusama, T. A useful method for deprotection of the protective allyl group at the anomeric oxygen of carbohydrate moieties using tetrakis(triphenylphosphine) palladium. *Chem. Pharm. Bull.* **1992**, *40*, 1718–1720; (b) Lüning, J.; Möller, U.; Debski, N.; Welzel, P. A new method for the cleavage of allyl glycosides. *Tetrahedron Lett.* **1993**, *34*, 5871–5874.

27. (a) Guchhait, G.; Misra, A.K. Total synthesis of the heptasaccharide repeating unit of the iron-binding exopolysaccharide secreted by *Klebsiella oxytoca* BAS-10. *Tetrahedron Asymm.* **2009**, *20*, 1791–1797; (b) Pandey, S.; Ghosh, S.; Misra, A.K. Synthesis of a trisaccharide and a tetrasaccharide from the cell-wall lipopolysaccharides of *Azospirillum brasilense* S17. *Synthesis* **2009**, 2584–2590; (c) Panchadhayee, R.; Misra, A.K. First synthesis of a pentasaccharide repeating unit of the *O*-antigenic polysaccharide from enterohaemorrhagic *Escherichia coli* O48:H21. *Tetrahedron Asymm.* **2009**, *20*, 1550–1555; (d) Mukherjee, C.; Misra, A.K. Total synthesis of a unique tetrasaccharide present in the human clotting factor IX and mammalian Notch 1 receptor. *Tetrahedron Asymm.* **2009**, *20*, 473–477; (e) Mukherjee, C.; Misra, A.K. Synthesis of a unique trisaccharide having an acetal linkage between open-chain and cyclic sugar found in the cell wall of *Proteus. Tetrahedron Asymm.* **2008**, *19*, 2746–2751.