

Available online at www.sciencedirect.com



Tetrahedron

Tetrahedron 61 (2005) 2481-2492

An efficient construction of bridged chiral tetracyclic indolidines, a core structure of asperparaline, via stereocontrolled catalytic Pauson–Khand reaction

Shinji Tanimori,* Tatsuya Sunami, Kouji Fukubayashi and Mitsunori Kirihata

Department of Applied Biochemistry, Graduate School of Agriculture and Life Sciences, Osaka Prefecture University, 1-1 Gakuen-cho, Sakai, Osaka 599-8531, Japan

Received 26 November 2004; revised 22 December 2004; accepted 24 December 2004

Available online 27 January 2005

Abstract—A reaction of chiral enyne 22 derived from L-proline with a catalytic amount of cobalt (0) octacarbonyl in the presence of N-methylmorphorine N-oxide gave tricyclic enone 24 in 54% yield (73% based on consumed starting material). Treatment of enone 11 with aqueous methylamine followed by silica gel afforded bridged tetracyclic indolidine 1, a common structural motif of natural metabolites, an asperparaline series of compounds and also a potential intermediate for the synthesis of a paralytic alkaloid, asperparaline C (4), in 70% yield.

© 2005 Elsevier Ltd. All rights reserved.

1. Introduction

In our previous report,¹ we briefly communicated the racemic synthesis of tetracyclic indolidine **1**, which constitutes the C3 and C10–C25 portions of a paralytic alkaloid, asperparaline C (**4**) (Fig. 1),² via Pauson–Khand cycloaddition reaction³ mediated by a stoichiometric amount of cobalt (0) octacarbonyl. In this paper, we wish to describe full details of the work which includes additional new work such as a catalytic version of the key Pauson–Khand reaction,⁴ chiral synthesis of **1** starting from L-proline using Seebach's protocol⁵ and also improvement of the yield of each step to result in a concise, enantio- and stereo-controlled synthesis of **1**.

The indolidine alkaloids, asperparalines A to C (2 to 4), were discovered by Hayashi and co-workers from 'okara' (the insoluble residue of whole soybeans) that has been fermented with *Aspergillus japonicus* JV-23.² Asperparalines showed interesting paralytic activity within 1 h against silkworms at a dose of 10 μ g of diet which lasted for 7 to 10 h upon oral administration. On the other hand, Everett and co-workers also isolated the same compound as asperparaline A (2) named aspergillimide (VM55598) along with SB202327 (5) possessing anthelmintic activity.⁶ There

are numerous other structurally related natural methabolites including brevinamides 6^7 paraherquamides 7^8 macrofortine 8^9 and sclerotamide 9^{10} which have a variety of biological activities such as anthelmintic,⁶ antinematodal,⁸ and antiinsectant¹⁰ activities.

Asperparalines and related compounds have a common unique diazabicyclo[2.2.2]octane skeleton formed from biosynthetic intramolecular [4+2] cycloaddition.¹¹ Asperparalines and asperdillimides have also 3-*spiro*-succinimide which is rare in naturally occurring products.¹² Previously, we reported the model studies for constructing spiro-succinimide from 2,2-dimethylcyclopentanone as shown in Scheme 1.¹³

Our retro-synthetic plan for asperparaline C (4) is shown in Figure 2. Following the results of our previous model studies for the construction of spirosuccinimide from cyclopentanone (Scheme 1),¹³ tetracyclic ketone 1 could be a precursor for introducing the spirosuccinimide. The energy minimized 3D-structure of α , β -unsaturated dinitrile **10**,¹⁴ which would be derived from ketone 1 by Knoevenagel condensation based on the model study,¹³ indicated that a Michael-type nucleophilic attack such as the cyanide anion to the β -position of an unsaturated system would occur from the convex face to produce the corresponding trinitrile possessing the correct stereo-chemistry at C-3 (asperparaline numbering) in accord with asperparaline. The ketone **1** could be transformed from enone **11** by stereoselective 1,4-addition of methylamine

Keywords: Asperparaline; Pauson–Khand reaction; Indolidine alkaloid; Paralytic activity; Spiro-succinimide.

^{*} Corresponding author. Tel.: +81 72 254 9469; fax: +81 72 254 9918; e-mail: tanimori@biochem.osakafu-u.ac.jp

^{0040–4020/\$ -} see front matter @ 2005 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.12.057



Figure 1. Asperparaline and related methabolites.



Scheme 1. Model studies for the synthesis of 3-spiro-succinimide.¹³

followed by intramolecular amide formation with an ester. The 3D structure of enone **11** suggested that the amine would attack from the same face with the ester group at C-13 (asperparaline numbering) to produce the desired stereochemistry at C-11. The precursor of **11** would be enyne **12** by the transformation using Pauson–Khand [2+2+1] cycloaddition reaction. The reaction would proceed via the stable conformer **13** other than **14** to produce **11**, as a major product. Enyne **12** could be prepared starting from L-proline as a chiral form by the use of Seebach's protocol⁵ for synthesizing chiral α -substituted proline.

2. Results and discussion

We initially intended to establish the synthetic route by the use of readily available racemic material. The racemic enyne **12** was prepared from *N*-Cbz-proline by the standard method involving the sequential alkylation of ester enolate, deprotection, followed by *N*-alkylation via **15** and **18** (Scheme 2). Disappointingly, the Pauson–Khand reaction of

12 under various conditions (Table 1) gave no desired product 11. Probably, the approach of the cobalt complex formed on alkyne to alkene would be prevented due to the steric bulk of the terminal geminal dimethy group. To confirm this speculation, two enynes, allyl proline 21 and crotyl proline 22, were synthesized along the same reaction sequences. Indeed, as shown in Table 2, allyl proline 21 yielded tricyclic enone 23 in high yield by the use of a stoichiometric amount of cobalt (0) octacarbonyl in THF in the presence of DMSO as promoter¹⁵ as a single stereoisomer. At this point, the stereochemistry of the newly produced C-20 position (asperparaline numbering) of enone 23 was tentatively estimated by molecular calculation as follows. A plausible mechanism of the Pauson-Khand reaction is shown in Scheme 3.3 The stereo-discrimination step should be the stage from complex 25 to metallacycle 26. Although the best way to predict the stereochemical outcome would be to calculate the transition state energy of every possible conformer of 25, it is necessary to use an accurate molecular calculation program. We conducted the calculation with 26 instead of transition state 25 based upon the hypothesis that the steric energy of 26 would reflect that



Scheme 2. Synthesis of racemic enone 11, 23 and 24.

Table 1. Reaction condition from enyne 12 to enone 11



Entry	Co ₂ (CO) ₈ (equiv)	Solvent	Conditions for the formation of cobalt–alkyne complex	Promoter	Conditions for the reaction of cobalt–alkyne complex with alkene	Yield (%)
1	1.05	THF	Ar, rt, 2 h	DMSO (6 equiv)	Ar, 50 °C, 72 h	0
2	1.05	CH_2Cl_2	Ar, rt, 2 h	NMO (9 equiv)	Ar, rt, 96 h	0
3	1.0	THF	Ar, rt, 2 h	NMO (6 equiv)	Ar, rt, 15 h	0
4	1.05	THF	Ar, rt, 2 h	TMANO $\cdot 2H_2O$ (6 equiv)	Ar, rt, 12 h	0
5	1.0	THF	Ar, rt, 2 h	TMANO (6 equiv)	Ar, rt, 24 h	0

Table 2. Reaction conditions for the synthesis of enone 23



of 25. The possible conformer of transition state 25 would be 25a to 25h (Scheme 4) and the corresponding metallacycle 26a to 26d should afford enone ($7aR^*,9R^*$)-23; on the other hand, 26e to 26h from 25e to 25h should provide diastereomer ($7aS^*,9R^*$)-23. Table 3 shows the calculated steric energy of 26a to 26h, the difference in energy between 26b to 26h and the most stable conformer 26a, the equilibrium constants based on Boltzmann equation, the calculated distribution of 26a to 26h, and the ratio of formation on ($7aR^*,9R^*$)-23 and ($7aS^*,9R^*$)-23 based on the MM2 calculation composed in Chem3D program.¹⁴ The carbonyl group on cobalt was replaced to hydrogen to simplify the calculation. These results well reflected the experimental results.

The formation of bridged lactam was studied using enone 23 (Table 4). Thus, enone 23 was treated with methyl amine in the presence of additives followed by acidic treatment to produce lactam 27 in excellent yield as a single diastereomer. The reaction pathway was assumed as follows shown in Scheme 5. The first Michael-type addition of methylamine to enone 23 would give intermediate 28 stereoselectively (not isolated), then spontaneous intramolecular lactam formation afforded imine 29. Acidic

treatment to hydrolyze imine **29** would produce the desired lactam **27**. Especially, silica gel in aqueous methanol was extremely effective (Table 4, entry 5). The relative stereochemistry of **27** and also the precursor enone **23** became undoubtedly apparent by this transformation, because another Michael adduct **30** from stereoisomer 7a*epi*-**23** could not cyclyze to lactam as shown in Scheme 5. It was assumed that the attack of methylamine to enone **23** would occur from the convex face of the molecule, predominantly. Consequently, the chirality of the quaternary carbon of enyne **21** was effectively transformed into C-1 and C-5 of the product **27** via the Pauson–Khand reaction followed by Michael-type addition.

Unfortunately, introduction of a geminal dimethyl group on enone 23 at C7 position was ineffective to yield the desired 11 in only 3% yield along with mono methylated 24 (14%) and recovered 23 (28%) by treatment of 23 with LDA and MeI in the presence of HMPA at -78 °C (Scheme 6). We next examined the cycloaddition of enyne 22 possessing a crotyl side chain. Although the efficiency rather declined in comparison to 21, the enone 24 was obtained in 62% yield in a ratio of 4:1 based on the stereochemistry of the secondary methyl group at C7 (Table 5). Introduction of a lacked



Scheme 3. Plausible mechanism of Pauson-Khand reaction.



Scheme 4. Reaction pathway from transition state 25 to enone 23 via metallacycle 26.

Table 3. Calculated product distribution based on steric energy of metallacyc	le 2	26
---	------	----

	26a	26b	26c	26d
Steric energy (kcal/mol)	299.55	301.62	300.97	302.66
Difference of steric energy to 26a (kcal/mol)	0	2.07	1.42	3.11
Equilibrium constant	1	3.97×10^{-2}	1.09×10^{-1}	7.90×10^{-3}
Calculated distribution	85.8	3.41	9.35	6.78×10^{-1}
Total distribution of $(7aR^*,9R^*)$ -23 (%)			99	
	26e	26f	26g	26h
Steric energy (kcal/mol)	302.58	304.63	307.35	305.54
Difference of steric energy to 26a (kcal/mol)	3.03	5.08	7.80	5.99
Equilibrium constant	8.87×10^{-3}	3.67×10^{-4}	5.28×10^{-6}	8.92×10^{-5}
Calculated distribution Total distribution of	7.61×10^{-1}	3.15×10^{-2}	4.53×10^{-4}	7.65×10^{-3}
(7a <i>S</i> *,9 <i>R</i> *)- 23 (%)				

Table 4. Reaction conditions for the synthesis of lactam 27 from enone 23



Entry	Conditions	Isolated yield (%)
1	(1) 40% MeNH ₂ aq (20 equiv), Na ₂ CO ₃ (0.2 equiv), rt, 24 h (2) Concd HCl, rt, 24 h \rightarrow 40 °C, 3 h	52
2	(1) 40% MeNH ₂ aq (20 equiv), Na ₂ CO ₃ (0.05 equiv), rt, 15 h (2) Concd HCl, 45 °C, 24 h	57
3	(1) 40% MeNH ₂ aq (20 equiv), NaHCO ₃ (0.1 equiv), rt, 13 h (2) Concd HCl, rt, 7 h	60
4	(1) 40% MeNH ₂ aq (20 equiv), MeNH ₃ Cl (0.2 equiv), rt, 21 h (2) Concd HCl, 45 °C, 13 h	69
5	(1) 40% MeNH ₂ aq (20 equiv), MeNH ₃ Cl (0.2 equiv), rt, 16 h (2) SiO ₂ , MeOH–H ₂ O, rt–45 °C, 7 h	95



Scheme 5. Reaction pathway from enone 23 to lactam 27.



Table 5. Reaction conditions for the synthesis of enone 24 from enyne 22



Table 6. Reaction conditions for the synthesis of lactam 1 from enone 11



Entry	Conditions	Isolated yield (%)
1	 (1) 40% MeNH₂ aq (20 equiv), MeNH₃Cl (0.2 equiv), rt, 16 h (2) Concd HCl, 45 °C, 18 h→rt, 41 h 	28
2	(1) 40% MeNH ₂ aq (20 equiv), MeNH ₃ Cl (0.5 equiv), rt, 96 h (2) SiO ₂ , MeOH, H ₂ O, reflux, 4 h	39
3	 (1) 40% MeNH₂ aq (20 equiv), MeNH₃Cl (0.5 equiv), rt, 48 h (2) SiO₂, MeOH, H₂O, rt, 18 h→reflux, 1 h 	39
4	(1) 40% MeNH ₂ aq (40 equiv), rt, 48 h \rightarrow reflux, 4 h (2) SiO ₂ , MeOH, H ₂ O, rt, 18 h	70

methyl group on **24** was successfully accomplished to give enone **11** in 77% yield by treatment with LDA and MeI (Scheme 6). The lactam formation of **11** needed stronger conditions (Table 6) to produce racemic lactam **1** in 70% yield again as a single diastereomer.

As the synthetic route to 1 was established as racemic form, we next examined the chiral synthesis of 1 and also a catalytic approach for the key Pauson–Khand reaction.

The optically active indolidine 1 was synthesized starting from chiral crotyl proline 34, prepared from bicyclic

compound 31^5 via 32 and 33 by the stereoselective crotylation, hydrolysis, followed by esterification (Scheme 7), by the same reaction sequence as the racemic series.

The catalytic Pauson–Khand reaction⁴ was successively introduced as shown in Table 7. The reaction was undertaken under a carbon monoxide atmosphere using a catalytic amount of $Co_2(CO)_8$ and NMO as promoter to yield **24** in moderate yield. The efficiency was almost the same as that of the stoichiometric series (see Table 5).



Table 7. Catalytic Pauson-Khand reaction of enyne 22 to enone 24



Entry	Co ₂ (CO) ₈ (equiv)	Solvent	Conditions	Promoter (equiv)	Isolated yield (%)
1	0.05	CH ₂ Cl ₂	CO, 22 h, rt	NMO (0.05)	23 (76) ^a
2	0.05	Benzene	CO, 22 h, 70 °C	NMO (0.05)	54 (26) ^a
3	0.05	Toluene	CO, 22 h, 120 °C	NMO (0.05)	49 (33) ^a

^a Values in parentheses represent recovery yields of compound 22.



Scheme 8. Knoevenagel reaction of lactams.

The next stage was set for the construction of 3-spirosuccinimide relying on our earlier study (Scheme 1).¹³ Unfortunately, the Knoevenagel reaction of ketone 1 with malononitrile under the same reaction condition as the model studies and also stronger conditions such as heating and using a Lewis acid catalyst¹⁶ gave no desired product.¹ The results would be attributed to the additional steric bulk by the bridged lactam on the convex face from which the nucleophile would approach and also the neighboring geminal dimethyl group and hydrogens, one of which would occupy pseudo axial position, respectively, to carbonyl group. Energy minimized conformation of 1 suggested that the *N*-methyl group would shield the convex face of the molecule and the pseudo axial methyl group and hydrogen would also prevent the nucleophilic attack (Scheme 8). To confirm this point, the Knoevenagel reaction of 27 and 36, derived from enone 24 in 73% yield by the same procedure as the synthesis of 1 and 27, was examined. Although unsaturated dinitrile 35 was obtained in 63% yield, 36 gave no condensed product.

3. Conclusion

In summary, a concise and stereocontrolled approach for the construction of tetracyclic indolidine 1, a common structural motif for the biologically active alkaloid asperparaline series of natural products and a potential intermediate for asperparaline C (4), has been demonstrated in 8 steps in 12% overall yield starting from L-proline as a chiral process. This protocol constitutes the following three key features: (i) an efficient synthesis of chiral enyne 22 using Seebach's protocol from L-proline; (ii) a stereocontrolled catalytic Pauson–Khand cycloaddition reaction (from 22 to 24); (iii) a novel facile formation of bridged lactam (from 11 to 1). The molecular modeling calculation using a popular program (Chem3D) satisfactorily predicted the

stereochemical outcome in the key metal-mediated cycloaddition step (from 21 to 23) and this assumption was confirmed later by the chemical transformation (from 23 to 27). Further study directed toward the total synthesis of asperparaline C (4) from tetracycle 1 by another approach is currently under investigation.

In addition, biological tests for most of the synthetic compounds described in this paper were conducted. As a result, all compounds have not revealed remarkable paralysis against silkworms at a dose of 0.1 mg of diet and also agrochemical profiles on the usual random screening program such as antifungal, insecticidal, and herbicidal activity at 100 ppm. The results suggested that the whole molecular architecture of asperparaline should be essential for exhibiting biological activities.

4. Experimental

4.1. General

Infrared (IR) spectra were recorded on a Perkin–Elmer FT-IR 1760X spectrometer. NMR spectra were recorded on a JEOL JNM-GX 270 spectrometer, operating at 270 MHz for ¹H NMR and 67.5 MHz for ¹³C NMR. Chemical shifts in CDCl₃ are reported on the δ scale relative to CHCl₃ (7.26 ppm for ¹H NMR and 77.00 ppm for ¹³C NMR) as an internal reference. The following abbreviations are used to multiplicities: 's' (singlet), 'd' (doublet), 't' (triplet), 'm' (multiplet), 'br' (broad). Optical rotations were measured on a JASCO DIP-360 polarimeter. Mass spectra were measured on a JEOL JNM-AX 500 mass spectrometer. Column chromatography was carried out with silica gel Merck 60 (230–400 mesh ASTM). Reactions were carried out in dry solvents under an argon atmosphere. Tetrahydrofuran (THF) was distilled from sodium benzophenone ketyl.

Dichloromethane (CH_2Cl_2) was distilled from calcium hydride. Other reagents were purified by usual methods.

2-(3-Methyl-but-2-enyl)-pyrrolidine-1,2-di-4.1.1. carboxylic acid 1-benzyl ester 2-methyl ester (15), 2-allyl-pyrrolidine-1,2-dicarboxylic acid 1-benzyl ester 2-methyl ester (16), and 2-but-2-enyl-pyrrolidine-1,2dicarboxylic acid 1-benzyl ester 2-methyl ester (17). 15% *n*-BuLi in hexane (18.8 mL, 30.4 mmol) was added slowly to a solution of diisopropylamine (3.08 g, 4.27 mL, 30.4 mmol) in anhydrous THF (100 mL) at -78 °C under nitrogen atmosphere and stirred for 30 min at 0 °C. After cooling to -78 °C, N-Cbz-proline methyl ester (5.0 g, 19.0 mmol) in anhydrous THF (25 mL) was added over 20 min. After stirring for 1 h, HMPA (6.81 g, 6.61 mL, 38.0 mmol) was added dropwise over 5 min and stirred for 10 min. Crotyl bromide (3.13 mL, 30.4 mmol) in anhydrous THF (25 mL) was added over 1 h and warmed to 0 °C over 30 min and stirred for 1 h at 0 °C. 1 N HCl solution (50 mL) was added to the reaction mixture and the organic layer was separated. The water layer was extracted with ethyl acetate (50 mL) and the combined organic layer was washed with satd aq NaHCO₃ (30 mL) and brine (30 mL) and dried over MgSO₄. Concentration in vacuo gave a crude product, which was purified by silica gel column chromatography (eluting with EtOAc-hexane = 1:9) to give crotyl proline methyl ester 17 (4.88 g, 81%) as a pale yellow oil. $R_f = 0.67$ (EtOAc-hexane = 1:2, I_2). IR (NaCl, film) ν_{max} cm⁻¹: 2954, 2881, 1742 (ester C=O), 1704 (carbamate C=O), 1408, 1357, 1169, 1120, 1025, 699. ¹H NMR δ (CDCl₃): 1.64 (3H, d, J=6.4 Hz, CH=CH-CH₃), 1.77-2.15 (4H, m), 2.48-3.07 (2H, m, CH₂-CH=CH), 3.47 (2H, s, O-CH₂-Ph), 3.70 (3H, s, O–CH₃), 5.11 (1H, dd, J=12.2, 47.3 Hz), 5.14 (1H, br s), 5.23-5.38 (1H, m), 5.38-5.58 (1H, m), 7.32-7.37 (5H, m, Ph). ¹³C NMR δ (CDCl₃): 18.2, 18.7, 23.3, 36.6, 48.3, 52.1, 67.0, 68.2, 125.0, 127.5, 127.5, 127.7, 127.8, 128.0, 130.3, 136.9, 154.0, 174.6. FAB MS m/z (%):318 (MH⁺ 19), 274 ([M-CO₂]⁺, 9), 182 ([M-Cbz]⁺, 16). HRMS (FAB) m/z (MH⁺): calcd for C₁₈H₂₄NO₄, 318.1705; found, 318.1694.

Compound **15**. Pale yellow oil. $R_f = 0.61$ (EtOAc-hexane = 1:2, I₂). IR (NaCl, film) ν_{max} cm⁻¹:2955, 2880, 1742 (ester C=O), 1705 (carbamate C=O), 1409, 1355, 1127, 1028, 699. ¹H NMR δ (CDCl₃): 1.58 (3H, s), 1.70 (3H, s), 1.74-2.08 (4H, m), 2.58-3.05 (2H, m, CH₂-CH=C(CH₃)₂), 3.46 (2H, s, O-CH₂-Ph), 3.71 (3H, s, O-CH₃), 4.97-5.24 (3H, m), 7.26-7.36 (5H, m, Ph). ¹³C NMR δ (CDCl₃) *cis + trans* rotamers: 18.1, 18.1, 22.8, 23.3, 26.1, 26.2, 31.9, 33.2, 35.7, 37.1, 48.4, 49.2, 52.1, 52.4, 66.6, 67.0, 68.0, 68.8, 118.3, 118.5, 127.5, 127.7, 127.8, 128.0, 128.3, 128.3, 135.1, 135.3, 136.3, 136.9, 154.0, 154.2, 174.7, 174.8. FAB MS m/z (%): 332 (MH⁺, 15), 288 ([M-CO₂]⁺, 10), 196 ([M-Cbz]⁺, 8). HRMS (FAB) m/z (MH⁺): calcd for C₁₉H₂₆NO₄, 332.1862; found, 332.1850.

Compound **16**.¹⁸ Pale yellow oil. $R_f = 0.59$ (EtOAchexane = 1:2, I₂).IR (NaCl, film) ν_{max} cm⁻¹: 2954, 2880, 1742 (ester C=O), 1704 (carbamate C=O), 1408, 1357, 1170, 1119, 1026, 699. ¹H NMR δ (CDCl₃): 1.82–2.16 (4H, m), 2.56–2.72 (1H, m, *H*–CH–CH=CH₂), 2.88–3.17 (1H, m, *H*–CH–CH=CH₂), 3.47 (2H, s, O–CH₂–Ph), 3.71 (3H, s, O–CH₃), 5.00–5.23 (4H, m), 5.62–5.80 (1H, m, CH=CH₂), 7.27–7.37 (5H, m, Ph). ¹³C NMR δ (CDCl₃): 18.0, 28.5, 35.3, 43.0, 50.7, 66.1, 69.0, 117.3, 127.3, 127.6, 128.1, 128.5, 128.6, 140.6, 140.9, 159.3, 174.1.

4.1.2. 2-(3-Methyl-but-2-enyl)-1-prop-2-ynyl-pyrrolidine-2-carboxylic acid methyl ester (12), 2-allyl-1-prop-2-ynyl-pyrrolidine-2-carboxylic acid methyl ester (21), 2-but-2-enyl-1-prop-2-ynyl-pyrrolidine-2-carand boxylic acid methyl ester (22). Under nitrogen atmosphere, sodium iodide (7.36 g, 49.1 mmol) was added to a solution of N-Cbz-crotyl proline 17 (2.0 g, 6.3 mmol) in anhydrous MeCN (20 mL) and cooled to 0 °C. TMSCI (3.20 mL, 25.2 mmol) was added to the mixture and the mixture was stirred for 6 h at room temperature. After cooling to 0 °C, 1 N HCl (20 mL) was added to the mixture. After stirring for 30 min, the phases were separaterd and the water layer was washed with hexane (20 mL \times 2). The water layer was converted to pH 9-10 by adding 50% K₂CO₃ and extracted with methylene chloride (20 mL \times 3). The organic phase was washed with brine, dried over MgSO₄, and concentrated in vacuo to give free amine 20.

A mixture of the above amine 20, propargyl bromide (0.71 mL, 9.4 mmol), sodium bicarbonate (1.59 g, 18.9 mmol), LiI (84.3 mg, 0.63 mmol) in MeCN (20 mL) was heated at 60 °C with stirring for 14 h. After cooling, water (20 mL) was added to the mixture and the two layers were separated. The water layer was extracted with ether $(20 \text{ mL} \times 2)$ and the combined organic phase was washed with brine, dried (MgSO₄) and evaporated. The residue was purified by silica gel column chromatography (EtOAchexane = 1:19) to give enyne 22 (1.12 g, 80%) as a colorless oil. $R_f = 0.72$ (EtOAc-hexane = 1:2, I_2). IR (NaCl, film) $\nu_{\text{max}} \text{ cm}^{-1}$: 3300 (C=C-H), 2950, 2856, 1728 (C=O), 1435, 1195, 970, 651. ¹H NMR δ (CDCl₃): 1.65 (3H, d, J =7.1 Hz, CH=CH-CH₃), 1.72-1.90 (3H, m), 2.16-2.26 (3H, m), 2.60 (1H, dd, J=7.0, 13.7 Hz, H-CH-CH=CH-CH₃), 2.72–2.84 (1H, m), 3.16–3.22 (1H, m), 3.34 (1H, dd, J=2.4, 16.8 Hz, *H*–CH–C \equiv CH), 3.61 (1H, dd, *J*=2.4, 16.8 Hz, *H*-CH-C≡CH), 3.68 (3H, s, O-CH₃), 5.30–5.41 (1H, m, CH=CH-CH₃), 5.46–5.59 (1H, m, CH=CH-CH₃). ¹³C NMR δ (CDCl₃): 18.2, 21.4, 33.9, 37.7, 38.2, 51.2, 51.3, $69.7, 71.3 \ (C \equiv CH), 80.9 \ (C \equiv CH), 125.8 \ (CH = CH - CH_3),$ 128.7 (CH=CH-CH₃), 173.7. FAB MS m/z (%): 222 $(MH^+, 13), 190 ([M-OMe]^+, 56), 162 ([M-CO_2Me]^+, 56))$ 100). HRMS (FAB) m/z (MH⁺): calcd for C₁₃H₂₀NO₂, 222.1494; found, 222.1499.

Compound **12**. $R_f=0.79$ (EtOAc-hexane = 1:2, I₂). IR (NaCl, film) ν_{max} cm⁻¹: 3298 (C=*C*-*H*), 2951, 1727 (C=O), 1435, 1195, 1175, 647. ¹H NMR δ (CDCl₃): 1.63 (3H, s), 1.70 (3H, s), 1.58–1.86 (3H, m), 2.19–2.24 (3H, m), 2.65 (1H, dd, *J*=7.6, 14.3 Hz, *H*-CH-CH=C(CH₃)₂), 2.74–2.83 (1H, m, H-C5), 3.17–3.24 (1H, m, H-C5), 3.34 (1H, dd, *J*=2.4, 16.8 Hz, *H*-CH-C=CH), 3.67 (3H, s, O-CH₃), 3.60–3.68 (1H, m), 5.02–5.08 (1H, m, *CH*=C(CH₃)₂). ¹³C NMR δ (CDCl₃):18.1, 21.4, 26.1, 33.6, 34.0, 37.8, 51.2, 51.3, 70.0 (C2), 71.3 (C=*C*H), 81.0 (*C*=CH), 119.0 (*C*H=C(CH₃)₂), 134.2 (CH=*C*(CH₃)₂), 173.7 (C=O). FAB MS *m*/*z* (%): 236 (MH⁺, 31), 176 ([M - CO₂Me]⁺, 38), 166 ([M - CH₂CH=C(CH₃)₂]⁺, 100). HRMS (FAB) *m*/*z* (MH⁺): calcd for C₁₄H₂₂NO₂, 236.1651; found, 236.1628. Compound **21**. $R_f=0.72$ (EtOAc-hexane=1:2, I₂). IR (NaCl, film) ν_{max} cm⁻¹: 3299, 2952, 2837, 1727 (C=O), 1641 (C=C), 1433, 1197, 1171, 989, 918, 649. ¹H NMR δ (CDCl₃): 1.81–1.88 (3H, m), 2.18–2.23 (2H, m), 2.32 (1H, dd, J=7.1, 14.3 Hz, H–CH–CH=CH₂), 2.65 (1H, dd, J= 7.3, 14.6 Hz, H–CH–CH=CH₂), 2.77–2.85 (1H, m), 3.17– 3.23 (1H, m), 3.35 (1H, dd, J=7.6, 15.6 Hz, H–CH– C≡CH), 3.61 (1H, dd, J=7.4, 15.4 Hz, H–CH–C≡CH), 3.68 (3H, s, O–CH₃), 5.06–5.13 (2H, m, CH=CH₂), 5.69– 5.85 (1H, m, CH=CH₂). ¹³C NMR δ (CDCl₃): 21.4, 33.9, 37.6, 39.3, 51.2, 51.3, 69.2, 71.3 (C≡CH), 80.8 (C≡CH), 118.0 (CH=CH₂), 133.5 (CH=CH₂), 173.6.

4.1.3. 6-Oxo-2,3,6,7,7a,8-hexahydro-1*H*,4*H*-3a-aza-sindacene-8a-carboxylic acid methyl ester (23) and 7-methyl-6-oxo-2,3,6,7,7a,8-hexahydro-1*H*,4*H*-3a-aza-sindacene-8a-carboxylic acid methyl ester (24).

4.1.3.1. Synthesis of 23 by a stoichiometric Pauson-Khand reaction (Table 2, entry 3). To a stirred solution of Co₂(CO)₈ (0.31 g, 0.91 mmol) in dry THF (9 mL) under Ar at room temperature was added dropwise a solution of envne **21** (0.19 g, 0.91 mmol) in THF (1 mL). After 2 h of stirring at room temperature, DMSO (0.39 mL, 5.46 mmol) was added in one portion. The reaction mixture was stirred for 26 h at 50 °C. After cooling, the mixture was filtered through Celite, which was thoroughly washed with acetone. The solvent was eliminated under reduced pressure, and the crude product was purified by silica gel column chromatography (eluting with hexane: EtOAc = 1:1) to give enone **23** (201 mg, 94%) as a pale yellow crystal. $R_{\rm f} = 0.25$ (EtOAc). Mp 80.2–81.1 °C. $[\alpha]_{D}^{18}$ + 34.1° (*c* 1.9, CHCl₃). IR (NaCl, film) ν_{max} cm⁻¹:2954, 1713 (C=O), 1630 (C=C), 1445, 1198, 1152. ¹H NMR δ (CDCl₃): 1.37 (1H, t, J =12.5 Hz, H-C8), 1.73-2.19 (5H, m), 2.57-2.66 (2H, m, H₂-C7), 2.70–2.77 (1H, m, H–C7a), 2.91 (1H, dd, J=7.3, 16.5 Hz, H-C3), 3.09-3.17 (1H, m, H-C3), 3.79 (3H, s, O-CH₃), 3.79–3.94 (2H, m, H₂–C4), 5.98 (1H, br s, H–C5). ¹³C NMR δ (CDCl₃): 21.0, 36.5, 38.1, 38.7, 42.0, 47.8, 49.9, 52.1, 67.3 (C8a), 128.4 (C5), 174.5 (C4a), 176.4 (CO₂Me), 207.7 (C6). FAB MS m/z (%): 236 (MH⁺, 69), 176 $([M - CO_2Me]^+, 100)$. HRMS (EI) m/z (M⁺): calcd for C₁₃H₁₇O₃N, 235.1209; found, 235.1220.

4.1.3.2. Synthesis of 24 by a catalytic Pauson-Khand reaction (Table 7, entry 2). To a stirred solution of $Co_2(CO)_8$ (37 mg, 0.11 mmol) in dry benzene (50 mL) under CO atmosphere at room temperature was added dropwise a solution of enyne 22 (500 mg, 2.26 mmol) in benzene (5 mL). After 2 h of stirring at room temperature, NMO (50% in water, 0.028 mL, 0.11 mmol) was added in one portion. The reaction mixture was stirred for 22 h at 70 °C. After cooling, the mixture was filtered through Celite, which was thoroughly washed with acetone. The solvent was eliminated under reduced pressure, and the crude product was purified by silica gel column chromatography (eluting with hexane–EtOAc = 1:1) to give enone 24 (304 mg, 54%) as a brown oil and recovered envne 22 (130 mg, 26%). $R_{\rm f}$ =0.30 (EtOAc, I₂). $[\alpha]_{\rm D}^{22}$ +44.9°(c 1.0, CHCl₃). IR (NaCl, film) $\nu_{\rm max}$ cm⁻¹: 2957, 2878, 1728 (ester C=O), 1708 (ketone C=O), 1632 (C=C), 1449, 1176. ¹H NMR δ (CDCl₃): 1.07 (3H, d, J=7.3 Hz, CH₃-C7), 1.36 (1H, t, J=12.9 Hz, H-8), 1.73–1.85 (1H, m, H–C1), 1.85– 1.95 (2H, m, H_2 –C2), 1.99 (1H, dq, J=2.9, 7.3 Hz, H–C7), 2.16 (1H, dt, J=12.2, 7.1 Hz, H–C1), 2.27–2.36 (1H, m, H–C7a), 2.67 (1H, dd, J=5.6, 12.9 Hz, H–C8), 2.93 (1H, dd, J=8.5, 15.3 Hz, H–C3), 3.12 (1H, dd, J=8.5, 15.3 Hz, H–C3), 3.68–3.92 (2H, m, H₂–C4), 3.79 (3H, s, O–CH₃), 5.95 (1H, br s, H–C5); ¹³C NMR δ (CDCl₃): 14.2, 21.0, 36.5, 37.8, 46.4, 47.8, 47.9, 50.0, 52.1, 67.3 (C8a), 127.2 (C5), 174.0 (C4a), 174.5 (CO₂Me), 209.9 (C6). EI-MS *m*/*z* (%): 249 (M⁺, 39), 190 ([M–CO₂Me]⁺, 32), 154 (100), 136 (83). HRMS (EI) *m*/*z* (M⁺): calcd for C₁₄H₁₉NO₃, 249.1365; found, 249.1382.

4.1.4. 7,7-Dimethyl-6-oxo-2,3,6,7,7a,8-hexahydro-1H, 4H-3a-aza-s-indacene-8a-carboxylic acid methyl ester (11). 15% n-BuLi in hexane (2.61 mL, 4.21 mmol) was added slowly to a solution of diisopropylamine (0.43 g, 0.59 mL, 4.21 mmol) in anhydrous THF (15 mL) at -78 °C under nitrogen atmosphere and stirred for 30 min at 0 °C. After cooling to -78 °C, enone **24** (350 mg, 1.40 mmol) in anhydrous THF (1.5 mL) was added to the above solution over 5 min. After stirring for 5 min, HMPA (0.50 g, 0.49 mL, 2.81 mmol) was added dropwise and stirred for 10 min. MeI (0.80 g, 0.35 mL, 5.61 mmol) was added and stirred for 1 h at -78 °C. Saturated aqueous NaCl (15 mL) was added and the organic layer was separated. The water layer was extracted with ethyl acetate (30 mL) and the combined organic layer was washed with brine (30 mL) and dried over MgSO₄. Concentration in vacuo gave a residue, which was purified by silica gel column chromatography (hexane-EtOAc = 3:2) to give enone 11 (285 mg, 77%) as a pale yellow oil. $R_{\rm f}$ =0.30 (EtOAc, I₂). $[\alpha]_{\rm D}^{22}$ + 18.1° (*c* 0.65, CHCl₃); IR (NaCl, film) $\nu_{\rm max}$ cm⁻¹: 2965, 2869, 1729 (ester C=O), 1708 (ketone C=O), 1633 (C=C), 1449, 1191, 1152. ¹H NMR δ (CDCl₃): 0.92 (3H, s, CH₃–C6), 1.03 (3H, s, CH₃-C6), 1.26-1.38 (1H, m, H-C8), 1.71-1.88 (3H, m), 2.04–2.13 (1H, m), 2.27–2.39 (2H, m), 2.92 (1H, dd, J=7.6, 15.6 Hz, H–C12), 3.12 (1H, dd, J=7.5, 15.4 Hz, H–C12), 3.72 (3H, s, O-CH₃), 3.79-3.87 (2H, m, H₂-C2), 5.83 (1H, br s, H–C4). ¹³C NMR δ (CDCl₃): 20.4, 21.0, 25.2, 33.8, 36.7, 46.3, 48.0, 49.7, 50.0, 52.1, 67.2, (C8a), 125.3 (C5), 174.0 (C4a), 174.5 (CO₂Me), 209.9 (C6). EI-MS m/z (%): 263 (M⁺, 100), 204 ([M-CO₂Me]⁺, 82), 154 (62), 136 (43). HRMS (EI) m/z (M⁺): calcd for C₁₅H₂₁NO₃, 263.1521; found, 263.1544.

4.1.5. 11,13-Diaza-4,4,13-trimethyltetracyclo- $[5.5.2.0^{1,5}.0^{7,11}]$ tetradecane-3,14-dione (1), 11,13-diaza-13-methyltetracyclo $[5.5.2.0^{1,5}.0^{7,11}]$ tetradecane-3,14dione (27), and 11,13-diaza-4,13-dimethyltetracyclo- $[5.5.2.0^{1,5}.0^{7,11}]$ tetradecane-3,4-dione (36). A solution of enone 11 (0.2 g, 0.76 mmol) in 40% aqueous methylamine (2.62 mL, 30.4 mmol) was stirred for 48 h at room temperature. Water (10 mL) was added to the mixture and the solution was heated at reflux for 4 h. After cooling, the mixture was concentrated in vacuo, the residue was dissolved in MeOH (6 mL) and water (1.2 mL), and SiO₂ (0.8 g) was added. The mixture was stirred for 18 h at room temperature. After cooling, the mixture was filterd and the filtrate was washed with methanol, and the solvent was concentrated in vacuo. The residue was purified by silica gel column chromatography (eluting with EtOAc-acetone = 1:1) to give lactam 1 (140 mg, 70%) as a brown oil. $R_{\rm f}$ =0.7 (acetone, I₂). $[\alpha]_{\rm D}^{22}$ -43.6°(c 0.96, CHCl₃). IR (NaCl, film) $\nu_{\rm max}$ cm⁻¹: 2991, 2887, 1766

(ketone C=O), 1660, (lactam C=O), 1650, 1388, 1095. ¹H NMR δ (CDCl₃): 0.97 (3H, s, CH₃–C4), 1.07 (3H, s, CH₃–C4), 1.38–1.49 (1H, m, H–C6), 1.65–1.74 (1H, m, H–C6), 1.83–1.99 (2H, m), 2.15–2.44 (4H, m), 2.47 (1H, dd, *J*=1.5, 2.0 Hz), 2.53–2.67 (2H, m), 2.99 (3H, s, CH₃–N13), 3.05–3.13 (1H, m, H–C10), 3.24 (1H, d, *J*=11.3 Hz, H–C12). ¹³C NMR δ (CDCl₃): 17.7, 20.9, 22.2, 23.1, 27.9, 29.8, 36.5, 43.9, 46.2, 52.2, 54.0, 56.0, 61.2, 173.0 (C14), 217.6 (C3). EI-MS *m*/*z* (%): 262 (M⁺, 1), 247 (1), 234 (1), 219 (8), 203 (80), 190 (6), 176 (7), 165 (8), 138 (17), 133 (33), 120 (13), 96 (27), 83 (12), 68 (15), 55 (25), 41 (100). HRMS (EI) *m*/*z* (M⁺): calcd for C₁₅H₂₂N₂O₂, 262.1681; found, 262.1679.

Compound **27**. $R_{\rm f}$ =0.43 (acetone). $[\alpha]_{\rm D}^{20}$ -149.6° (*c* 1.1, CHCl₃). IR (KBr, disk) $\nu_{\rm max}$ cm⁻¹:3440 (br), 2923, 1750 (ketone C=O), 1666 (lactam C=O), 1651, 1390, 1097. ¹H NMR δ (CDCl₃): 1.35–1.46 (1H, m, H–C6), 1.69 (1H, dd, J=5.5, 12.5 Hz, H–C6), 1.82–1.95 (2H, m), 2.16–2.36 (5H, m), 2.48–2.68 (4H, m), 2.98 (3H, s, CH₃–N13), 3.06–3.13 (1H, m, H–C10), 3.18–3.22 (1H, d, J=11.6 Hz, H–C12). ¹³C NMR δ (CDCl₃): 22.2, 26.9, 27.8, 34.3, 40.9, 42.6, 45.5, 53.7, 53.8, 62.8, 66.0 (C7), 172.8 (C14), 211.4 (C3). EI MS *m*/*z* (%): 234 (M⁺, 5), 206 (3), 175 (100), 149 (51), 137 (21), 108 (12), 96 (20). HRMS (EI) *m*/*z* (M⁺): calcd for C₁₃H₁₈N₂O₂, 234.1368; found, 234.1349.

Compound **36**. R_f =0.49 (acetone, I₂). Mp 116–118.2 °C. IR (KBr, disk) ν_{max} cm⁻¹: 2980, 2875, 2798, 1746 (ketone C=O), 1662 (lactam C=O), 1651, 1390, 1324, 1098. ¹H NMR δ (CDCl₃): 1.15 (3H, d, *J*=7.0 Hz, CH₃–C4), 1.36–1.48 (1H, m, H–C6), 1.64–1.73 (1H, m, H–C6), 1.84–1.96 (3H, m), 2.18–2.39 (4H, m), 2.51–2.64 (3H, m), 2.98 (3H, s, CH₃–N13), 3.05–3.13 (1H, m, H–C10), 3.24 (1H, d, *J*=11.3 Hz, H–C12). ¹³C NMR δ (CDCl₃): 12.8, 22.1, 26.6, 27.6, 33.4, 44.1, 47.9, 48.3, 53.7, 54.7, 60.9, 65.8, 172.6 (C14), 213.4 (C3). EI-MS *m*/*z* (%): 249 (MH⁺, 56), 189 (37), 154 (100), 136 (89). HRMS (EI) *m*/*z* (M⁺): calcd for C₁₄H₂₀N₂O₂, 248.1525; found, 248.1521.

4.1.6. 7a-But-2-enyl-3-tert-butyl-tetrahydro-pyrrolo-[1,2-c]oxazol-1-one (32). 15% *n*-BuLi in hexane (28.71 mL, 46.4 mmol) was added slowly to a solution of diisopropylamine (6.52 mL, 46.4 mmol) in anhydrous THF (100 mL) at -78 °C under nitrogen atmosphere and stirred for 30 min at 0 °C. After cooling to -78 °C, oxazolidone **31** (5.00 g, 27.3 mmol) in anhydrous THF (25 mL) was added over 20 min. After stirring for 1 h, HMPA (9.49 mL, 54.6 mmol) was added dropwise over 5 min and stirred for 10 min. Crotyl bromide (4.77 mL, 46.4 mmol) in anhydrous THF (25 mL) was added over 1 h and stirred for 1 h at -78 °C. The mixture was warmed to -30 °C over 1 h and saturated aqueous NaHCO₃ (50 mL) was added. The organic layer was separated. The water layer was extracted with ethyl acetate (50 mL) and the combined organic layer was washed with satd aq NaHCO₃ (50 mL) and brine (50 mL) and dried over MgSO₄. Concentration in vacuo gave a residue, which was purified by distillation under reduced pressure to give crotyl Oxazolidinone 32 (3.78 g, 64%) as a pale yellow oil. $R_f = 0.60$ (EtOAc-C₆H₆=1:200). Bp₃: 85–90 °C. [α]_D²² + 13.5°(*c* 1.0, CHCl₃). IR (NaCl, film) ν_{max} cm⁻¹: 2963, 2871, 1780 (C=O), 1633, 1191. ¹H NMR δ (CDCl₃): 0.92 (9H, s), 1.64 (3H, d, *J*= 7.1 Hz, CH=CH-CH₃), 1.73-1.90 (4H, m), 2.35 (2H, d,

J=7.4 Hz, *CH*₂–CH=CH), 2.74–2.99 (2H, m), 4.24 (1H, s), 5.46–5.58 (2H, m). ¹³C NMR δ (CDCl₃): 17.3, 21.1, 21.2, 21.3, 28.8, 34.3, 36.8, 49.5, 107.1, 124.8, 133.1, 174.5 (C=O). FAB MS *m*/*z* (%): 238 (MH⁺, 4), 180 ([M–*t*-Bu]⁺,100), 135 (89). HRMS (FAB) *m*/*z* (MH⁺): calcd for C₁₄H₂₄NO₂, 238.1807; found, 238.1834.

4.1.7. 2-But-2-enyl-pyrrolidine-2-carboxylic acid (33). A mixture of crotyl oxazolidone 32 (3.0 g, 13.4 mmol) and silica gel (SiO₂, 3.0 g) in MeOH/H₂O (6:1, 30 mL) was stirred for 48 h at room temperature. After filtration, the residue was washed with MeOH and the filtrate was concentrated in vacuo. The residue was dissolved in CHCl₃/MeOH (20:1, 30 mL) and again filtered. The residue was washed with CHCl₃/MeOH (20:1) and the filtrate was concentrated in vacuo. The residue was washed with ether and dried over P₂O₅ under reduced pressure to give carboxylic acid 33 (2.0 g, 88%) as a colorless crystal. $R_{\rm f} = 0.23$ (MeOH–CHCl₃=1:4, ninhydrin).Mp 280–285 °C. $[\alpha]_{\rm D}^{20}$ -70.4°(c 1.0, CH₃OH). IR (KBr, disk) $\nu_{\rm max}$ cm⁻¹: 3085, 1628 (C=O), 1390, 933. ¹H NMR δ (D₂O): 1.64 (3H, d, J = 6.4 Hz, CH=CH-CH₃), 1.73-1.90 (4H, m), 2.24 (1H, dd, J=7.1, 14.6 Hz, H-CH-CH=CH), 2.69 (1H, dd, J= 7.3, 14.6 Hz, H-CH-CH=CH), 2.88-3.03 (2H, m), 5.46-5.58 (2H, m). ¹³C NMR δ (D₂O): 17.3, 22.6, 31.3, 37.8, 42.5, 77.1, 123.8, 133.2, 179.5 (CO₂H). FAB MS m/z (%):170 (MH⁺, 100), 124 ($[M - CO_2H]^+$, 100). HRMS (FAB) m/z (MH⁺): calcd for C₉H₁₆NO₂, 170.1181; found, 170.1100.

4.1.8. (R)-2-(-)-Crotylproline methyl ester hydrochloride (34). Thionyl chloride (4.31 mL, 59.1 mmol) was added dropwise to a solution of α -crotyl proline **33** (2.00 g, 11.82 mmol) in anhydrous MeOH (50 mL) at 0 °C and the mixture was stirred for 48 h at room temperature. After refluxing for 1 h, the mixture was concentrated in vacuo. The residue was crystallized in MeOH/EtOAc followed by drying over P₂O₅ under reduced pressure to give methyl ester 34 (2.34 g, 90%) as a colorless crystal. $R_{\rm f}=0.74$ (MeOH–CHCl₃=1:4, ninhydrin). Mp 166.5–170.0 °C. $[\alpha]_{D}^{22}$ -48.7°(c 1.0, CH₃OH). IR (KBr, disk) ν_{max} cm⁻¹: 2880, 2711, 2490, 1759 (C=O), 1644 (C=C), 1588, 1341, 1220, 1154, 946. ¹H NMR δ (CD₃OD): 1.73 (3H, d, J= 6.4 Hz, CH=CH-CH₃), 1.88-2.50 (4H, m), 2.71 (1H, dd, J=7.1, 14.6 Hz, H-CH-CH=CH), 2.93 (1H, dd, J=7.1, 14.6 Hz, H-CH-CH=CH), 3.40-3.48 (2H, m), 3.89, (3H, s, O-CH₃), 5.55-5.78 (2H, m). ¹³C NMR δ (CD₃OD): 17.6, 20.8, 31.3, 37.8, 42.2, 50.7, 67.7, 123.1, 133.0, 174.5.

4.1.9. 11,13-Diaza-3-(dicyanomethylene)-13-methyltetracyclo-[5.5.2.0^{1,5}.0^{7,11}]tetradecan-14-one (35). The lactam **27** (90.3 mg, 0.385 mmol) and malononitrile (51 mg, 0.77 mmol) were added to benzene (10 mL) containing piperidine (7.99 mg, 0.039 mmol) and benzoic acid (18.8 mg, 0.154 mmol), and the mixture was heated to reflux with azeotropic removal of water by Dean–Stark trap for 28 h. After cooling, the mixture was diluted with diethyl ether (20 mL), and successively washed with water (10 mL), a 10% NaHCO₃ solution (10 mL) and brine (10 mL). The organic phase was dried (MgSO₄) and concentrated in vacuo. The residue was purified by flash chromatography over silica gel (eluting with EtOAc) to give unsaturated dinitrile **35** (68.5 mg, 63%) as an pale yellow crystal. $R_{\rm f}$ =0.70 (acetone, I₂). Mp 151.6–153.0 °C. IR (KBr, disk) $\nu_{\rm max}$ cm⁻¹:3437 (br), 2946, 2806, 2234 (C=N), 1661 (amide C=O), 1620 (C=C), 1386, 1320, 1103. ¹H NMR δ (CDCl₃): 1.34–1.46 (1H, m, H–C6), 1.68 (1H, dd, J=6.3, 13.2 Hz, H–C6), 1.80-1.92 (2H, m), 2.13–2.39 (5H, m), 2.48–2.72 (4H, m), 2.96 (3H, s, CH₃–N13), 3.01–3.15 (1H, m, H–C10), 3.21 (1H, dd, J=7.8, 20.0 Hz, H–C12). ¹³C NMR δ (CDCl₃): 22.3, 27.0, 27.7, 33.2, 37.9, 40.3, 43.0, 53.6, 53.7, 64.7, 66.0, 84.9 (C(CN)₂), 110.5 (CN), 110.9 (CN), 172.5 (C14), 184.4 (C3). FAB MS m/z (%): 283 (MH⁺, 100), 223 (53). HRMS (FAB) m/z (MH⁺): calcd for C₁₆H₁₉N₄O, 283.1559; found, 283.1548.

Acknowledgements

We thank Professor H. Hayashi of Graduate School of Agriculture and Life Sciences at Osaka Prefecture University for his kind cooperation with this project and helpful advice.

References and notes

- Tanimori, S.; Fukubayashi, K.; Kirihata, M. *Tetrahedron Lett.* 2001, 42, 4013–4016.
- Hayashi, H.; Nishimoto, Y.; Nozaki, H. *Tetrahedron Lett.* 1997, 38, 5655–5658. Hayashi, H.; Nishimoto, Y.; Akiyama, K.; Nozaki, H. *Biosci. Biotech. Biochem.* 2000, 64, 111–115.
- A review for Pauson–Khand reaction, see: Shore, N. E. Org. React. 1991, 40, 1–90. Shore, N. E. In Trost, B. M., Fleming, I., Pattanden, G., Eds.; Comprehensive Organic Synthesis; Pergamon: Oxford, 1991; Vol. 5, pp 1037–1064. Ching, Y. K. Coord. Chem. Rev. 1999, 188, 297–341. Brummond, K. M.; Kent, J. L. Tetrahedron 2000, 56, 3263–3283.
- 4. Recent review for catalytic Pauson–Khand reaction, see: Gibson, S. E.; Stevenazzi, A. *Angew. Chem., Int. Ed.* **2003**, *42*, 1800–1810.

- Seebach, D.; Boes, M.; Naef, R.; Schweizer, W. B. J. Am. Chem. Soc. 1983, 5390–5398.
- Banks, R. M.; Blanchflower, S. E.; Everett, J. R.; Manger, B. R.; Reading, C. J. Antibiot. 1997, 50, 840–846.
- 7. Paterson, R. R. M.; Hawksworth, D. L. *Trans. Br. Mycol. Soc.* **1985**, *85*, 95–100 and references cited therein.
- Blanchflower, S. E.; Banks, R. M.; Everett, J. R.; Reading, C. J. Antibiot. 1993, 46, 1355–1363 and references cited therein.
- (a) Polonsky, J.; Merrien, M. A.; Prangee, T.; Pascard, C.; Moreau, S. J. Chem. Soc., Chem. Commun. 1980, 601–602. (b) Prange, T.; Bullion, M.-A.; Vuilhorgne, M.; Pascard, C.; Polosky, J. Tetrahedron Lett. 1980, 22, 1977–1980.
- 10. Whyte, A. C.; Gloer, J. B. J. Nat. Prod. 1996, 59, 1093-1095.
- 11. Williams, R. M. *Chem. Pharm. Bull.* **2002**, *50*, 711–740 and references cited therein.
- Biosynthetic studies of the spirosuccinimide ring system, see: Gray, C. R.; Sanz-Cervera, J. F.; Silks, L. A.; Williams, R. M. J. Am. Chem. Soc. 2003, 125, 14692–14693.
- Tanimori, S.; Fukubayashi, K.; Kirihata, M. Biosci. Biotechnol. Biochem. 2000, 64, 1758–1760. Earlier studies on spirosuccinimide ring system, see: Gonzalez, F.; Sanz--Cervera, J. F.; Williams, R. M. Tetrahedron Lett. 1999, 40, 4519–4522.
- 14. All calculations were carried out using the MM2 in CS Chem3D version 5.0, Cambridge Soft Corporation.
- Effect of promoters on Pauson–Khand reaction, see: Chung, Y. K.; Lee, B. Y.; Jeong, N.; Hudecek, M.; Pauson, P. L. Organomettalics 1993, 12, 220–223.
- For Lewis acid-catalyzed Knoevenagel reaction, see: Mori, K.; Hara, T.; Mizugaki, T.; Ebitani, K.; Kaneda, K. J. Am. Chem. Soc. 2003, 125, 11460–11461.
- 17. Only a small amount of decomposed ring-opening and/or retro-Michael products was obtained.
- (a) Dumas, J.-P.; Germanas, J. P. *Tetrahedron Lett.* **1994**, *35*, 1493–1496.
 (b) Kim, K.; Dumas, J.-P.; Germanas, J. P. *J. Org. Chem.* **1996**, *61*, 3138–3144.
 (c) Pfeifer, M. E.; Linden, A.; Robinson, J. A. *Helv. Chim. Acta* **1997**, *80*, 1513–1527.