

DEVELOPMENT OF A NEW N^π -PROTECTING GROUP FOR HISTIDINE, N^π -1-ADAMANTYLOXYMETHYLHISTIDINE

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N^π -1-Adamantyloxymethylhistidine, His(N^π -1-Adom) was prepared, and the properties of the 1-Adom group were examined. 1-Adom group can be easily removed by TFA; it is stable to 20% piperidine/DMF and 1N NaOH. His(N^π -1-Adom) derivatives can suppress racemization during coupling reaction. TRH was synthesized using His(N^π -1-Adom), successfully.

KEY WORDS histidine; protecting group; 1-adamantyloxymethyl; racemization

Various kinds of protecting groups for imidazole nitrogen of histidine residue have been developed in peptide synthesis. It is well known that protecting groups on π -nitrogen of imidazole function are more successful than those on τ -nitrogen for the prevention of racemization during peptide synthesis. Previously, N^π -benzyloxymethylhistidine, His(N^π -Bom), was developed.³⁾ Bom group is stable to CF_3COOH (TFA) and cleaved by hydrogenation over Pd catalyst or HF.⁴⁾ Therefore, His(N^π -Bom) can be applied for peptide synthesis by Boc strategy both in solution and in solid phase methods. N^π -*tert*-Butyloxymethylhistidine, His(N^π -Bum), was also developed in order to suppress racemization.⁵⁾ Bum group can be removed by TFA and is stable under alkaline conditions. Therefore, His(N^π -Bum) is applied for peptide synthesis by Fmoc (9-fluorenylmethyloxycarbonyl) strategy in solid phase method. However, it was reported that Fmoc-His(N^π -Bum)-OH had poor solubility in dichloromethane.⁶⁾ From these points of view, our studies were directed to the development of novel N^π -protecting groups with the objectives of suppressing side reactions, preventing racemization and increasing the solubility of His-containing peptide intermediates in organic solvents. Previously, it was reported that 1-adamantyl (Ada) ester could be removed by TFA and was stable to 20% piperidine/DMF⁷⁾ and that adamantyl ester derivatives exhibited high solubility in organic solvents.⁸⁾ These results gave us the idea of designing a new N^π -protecting group.

This paper deals with the synthesis of His(N^π -1-Adom) (**1**) (Fig. 1), its properties and its application to the synthesis of thyrotropin-releasing hormone (TRH). According to Chart 1, 1-adamantyloxymethyl chloride (1-Adom-Cl),⁹⁾ which was prepared from 1-adamantyloxymethyl methyl sulfide and sulfuryl chloride, is completely involatile and is easier to be purified than Bum-Cl. Z-His-OMe, is acetylated with acetic anhydride to give Z-His(N^τ -Ac)-OMe¹⁰⁾ (**2**), which was reacted with 1-Adom-Cl, followed by treatment with NaHCO_3 to afford Z-His(N^π -1-Adom)-OMe in

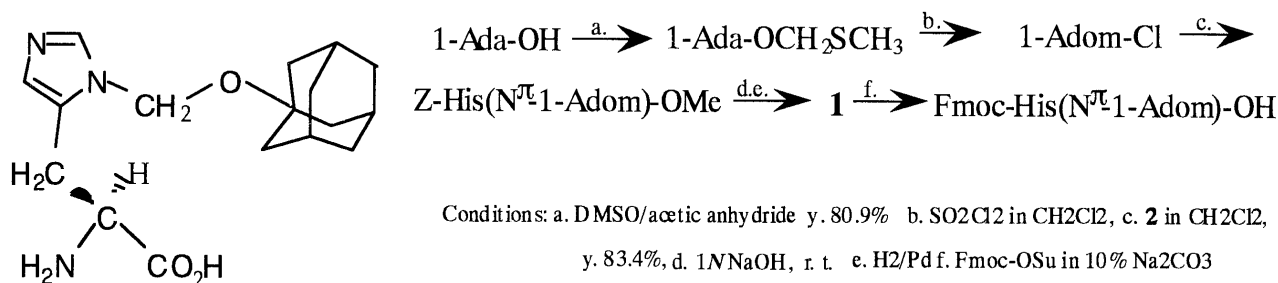


Fig. 1. Structure of **1**

Chart 1. Synthesis of **1** and Its Derivatives

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high yield compared with Bum derivative. Z-His(N π -1-Adom)-OMe was saponified with 1N NaOH, followed by hydrogenation over Pd catalyst to give **1** (Fig. 1).¹¹⁾ The stability and susceptibility of the 1-Adom group to various acids and bases were examined, and the results are summarized in Table 1. The 1-Adom group is easily cleaved by TFA and stable to 20% piperidine/DMF at room temperature for up to 48 h. Therefore, His(N π -1-Adom) can be used for peptide synthesis in combination with Fmoc as N α -protecting group. Fmoc-His(N π -1-Adom)-OH¹²⁾ was prepared from His(N π -1-Adom) and Fmoc-OSu in a good yield, and it is more soluble in organic solvents than Fmoc-His(N π -Bum)-OH.⁵⁾

Table 1. Properties of **1** to Acids and Bases

Conditions	%Histidine regenerated						
	15min	30min	45min	60min	12h	24h	48h
TFA (200eq, 5eq anisole)	73.9	88.1	100	100			
0.1N HCl (300eq)	0	0	0	0	0	0	0
1N NaOH (100eq)	0	0	0	0	0	0	0
20% Piperidine/DMF (200eq)	0	0	0	0	0	0	0

Next, the efficiency of N π -1-Adom group in the prevention of side-chain induced racemization was examined. Z-D-His(N π -1-Adom)-OH was prepared by the same method as described above. Z-D-His(N π -1-Adom)-L-Phe-OMe was well separated from Z-L-His(N π -1-Adom)-L-Phe-OMe on HPLC.¹³⁾ Therefore, this sequence was employed for model study on racemization. Z-L-His(N π -1-Adom)-OH was coupled with H-L-Phe-OMe by N,N'-dicyclohexylcarbodiimide (DCC), DCC/1-hydroxybenzotriazole (HOBt), benzotriazol-1-yloxytris(dimethylamino) phosphonium hexafluorophosphate (BOP),¹⁴⁾ 2-(1H-benzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU)¹⁵⁾ or diphenylphosphoryl azide (DPPA),¹⁶⁾ and then the crude product was analyzed by HPLC. The results summarized in Table 2 show that the formation of D-L dipeptide was particularly low in all coupling methods so far examined.

Table 2. Racemization Rate During the Coupling of Z-His(N π -1-Adom)-OH and H-Phe-OMe

Coupling method	D-L/(D-L+L-L) %
DCC	2.74
DCC/HOBt	0.55
BOP	0.60
HBTU	0.71
DPPA	0.50

Finally, Z-His(N π -1-Adom)-OH was employed to synthesize the thyrotropin-releasing hormone (TRH). Z-His(N π -1-Adom)-OH was coupled with H-Pro-NH₂ by BOP reagent to afford Z-His(N π -1-Adom)-Pro-NH₂. After removal of Z group by catalytic hydrogenation, the resultant amine was coupled with *t*-butyloxycarbonylpyroglutamic acid (Boc-Pyr-OH) by BOP reagent to give Boc-Pyr-His(N π -1-Adom)-Pro-NH₂. The protected tripeptide was purified by silica gel column chromatography, and the purified tripeptide was treated with TFA/thioanisole at room temperature for 1 h. After washings with ether and transformation to acetate form by treating with Amberlite IRA-93ZU resin, TRH¹⁷⁾ thus obtained exhibited a single peak on analytical HPLC at the same retention time as that of authentic TRH.¹⁸⁾

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REFERENCES AND NOTES

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- 3) Brown T., Jones J. H., Richards, J.D., *J. Chem. Soc., Perkin Trans. 1*, **1982**, 1553-1561.
- 4) Sakakibara S., Shimonishi Y., Kishida Y., Okada M., Sugihara U., *Bull. Chem. Soc. Japan* **40**, 2164-2167 (1967).
- 5) Colombo R., Colombo F., Jones J. H., *J. Chem. Soc., Chem. Commun.* **1984**, 292-293.
- 6) Fields G. B., Noble R. L., *Int. J. Peptide Protein Res.*, **35**, 161-214 (1990).
- 7) Okada Y., Iguchi S., *J. Chem. Soc. Perkin Trans. 1*, **1988**, 2129-2136.
- 8) Nishiyama Y., Shintomi N., Kondo Y., Okada Y., *J. Chem. Soc. Perkin Trans. 1*, **1994**, 3201-3207.
- 9) Jones J.H., Thomas D.W., Thomas R.W., Wood M.E., *Synth. Commun.* **16**, 1607-1610, (1986)
- 10) Jones J. H., Rathbone D.L., Wyatt. P.B., *Synthesis*, **1987** 1110-1113.
- 11) m.p.228-229°C (dec.) $[\alpha]_D^{25} = -8.3^\circ$ ($c = 0.5$ in MeOH) (*Anal.* Calcd for $C_{17}H_{25}N_3O_3$: C, 63.9; H, 7.89; N, 13.2. Found: C, 63.8; H, 7.88; N, 13.2.), 1H -NMR spectra was measured with a Bruker ARS 500 spectrometer operating at a frequency of 500 MHz. 1H -NMR($CDCl_3$) δ =1.64 –1.87 (12H, m, adamantyl), 2.19 (3H, s, adamantyl), 3.09-3.45 (2H, m, CH_2CH), 3.85-3.87 (1H, m, $CH-CH_2$), 5.46 (2H, s, NCH_2O), 6.93 (1H, s, 5^{im} -H), 7.68 (1H, s, 2^{im} -H).
- 12) m.p. 160-161°C. $[\alpha]_D^{25} = +5.4^\circ$ ($c = 0.5$ in MeOH)
- 13) HPLC condition: Cosmosil pack 5C 18-AR (4.6x250mm); eluent: A (0.05% TFA in H_2O), B (0.05% TFA in CH_3CN), from 69/31 to 55/45 in 50 min, and to 69/31 in 5 min; flow rate: 1 ml/min.
- 14) Castro B., Dormoy J. R., Evin G., Selve C., *Tetrahedron Lett.*, **14**, 1219-1222 (1975).
- 15) Knorr R., Trzeciak A., Bannwarth W., Gillessen D., *Tetrahedron Lett.*, **30**, 1927-1930 (1989).
- 16) Shioiri T., Ninomiya K, Yamada S., *J. Am. Chem. Soc.*, **94**, 6203-6205 (1972).
- 17) $[\alpha]_D^{25} = -62.2^\circ$ ($c = 1.0$ in H_2O)
- 18) Authentic TRH was purchased from Peptide Institute(Osaka, Japan) $[\alpha]_D^{25} = -61.3^\circ$ ($c = 1.0$ in H_2O)

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