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## Efficient Synthesis of D-[5-<sup>13</sup>C]Ribose from D-Ribose and Its Conversion into [5'-<sup>13</sup>C]Nucleosides

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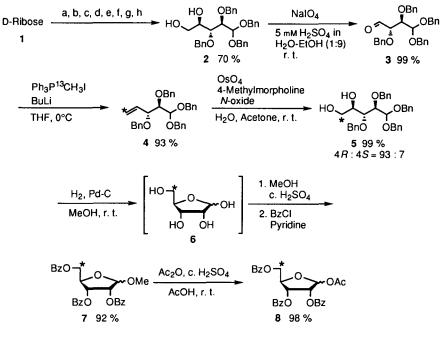
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Abstract: An approach to the synthesis of  $[5^{-13}C]$ ribose (6) was achieved by a sequence reaction as follows: periodate oxidation of 2,3-di-O-benzyl-D-ribose dibenzyl acetal (2), which was derived from D-ribose (1) in 70% overall yield in 8 steps, followed by the introduction of <sup>13</sup>C into the 5-position of D-ribose by Wittig reaction using Ph<sub>3</sub>Pl<sup>3</sup>CH<sub>3</sub>I - BuLi, highly diastereoselective hydroxylation with OsO<sub>4</sub>, and then debenzylation. Compound 6 with the necessary protecting groups and a leaving group was derived into [5<sup>-13</sup>C]ribonucleoside derivatives by the coupling reaction with persilylated nucleobases. Copyright © 1996 Elsevier Science Ltd

The conformational diversity of the sugar-phosphate backbone and/or the sugar moieties in nucleic acid is considered to be important in the elucidation of nucleic acid - protein or - drug recognition processes. The utility of <sup>13</sup>C labeled RNAs and DNAs for the heteronuclear multidimensional NMR studies for nucleic acids has been shown in several reports.<sup>1-8</sup> Based on this background, the title nucleosides should be significantly useful for the conformational analysis of the sugar-phosphate backbone structure in nucleic acids. Incidentally, the chemical syntheses of [5'-<sup>13</sup>C]adenosine and D-[5-<sup>13</sup>C]ribose have been reported by Matwiyoff *et al.*<sup>9</sup> and by Serianni *et al.*<sup>10</sup> Matwiyoff *et al.* prepared [5'-<sup>13</sup>C]adenosine<sup>9</sup> by the coupling reaction of an adenine derivative with a [5-<sup>13</sup>C]ribose derivative which was synthesized by way of L-[1-<sup>13</sup>C]ribonic acid obtained by the reaction of L-erythrose with [<sup>13</sup>C]KCN, followed by hydrolysis under acidic conditions. Serianni *et al.*, on the other hand, synthesized D-[5-<sup>13</sup>C]ribose from D-[6-<sup>13</sup>C]glucose which was obtained by the reaction of 1,2-*O*-isopropylidene- $\alpha$ -*D-xylo*-pentidialdo-1,4-furanoside with [<sup>13</sup>C]KCN.<sup>10</sup> These synthetic methods, however, inevitably involved multiple steps after the introduction of <sup>13</sup>C label, to attain the final product and, were therefore judged impractical. Thus, more efficient processes were sought to make the nucleosides more easily available.

Therefore, the present synthetic study on D-[5-<sup>13</sup>C]ribose (6) from D-ribose (1) was undertaken to overcome the drawback described above, and the results thus obtained will be described herein. Chemical conversion of D-ribose (1) into D-[5-<sup>13</sup>C]ribose (6) was efficiently performed by the sequence of reactions shown in Scheme 1. Compound 1 was successfully derived into 2,3-di-*O*-benzyl-D-ribose dibenzyl acetal (2) in 70% overall yield in 8 steps. After periodate oxidation of 2, the resulting 4-*aldehydo*-2,3-di-*O*-benzyl-D-erythrose dibenzyl acetal (3) (99% yield) was then subjected to Wittig reaction<sup>11</sup> with Ph<sub>3</sub>P<sup>13</sup>CH<sub>3</sub>I (99 atom % <sup>13</sup>C) - BuLi in THF at room temperature, which is the first key step in introducing <sup>13</sup>C labeling as the 5-position of a pentose, to give 2,3-di-*O*-benzyl-4,5-didehydro-4,5-dideoxy-D-[5-<sup>13</sup>C]ribose dibenzyl acetal (4) in 93% yield.<sup>12</sup> The second key step is to perform a diastereoselective hydroxylation of **4** with OsO4;<sup>13</sup>

the treatment of 4 with OsO<sub>4</sub> (0.05 molar equiv.) in the presence of 4-methylmorpholine *N*-oxide as a cooxidizing agent gave a 93:7 mixture of 2,3-di-*O*-benzyl-D-[5-<sup>13</sup>C]ribose dibenzyl acetal (5) and the corresponding L-lyxo derivative in 99% yield. Such a high diastereoselectivity (4R/4S = 93/7) was confirmed by 400 MHz <sup>1</sup>H NMR spectroscopy after 4,5-di-*O*-benzoylation. The stereochemistry to give the excellent diastereoselectivity might be explained in terms of the concept described by Kishi *et al.*<sup>13</sup> Subsequent debenzylation of 5 in methanol by the catalytic hydrogenation on palladium charcoal gave D-[5-<sup>13</sup>C]ribose (6), which was then converted into 1-*O*-Ac-2,3,5-tri-*O*-benzoyl-D-[5-<sup>13</sup>C]ribose (8) *via* methyl 2,3,5-tri-*O*benzoyl-D-[5-<sup>13</sup>C]ribofuranoside (7) by the usual method. The efficiency of the present approach from 1 to 8 is characterized by the 62% overall yield of 8 from Ph<sub>3</sub>Pl<sup>3</sup>CH<sub>3</sub>I, *i.e.*, after introducing the <sup>13</sup>C labeling by Witting reaction, in contrast with the 4% overall yield of 8 from [<sup>13</sup>C]KCN reported by Matwiyoff *et al.*<sup>9</sup> and 25% overall yield of 6 from [<sup>13</sup>C]KCN reported by Serianni *et al.*<sup>10</sup> after introducing the <sup>13</sup>C label.



a) MeOH, c. H<sub>2</sub>SO<sub>4</sub>, 0°C; b) DMTrCl, Pyridine, r. t,; c) BnBr, NaH, DMF, r. t.;

d) 3% CCI<sub>3</sub>COOH in CHCI<sub>3</sub>, r. t.; e) EtSH, c. HCI, 0°C;<sup>14</sup> f) Ac<sub>2</sub>O, DMAP, CH<sub>2</sub>CI<sub>2</sub>, r. t.;

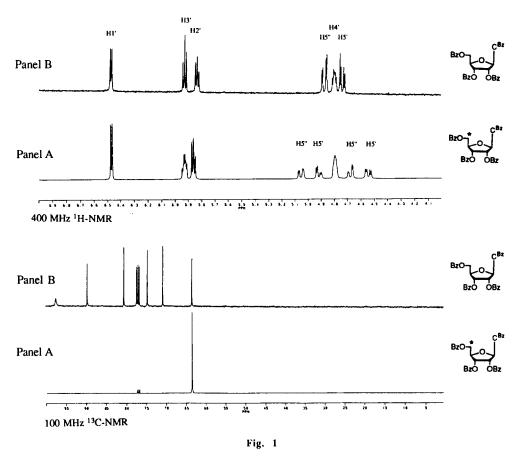
g) HgO, HgCl<sub>2</sub>, CaSO<sub>4</sub>, BnOH, 70°C;<sup>15</sup> h) 2.0 M NaOH aq., MeOH, r. t

## Scheme 1

Compound 8 was finally subjected to a coupling reaction<sup>16</sup> with a persilylated nucleobase (persilylated  $N^6$ -benzoyladenine,<sup>17</sup>  $N^2$ -acetyl- $O^6$ -diphenylcarbamoylguanine,<sup>17</sup>  $N^4$ -benzoylcytosine,<sup>18</sup>

thymine, <sup>19</sup> and uracil<sup>18</sup>) to give  $[5'-^{13}C]$ ribonucleoside derivatives in good yields (80%, 79%, 96%, and 78%, respectively).

The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of  $N^4$ ,  $O^2$ ,  $O^3$ ,  $O^5$ -tetrabenzoyl[5'-<sup>13</sup>C]cytidine (Panel A) and  $N^4$ ,  $O^2$ ,  $O^3$ ,  $O^5$ -tetrabenzoylcytidine (panel B) for comparison are illustrated in Fig. 1.



An approach to the synthesis of  $[5'-1^3C]$ ribonucleosides was thus achieved by the coupling reaction of persilylated nucleobases with D- $[5-1^3C]$ ribose derivative (9). The products were derived into the corresponding 2'-deoxy $[5'-1^3C]$ ribonucleoside derivatives by the established method.<sup>20</sup>

## **References and Notes**

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- 12. 2,3-di-O-benzyl-4,5-didehydro-4,5-dideoxy-D-[5-<sup>13</sup>C]ribose dibenzylacetal(4) <sup>1</sup>H-NMR (CDCl<sub>3</sub>) : δ 7.2-7.4(20H, m, Ph-H),
  5.96(1H, ddd, J<sub>3,4</sub> = 8.0 Hz, J<sub>4,5</sub> = 17.4 Hz, J<sub>4,5</sub> = 10.4 Hz, H4), 5.29(1H, ddd, J<sub>4,5</sub> = 10.4 Hz, J<sub>5,5</sub> = 1.9 Hz, J<sub>5,C5</sub> = 158.4 Hz, H5'), 5.17(1H, ddd, J<sub>4,5</sub> = 17.4 Hz, J<sub>5,5</sub> = 1.9 Hz, J<sub>5,C5</sub> = 155.2 Hz, H5), 4.87(1H, d, J = 11.5 Hz, Bn-CH), 4.78(1H, d, J = 11.5 Hz, Bn-CH), 4.70(1H, d, J<sub>1,2</sub> = 6.4 Hz, H1), 4.58-4.68(4H, m, Bn-CH X 4), 4.49(1H, d, J = 11.7 Hz, Bn-CH),
  4.32(1H, d, J = 12.0 Hz, Bn-CH), 4.10(1H, dd, J<sub>2,3</sub> = 3.3 Hz, J<sub>3,4</sub> = 8.0 Hz, J<sub>3,C5</sub> = 4.3 Hz, H3), 3.98(1H, dd, J<sub>1,2</sub> = 6.4 Hz, J<sub>2,3</sub> = 3.3 Hz, H2). <sup>13</sup>C-NMR (CDCl<sub>3</sub>) : δ 138.80, 138.68, 138.21, 137.72, 134.99(d, J<sub>4,5</sub> = 68.8 Hz), 128.36, 128.32, 128.26, 12, 127.93, 127.83, 1.66, 127.54, 127.38, 127.34, 119.30, 101.61, 81.69, 81.24, 74.41, 70.16, 69.51, 69.03. Anal. Calcd. for C<sub>32</sub><sup>13</sup>CH<sub>36</sub>O<sub>6</sub>.0.1H<sub>2</sub>O: C, 79.85 H, 6.95. Found: C, 79.75 H, 7.03.
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