Bioactive Surfactants containing a β-Lactam Group: Synthesis and Properties

Laurence Molina,^a Angelo Perani,^b Maria-Rosa Infante,^c Maria-Angeles Manresa,^d Michel Maugras,^b Samuel Achilefu,^a Marie-José Stebe^a and Claude Selve^{*a}

^a Université Henri Poincaré-Nancy I-Laboratoire d-Etudes des Systèmes Organiques et Colloïdaux INCM-CNRS FU0008-Lesoc URA CNRS 406-BP 239-54506-Vandoeuvre les Nancy Cedex, France

^b Groupe de Recherches sur les Intéractions Moléculaires aux Interfaces, Université Henri Poincaré-Nancy I-Bâtiment Inserm, CO 10-Plateau de Brabois 54511, Vandoeuvre les Nancy Cedex, France

^c Centro de Investigacion y Desarrollo (CSIC), Departamento de Tensioactivos c/Jorge Girona 18-26,

^d Laboratori de Microbiologia, Facultat de Farmacia, Universitat de Barcelona, 08028-Barcelona, Spain

e Mallinckrodt Medical, Inc., 675 McDonnell Boulevard, PO Box 5840, St. Louis, MO 63134, USA

Selective activation of 3-hydroxy-2-hydroxymethyl-2-methyl propanamide with $P(NMe_2)_3$ -CCl₄, and subsequent intramolecular cyclisation yields β -lactam derivatives that have both surface and antibiotic activity.

Little is known about the correlation between the surfactant properties of β -lactams and their biological activity. Here we report the synthesis and properties of a new class of β -lactam that are both bio- and surface-active materials (i.e. bioactive surfactants). In previous work,1 we described the synthesis and properties of fatty amide derivatives of 3-hydroxy-2-hydroxymethyl-2-methyl propanoic acid (A, Scheme 1). In this study, we planned to activate only one of the hydroxy functions with P(NMe₂)₃-CCl₄ couple. The resulting alkoxy tris(dimethylamino)phosphonium salt (ATDP salt B, Scheme 1)^{2,3} was obtained in good yield. A standard procedure for β-lactam synthesis involves the intramolecular reaction, under alkaline condition, of a primary or secondary amide with a nucleofugebearing γ -carbon.⁴ Applying this cyclisation procedure, **B** was transformed into a β -lactam derivative by the use of anhydrous potassium carbonate either in acetone or dioxane, as shown below. No neopentyl rearrangement was observed.3

β-Lactam derivatives (type I) were obtained in good yield (50–55%). We also applied this procedure to the synthesis of mimics of nocardicine A, a natural β-lactam with antibiotic activity.^{5–7} Thus, reaction of phenylalanine (an α-amino acid)³ with a long-chain alcohol or amine gave the fatty ester or amide derivatives respectively. Reaction of these with 3-hydroxy-2-hydroxymethyl-2-methyl propanoic acid afforded the amidodiols, A, which were subsequently cyclised to type II β-lactams in good yields (55–65%). Both types of β-lactams have a hydrophobic long alkyl chain and a hydrophilic β-lactam ring, a condition necessary for surface activity.

Although all the compounds are only slightly soluble (*ca*. 10^{-5} mol dm⁻³) in polar media such as water and formamide,⁸ they exhibit good surfactant properties in these solvents at room temperature. Saturated aqueous solutions of both types of compounds reduce the surface tension (γ_{sat}) of water from 72



Type I β -Lactams: R = C₈H₁₇ 1, C₁₀H₂₁ 2, C₁₂H₂₅ 3, C₁₄H₂₉ 4, C₆F₁₃CH₂CH₂ 5, C₈F₁₇CH₂CH₂ 6

Type II β -Lactams: R = R¹-Z-C(0)-CHCH₂Ph: Z = O, R¹ = C₈H₁₇7; Z = NH, R¹ = C₈H₁₇8; Z = O, R¹ = C₁₀H₂₁9; Z = O, R¹ = C₁₂H₂₅10; Z = O, R¹ = PhCH₂11

Scheme 1. Activation of amido-diols of 3-hydroxy-2-hydroxymethyl-2-methylpropanoic acid A by the formation of ATDP monosalt derivatives B and β -lactam cyclisation

 $mN m^{-1}$ to about 35 mN m⁻¹. The solubility of 1 and 5 in water was sufficient to obtain a γ vs. log c (concentration) plot that was used to determine their critical micelle concentrations (CMCs). At room temperature, 1 and 5 have CMCs of 0.04 mmol dm⁻³ ($\gamma_{CMC} = 2\hat{8}$ mN m⁻¹) and 0.09 mmol dm⁻³ (γ_{CMC} = 16 mN m⁻¹) respectively. These CMCs are slightly lower than those of the corresponding derivatives of A that we reported in a previous paper.¹ The solubility of the compounds 1–11 in formamide, though higher than that in water, is only moderate (about 1 mmol dm⁻³). However, the surface tensions of the saturated solutions in formamide, (γ_{sat}) about 33 mN m⁻¹, are comparable to those obtained in water. The compounds of the type I compounds, and two homologues (7, 8)of the type II lactams are relatively more soluble in formamide than 9, 10 and 11. Since the amphiphilic surfactants with higher hydrophobic character are usually more soluble in formamide than in water, is expected that their CMCs are generally higher in the former.^{8,9} Examination of the γ vs. log c plot for solutions of 7 in formamide gave a CMC of 178 mmol dm⁻³ and γ_{CMC} of 29 mN m⁻¹. Note that this compound does not form micelles in aqueous medium due to its low solubility $(10^{-6} \text{ mol dm}^{-3})$.

Hydrolysis of synthetic β -lactams in aqueous medium is a potential problem. Consequently, we examined the possible hydrolysis of the type II compounds that have both β -lactam ring and an ester function (7, 9, 10 and 11). A standard experiment was carried out by adding 7 to a mixture of $(CD_3)_2$ SO-water (3:2) and both the ester and ring carbonyls were monitored by ¹³C NMR spectroscopy. The carbonyl groups remained unchanged after 48 hours at room temperature. On the basis of the apparent stability of these compounds to rapid hydrolysis, we evaluated some of their biological properties. In haemocytes, saturated solutions of both type I and II lactams did not cause any haemolysis, except for $\overline{7}$ which produced more than 80% haemolysis. The acute toxicity of 1 was evaluated by intravenous injection in the caudal vein of five 20 g Swiss mice. Injection of 0.2 cm³ of a 60 g dm⁻³ solution of 1 in physiological serum did not kill any of the Swiss mice. We, therefore, established that the mean lethal dose (LD_{50}) of 1 is greater than 60 mg kg⁻¹ (0.27 mmol kg⁻¹). Further, we examined the aggressiveness of some of our compounds on hybridoma cells HF2 \times 653. These cells are not naturally apoptotic¹⁰ but the addition of β -lactam surfactants may alter the cell physiology. Consequently, we studied the viability and possible apoptotic or necrotic effects^{10,11} of 1, 5, 7, 8, 10 and 11 on hybridoma cells incubated for one hour at different concentrations. The following trends were observed: (i) neither apoptosis nor necrosis was observed with 10. This compound is completely biocompatible at the concentrations studied. (ii) For 1, 5 and 11, a decrease in the number of viable cells with increase in the concentration of surfactants was observed. It appears that apoptosis was the major cause of death because the number of necrotic cells was negligible. (iii) At high concentrations of 7 and 8, a high proportion of necrotic cells was observed while the number of apoptotic cells was small.

⁰⁸⁰³⁴⁻Barcelona, Spain

The antibiotic activity on basis to the Minimum Inhibitory Concentration (MIC)¹² of some of the compounds was tested on different strains of gram-positive and -negative bacteria. All the compounds examined had antibiotic activity, though less than commercial monolactams used in chemotherapy.¹³ The compounds **7**, **8**, **10** and **11** were required in concentrations > 32 mg dm⁻³ in order to inhibit the growth of a range of bacteria. We note that compound **7** has a higher activity than the compounds **8**, **10** and **11**.

In summary, we have established a simple and efficient procedure for the synthesis of surfactant molecules containing a β -lactam ring, and showed that their surface properties are similar to those of known hydrophobic surfactants with higher hydrophobic character.¹⁴ We also demonstrated that most of the compounds tested are biocompatible and that **1** has an LD₅₀ > 60 mg kg⁻¹ (intravenous). The search for surfactants with excellent antibiotic activity is now in progress. Those that form micelles in aqueous solutions could be used as adjuvants in the delivery of lipophilic drugs, where complementary antibiotic activity is needed.

This work was generously supported by CNRS and CSIC 'Cooperation project 1940'.

Received, 23rd March 1995; Com. 5/01861G

References

1 C. Selve, C. Delestre, S. Achilefu, M. Maugras and F. Attioui, J. Chem. Soc., Chem., Commun., 1991, 863.

J. CHEM. SOC., CHEM. COMMUN., 1995

- 2 B. Castro and C. Selve, Bull. Soc. Chim. Fr., 1971, 6, 2296; B. Castro, Organic Reaction, ed. W. Dauben, Wiley, 1983, vol. 29, pp. 1–162.
- 3 L. Molina, D. Papadopoulos and C. Selve, New J. Chem., in the press.
- 4 N. S. Isaacs, Chem. Soc. Rev., 1976, 5, 181.
- 5 R. B. Sykes, C. M. Cimarusti, D. P. Bonner, K. Busch, D. M. Floyd, N. H. Georgopapadakou and J. S. Well, *Nature*, 1981, **291**, 489.
- 6 M. Hashimoto, T-A. Kamori and T. Kamiya, J. Am. Chem. Soc., 1976, 98, 3023; T. Kamiya, M. Hashimoto, O. Nakaguchi and T. Oku, Tetrahedron, 1979, 35, 323.
- 7 I. Kawamoto and M. Miyauchi, Antibiotics I—β-lactams and other antimicrobial agents, Gordon and Breach Sc. Publ., Tokyo (Japan), 1992, vol. 2/2, p. 121.
- 8 I. Rico and A. Lattes, New J. Chem., 1984, 8, 429, J. Colloid Interf. Sci., 1984, 102, 285.
- 9 I. Rico, N. Hajjaji-Shriri, B. Escoula, T. N. Decastro Dantas and A. Lattes, *New J. Chem.*, 1986, **10**, 25; I. Rico, H. Duplaa, N. Hajjaji-Shriri, T. N. Decastro Dantas, C. Cecutti and A. Lattes, *J. Commun. Esp. Deterg.*, 1986, **17**, 535.
- 10 G. B. Corcoran, L. Fix, D. P. Jones, M. T. Moslen, P. Nicotera, F. A. Oberhammer and R. Buttyan, *Toxicol. Appl. Pharmacol.*, 1994, 128, 169.
- 11 A. Perani, L. Molina, G. Stacey, R. Infante, C. Selve and M. Maugras, *FEBS Lett.*, submitted for publication.
- 12 D. F. Sahm and J. A. Washington, Antibacterial susceptibility test: Dilution methods, Manual of Clinical Microbiology, 5th edn., ed. A. Balows, W. J., Hausler, K. L., Herrman and H. J. Shadomy, Am. Soc. Microbiol., Washington DC, 1991, p. 1105.
- 13 I. Kawamoto, M. Miyauchi, Antibiotics I—β-lactams and other antimicrobials agents, Gordon and Breach Sc. Publ., 1992, vol. 2/2, pp. 123 and 124.
- 14 B. A. Bergenstähl and P. Stenius, J. Phys. Chem., 1987, 91, 5944.