

## SHORT COMMUNICATIONS

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**Uridine diphosphate  $\beta$ -glucuronic acid. A new substrate for  $\beta$ -glucuronidase**

The glucosiduronic acids which are isolated from urine are always of the  $\beta$ -configuration with reference to the oside linkage. Yet the accepted glucuronyl donor, uridine diphosphate glucuronic acid (UDPGlcUA) is of the opposite,  $\alpha$ -configuration. It is tacitly, therefore, assumed that glucuronyltransferase produces inversion of the oside linkage as the glucuronic acid combines with aglycone acceptor<sup>1</sup>. The latter mechanism has never been clarified.

One need not postulate that the same enzyme was catalyzing both an inversion and a group transfer at the same time if the UDPGlcUA was of the  $\beta$ -oside configuration to begin with. Such a compound, uridine diphosphate  $\beta$ -glucuronic acid ( $\beta$ -UDPGlcUA), has so far not been isolated from natural sources but has been synthesized<sup>2</sup> and was found in a preliminary experiment to be unsatisfactory as a glucuronyl donor using a microsomal suspension and *p*-nitrophenol as the acceptor.

On the other hand,  $\beta$ -UDPGlcUA could be a substrate for  $\beta$ -glucuronidase and would then serve as a glucuronyl donor in a transfer reaction.

In order to examine these possibilities, it was first necessary to prepare  $\beta$ -UDPGlcUA of high purity, to characterize it, to devise conditions for separating it from uridine diphosphate  $\alpha$ -glucuronic acid ( $\alpha$ -UDPGlcUA) and to carry out hydrolytic reactions with  $\beta$ -glucuronidase. This paper describes the results of these experiments.

**Materials.** Starting materials for the synthesis of  $\beta$ -UDPGlcUA included 1,3-dicyclohexyl guanidinium uridine 5'-phosphoramidate (graciously supplied by Takeda Chemical Industries, Osaka, Japan) and the triethyl ammonium salt of  $\beta$ -glucuronic acid 1-phosphate ( $\beta$ -GlcUA-1-*P*) prepared in this laboratory,  $\alpha$ -UDPGlcUA, UDP, UMP used as standards were obtained from Sigma Chemical Co., U.S.A. Silica gel (F<sub>254</sub>, 10 cm  $\times$  20 cm) plates used for thin-layer chromatography were supplied by E. Merck, Germany. Other reagents were obtained from commercial sources and were of reagent grade.

Mouse urinary  $\beta$ -glucuronidase purified according to PETTENGILL AND FISHMAN<sup>4</sup> of specific activity 50000, was the enzyme source.

**Synthesis of  $\beta$ -UDPGlcUA.** The method of HONJO *et al.*<sup>2</sup>, was employed to synthesize the  $\beta$ -UDPGlcUA with modifications. Thus, 1 g of the cyclohexylamine salt of  $\beta$ -GlcUA-1-*P*<sup>5,6</sup> was converted to its triethylammonium salt and 1,3-dicyclohexyl guanidinium uridine 5'-phosphoramidate (1.6 g) in pyridine (200 ml) was added to it. The pyridine mixture was incubated at 37° for 8 days with occasional stirring. Product formation was checked periodically on thin-layer chromatographic plates with an aliquot of the reaction mixture by developing them in the solvent 0.5 M ammonium acetate buffer (pH 3.8)–ethanol (2:5, v/v) with  $\alpha$ -UDPGlcUA as standard. The spots

Abbreviations:  $\beta$ -GlcUA-1-*P*,  $\beta$ -glucuronic acid 1-phosphate;  $\alpha$ -UDPGlcUA, uridine diphosphate  $\alpha$ -glucuronic acid;  $\beta$ -UDPGlcUA, uridine diphosphate  $\beta$ -glucuronic acid.

were identified under ultraviolet light and developed with a conc.  $\text{H}_2\text{SO}_4$ -ethanol (1:1, v/v) spray with heating. To enhance the purity of  $\beta$ -UDPGlcUA the product was passed through a second charcoal column and the fractions of  $\beta$ -UDPGlcUA were pooled, evaporated to dryness at  $20^\circ$ , redissolved in small amounts of water, and analysed.

*Observations.* The  $R_F$  values obtained in descending paper chromatography and thin-layer chromatographic plates are presented in Table I which shows that there is a good separation of the uridine phosphates and 1,3-dicyclohexyl guanidinium uridine 5'-phosphoramidate which are the probable impurities of the synthesized  $\beta$ -UDPGlcUA. However, thin-layer chromatography and not paper chromatography was able to separate  $\alpha$ -UDPGlcUA and  $\beta$ -UDPGlcUA.

TABLE I

$R_F$  VALUES FOR  $\beta$ -UDPGlcUA,  $\alpha$ -UDPGlcUA AND THEIR PRECURSORS

Solvent for paper chromatography: ammonium acetate-ethanol (3:7.5, v/v)<sup>6</sup>, 8 h. Solvent for thin-layer chromatography: 0.5 M ammonium acetate buffer (pH 3.8)-ethanol (2:5, v/v).

Compound	$R_F$ values	
	Paper chromatography	Thin-layer chromatography
UMP	0.31	0.35
UDP	0.40	0.06
$\alpha$ -UDPGlcUA	0.28	0.10
$\beta$ -UDPGlcUA	0.30	0.25
UMP-NH <sub>2</sub> *	0.45	0.50

\* 1,3-dicyclohexyl guanidinium uridine 5'-phosphoramidate.

The uridine absorption in ultraviolet light was compared with the standard  $\alpha$ -UDPGlcUA in solution at pH 1.0 (1 M HCl) and was read in a DU-spectrophotometer from 248 to 280 nm. The products were also developed and compared by descending paper chromatography with the solvent ammonium acetate-ethanol (3:7.5, v/v)<sup>2</sup> with the standards of  $\alpha$ -UDPGlcUA, UDP, 1,3-dicyclohexyl guanidinium uridine 5'-phosphoramidate, UMP.

*Conditions for  $\beta$ -UDPGlcUA as substrate for  $\beta$ -glucuronidase (Table II).* The enzyme used was purified mouse urinary  $\beta$ -glucuronidase<sup>4</sup>. The substrates were standard  $\alpha$ -UDPGlcUA and synthesized  $\beta$ -UDPGlcUA in 0.2 M acetate buffer (pH 4.5), saccharolactone (0.67 mM) was used as an inhibitor. The reaction mixture was incubated at  $37^\circ$  for 24 h, chilled to stop the reaction and the FISHMAN AND GREEN<sup>7</sup> method was applied directly.

The FISHMAN AND GREEN<sup>7</sup> test for bound glucuronic acid of the synthesized compound recovers the amount of glucuronic acid expected from the companion measurements of the ultraviolet absorption at 260 nm, made on  $\beta$ -UDPGlcUA.

The results of hydrolysis of  $\beta$ -UDPGlcUA was presented in Table II which shows that  $\beta$ -UDPGlcUA is a substrate for  $\beta$ -glucuronidase since 12 out of 13  $\mu\text{g}$  of its glucuronic acid were liberated in the enzyme digest. Saccharolactone inhibited hydrolysis by 75%. On the other hand, no glucuronic acid was released from  $\alpha$ -UDPGlcUA by  $\beta$ -glucuronidase.

The ultraviolet absorption spectra of  $\beta$ -UDPGlcUA at 262 nm is indistinguishable from that of  $\alpha$ -UDPGlcUA. It has been found (Table I) that  $\beta$ -UDPGlcUA travels close to  $\alpha$ -UDPGlcUA on paper but can be separated from it by thin-layer chromatography. The above results indicate that the  $\beta$ -UDPGlcUA synthesized was sufficiently pure to be used as a substrate for  $\beta$ -glucuronidase. Indeed, it is a valid substrate (Table II) for  $\beta$ -glucuronidase which was also verified by the well known saccharolactone inhibition of  $\beta$ -glucuronidase.

TABLE II

HYDROLYSIS OF  $\alpha$ -UDPGlcUA AND  $\beta$ -UDPGlcUA BY  $\beta$ -GLUCURONIDASE

Test substrate	Enzyme	Substrate	Saccharo- lactone	Glucuronic acid in the incubation mixture ( $\mu$ g)		
				Total	Bound	Free
$\alpha$ -UDPGlcUA	+	+	—	10.0	10.0	0.0
	—	+	—	10.0	10.0	0.0
	+	+	+	10.0	10.0	0.0
	+	—	—	0.0	0.0	0.0
$\beta$ -UDPGlcUA	+	+	—	13.0	1.0	12.0
	—	+	—	13.0	13.0	0.0
	+	+	+	13.0	10.0	3.0
	+	—	—	0.0	0.0	0.0

One wonders whether  $\beta$ -UDPGlcUA may be a constituent of normal mammalian cells. In this regard, attention may be directed to the endogenous glucuronide synthesis of homogenates<sup>8</sup> unrelated to UDPGlcUA glucuronyltransferase as a likely location of  $\beta$ -UDPGlcUA.

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