

CHEMICAL STUDIES OF 10-DEACETYL BACCATIN III.
HEMISYNTHESIS OF TAXOL DERIVATIVES.

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Abstract - The chemical reactivities of 10-deacetyl baccatin III and of baccatin III, two natural products extracted from *Taxus baccata* L., were studied with the aim of synthesizing taxol analogues having a modified side-chain at C-13, thereby restoring good binding to tubulin.

In 1971 taxol 1 was isolated from *Taxus brevifolia* Nutt and was the first taxane diterpene shown to exhibit cytotoxic activity.¹ *In vivo*, taxol has antileukemic and tumor inhibiting properties^{1,2} and it is currently in clinical trials in France and in the USA. The biological activity has been related to the *in vitro* interaction with microtubule proteins.^{3,4} In contrast with other spindle poisons such as vinblastine and colchicine which prevent the assembly of tubulin^{5,6} taxol 1 promotes the assembly of microtubules and inhibits the depolymerisation process of tubulin. In addition to taxol, other taxane derivatives showing similar biological activity have been isolated from various species of yew tree.⁷⁻⁹ Because of its unique mode of action taxol may be the prototype of a new class of chemotherapeutic drugs. However, one of the disadvantages of taxol is associated with its limited availability from natural sources: it is extracted in low yield from the stem bark of the very slow-growing yew tree. To circumvent this major problem several attempts to synthesize the unusual taxane skeleton have been described¹⁰ but to date no total synthesis of taxol has been reported. One other way to prepare this compound is to use simpler taxane derivatives which could be used as precursors in a taxol hemisynthesis.

Baccatin III 2 has been isolated from an alcohol extract of heart wood⁸ and 10-deacetyl baccatin III 3 was easily extracted from the annual cut of the yew leaves.⁹ These two compounds are not as active as taxol 1 both *in vitro* and *in vivo*, but they can be used as raw materials for the preparation of taxol and derivatives. In this paper we wish to report some chemical properties of these compounds and the preparation of new taxane diterpenes which could be used as intermediates in the hemisynthesis of taxol itself.* The compounds obtained in this study have been submitted for *in vitro* antitubulin evaluation which will allow establishment of structure-activity relationships in this series.

(*) Some of this work has been the subject of the PhD Thesis of one of us¹² and has been partly described in a short communication.¹⁴ We thank Professor D. Kingston for a personal communication concerning his study of baccatin III.

The unusual taxane skeleton of 2 and 3 has a very folded structure (figure 1) in which the α hydroxyl group at C-13 is in a hindered position and furthermore, it can form a hydrogen bond with the 4α acetyl group. It is also important to remember that the 7β hydroxyl can easily epimerize into the 7α isomer via a retro aldol mechanism. The presence of a hydrogen bond with the 4α acetyl group stabilizes the α isomer during the aldol condensation.¹¹

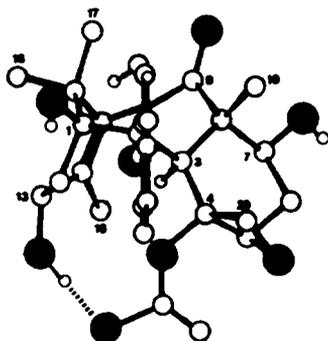


Figure 1.

The other functionalities of tetraol 2 seem stable enough to our experimental conditions, except for the 1-hydroxyl group and the oxetan ring which can be rearranged in acidic media.^{12,13}

I. Reactivity of 10-deacetyl baccatin III 3 with acylating agents.

Acetylation of 3 with acetic anhydride yielded three acetylation products depending on the experimental conditions (Table 1). Structure elucidation of these compounds was obtained by considering the chemical shifts of the three protons at C-7, C-10 and C-13 in their proton NMR spectra (see Experimental Part). These data thus show that there is no selectivity between C-7 and C-10 hydroxyl groups toward acylating agents and that the C-13 hydroxyl group is the least reactive, as expected.

Considering these results, we next undertook to protect the two most reactive hydroxyl groups.

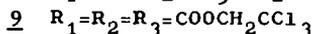
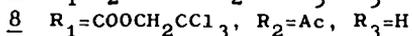
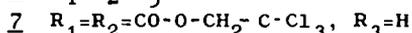
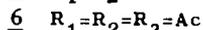
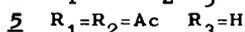
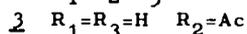
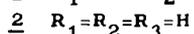
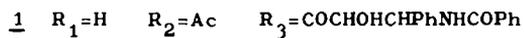
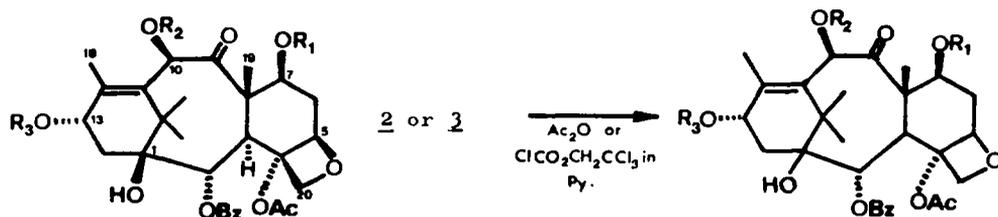


Figure 2.

Table 1.

	Time (h)	Temp. (°C)	<u>4</u>	<u>5</u>	<u>6</u>
1	24	20	48%	48%	-
2	48	60	-	49%	49%
3	24	80	-	-	95%

2. Protection and Deprotection.

Taking into consideration the instability of taxol in basic medium¹ we thought that 2,2,2-trichloroethyl chloroformate could be a good protective group, since it can be removed under very mild conditions. We thus obtained compound 7 in good yield from 10-deacetyl baccatin III 2. With an excess of the acid chloride compound 9 was also prepared. Alkyl 2,2,2-trichloroethyl carbonate can be cleaved by β -elimination with zinc dust in methanol or acetic acid.¹⁵ Compound 7 was cleaved as expected with zinc dust in acetic acid to give the starting material in quantitative yield.

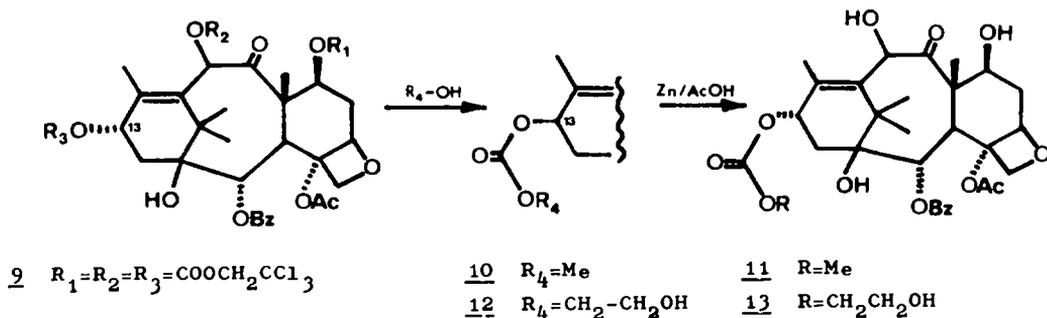


Figure 3.

The same experiments have been done with the 7,10,13-tri(2,2,2-trichloroethoxycarbonyl)-10-deacetyl baccatin III 9. Treatment of this compound with zinc dust in methanol yielded a new derivative 10. Deprotection of the 7 and 10-positions by reductive cleavage in acetic acid gave a quantitative yield of 13-methoxycarbonyl-10-deacetyl baccatin III 11. This product was also obtained by direct methanolysis of 9 after deprotection of the 7 and 10-positions. These results led us to try other nucleophilic agents. Thus preparation of compounds 12 and 13 was achieved by treatment of 9 with ethylene glycol. Unfortunately the *in vitro* activities on microtubule assembly of 11 and 13 were less than that of taxol 1.

Baccatin III 2 was protected in the same way to give 8.

3. Hemisynthesis of taxol derivatives from 7 and 8.

Various *Taxus* species contain a mixture of alkaloids named "taxine".¹⁶ The basic property of these compounds is due to the Winterstein's acid (3-dimethyl amino-3-phenyl propanoic acid). Biosynthetic study of this acid has shown that it arises from phenylalanine by a β -amination of cinnamic acid.¹⁷ It is also well known that cinnamic esters of taxane diterpenes have been isolated from different species of yew trees. Cinnamic acid is thus an attractive candidate for esterification of the free C-13 hydroxyl group of compound 7.

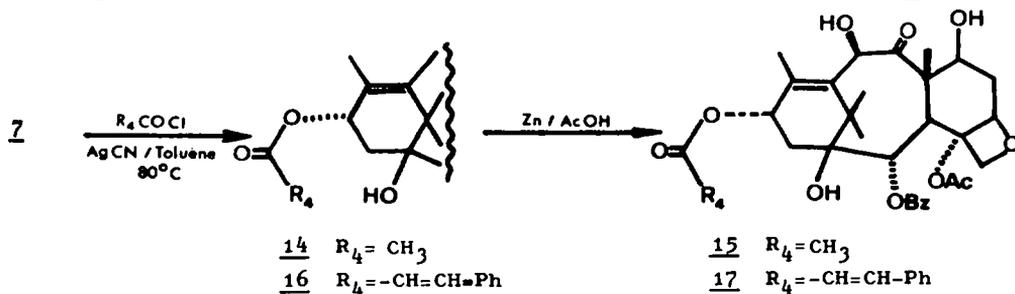


Figure 4.

In contrast to the relatively easy acetylation of 7 leading to 14 and 15 after deprotection, acylation with cinnamoyl chloride has proved to be a difficult reaction under the usual experimental conditions. Preparation of 13-cinnamoyl-10-deacetyl baccatin III 17 was finally achieved by the coupling of cinnamoyl chloride and 7 in the presence of silver cyanide at 110°C in toluene, followed by the deprotection of the 10 and 7 positions of 16 by treatment with zinc in acetic acid. The use of more complex acid chlorides such as β phenyl isoserine (Threo and Erythro) to prepare taxol 1 was not successful in our hands. Therefore, our next approach was to investigate some addition reactions on the double bond of the cinnamoyl ester 16.

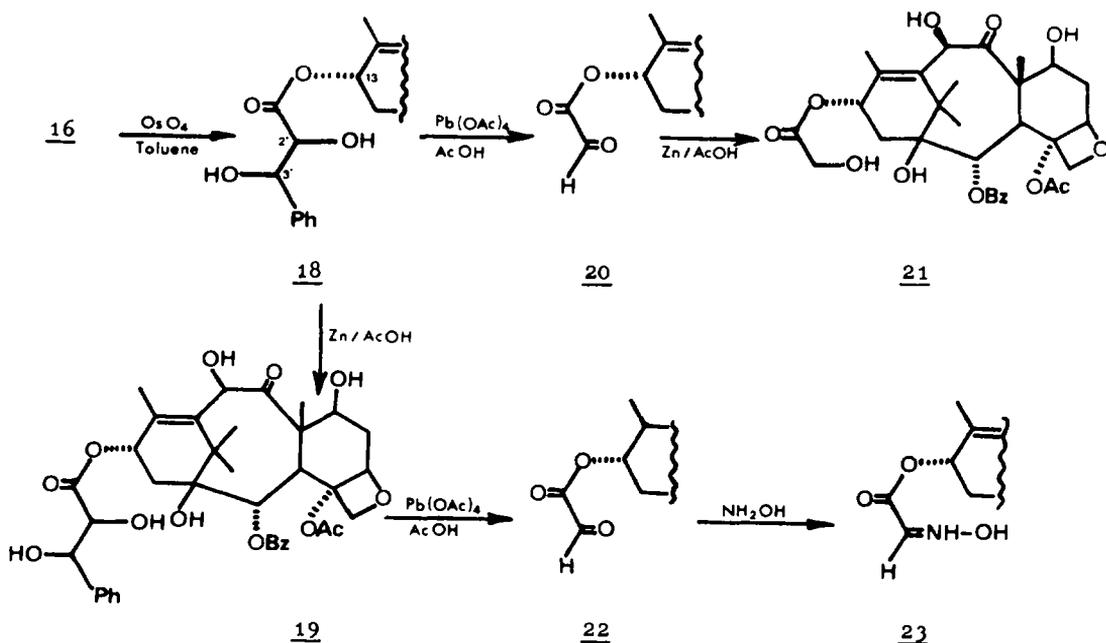


Figure 5.

The relative configuration of the 2' and 3' carbons in taxol and the *trans* configuration of the cinnamate ester 16 require a *cis* addition on the 2',3' double bond. The reaction of 16 with osmium tetroxide in pyridine led to the rapid formation of the 2',3'-dihydroxy derivative 18 as a mixture of diastereoisomers (2'S, 3'R and 2'R, 3'S) which could be purified by HPLC of the deprotected mixture 19 (19a and 19b). The ^1H NMR spectrum of 19a and 19b showed two new doublets ($J = 2$) corresponding to the C-3' and C-2' protons. The FAB mass spectrum gave peaks at m/z 709 (MH^+) corresponding to the addition of two hydroxyl groups.

Oxidation of 18 with lead tetraacetate followed by deprotection of the resulting aldehyde 20 with zinc in acetic acid gave 13-hydroxyacetyl-10-deacetyl baccatin III 21. In a similar manner oxidation of the mixture 19 gave the aldehyde intermediate 22 which was characterized as its oxime 23.

We also tried acylation of baccatin III with crotonyl chloride in order to evaluate the influence of the C-3' phenyl group in 17 on the *in vitro* activity. Treatment of 7-(2,2,2-trichloroethoxycarbonyl) baccatin III 8 with crotonyl chloride gave the C-13 ester 24 which yielded 25 after deprotection of the C-7 position.

Hydroxylation of 24 with osmium tetroxide followed by deprotection of the resulting dihydroxy derivative 26 with zinc in acetic acid gave the ester 27

resulting dihydroxy derivative 26 with zinc in acetic acid gave the ester 27 which is less active on tubulin than the esters 19 containing a phenyl group at C-3'.

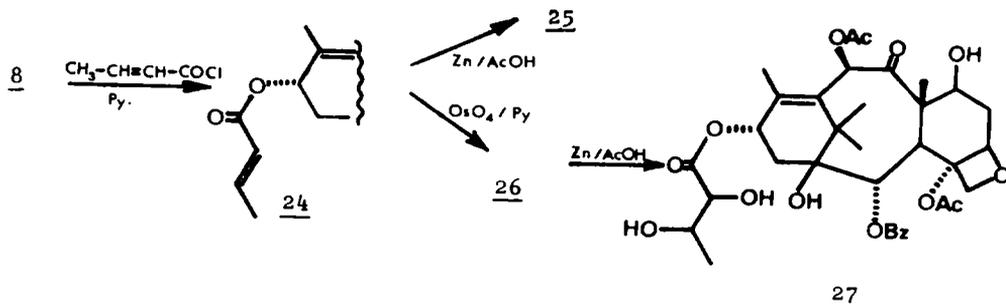


Figure 6.

Structure-activity relationships of these new taxol derivatives will be discussed in a subsequent publication but it is already interesting to note that, in contrast to the phenyl group, a methyl group at C-3' 25 destroys the *in vitro* activity and that hydroxyl groups at the 2' and 3' positions 19 increase the activity in comparison to the cinnamate ester 17.

Conclusion.

The results obtained in our work show that it is possible to carry out esterification of the extremely hindered C 13-hydroxy group of baccatin III and 10-deacetyl baccatin III. In particular, hemisynthesis of cinnamate ester 16 allowed us to prepare some taxol derivatives. This compound would seem to be a good precursor of taxol itself using Sharpless hydroxyamination¹⁸ as described previously.¹⁴

EXPERIMENTAL SECTION

Due to the complexity of the molecules and the small size of the samples available, no elemental analysis is given. Purity of the samples was determined by chromatographic homogeneity and careful analysis of NMR spectra (200 MHz or 400 MHz). Preparative T.L.C. was performed on Merck Silica Gel PF-254 plates. Melting points were observed on a Kofler apparatus, optical rotations measured (c, g/100 ml) on a Perkin-Elmer 141 MC, infrared spectra ($\nu_{\text{cm}^{-1}}$, CHCl_3) on a Perkin-Elmer 257, ultraviolet spectra [EtOH , λ_{max} nm (ϵ)] on a Jobin-Yvon duospac 203. ^1H NMR spectra were obtained at 200 MHz or at 400 MHz (Bruker AM 200 or AM 400) using TMS as internal standard (coupling constants (J) are given in Hertz (Hz); s, d, t, dd and m indicates singlet, doublet, triplet, doublet of doublets and multiplet, respectively). Mass spectra were measured on an AEI MS 9 (CI) or a Kratos MS 80 (FAB). ^{13}C NMR Spectra of taxol derivatives will be presented and discussed in a further publication.

Acetylation of 10-Deacetyl baccatin III 2

Exp. 1 : Acetic anhydride (0.1 ml) was added to a solution of 2 (17 mg) in pyridine (1 ml) with stirring at room temperature for 21 h. Work up by standard methods and purification by preparative TLC (solvent : CH_2Cl_2 -MeOH, 93-7) yielded 7-acetyl-10-deacetyl baccatin III 4 (10 mg) and 7-acetyl baccatin III 5 (8.4 mg).

Exp. 2 : Acetic anhydride (0.18 ml) was added to a solution of 2 (30 mg) in pyridine (1 ml) with stirring at 60°C for 48 h. Usual work-up and isolation of the products by preparative TLC (solvent : CH_2Cl_2 -MeOH, 97-3) gave 5 (17 mg) and 7,13-diacetyl baccatin 6 (18 mg).

Exp. 3 : Acetic anhydride (0.25 ml) was added to a solution of 2 (25.5 mg) in pyridine (1 ml) with stirring at 80°C for 24 h. Usual work up yielded 7,13-diacetyl baccatin III 6 (30 mg).

7-Acetyl-10-deacetyl baccatin III 4

Mp 265-266°C (MeOH-H₂O); $[\alpha]_{\text{D}}^{23} = -56^\circ$ (c = 0.53, CHCl_3); UV : 231(16100), 275(1090), 280(920); IR : 3400, 1730, 1710; ^1H NMR(CDCl_3) δ : 1.04 (3H, s, C₁₉-H₃), 1.09 (3H, s, C₁₆H₃), 1.83 (3H, s, C₁₉H₃), 1.97 (3H, s, C₁₈H₃), 2.08 and 2.32 (2 x 3H, 2s, 2 x OAc), 4.06 (1H, d, $J = 7$, C₃-H), 4.20 and 4.30 (2H, 2d, $J = 9$, C₂₀H₂), 4.80 (1H, t, $J = 9$, C₁₃-H), 4.95 (1H, d, $J = 9$, C₅-H), 5.33 (1H, s, C₁₀-H), 5.59 (1H, m, C₇-H), 5.61 (1H, d, $J = 7$, C₂-H), 7.45, 7.58 and 8.08 (5H, OBz); MS(C.I., m/z : 587(MH⁺), 569, 551, 509, 387, 327, 309, 123.

7-Acetyl baccatin III 5.

Mp 249-251°C (MeOH-H₂O) ; $[\alpha]_D^{23} = -79^\circ$ (c = 0.6, CHCl₃) ; UV : 230(19500), 276(1150), 282(1040) ; IR : 3400, 1730, 1710 ; ¹H NMR (CDCl₃) δ : 1.08 (3H, s, C₁₇H₃), 1.13 (3H, s, C₁₆H₃), 1.79 (3H, s, C₁₉H₃), 2.03 (3H, s, C₁₈H₃), 2.10, 2.16 and 2.29 (3 x 3H, 3s, 3 OAc), 2.60 and 1.80 (2H, C₆H₂), 2.28 (m, C₁₄H₂), 4.00 (1H, d, J = 7, C₃-H), 4.14 and 4.31 (2H, 2d, J = 9, C₂₀H₂), 4.86 (1H, t, J = 9, C₁₃-H), 4.97 (1H, d, J = 9, C₅-H), 5.59 (1H, m, C₇-H), 5.62 (1H, d, J = 7, C₂-H), 6.26 (1H, s, C₁₀-H), 7.46, 7.58 and 8.08 (5H, OBz), MS(CI), m/z : 629 (MH⁺), 611, 569, 551, 509, 491, 387, 327, 309, 123.

7,13-Diacetyl baccatin III 6.

Mp 236-237°C (MeOH-H₂O) ; $[\alpha]_D^{23} = -80^\circ$ (c = 0.52, CHCl₃) ; UV : 232(19500), 275(1380), 282(1250) ; IR : 3400, 1730, 1710 ; ¹H NMR (CDCl₃) δ : 1.17 (3H, s, C₁₇H₃), 1.22 (3H, s, C₁₆H₃), 1.81 (3H, s, C₁₉H₃), 1.97 (3H, s, C₁₈H₃), 1.80 and 2.60 (m, 2H, C₆H₂), 2.03, 2.17, 2.21 and 2.34 (4 x 3H, 4s, 4 OAc), 2.25 (m, 2H, C₁₄H₂), 3.97 (1H, d, J = 7, C₃-H), 4.16 and 4.33 (2H, 2d, J = 9, C₂₀H₂), 4.98 (1H, d, J = 9, C₅-H), 5.59 (1H, m, C₇-H), 5.66 (1H, d, J = 7, C₂-H), 6.18 (1H, t, J = 9, C₁₃-H), 6.26 (1H, s, C₁₀-H), 7.48, 7.61 and 8.08 (5H, OBz) ; MS(CI) m/z 671 (MH⁺), 653, 611, 593, 551, 491, 489, 429, 369, 309, 123.

7,10-di(2,2,2-Trichloroethyloxycarbonyl)-10-deacetyl baccatin III 7.

2,2,2-Trichloroethylchloroformate (0.085 ml) was added to a solution of 10-deacetyl baccatin III 2 (100 mg) in pyridine (2 ml) with stirring at 80°C. After 5 min the reaction mixture was treated with H₂O and extracted with CH₂Cl₂. Usual work-up and purification by preparative TLC (solvent : CH₂Cl₂-MeOH, 95-5) yielded 7 (153 mg). mp 233-234°C (MeOH-H₂O) ; $[\alpha]_D^{23} = -58^\circ$ (c = 0.465, CHCl₃) ; UV : 232(19000), 276(990), 283(810) ; IR : 3420, 1765, 1730, 1720 ; ¹H NMR (CDCl₃) δ : 1.12 (3H, s, C₁₇H₃), 1.16 (3H, s, C₁₆H₃), 1.85 (3H, s, C₁₉H₃), 2.16 (3H, s, C₁₈H₃), 2.30 (3H, s, OAc), 2.30 (m, C₁₄H₂), 2.05 and 2.65 (2m, C₆H₂), 4.00 (1H, d, J = 7, C₃-H), 4.18 and 4.35 (2H, 2d, J = 9, C₂₀H₂), 4.63 and 4.92 (2H, 2d, J = 12, CH₂ of the protecting group), 4.76 and 4.80 (2H, 2d, J = 12, CH₂ of the protecting group), 4.92 (1H, t, J = 9, C₁₃-H), 5.00 (1H, d, J = 9, C₅-H), 5.61 (1H, m, C₇-H), 5.66 (1H, d, J = 7, C₂-H), 6.30 (1H, s, C₁₀-H), 7.50, 7.64 and 8.13 (2H, 1H and 2H, 2t and 1d, J = 7, OBz) ; MS(CI), m/z 893(MH⁺), 875, 701, 683, 579, 387, 327, 309, 123.

7,10,13-tri(2,2,2-Trichloroethyloxycarbonyl)-10-deacetyl baccatin III 9.

2,2,2-Trichloroethylchloroformate (0.13 ml) was added to a solution of 2 (100 mg) in pyridine (1.4 ml) with stirring at 80°C. After 30 min, water was added and the reaction mixture was extracted with CH₂Cl₂. Work-up by usual methods and purification by preparative TLC gave 9 (185 mg). mp 229-231°C (MeOH-H₂O) ; $[\alpha]_D^{23} = -55^\circ$ (c = 0.56, CHCl₃) ; UV : 232 nm (19800), 275(1600), 282(1280) ; IR : 3400, 1765, 1730, 1720 ; ¹H NMR (CDCl₃) δ : 1.19 (3H, s, C₁₇H₃), 1.21 (3H, s, C₁₆H₃), 1.85 (3H, s, C₁₉H₃), 2.12 (3H, s, C₁₈H₃), 2.42 (3H, s, OAc), 2.40 (m, C₁₄H₂), 2.06 and 2.65 (m, C₆H₂), 4.00 (1H, d, J = 7, C₃-H), 4.15 and 4.36 (2H, 2d, J = 9, C₂₀H₂), 4.61 and 4.92 (2d, J = 12), 4.77 and 4.80 (2d, J = 12) and 4.88 (s) (3 x CH₂ of the protecting groups), 4.98 (1H, d, J = 9, C₅-H), 5.69 (1H, d, J = 7, C₂-H), 5.64 (1H, m, C₇-H), 6.03 (1H, t, J = 9, C₁₃-H), 6.29 (1H, s, C₁₀-H), 7.51, 7.63 and 8.10 (2H, 1H and 2H, 2t and 1d, J = 7, OBz) ; MS (CI), m/z 1067 (MH⁺), 875, 683, 491, 431, 309, 123.

7,10-di(2,2,2-Trichloroethyloxycarbonyl)-13-methyloxycarbonyl-10-deacetyl baccatin III 10.

Zinc dust (20 mg) was added to a solution of 9 (30 mg) in methanol (2 ml) with stirring at 60°C. After 1h 30, the reaction mixture was filtered and concentrated to dryness. Purification by preparative TLC (solvent = CH₂Cl₂-MeOH, 98-2) gave 10 (23 mg). ¹H NMR (CDCl₃) δ : 1.18 (3H, s, C₁₇H₃), 1.21 (3H, s, C₁₆H₃), 1.85 (3H, s, C₁₉H₃), 2.08 (3H, s, C₁₈H₃), 2.36 (3H, s, OAc), 2.36 (m, C₁₄H₂), 2.06 and 2.65 (2m, C₆H₂), 3.88 (3H, s, CH₃O), 3.96 (1H, d, J = 7, C₃-H), 4.15 and 4.35 (2H, 2d, J = 9, C₂₀H₂), 4.60 and 4.90 (2H, 2d, J = 12), 4.75 and 4.78 (2H, 2d, J = 12) (2 x CH₂ of the protecting groups), 4.98 (1H, d, J = 9, C₅-H), 5.62 (1H, m, C₇-H), 5.68 (1H, d, J = 7, C₂-H), 5.95 (1H, t, J = 9, C₁₃-H), 6.26 (1H, s, C₁₀-H), 7.50, 7.63, 8.08 (2H, 1H and 2H, 2t and 1d, J = 7, OBz) ; MS (CI) m/z 951 (MH⁺), 933, 759, 683, 491, 369, 309, 123.

13-Methyloxycarbonyl-10-deacetyl baccatin III 11.

Zinc dust (20 mg) was added to a solution of 10 (23 mg) in acetic acid (1 ml) with stirring at 40°C for 2 h. The reaction mixture was filtered, concentrated and extracted with ethyl acetate. The organic layer was washed with water, brine and dried (MgSO₄). Filtration and concentration gave a quantitative yield of 11. mp 208-210°C (MeOH-H₂O) ; $[\alpha]_D^{23} = -70^\circ$ (c = 0.38, CHCl₃) ; UV : 231(16600), 275(1130), 282(955, EtOH) ; IR : 3450, 1740, 1720, 1705 ; ¹H NMR (CDCl₃) δ : 1.08 (3H, s, C₁₇H₃), 1.11 (3H, s, C₁₆H₃), 1.70 (3H, s, C₁₉H₃), 1.95 (3H, s, C₁₈H₃), 2.27 (3H, s, OAc), 2.27 (m, C₁₄H₂), 1.94 and 2.46 (2m, C₆H₂), 3.81 (3H, s, OCH₃), 3.88 (1H, d, J = 7, C₃-H), 4.18 (1H, m, C₇-H), 4.13 and 4.26 (2H, 2d, J = 9, C₂₀H₂), 4.92 (1H, d, J = 9, C₅-H), 5.19 (1H, s, C₁₀-H), 5.56 (1H, d, J = 7, C₂-H), 5.84 (1H, t, J = 9, C₁₃-H), 7.41, 7.54 and 7.98 (2H, 1H and 2H, 2t and 1d, J = 7, OBz) ; MS(CI), m/z 603 (MH⁺), 585, 509, 387, 345, 327, 309, 123.

Compound 12.

Ethylene glycol (1.2 ml) was added to a solution of 9 (43 mg) in DMF (0.6 ml) with stirring at 80°C for 22 h. The reaction mixture was concentrated and purified by preparative TLC (solvent : CH₂Cl₂-MeOH, 97.5-2.5) to give 12 (27.5 mg). ¹H NMR (CDCl₃) : 1.19 (3H, s, C₁₇H₃), 1.21 (3H, s, C₁₆H₃), 1.84 (3H, s, C₁₉H₃), 2.08 (3H, s, C₁₈H₃), 2.36 (3H, s, OAc), 2.36 (m, C₁₄H₂), 2.04 and 2.62 (m, C₆H₂), 3.91 and 4.36 (2 x 2H, m, HOCH₂CH₂O), 3.94 (1H, d, J = 7, C₃-H), 4.14 and 4.33 (2H, 2d, J = 9, C₂₀H₂), 4.60 and 4.90 (2H, 2d, J = 12), 4.77 and 4.79 (2H, 2d, J = 12) (2 x CH₂ of the protecting groups), 4.98 (1H, d, J = 9, C₅-H), 5.60 (1H, m, C₇-H), 5.66 (1H, d, J = 7, C₂-H), 5.94 (1H, t, J = 9, C₁₃-H), 6.27 (1H, s, C₁₀-H), 7.50, 7.62 and 8.08 (2H, 1H, 2H, 2t and 1d, J = 7, OBz); MS(Cl), m/z 981 (MH⁺), 963.

13-(2-Hydroxyethyloxycarbonyl)-10-deacetyl baccatin III 13.

Zinc dust (30 mg) was added to a solution of 12 in acetic acid (1 ml) at 40°C for 4 h, with stirring. Filtration, concentration and purification of the crude mixture by preparative TLC (solvent : CH₂Cl₂-MeOH, 90-10) gave 12 mg of 13 (68%); [α]_D²³ = -72° (c = 0.33, CHCl₃); UV : 231(18200), 275(16200), 282(13600); IR : 3450, 1740, 1720, 1705; ¹H NMR (CDCl₃) δ : 1.14 (3H, s, C₁₇H₃), 1.19 (3H, s, C₁₆H₃), 1.76 (3H, s, C₁₉H₃), 2.02 (3H, s, C₁₈H₃), 2.36 (3H, s, OAc), 2.02 and 2.62 (2m, C₆H₂), 2.34 (m, C₁₄H₂), 3.93 and 4.36 (2 x 2H, m, HOCH₂-CH₂O), 3.99 (1H, d, J = 7, C₃-H), 4.18 and 4.30 (2H, 2d, J = 9, C₂₀H₂), 4.30 (1H, m, C₇-H), 4.99 (1H, d, J = 9, C₅-H), 5.27 (1H, s, C₁₀-H), 5.67 (1H, d, J = 7, C₂-H), 5.93 (1H, t, J = 9, C₁₃-H), 7.50, 7.63 and 8.09 (2H, 1H and 2H, 2t and 1d, J = 7, OBz); MS(Cl), m/z 633 (MH⁺), 615, 597, 527, 509, 405, 345, 123, 89.

7,10-di(2,2,2-Trichloroethyloxycarbonyl)-13-acetyl-10-deacetyl baccatin III 14.

A solution of 7 (65 mg) in toluene was stirred at 80°C. Acetyl chloride (0.055 ml) and silver cyanide (42 mg) were added to the reaction mixture. After 3 h, water was added and the reaction was extracted with CH₂Cl₂. Usual work-up and purification by preparative TLC (solvent : CH₂Cl₂-MeOH) gave 14 (46 mg); [α]_D²³ = -48° (c = 0.35, CHCl₃); UV : 232(16200), 275(13200), 282(12100); IR : 3450, 1765, 1730, 1715 cm⁻¹; ¹H NMR (CDCl₃) δ : 1.17 (3H, s, C₁₇H₃), 1.24 (3H, s, C₁₆H₃), 1.84 (3H, s, C₁₉H₃), 2.03 (3H, s, C₁₈H₃), 2.21 and 2.36 (2 x 3H, 2s, 2 x OAc), 3.94 (1H, d, J = 7, C₃-H), 4.16 and 4.33 (2H, 2d, J = 9, C₂₀H₂), 4.61 and 4.91 (2H, 2d, J = 12), 4.79 (2H, s) (2 x CH₂ of the protecting groups); 4.99 (1H, d, J = 9, C₅-H), 5.59 (1H, m, C₇-H), 5.70 (1H, d, J = 7, C₂-H), 6.21 (1H, t, J = 9, C₁₃-H), 6.27 (1H, s, C₁₀-H), 7.50, 7.62 and 8.08 (2H, 1H and 2H, 2t and 1d, J = 7, OBz); MS(Cl), m/z 935 (MH⁺), 743, 683, 623, 561, 369, 309, 291, 123.

12-Acetyl-10-deacetyl baccatin III 15.

Zinc dust (20 mg) was added to a solution of 14 (46 mg) in acetic acid (1.5 ml) at 40°C with stirring. After 2h 30, the reaction was filtered and extracted with ethyl acetate. Purification by preparative TLC (solvent : CH₂Cl₂-MeOH, 95-5) gave 20 mg of 15. [α]_D²³ = -69° (c = 0.67, CHCl₃); UV : 232(16100), 276(12900), 282(12000); IR : 3450, 1730, 1715; ¹H NMR (CDCl₃) δ : 1.11 (3H, s, C₁₇H₃), 1.21 (3H, s, C₁₆H₃), 1.74 (3H, s, C₁₉H₃), 1.94 (3H, s, C₁₈H₃), 1.86 and 2.59 (2m, C₆H₂), 2.23 and 2.32 (2 x 3H, 2s, 2 x OAc), 2.23 (m, C₁₄H₂), 3.95 (1H, d, J = 7, C₃-H), 4.18 and 4.32 (2H, 2d, J = 9, C₂₀H₂), 4.26 (m, C₇-H), 4.98 (1H, d, J = 9, C₅-H), 5.22 (1H, s, C₁₀-H), 5.67 (1H, d, J = 7, C₂-H), 6.16 (1H, t, J = 9, C₁₃-H), 7.48, 7.60 and 8.07 (2H, 1H and 2H, 2t and 1d, J = 7, OBz); MS (Cl) : m/z : 587 (MH⁺), 569, 551, 527, 509, 405, 387, 345, 327, 123.

7,10-di(2,2,2-Trichloroethyloxycarbonyl)-13 cinnamoyl-10 deacetyl baccatin III 16.

Oxalyl chloride (0.057 ml) was added to a solution of cinnamic acid (50 mg) in dry toluene (1 ml) with stirring at 60°C for 30 min. After distillation of oxalyl chloride, 2 ml of dry toluene, compound 7 (60 mg) and silver cyanide (40 mg) were added and the reaction mixture was heated at 110°C for 20h. After filtration, extraction with ethyl acetate and usual work-up, the product 16 (42 mg) was isolated by preparative TLC (solvent : CH₂Cl₂-MeOH, 99.5-0.5). 16 : [α]_D²³ = -56° (c = 0.57, CHCl₃); UV : 217(26800), 222(26900), 232(16100), 276(24400); IR : 3420, 1760, 1725, 1710, 1635; ¹H NMR (CDCl₃) δ : 1.29 (3H, s, C₁₇H₃), 1.29 (3H, s, C₁₆H₃), 1.88 (3H, s, C₁₉H₃), 2.16 (3H, s, C₁₈H₃), 2.31 (3H, s, OAc), 3.99 (1H, d, J = 7, C₃-H), 4.20 and 4.34 (2H, 2d, J = 9, C₂₀H₂), 4.62 and 4.93 (2H, 2d, J = 12), 4.79 (2H, s) (2 x CH₂ of the protecting groups), 5.02 (1H, d, J = 9, C₅-H), 5.62 (1H, m, C₇-H), 5.73 (1H, d, J = 7, C₂-H), 6.21 (1H, t, J = 8, C₁₃-H), 6.30 (1H, s, C₁₀-H), 6.53 and 7.89 (2H, 2d, J = 16, C₂-H and C₃-H); 7.45 and 7.60 (5H, Ph); 7.45, 7.60 and 8.07 (5H, OBz); MS (C.I.), m/z 1023 (MH⁺), 1005, 831, 813, 683, 665, 491, 431, 369, 309, 291, 149, 131, 123.

13-Cinnamoyl-10 deacetyl baccatin III 17.

Zinc dust (15 mg) was added to a solution of 16 (30mg) in acetic acid (1.5ml) at 40°C for 2 h. Filtration, concentration, extraction with ethyl acetate and purification by preparative TLC (solvent : CH₂Cl₂-MeOH, 93-7) gave 14 mg of 17. mp = 229-231°C (MeOH); [α]_D²³ = -104° (c = 0.45, CHCl₃); UV : 217(26200), 221(26100), 232(16400), 276(24600); IR : 3440, 1725, 1710, 1635; ¹H NMR (CDCl₃) δ : 1.15 (s, 3H, C₁₇H₃), 1.25 (3H, s, C₁₆H₃), 1.78 (3H, s, C₁₉H₃), 2.06 (3H, s, C₁₈H₃).

2.28 (3H, s, OAc), 4.00 (1H, d, J = 7, C₃-H), 4.31 (m, C₇-H), 4.20 and 4.31 (2H, 2d, J = 9, C₂₀H₂), 4.99 (1H, d, J = 9, C₅-H), 5.28 (1H, s, C₁₀-H), 5.70 (1H, d, J = 7, C₇-H), 6.18 (1H, t, J = 8, C₁₃-H), 6.51 and 7.85 (2H, 2d, J = 16, C₂-H and C₃-H); 7.45 and 7.58 (5H, C₃-Ph), 7.45, 7.58 and 8.07 (5H, OBz); MS (C.I.), m/z 675 (MH⁺), 657, 639, 527, 509, 491, 405, 387, 345, 327, 149, 131, 123.

Compound 18.

Osmium tetroxide (54 mg) was added to a solution of 16 (140 mg) in pyridine (3.6 ml) at room temperature with stirring. After 1h, sodium bisulfite (245 mg) in water (3.6 ml) was added. The reaction mixture was left at room temperature for 2h and extracted with ethyl acetate. Usual work-up and purification by preparative TLC (solvent: CH₂Cl₂-MeOH, 94-6) gave 18 (115 mg) (mixture of two diastereoisomers). IR: 3500, 1760, 1730 and 1715 cm⁻¹; ¹H NMR (CDCl₃) δ: 1.19 (s, C₁₇H₃), 1.26 (s, C₁₆H₃), 1.84 (s, C₁₉H₃), 2.02 (s, C₁₈H₃), 2.24 (s, OAc), 3.88 (d, J = 7, C₃-H), 4.17 and 4.31 (2d, J = 9, C₂₀H₂), 4.40 and 4.46 (1d, J = 3 and brs, C₂-H), 4.96 (d, J = 9, C₅-H), 5.09 and 5.21 (1d, J = 3 and brs, C₃-H), 4.61, 4.62 and 4.79 (2d, J = 12 and 1s, CH₂ of protecting groups), 5.56 (m, C₇-H), 5.69 (d, J = 7, C₂-H), 6.22 (t, J = 9, C₁₃-H), 6.26 (s, C₁₀-H), 7.44 and 7.53 (5H, C₃-Ph), 7.53, 7.66 and 8.07 (5H, OBz); MS (C.I.), m/z: 875 (MH⁺ - side chain), 683, 491, 369, 309, 165, 137, 123.

13-(2',3'-dihydroxy-3' phenyl propanoyl)-10-deacetyl baccatin III 19a and 19b (2'R, 3'S and 2'S, 3'R):

Zinc dust (20 mg) was added to a solution of 18 (51 mg) in acetic acid (2 ml) at 40°C for 5 h. After filtration and concentration, the reaction mixture was extracted with ethyl acetate. Purification by preparative TLC (solvent: CH₂Cl₂-MeOH, 92.5-7.5) gave 21 mg of 19a and 4 mg of 19b.

19a: [α]_D²³ = -59° (c = 0.44, CHCl₃); UV: 219(14400), 230(15700), 275(1210), 282(1100); IR: 3450, 1735, 1720; ¹H NMR (CDCl₃) δ: 1.12 (3H, s, C₁₇H₃), 1.23 (3H, s, C₁₆H₃), 1.74 (3H, s, C₁₉H₃), 1.89 (3H, s, C₁₈H₃), 2.20 (3H, s, OAc), 3.87 (1H, d, J = 7, C₃-H), 4.16 and 4.19 (2H, 2d, J = 9, C₂₀H₂), 4.22 (1H, m, C₇-H), 4.38 (1H, d, J = 2, C₂-H), 4.93 (1H, d, J = 9, C₅-H), 5.05 (1H, d, J = 2, C₃-H), 5.21 (1H, s, C₁₀-H), 5.66 (1H, d, J = 7, C₂-H), 6.19 (1H, t, J = 9, C₁₃-H), 7.43 and 7.53 (5H, C₃-Ph), 7.53, 7.66 and 8.08 (5H, OBz); MS (FAB), m/z 709 (MH⁺).

19b: [α]_D²³ = -73° (c = 0.33, CHCl₃); UV: 220(15000), 230(17400), 275(1650), 282(1400); IR: 3450, 1735, 1715; ¹H NMR (CDCl₃) δ: 1.12 (3H, s, C₁₇H₃), 1.22 (3H, s, C₁₆H₃), 1.76 (3H, s, C₁₉H₃), 1.88 (3H, s, C₁₈H₃), 2.27 (3H, s, OAc); 3.94 (1H, m, C₃-H), 4.18 and 4.31 (2H, 2d, J = 9, C₂₀H₂), 4.24 (1H, m, C₇-H), 4.43 (1H, d, J = 2, C₂-H), 4.96 (1H, d, J = 9, C₅-H), 5.17 (1H, d, J = 2, C₃-H), 5.23 (1H, s, C₁₀-H), 5.68 (1H, d, J = 7, C₂-H), 6.26 (1H, t, J = 9, C₁₃-H), 7.42 and 7.48 (5H, C₃-Ph), 7.48, 7.62 and 8.05 (5H, OBz); MS (FAB), m/z 709 (MH⁺).

13-Hydroxy acetyl-10-deacetyl baccatin III 21.

Lead tetraacetate (230 mg) was added to a solution of 18 (360 mg) in acetic acid (5 ml) with stirring at room temperature for 30 min. Usual work-up and purification by preparative TLC (solvent: CH₂Cl₂-MeOH, 95-5) gave 20 (309 mg). Compound 20 (30 mg) was treated with zinc dust (15 mg) in acetic acid (1 ml) at 60°C for 1 h. Filtration, usual work-up and purification by preparative TLC (solvent: CH₂Cl₂-MeOH, 90-10) yielded 21 (20 mg); mp = 188-190°C (MeOH); [α]_D²³ = -48° (c = 0.4, MeOH); IR: 3560, 3450, 1730, 1715; ¹H NMR (CDCl₃-CD₃OD) δ: 1.12 (3H, s, C₁₇H₃), 1.19 (3H, s, C₁₆H₃), 1.72 (3H, s, C₁₉H₃), 1.92 (3H, s, C₁₈H₃), 2.23 (2H, C₁₄H₂), 2.29 (3H, s, OAc), 2.5 (1H, C₆-H), 3.88 (1H, d, J = 7, C₃-H), 4.18-4.28 (5H, C₂₀H₂, C₇-H and C₂H₂), 4.18 and 4.28 (2H, 2d, J = 8, C₂₀H₂), 4.18 (1H, m, C₇-H), 4.24 (2H, d, J = 7.5, C₂H₂), 4.95 (1H, d, J = 9, C₅-H), 5.21 (1H, s, C₁₀-H), 5.63 (1H, d, J = 7, C₂-H), 6.23 (1H, t, J = 9, C₁₃-H), 7.45, 7.58 and 8.01 (5H, OBz); MS (C.I.), m/z 603 (MH⁺), 585, 567, 527, 509, 405, 387, 345, 327, 123, 105, 77.

Compound 23.

Lead tetraacetate (28 mg) was added to a solution of 19 (a + b) (30 mg) in acetic acid (1 ml) with stirring at room temperature for 30 min. Usual work-up and purification by preparative TLC (solvent: CH₂Cl₂-MeOH, 90-10) gave 22 (19 mg). Refluxing a mixture of 22 (9 mg) and hydroxylamine hydrochloride (4 mg) in pyridine (1 ml) for 20 min. yielded 23 in quantitative yield after usual work-up; [α]_D²³ = -51° (c = 0.14, MeOH); ¹H NMR (CDCl₃-CD₃OD) δ: 1.13 (3H, s, C₁₇H₃), 1.21 (3H, s, C₁₆H₃), 1.76 (3H, s, C₁₉H₃), 2.00 (3H, s, C₁₈H₃), 2.31 (3H, s, OAc), 3.93 (1H, d, J = 7, C₃-H), 4.26 (1H, m, C₇-H), 4.28 (2H, d, J = 9, C₂₀H₂), 5.01 (1H, d, J = 9, C₅-H), 5.29 (1H, s, C₁₀-H), 5.69 (1H, d, J = 7, C₂-H), 6.28 (1H, t, J = 9, C₁₃-H), 7.66 (1H, s, C₂-H), 7.52, 7.64 and 8.11 (5H, OBz); MS (FAB), m/z 616 (MH⁺), 598, 580, 527, 509.

7-(2,2,2-Trichloroethyloxy carbonyl)-13-crotonoyl baccatin III 24.

To a solution of 8 (200 mg) in dry toluene (8 ml) was added silver cyanide (455 mg) and crotonyl chloride (0.76 ml). The reaction mixture was stirred at 110°C for 24 h. Filtration, extraction with CH₂Cl₂ and concentration gave a

mixture which was purified by preparative TLC (solvent : CH_2Cl_2 -MeOH, 99-1) to give 24 (120 mg). $^1\text{H NMR}$ (CDCl_3) δ : 1.17 (3H, s, C_{17}H_3), 1.22 (3H, s, C_{16}H_3), 1.88 (3H, s, C_{19}H_3), 1.98 (3H, dd, $J = 7$ et $J = 2$, $\text{C}_4\text{-H}_3$), 2.04 (3H, s, C_{18}H_3), 2.18 and 2.30 (2 x 3H, 2s, 2 OAc), 4.00 (1H, d, $J = 7$, $\text{C}_3\text{-H}$), 4.19 and 4.33 (2H, 2d, $J = 9$, C_{20}H_2), 4.99 (1H, d, $J = 9$, $\text{C}_5\text{-H}$), 4.65 and 5.41 (2H, 2d, $J = 12$, CH_2 of the protecting group), 5.61 (1H, m, $\text{C}_7\text{-H}$), 5.69 (1H, d, $J = 7$, $\text{C}_2\text{-H}$), 5.96 (1H, dd, $J = 15$ and $J = 2$, $\text{C}_2\text{-H}$), 6.13 (1H, t, $J = 9$, $\text{C}_{13}\text{-H}$), 6.40 (1H, s, $\text{C}_{10}\text{-H}$), 7.18 (1H, dd, $J = 15$ and $J = 7$, $\text{C}_3\text{-H}$), 7.48, 7.60 and 8.06 (5H, OBz).

13-Crotonoyl baccatin III 25.

Zinc dust (50 mg) was added to a solution of 24 (50 mg) in acetic acid with stirring at 40°C for 3 h. Filtration, concentration of the solution and purification by preparative TLC (solvent : CH_2Cl_2 -MeOH, 97-3) gave 25 (25 mg). $\alpha_D^{23} = -62^\circ$ ($c = 0.56$, EtOH); UV : 226(20200), 274(1750), 282(1510, EtOH); IR : 3500, 1730, 1650; $^1\text{H NMR}$ (CDCl_3) δ : 1.13 (3H, s, C_{17}H_3), 1.25 (3H, s, C_{16}H_3), 1.68 (3H, s, C_{19}H_3), 1.95 (3H, s, C_{18}H_3), 1.97 (3H, dd, $J = 7$ and $J = 2$, $\text{C}_4\text{-H}_3$), 2.25 and 2.28 (2 x 3H, 2s, 2 OAc), 3.83 (1H, d, $J = 7$, $\text{C}_3\text{-H}$), 4.18 and 4.31 (2H, 2d, $J = 9$, C_{20}H_2), 4.45 (1H, m, $\text{C}_7\text{-H}$), 5.00 (1H, d, $J = 9$, $\text{C}_5\text{-H}$), 5.68 (1H, d, $J = 7$, $\text{C}_2\text{-H}$), 5.95 (1H, dd, $J = 15$ and $J = 2$, $\text{C}_2\text{-H}$), 6.15 (1H, t, $J = 9$, $\text{C}_{13}\text{-H}$), 6.33 (1H, s, $\text{C}_{10}\text{-H}$), 7.16 (1H, dd, $J = 15$ and $J = 7$, $\text{C}_3\text{-H}$), 7.50, 7.61 and 8.07 (5H, OBz). MS (FAB) m/z 655 (MH^+), 637, 595, 577, 509, 491, 449, 349, 327, 309.

Compound 26 (2'S, 3'R and 2'R, 3'S).

Osmium tetroxide (47 mg) was added to a solution of 24 (100 mg) in pyridine (3 ml) at room temperature with stirring for 1 h. Sodium bisulfite in water was added. Extraction with ethyl acetate and purification by preparative TLC (solvent Hexane-EtOAc, 40-60) gave 26 (43 mg, mixture of diastereoisomers). IR : 3500, 1765, 1730; $^1\text{H NMR}$ (CDCl_3) δ : 1.18 (s, C_{17}H_3), 1.26 (s, C_{16}H_3), 1.40 and 1.36 (2d, $J = 6$, $\text{C}_4\text{-H}_3$), 1.85 (s, C_{19}H_3), 2.06 and 2.01 (2s, C_{18}H_3), 2.17 (s, OAc), 2.30 (s, OAc), 3.98 (d, $J = 7$, $\text{C}_3\text{-H}$), 4.12 (m, $\text{C}_2\text{-H}$), 4.33 and 4.18 (d, $J = 9$, C_{20}H_2), 4.65 (s, CH_2 of the protected group), 4.96 (d, $J = 9$, $\text{C}_5\text{-H}$), 5.58 (m, $\text{C}_7\text{-H}$), 5.68 (d, $J = 7$, $\text{C}_2\text{-H}$), 6.33 and 6.23 (t, $J = 9$, $\text{C}_{13}\text{-H}$), 6.50 (s, $\text{C}_{10}\text{-H}$), 8.06, 7.65 and 7.51 (OBz); MS(I.C) m/z : 863 (MH^+), 803, 785, 743, 683, 621, 123, 121 ($\text{CH}_3\text{CHOH-CHOH-COOH} + \text{H}^+$).

Compound 27 (2'S, 3'R and 2'R, 3'S).

Zinc dust (20 mg) was added to a solution of 26 in acetic acid (2 ml) at 40°C for 2 h. Filtration, concentration, extraction with ethyl acetate and purification by preparative TLC (solvent : CH_2Cl_2 -MeOH, 97-3) gave 10 mg of 27. UV(EtOH) 231(13360), 276(1410), 283(1420). IR : 3300, 3500, 1700. $^1\text{H NMR}$ (CDCl_3) δ : 1.18 (s, C_{17}H_3), 1.25 (s, C_{16}H_3), 1.41 and 1.36 (2d, $J = 6$, $\text{C}_4\text{-H}_3$), 1.68 (s, C_{19}H_3), 1.98 and 1.93 (2s, C_{18}H_3), 2.23 (s, OAc), 2.30 (s, OAc), 3.83 (d, $J = 7$, $\text{C}_3\text{-H}$), 4.08 (m, $\text{C}_2\text{-H}$), 4.30 and 4.16 (d, $J = 9$, C_{20}H_2), 4.47 (m, $\text{C}_7\text{-H}$), 4.95 (d, $J = 9$, $\text{C}_5\text{-H}$), 5.66 (d, $J = 7$, $\text{C}_2\text{-H}$), 6.33 and 6.26 (2t, $J = 9$, $\text{C}_{13}\text{-H}$), 6.32 (s, $\text{C}_{10}\text{-H}$), 8.05, 7.61, 7.48 (OBz). MS (FAB) m/z : 689 (MH^+), 671, 629, 611, 569, 551, 527, 509, 449, 121 ($\text{CH}_3\text{-CHOH-CHOH-COOH} + \text{H}^+$).

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REFERENCES

1. M.C. Wani, H.L. Taylor, M.E. Wall, P. Coggon, A.T. McPhail, J. Amer. Chem. Soc., **93**, 2325 (1971).
2. M. Jacrot, J. Riodel, F. Picot, D. Leroux, C. Mouriquand, H. Bériel, P. Potier, C. R. Acad. Sc., **297**, série III, 597 (1983).
3. P.B. Schiff, J. Fant, S.B. Horwitz, Nature, **277**, 665 (1979).
4. P.B. Schiff, S.B. Horwitz, Proc. Natl. Acad. Sci. USA, **77**, 1561 (1980).
5. J.A. Snyder, R.J. McIntosh, Annu. Rev. Biochem., **45**, 699 (1976).
6. J.B. Olmsted, G.G. Borisy, Biochemistry, **12**, 4282 (1973).
7. J. Parness, D.G.I. Kingston, R.G. Powell, C. Harracksingh, S.B. Horwitz, Biochem. Biophys. Res. Comm., **105**, 1082 (1982) and references cited therein.
8. V. Sénilh, S. Blechert, M. Colin, D. Guénard, F. Picot, P. Potier, P. Varenne J. Nat. Prod., **47**, 131 (1984). D.P. Della Casa de Marcano, T.G. Halsall, J. Chem. Soc. Chem. Comm., 365 (1975).
9. G. Chauvière, D. Guénard, F. Picot, V. Sénilh, P. Potier, C. R. Acad. Sc., **293**, série II, 501 (1981).
10. H. Neh, S. Blechert, W. Schnick, M. Jansen, Angew. Chem. Int. Ed. Engl., **23**, 905 (1984).

11. J.R. McLaughlin, R.W. Miller, R.G. Powell, C.R. Smith, J. Nat. Prod., 44, 312 (1981).
12. V. Sénilh, Doctorat Thesis, Orsay, (1984).
13. D.G.I. Kingston, private communication.
14. V. Sénilh, F. Guéritte, D. Guénard, M. Colin, P. Potier, C. R. Acad. Sc., 299, série II, 1039 (1984).
15. T.B. Windholz, D.B.R. Johnston, Tetrahedron Lett., 2555 (1967).
16. B. Lythgoe, in "the Alkaloids" (Manske), 10, 597 (1968).
17. J.W. Harrison, R.M. Scrowston, B. Lythgoe, J. Chem. Soc., 1966, 1933 (1966).
18. E. Herranz, S.A. Biller, K.B. Sharpless, J. Amer. Chem. Soc., 100, 3596 (1978).