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Synthesis of C-disaccharide analogues of the α -D-arabinofuranosyl- $(1 \rightarrow 5)$ - α -D-arabinofuranosyl motif of mycobacterial cell walls via alkynyl intermediates

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Abstract—Two *C*-glycosides related to the recurrent α -D-arabinofuranosyl-(1 \rightarrow 5)- α -D-arabinofuranosyl structural motif of mycobacterial arabinan have been prepared by routes involving acetylenic intermediates. © 2005 Elsevier Ltd. All rights reserved.

Tuberculosis remains one of the major infectious diseases worldwide, responsible for around two million deaths annually, particularly in the developing world.¹ Additionally, the synergism of the disease with HIV and the appearance of multi-drug resistant strains of the causative agent, *Mycobacterium tuberculosis*, raises the prospect that tuberculosis may become prevalent as a disease only curable with great difficulty in developed nations.²

In a search for new drug targets, the mycobacterial cell envelope³ is attractive. Major components of the cell wall are two polysaccharide structures, lipoarabinomannan (LAM) and arabinogalactan (AG), this latter structure being covalently linked to the bacterial peptidoglycan and esterified as the non-reducing termini of the arabinan component with long-chain fatty acids (mycolic acids).

In seeking to block the biosynthesis of AG^4 and/or LAM,⁵ the glycosyl transferases responsible for assembly of the arabinan components of both these polymers⁶ are appealing targets for inhibitor design, given that the D-arabinose units are exclusively in their furanose form, and the xenobiotic status of the furanose form of D-arabinose. It is known that arabinofuranosyl transfer to the growing arabinan from the sugar donor β -D-ara-

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binofuranosyl-1-monophosphoryldecaprenol is inhibited by one major anti-tuberculosis agent in clinical use, ethambutol.⁷

The most common recurrent structural motif in mycobacterial arabinan is the α -D-arabinofuranosyl-(1 \rightarrow 5)- α -D-arabinofuranosyl unit **1**. *C*-Glycosides are close analogues of oligosaccharide structures, which are resistant to chemical and enzymic degradation.⁸ We here describe direct routes for the synthesis of two *C*-glycosides **2** and **3** related to motif **1**, by the use of acetylenic intermediates. An alternative route to **2** using a Wittig-based approach had recently been reported,⁹ whilst a synthesis of **3** using the Henry reaction as a key step has been claimed.¹⁰

For the synthesis of 2 (see Scheme 1) the lactone 4, prepared by oxidation¹¹ of commercially available tri-O-benzyl-D-arabinofuranose, was treated with lithio(trimethylsilyl)ethyne at low temperature to give the lactol 5 (66%). Treatment of this with triethylsilane and BF_3 . Et_2O gave the separable isomers 6α (60%) and 6β (13%).¹² That the major isomer 6α was the expected ¹³ isomer in which hydride delivery had occurred cis to the vicinal benzyloxy group was confirmed by the observation of an NOE interaction between H-1' and H-3', which was absent in the spectrum of 6β , which however showed a strong interaction between H-1' and H-2'. Desilylation of 6α gave 7 (97%), which again showed an NOE interaction between H-1' and H-3', confirming the stereochemistry of this material. The synthesis could then be completed by an iterative route. Treatment of 7 with *n*-BuLi, followed by addition of lactone 4 gave the



Scheme 1. Reagents and conditions: (a) TMS-acetylene, *n*-BuLi, THF, -78 °C; (b) BF₃·Et₂O, Et₃SiH, DCM, -78 °C; (c) K₂CO₃, MeOH; (d) BuLi, THF, -78 °C, then 4; (e) Pd(OH)₂/C, H₂ (1 atm), MeOH–EtOAc (5:1).

hemiacetal 8 (85%), which on reduction with Et₃SiH and BF₃:Et₂O gave the disubstituted alkyne 9 (83%) as the major product. The stereochemistry of 9 was clear from its NMR spectra,¹⁴ which indicated C_2 symmetry, and the observation of NOEs between H-5/8 and both H3/ 10 and (weakly) H-1/12. Reduction-hydrogenolysis of 9 then gave the *C*-disaccharide 2^{15} in near-quantitative yield.

For the synthesis of the methyl glycoside **3** (see Scheme 2), methyl 2,3-di-*O*-benzyl- α -D-arabinofuranoside (**10**)¹⁶ was prepared by modifications of the original route.¹⁷ Swern oxidation of **10**, followed by treatment of the aldehyde with CBr₄–Ph₃P gave the dibromoalkene **11** (73% overall), which on treatment with *n*-BuLi gave alkyne **12** (75%).¹⁸ Reaction of the lithio-derivative of **12** with lactone **4** gave hemiacetal **13** (87%), which on treatment with Et₃SiH and BF₃·Et₂O gave **14** α (66%) and **14** β (20%),¹⁹ separable by chromatography. The stereochemistry of the major product **14** α was indicated by an NOE between H-7 and H-9; by comparison, the minor prod-

uct 14 β showed an NOE between H-7 and H-10. Additionally, the signal for C-8 in 14 β was significantly shielded (δ 83.4) as compared with the equivalent signal for 14 α (δ 88.8), a correlation, which was also found for alkynes 6 α and 6 β ,¹² and for 7 and its β -isomer. Both 14 α and 14 β showed C-3 in the region of δ 88. For 14 α , the signals for H-8 (δ 4.23), and to a smaller extent, H-10 (δ 4.25), were also deshielded by the anisotropic effect of the alkyne as compared with the equivalent signals for 14 β (δ 4.00 and 4.06, respectively). It should also be noted that alkyne 9 showed signals for positions 4/9 at $\delta_{\rm C}$ 88.8 and $\delta_{\rm H}$ 4.23.

Catalytic hydrogenation of 14α gave the *C*-disaccharide **3** in quantitative yield. The data for 3^{20} was similar to, but significantly different from, that noted in the previous report¹⁰ on the synthesis of this compound. However, in the ¹H NMR data of **3**, NOEs were observed between the multiplet for H-5/6 and both H-8 and H-3, in support of its stereostructure. Given these discrepancies, 14α was subjected to diimide reduction to give



Scheme 2. Reagents and conditions: (a) i. DMSO, $(COCl)_2$, DCM, $-78 \circ C$, then Et_3N ; ii. CBr_4 , Ph_3P , DCM, $0 \circ C$; (b) *n*-BuLi, THF, $-78 \text{ to } 0 \circ C$; (c) *n*-BuLi, THF, $-78 \circ C$, then 4; (d) $BF_3 \cdot Et_2O$, Et_3SiH , DCM, $-78 \circ C$; (e) $Pd(OH)_2/C$, H_2 (1 atm), MeOH; (f) dipotassium azodicarboxylate, MeOH–pyridine, then HOAc.

the saturated compound 15^{21} (83%), which had also been reported by the previous workers.¹⁰ This compound, as prepared here, had spectroscopic data in full agreement with its structure, but at considerable variance from the data reported earlier.¹⁰ In the ¹H NMR data of 15, NOEs were again observed between H-5/6 and both H-8 and H-3. We thus must question the previous report on the synthesis of 3, and suggest that our work constitutes the first definitive synthesis of this compound.

Acknowledgements

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- 12. Selected data: 6α : $[\alpha]_D$ +43.4 (*c* 1.0, CHCl₃); δ_H (400 MHz, CDCl₃): 0.20 (9H, s, SiMe₃), 3.58 (1H, dd, *J* 10.6, 4.9, H-5'a), 3.63 (1H, dd, *J* 10.6, 4.3, H-5'b), 4.04 (1H, dd, *J* 5.8, 3.5, H-3'), 4.20-4.27 (2H, m, H-2', H-4'), 4.49-4.60 (5H,

m, OCH₂Ph), 4.68 (1H, d, J 11.6, OCH₂Ph), 4.70 (1H, d, J 4.2, H-1'), 7.20–7.40 (15H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃): -0.2 (SiMe₃), 69.7 (C-5'), 72.05, 72.1 (2×OCH₂Ph), 72.3 (C-1'), 73.4 (OCH₂Ph), 81.2 (C-4'), 84.2 (C-3'), 88.8 (C-2'), 91.8, 102.8 (alkyne), 127.6-128.4 (Ph), 137.4, 137.9. 138.1 (Ph, q). Compound **6** β : [α]_D – 39.8 (*c* 1.3, CHCl₃); δ _H (400 MHz, CDCl₃): 0.10 (9H, s, SiMe₃), 3.59 (1H, dd, J 10.0, 6.6, H-5'a), 3.69 (1H, dd, J 10.0, 5.9, H-5'b), 3.93 (1H, dd, $J_{3',4'}$ 3.7, $J_{3',2'}$ 1.9, H-3'), 3.98 (1H, dd, $J_{2',1'}$ 4.1, $J_{2',3'}$ 1.8, H-2'), 4.02 (1H, ddd, J 6.6, 6.0, 3.7, H-4'), 4.40-4.62 (5H, m, OCH₂Ph), 4.68 (1H, d, J 4.1, H-1'), 4.69 (1H, d, J 12.4, OCH₂Ph), 7.20–7.40 (15H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃): -0.3 (SiMe₃), 70.3 (C-5'), 71.66 (OCH_2Ph) , 71.7 (C-1'), 72.0, 73.3 $(2 \times OCH_2Ph)$, 82.3 (C-4'), 83.3 (C-2'), 84.6 (C-3'), 93.0, 100.1 (alkyne), 127.6-128.4 (Ph), 137.6, 137.9, 138.2 (Ph, q).

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- 14. Selected data for **9**: $\delta_{\rm H}$ (400 MHz, CDCl₃): 3.59 (2H, dd, J_{gem} 10.6, $J_{1a,2}/J_{12a,11}$ 5.0, H-1a/12a), 3.70 (2H, dd, J_{gem} 10.6, $J_{1b,2}/J_{12b,11}$ 4.4, H-1b/12b), 4.03 (2H, dd, $J_{3,2}/J_{10,11}$ 5.6, $J_{3,4}/J_{10,9}$ 2.9, H-3/10), 4.23 (2H, t, $J_{4,3}/J_{9,10} \sim J_{4,5}/J_{9,8} \sim 3.2$, H-4/9), 4.26 (2H, dt, $J_{2,3}/J_{11,10}$ 5.6, $J_{2,1a}/J_{11,12a} \sim J_{2,1b}/J_{11,12b} \sim 4.9$, H-2/11), 4.48 (2H, d, J 11.7, OCH₂Ph), 4.49 (2H, d, J 12.1, OCH₂Ph), 4.53 (2H, d, J 12.1, OCH₂Ph), 4.54 (2H, s, OCH₂Ph), 4.56 (2H, s, OCH₂Ph), 4.61 (2H, d, J 11.7, OCH₂Ph), 4.78 (2H, d, J 3.2, H-5/8), 7.23-7.36 (30H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃): 69.7 (C-1/12), 72.0 (OCH₂Ph), 72.1 (OCH₂Ph), 72.2 (C-5/8), 73.4 (OCH₂Ph), 81.5 (C-2/11), 84.20 (C-3/10), 84.24 (C-6/7), 88.8 (C-4/9), 127.5-128.40 (Ph), 137.38, 137.81, 138.05 (Ph, q); m/z (ES) 848.4150 [(M+NH₄)⁺; C₅₄H₅₈NO₈ requires 848.41].
- 15. Selected data for 2: $\delta_{\rm H}$ (400 MHz, D₂O): 1.78 (4H, m, H-6/7), 3.69 (2H, dd, J_{gem} 12.3, $J_{1a,2}/J_{12a,11}$ 5.9, H-1a/12a), 3.75 (2H, dd, J_{gem} 12.3, $J_{1b,2}/J_{12b,11}$ 3.5, H-1b/12b), 3.83–3.90 (2H, br m, H-5/8), 3.89 (2H, td, $J_{2,3}/J_{11,10} \sim J_{2,1a}/J_{11,12a} \sim 6.2$, $J_{2,1b}/J_{11,12b}$ 3.5, H-2/11), 3.93 (2×1H, t, $J_{4,5}/J_{9,8} \sim J_{2,3}/J_{9,10} \sim 6.2$, H-4/9), 4.03 (2H, t, $J_{3,2}/J_{10,11} \sim J_{3,4}/J_{10,9}$ 6.2, H-3/10); $\delta_{\rm C}$ (100 MHz, D₂O): 28.68 (C-6/7), 61.65 (C-1/12), 77.24 (C-3/10), 80.67 (C-4/9), 82.20 (C-5/8), 82.55 (C-2/11); m/z (ES) 295.1387 [(M+H)⁺; C₁₂H₂₃O₈ requires 295.1387].
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- 18. Selected data for **12**: $[\alpha]_{D}$ +60.8 (*c* 1.085, DCM); δ_{H} (200 MHz, CDCl₃): 2.62 (1H, d, *J* 2.1, H-6), 3.40 (3H, s, OCH₃), 3.98 (1H, dd, $J_{2,3}$ 3.3, $J_{2,1}$ 1.2, H-2), 4.15 (1H, dd, $J_{3,4}$ 6.6, $J_{3,2}$ 3.3, H-3), 4.48 (1H, d, *J* 11.6, Bn), 4.55 (1H, dd, *J* 11.6, Bn), 4.60 (1H, d, *J* 11.6, Bn), 4.65 (1H, dd, $J_{4,3}$ 6.6, $J_{4,6}$ 2.1, H-4), 4.71 (1H, d, *J* 11.6, Bn), 4.96 (1H, d, *J* 1.2, H-1), 7.25–7.45 (10H, m, Ph); δ_{C} (50 MHz, CDCl₃): 55.2 (OCH₃), 70.6 (C-6), 72.0 (OCH₂Ph), 72.5 (OCH₂Ph), 75.1 (C-5), 80.7 (CH), 87.8 (CH), 87.9 (CH), 107.2 (C-1), 127.8,

128.4 (Ph), 137.29 (Ph, q); m/z (ES) 356.1847 [(M+NH₄)⁺; C₂₁H₂₆NO₄ requires 356.1856].

- 19. Selected data: 14 α : $\delta_{\rm H}$ (400 MHz, CDCl₃): 3.37 (3H, s, OCH₃), 3.59 (1H, dd, J_{gem} 10.6, J_{11a,10} 5.0, H-11a), 3.63 (1H, dd, J_{gem} 10.6, $J_{11b,10}$ 5.0, H-11b), 3.94 (1H, dd, $J_{2,3}$ $3.5, J_{2,1} = 1.5, H-2$, $4.02 (1H, dd, J_{9,10} 5.9, J_{9,8} 3.2, H-9)$, 4.09 (1H, ddd, J_{3,4} 7.1, J_{3,2} 3.5, J_{3,1} 0.6, H-3), 4.23 (1H, t, $J_{8,7} \sim J_{8,9} \sim 3.2$, H-8), 4.25 (1H, t, $J_{10,9} \sim J_{10,11} \sim 5.0$, H-10), 4.44–4.65 (10H, m, Bn), 4.67 (1H, dd, J_{4,3} 7.1, J_{4,7} 1.5, H-4), 4.80 (1H, dd, $J_{7,8}$ 3.5, $J_{7,4}$ 1.5, H-7), 4.92 (1H, dd, $J_{1,2}$ 1.5, $J_{1,3}$ 0.5, H-1), 7.22–7.36 (25H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃): 55.2 (OCH₃), 69.7 (C-11), 70.8 (C-4), 72.0, 72.1 (OCH₂Ph), 72.2 (C-7), 72.6, 73.4 (OCH₂Ph), 81.6 (C-10), 83.8 (alkyne), 84.2 (C-9), 84.4 (alkyne), 87.9 (C-3), 88.2 (C-2), 88.8 (C-8), 107.2 (C-1), 127.6-128.5 (Ph), 137.35, 137.43, 137.47, 137.8, 138.1 (Ph, q); m/z (ES) 758.3693 $[(M+NH_4)^+; C_{47}H_{52}NO_8 \text{ requires } 758.3687].$ Compound 14 β : $\delta_{\rm H}$ (400 MHz, CDCl₃): 3.36 (3H, s, OCH₃), 3.59 (1H, dd, J_{gem} 10.0, J_{11a,10} 6.8, H-11a), 3.68 (1H, dd, J_{gem} 10.0, J_{11b,10} 5.9, H-11b), 3.93-3.96 (2H, m, H-2 and H-9), 4.00 (1H, dd, J_{8,7} 4.1, J_{8,9} 2.1, H-8), 4.06 (1H, dt, $J_{10,11a} \sim J_{10,11b} \sim 6.2$, $J_{10,9}$ 3.8, H-10), 4.12 (1H, ddd, J_{3,4} 6.8, J_{3,2} 3.5, J_{3,1} 0.6, H-3), 4.44–4.68 (10H, m, Bn), 4.71 (1H, dd, J_{4,3} 6.8, J_{4,7} 1.5, H-4), 4.76 (1H, dd, J_{7,8} 4.1, $J_{7,4}$ 1.5, H-7), 4.91 (1H, dd, $J_{1,2}$ 1.5, $J_{2,3}$ 0.6, H-1), 7.21–7.36 (25H, m, Ph); $\delta_{\rm C}$ (100 MHz, $\overline{\rm CDCl_3}$): 55.1 (OCH₃), 70.4 (C-11), 71.0 (C-4), 71.4 (C-7), 71.7, 72.1, 72.3, 72.4, 73.3 (OCH₂Ph), 82.1 (alkyne), 82.3 (C-10), 83.4 (C-8), 84.3 (C-9), 84.8 (alkyne), 87.6 (C-3), 88.1 (C-2), 107.2 (C-1), 127.6-128.4 (Ph), 137.44, 137.45, 137.6, 137.7, 138.1 (C).
- 20. Selected data for 3: $\delta_{\rm H}$ (400 MHz, D₂O): 1.76–1.85 (4H, m, H-5, H-6), 3.42 (3H, s, OCH₃), 3.70 (1H, dd, J_{gem} 12.3, $J_{11a,10}$ 5.8, H-11a), 3.77 (1H, dd, J_{gem} 12.3, $J_{11b,10}$ 3.4, H-11b), 3.84 (1H, ddd, $J_{3,4}$ 6.0, $J_{3,2}$ 3.3, $J_{3,1}$ 0.4, H-3), 3.86–3.90 (1H, m, H-7), 3.89–3.93 (1H, m, H-10), 3.94 (1H, dd, $J_{8,7}$ 6.7, $J_{8,9}$ 5.7, H-8), 3.40–4.01 (1H, m, H-4), 4.04 (1H, dd, $J_{9,10}$ 6.5, $J_{9,8}$ 5.8, H-9), 4.05 (1H, dd, $J_{2,3}$ 3.3, $J_{2,1}$ 1.6, H-2), 4.91 (1H, dd, $J_{1,2}$ 1.6, $J_{1,3}$ 0.4, H-1); $\delta_{\rm C}$ (100 MHz, D₂O) 28.8 (CH₂), 28.9 (CH₂), 55.2 (OCH₃), 61.7 (C-11), 77.3 (C-9), 80.4 (C-3), 80.7 (C-8), 81.5 (C-2), 82.1 (C-7), 82.6 (C-10), 83.5 (C-4), 108.4 (C-1); m/z (ES) 312.1658 [(M+NH₄)⁺; C₁₂H₂₆NO₈ requires 312.1653].
- 21. Selected data for **15**: $\delta_{\rm H}$ (400 MHz, CDCl₃): 1.62–1.89 (4H, m, H-5, H-6), 3.35 (3H, s, OMe), 3.52 (1H, dd, J_{gem} 10, $J_{11a,10}$ 5.9, H-11a), 3.59 (1H, dd, J_{gem} 10, $J_{11b,10}$ 5.6, H-11b), 3.63 (1H, dd, $J_{3,4}$ 6.8, $J_{3,2}$ 2.9, H-3), 3.81 (1H, dd, J 4.1, 2.6, H-8), 3.93 (1H, dd, $J_{2,3}$ 2.9, $J_{2,1}$ 0.9, H-2), 3.98–4.03 (3H, m, H-7, H-9, H-4), 4.19 (1H, dt, $J_{10,11a} \sim J_{10,11b} \sim 5.3$, $J_{10,9}$ 4.1, H-10), 4.45–4.60 (10H, m, $5 \times OCH_2$ Ph), 4.88 (1H, s, H-1), 7.22–7.38 (25H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃): δ 29.39 (CH₂), 29.45 (CH₂), 54.6 (OCH₃), 70.3 (C-11), 71.7, 71.8 (OCH₂Ph), 72.0, 72.2, 73.3 (OCH₂Ph), 80.5 (C-4), 81.2 (C-10), 82.1 (C-9), 85.4 (C-7), 87.2 (C-3), 87.8 (C-8), 88.6 (C-2), 106.7 (C-1), 127.6–128.42 (Ph), 137.6, 137.8, 137.91, 137.94, 138.2 (Ph, q); *m*/z (ES) 762.4002 [(M+NH₄)⁺; C₄₇H₅₆NO₈ requires 762.4000].