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Note

Synthesis of a jojoba bean disaccharide

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Abstract

A synthesis of the disaccharide recently isolated from jojoba beans, $2-O-\alpha$ -D-galactopyranosyl-D-*chiro*-inositol, has been achieved. The suitably protected *chiro*-inositol unit was prepared by an enantiospecific synthesis from L-xylose utilizing SmI₂-mediated pinacol coupling as a key step. © 1998 Elsevier Science Ltd. All rights reserved

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Naturally occurring galactosylcyclitols are believed to play an important role in plant physiology, particularly in seed desiccation tolerance [1]. Several of these natural products incorporate the optically active inositol isomer D-*chiro*-inositol. Examples include ciceritol (1) [2], galactopinitol (2) [3], and the disaccharide recently isolated from jojoba beans, 2-O- α -D-galactopyranosyl-D-*chiro*-inositol (3) [4].

The identification of glycosylated *chiro*-inositol in GPI protein membrane anchors [5] and the demonstration of the insulin mimetic activity of some of these compounds [6] have provided additional impetus to develop practical strategies for the synthesis of *chiro*-inositol-containing oligosaccharides [7]. However, the poor availability of differentially protected *chiro*-inositols, has impeded such efforts, and, therefore, has been addressed by several groups including ours [8]. Herein, we demonstrate the utility of our approach by applying it to the first synthesis of the disaccharide **3** from jojoba beans (Scheme 1).

Octadiene 4, prepared in seven steps from commercially available L-xylose [8], was subjected to

ozonolysis followed by SmI2-mediated pinacol cyclization to obtain chiro-inositol 5 as the sole cyclitol product detected in the reaction mixture (40% from 4). The structure of 5 was assigned by ¹H NMR analysis of its diacetyl derivative. The high diastereoselectivity of this cyclization is consistent with previously established patterns [8] and further demonstrates the strength of the technique. Selective *p*-methoxybenzylation of the equatorial hydroxyl group via the dibutylstannyl ester of 5 afforded 6 without any detectable amount of the axially alkylated isomer (77%). The structure of **6** was established by ¹H NMR analysis of its acetyl derivative. Benzylation of 6 followed by oxidative removal of the *p*-methoxybenzyl group provided acceptor 7 (59% for two steps). To confirm the assigned structures of 5 and 6 a small sample of 7 was methylated to produce the enantiomer of the previously synthesized pentabenzyl-(-)-quebrachitol (10) [8]. Synthetic 10 was identical to that obtained from the natural material by ¹H NMR.

Glycosylation of 7 was accomplished by treatment with trichloroacetimidate 8 [9] (2 equivalent) in the presence of 0.15 equivalent of Me₃SiOTf, to produce the α -disaccharide 9 in 70% yield

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(together with 18% recovered 7). No β anomer was found in the reaction mixture. Finally, hydrogenolysis afforded pure 3. Synthetic 3 was identical to a sample of the natural material by ¹H and ¹³C NMR.

1. Experimental

General methods.—All reactions, with the exception of ozonolysis and hydrogenolysis, were

performed under an atmosphere of argon. Solvents were removed in vacuo on a Büchi rotary evaporator. Solvents and reagents obtained from commercial sources were used without further purification with the following exceptions. Tetrahydrofuran (THF) and ether were distilled prior to use from sodium benzophenone ketyl; CH₂Cl₂ was distilled from CaSO₄; pyridine and benzene were distilled from CaH₂; Ac₂O was fractionally distilled. Anhydrous reactions were performed with material dried by repeated coevaporation with



Scheme 1. (a) O₃, CH₂Cl₂, Py, -78 °C, 15 min; DMS, rt, 5 h; (b) SmI₂, *t*-BuOH, THF, -78°C, 3 h; (c) Bu₂SnO, benzene, reflux; PMBCl, TBABr, reflux, 45 min; (d) NaH, BnBr, DMF, 12 h; (e) DDQ, CH₂Cl₂-H₂O, rt, 1.5 h; (f) TMSOTf, ether, -78 °C, 10 min; (g) H₂ (40 psi), Pd black, THF–ethanol–H₂O, 30 h; (h) NaH, MeI, DMF, 1 h.

toluene. TLC and preparative TLC were performed using Baker glass-backed silica gel plates (0.25 mm thickness) with 254-nm fluorescent indicator. The chromatograms were visualized by (a) ultraviolet illumination and (b) dipping in the Hanes–Isherwood solution $(1 \text{ g of } (\text{NH}_4)_6 \text{Mo}_7)$ O_{24} ·4H₂O, 10 mL of 1N HCl, 3 mL of HClO₄ in 90 mL of H₂O) followed by heating. Flash chromatography was performed on Baker silica gel (40 mesh). Ozone was generated using an ozonator purchased from Ozone Pure Water, Inc. (model 2HD). NMR spectra were recorded on a Bruker AM300 spectrometer using Me₄Si as an internal standard for ¹H in CDCl₃, DOH (4.65 ppm) for ¹H in D₂O, and 1,4-dioxane (67.4 ppm) for ${}^{13}C$ in D₂O. Solutions of SmI_2 were titrated with I_2 prior to use.

1,2,3,4-Tetra-O-benzyl-D-chiro-inositol (5).—Due to the low ozone flux of the ozonator 400 mg (0.75 mmol) of **4** were ozonolyzed in eight portions. In each case a solution containing 50 mg of 4 and $20\,\mu\text{L}$ of pyridine in $3\,\text{mL}$ of CH_2Cl_2 was treated with ozone at -78 °C until TLC (8:2 hexane-EtOAc) showed complete disappearance of the starting material ($R_f 0.6$), at which point 200 μ L of Me₂S were added. All eight solutions were kept at rt for 5h, pooled and treated with water (15mL). After the separation of the organic layer the aqueous phase was extracted with additional CH₂Cl₂ (15 mL). The combined organic layers were dried (MgSO₄) and evaporated to dryness. The residue was diluted with t-butanol (0.22 mL, 2.25 mmol) in THF (25 mL) and added dropwise over a period of 30 min to a cold (-78 °C) solution of SmI₂ (5.1 mmol) in THF (85 mL). The mixture was stirred for 3 h at -78 °C and then overnight at rt. Sat. NaHCO₃ solution (30 mL) was added and the white slurry was extracted with EtOAc ($2 \times 50 \text{ mL}$). The organic layer was washed with 10% Na₂S₂O₃ (50 mL), sat. NaCl (50 mL), and dried (MgSO₄). Evaporation of the solvent and flash chromatography (7:3 hexane-EtOAc) afforded pure 5 (160 mg, 40% yield); ¹H NMR (CDCl₃): δ 7.4–7.22 (m, 20 H, aromatic), 5.02, 4.81 (2d, 2 H, J_{gem} 11.3 Hz, CH₂Ph), 4.99, 4.65 (2d, 2 H, J_{gem} 10.5 Hz, CH₂Ph), 4.81, 4.65 (2d, 2 H, J_{gem} 10.7 Hz, CH₂Ph), 4.72, 4.61 (2d, 2 H, J_{gem} 11.7 Hz, CH₂Ph), 4.07-3.86 (m, 5 H, CH-OR), 3.64 (*\psi*t, 1 H, J_{3.4} 9.1 Hz, H-3 or H-4), 2.31, 2.32 (2 br d, 2 OH); FAB HRMS (NBA/NaI) m/z 563.2434, M + Na⁺ calcd for $C_{34}H_{36}O_6$ 563.2411. A sample of 5 (5 mg) was treated with Ac₂O (10 μ L) and pyridine (0.2 mL) for 2h at rt. After aqueous work-up, 5mg of

sufficiently pure diacetate **5** was obtained; ¹H NMR (CDCl₃, inositol numbering is the same as in **5**): δ 7.42–7.28 (m, 20 H, aromatic), 5.41 (ψ t, 1 H, $J_{5,6}$, $J_{6,1}$ 3.6 Hz, H-6), 5.34 (dd, 1 H, $J_{4,5}$, 9.5 Hz, $J_{5,6}$ 3.6 Hz, H-5), 4.93, 4.82 (2d, 2 H, J_{gem} 10.4 Hz, CH₂Ph), 4.86, 4.70 (2d, 2 H, J_{gem} 11.6 Hz, CH₂Ph), 4.73 (s, 2 H, CH₂Ph), 4.69, 4.57 (2d, 2 H, J_{gem} 11.7 Hz, CH₂Ph), 4.04 (ψ t, 1 H, $J_{2,3}$, $J_{3,4}$ 9.5 Hz, H-3), 3.79 (ψ t, 1 H, $J_{3,4}$, $J_{4,5}$ 9.5 Hz, H-4), 3.72 (dd, 1 H, $J_{1,2}$ 3.6, $J_{2,3}$ 9.5 Hz, H-2), 3.70 (ψ t, 1 H, $J_{6,1}$, $J_{1,2}$ 3.6 Hz, H-1), 2.00 (s, 3 H, OAc), 1.97 (s, 3 H, OAc).

1,2,3,4-Tetra-O-benzyl-5-O-p-methoxybenzyl-Dchiro-inositol (6).—A suspension of 5 (20 mg, $37 \,\mu \text{mol}$) and dibutyltin oxide (10 mg, 41 μmol) in 20 mL of benzene was fitted with a distillation head and placed in an oil bath at 110 °C until most of the benzene had distilled. An additional portion of benzene (10 mL) was added to the residue and the mixture was heated until again most of the benzene had distilled. The reaction mixture was cooled and treated with *p*-methoxybenzyl chloride $(7.5 \,\mu\text{L},$ $55 \,\mu \text{mol}$) and tetrabutylammonium bromide $(13 \text{ mg}, 41 \,\mu\text{mol})$. The mixture was then heated at reflux for 45 min more, then NaHCO₃ (2 mL of 1 M solution) was added and the mixture was extracted with CH_2Cl_2 (3×5mL). The combined organic extracts were dried (MgSO₄) and evaporated to dryness. Preparative TLC (1:1 hexanesether) gave 18.7 mg of 6 (77% yield); ¹H NMR (CDCl₃): δ 7.35–7.2 (m, 22 H, aromatic), 6.85–6.78 (d, 2 H, aromatic), 4.94–4.47 (m, 10 H, CH₂Ar), 3.94 (dd, 1 H, J 9.3, 8.8 Hz, H-3 or H-4), 3.95–3.90 (m, 2 H, CHOR), 3.85 (dd, 1 H, J 10.3, 2.6 Hz, H-2 or H-5), 3.80–3.72 (m, 2 H, CHOR), 3.79 (s, 3 H, OCH₃), 2.41 (br s, 1 H, OH); Anal calcd for C₄₂H₄₄O₇: C, 76.34; H, 6.71. Found: C, 75.98; H, 6.84. A solution of 6 (3.3 mg, 5 μ mol) in pyridine (192 μ L), was treated with Ac₂O (3.3 μ L, 35 μ mol). After aqueous work-up, 2.7 mg of acetylated product was obtained; ¹H NMR (CDCl₃, inositol numbering is the same as in 6): δ 7.35–7.2 (m, 22 H, aromatic), 6.85–6.78 (d, 2 H, aromatic), 5.32 (\u03c6t, 1 H, J_{6,1} and J_{5,6} 3.4 Hz, H-6), 4.93–4.40 (m, 10 H, CH₂Ar), 3.97–3.85 (m, 2 H, CHOR), 3.77 (s, 3 H, OCH₃), 3.78–3.68 (m, 4 H, CHOR), 2.0 (s, 3 H, OAc).

1,2,3,4,6-Penta-O-benzyl-D-chiro-inositol (7).—A solution of **6** (14.2 mg, 21.5 μ mol) in DMF (190 μ L) at 0 °C was treated with NaH (3.4 mg of a 60% oil dispersion, 86 μ mol). After 0.5 h, the mixture was treated with benzyl bromide (7.7 μ L, 65 μ mol) and allowed to warm to room temperature.

After overnight stirring, the mixture was cooled to 0 °C, and water (1 mL) was added. The aqueous layer was extracted with CH_2Cl_2 (3×5mL) and the combined organic extracts were dried (MgSO₄) and concentrated under diminished pressure. To the residue was added CH_2Cl_2 (544 μ L), H_2O (60 μ L) and DDQ (5.2 mg, 22 μ mol) and the mixture was stirred at room temperature for 1.5 h. Sat. NaHCO₃ solution (1 mL) was added, the layers were separated and the aqueous phase was extracted with CH_2Cl_2 (3×5mL) the combined organic extracts were dried (MgSO₄) and evaporated to dryness. Preparatative TLC (2:1 hexanes-ether) gave 7.9 mg of 7 (58% yield for two steps); ¹H NMR (CDCl₃): δ 7.35–7.2 (m, 25 H, aromatic), 4.96–4.29 (m, 10 H, CH₂Ph), 3.97 (dd, 1 H, J 3.4, 9.2 Hz, H-2 or H-5), 3.93 (dd, 1 H, J 9.3, 9.2 Hz, H-3 or H-4), 3.79 (dd, 1 H, J 2.8, 9.3 Hz, H-2 or H-5), 3.75 (dd, 1 H, J 3.8, 3.4 Hz, H-1 or H-6) 3.71 (dd, 1 H, J 3.8, 2.8 Hz, H-1 or H-6), 3.65 (dd, 1 H, J 9.3, 9.2 Hz, H-3 or H-4); FAB HRMS (NBA/NaI) m/z 653.2883, M + Na⁺ calcd for $C_{41}H_{42}O_6$ 653.2880.

To confirm unequivocally the structure of 7, 2.6 mg of 7 was treated with NaH (1 mg) and MeI (12 μ L) in DMF (0.1 mL) for 1 h. Aqueous workup gave 2.6 mg of **10**; ¹H NMR (CDCl₃): δ 7.5–7.12 (m, 25 H, aromatic), 4.93–4.76 (m, 5 H, CH₂Ph), 4.67, 4.48 (2d, 2 H, J_{gem} 11.8 Hz, CH₂Ph), 4.63, 4.41 (2d, 2 H, J_{gem} 12.3 Hz, CH₂Ph), 4.32 (d, 1 H, J_{gem} 12.2 Hz, CH₂Ph), 3.92–3.77 (m, 3 H, CH-OR), 3.71 (dd, 1 H, J 3.0, 4.0 Hz, CH-OR), 3.66 (dd, 1 H, J 2.7, 4.0 Hz, CH-OR), 3.55 (dd, 1 H, J 3.0, 9.7 Hz, CH-OR), 3.40 (s, 3 H, OCH₃).

1,3,4,5,6-Penta-O-benzyl-2-O-(2,3,4,6-tetra-Obenzyl- α -D-galactopyranosyl)-D-chiro-inositol (9). To a cold (-78 °C) solution of 7 (15 mg, 24 μ mol) and 8 (35 mg, 51 μ mol) in dry ether (1.5 mL) was added trimethylsilyl triflate (0.65 μ L, 3.6 μ mol). The mixture was stirred at -78 °C for 10 min and allowed to warm to room temperature. Sat. aqueous NH₄Cl (1 mL) was added and the mixture was extracted with ether $(2 \times 1.5 \text{ mL})$. The ethereal layers were combined, dried (MgSO₄), and evaporated to dryness. The residue was purified by preparative TLC (4:1 hexane–EtOAc) to give 19 mg (70%) yield) of pure 9 (R_f 0.5) and 2.5 mg of recovered 7 $(R_f 0.3)$; ¹H NMR (CDCl₃) δ 7.35–7.2 (m, 43 H, aromatic), 7.0-6.96 (m, 2 H, aromatic), 5.07 (d, 1 H, $J_{1'2'}$ 3.5 Hz, H-1'), 4.91–4.40 (m, 18 H, CH₂Ph), 4.39–3.42 (m, 12 H, CH-OR); Anal calcd for C₇₅H₇₆O₁₁: C, 78.10; H, 6.64. Found: C, 77.64; H, 6.74.

2-O- α -D-Galactopyranosyl-D-chiro-inositol (3).— A solution of **9** (15 mg, 13 μ mol) in THF–ethanol– H₂O (1:1:1) was hydrogenolyzed with Pd black (28 mg) at 40 psi for 30 h. The mixture was filtered through Celite and concentrated to produce **3** (4.4 mg, 100% yield); m.p. 140–180 °C (dec) [lit. 165–170 (dec)] [4], ¹H and ¹³C NMR (D₂O) were identical of those of an authentic sample of natural **3** [4], generously provided by Professor S. Kondo.

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References

- [1] R.L. Obendorf, Seed Sci. Res., 7(2) (1997) 63-74.
- [2] M. Bernabe, R. Fenwick, J. Frias, J. Jimenez-Barbero, K. Price, S. Valverde, and C. Vidal-Valverde, J. Agr. Food. Chem., 41(6) (1993) 870– 872.
- [3] C.T. Chien, T.P. Lin, C.G. Juo, and G.R. Her, *Plant Cell Phys.*, 37(4) (1996) 539–544.
- [4] K. Ogawa, T. Watanabe, Y. Ikeda, and S. Kondo, *Carbohydr. Res.*, 302 (1997) 219–221.
- [5] Y. Pak and J. Larner, Biochem. Biophys. Res. Commun., 184 (1992) 1042–1047.
- [6] J.M. Mato, K.L. Kelly, A. Abler, L. Jarett, B.E. Corkey, J.A. Cashel, and D. Zopf, *Biochem. Biophys. Res. Commun.*, 146 (1987) 764–770; J. Larner, L.C. Huang, C.F.W. Schwartz, A.L. Oswald, T.Y. Shen, M. Kinter, G. Tang, and K. Zeller, *Biochem. Biophys. Res. Commun.*, 151 (1988) 1416–1426; I. Asplin, G. Galasko, and J. Larner, *Proc. Natl. Acad. USA*, 90 (1993) 5924– 5928; R.V. Farese, M.L. Standaert, K. Yamada, L.C. Huang, C. Zhang, D.R. Cooper, Z. Wang, Y. Yang, S. Suzuki, T. Toyota, and J. Larner, *Proc. Natl. Acad. USA*, 91 (1994) 11040–11044; M.C. Fonteles, L.C. Huang, and J. Larner, *Diabetologia*, 39 (1996) 731–734.
- [7] C. Jaramillo, J.L. Chiara, and M. Martin-Lomas, J. Org. Chem., 59 (1994) 3135–3141; K.K. Reddy, J.R. Falck, and J. Capdevila, Tetrahedron. Lett., 34 (1993) 7869–7872; M.M. Silva, J. Cleophax, A.A. Benicio, M.V. Almeida, J.M. Delaumeny, A.S. Machado, and S.D. Gero, Synlett, 8 (1996) 764.
- [8] A. Kornienko and M. d'Alarcao, *Tetrahedron Lett.*, 38 (1997) 6497–6500, and references cited therein.
- [9] B. Wegmann and R.R. Schmidt, J. Carbohydr. Chem., 6 (1987) 357–375.