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Graphical Abstract

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Synthesis of glycosphingolipids from the fungus *Hirsutella rhossiliensis*

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Synthesis of glycosphingolipids from the fungus *Hirsutella rhossiliensis*

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ABSTRACT

The total synthesis of two neutral glycosphingolipids (GSLs) from the fungus *Hirsutella rhossiliensis* has been achieved. The GSLs possess a common neogala-core (Gal β 1-6Gal) and have the following sequence: α -D-Man $p(1\rightarrow 3)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\leftrightarrow 1)$ Cer (1) and

 α -D-Man $p(1 \rightarrow 3)$ - β -D-Gal $p(1 \rightarrow 6)$ [α -D-Glc $p(1 \rightarrow 4)$]- β -D-Gal $p(1 \rightarrow 6)$ - β -

1. Introduction

Many glycosphingolipids (GSLs) sequences isolated from vertebrates terminate in sialic acid residues. In recent years, the synthesis of these GSLs and the understanding of the biological functions has attracted a lot of attention by various research groups¹. In contrast, research on GSLs isolated from invertebrates has been neglected. There are only limited reports which describe naturally occurring sequences of GSLs in invertebrates²⁻²¹. These structures are significantly different from GSLs isolated from vertebrates. Moreover, the biological function of GSLs in invertebrates and fungi are unknown. For this reason we have been interested in the synthesis of glycolipids derived from various invertebrate sources in order to clarify their biological functions²²⁻³⁷.

Isolation and purification of glycolipids from invertebrate species are very difficult and the amount available is extremely small. As a result, controlled regio- and stereoselective synthetic approaches are required to produce sufficient amounts of homogenous material to explore biological structure/function relationships. Once the synthetic methodology has been developed this approach can also be applied to produce non-natural glycolipid analogs to exploit or manipulate their biological functions.

Tani et al. isolated and characterized three kinds of new GSLs, α -D-Man $p(1\rightarrow 3)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\leftrightarrow 1)$ Cer (**1**), α -D-Man $p(1\rightarrow 3)$ - β -D-Gal $p(1\rightarrow 6)[\alpha$ -D-Glc $p(1\rightarrow 4)$]- β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\leftrightarrow 1)$ Cer r (**2**), and α -D-Glc $p(1\rightarrow 2)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\leftrightarrow 1)$ Cer (**3**), which have neogala-series, β -D-Galp(1-6)- β -D-Galp(1-6)- β -D-Gal $p(1\leftrightarrow 1)$ Cer (**3**), which have neogala-series, β -D-Galp(1-6)- β -D-Galp(1-6)- β -D-Galp, as core structure from the nematophagous fungus *Hirsutella rhossiliensis*³⁸. Compound **3** was also isolated from mold, *Neurospora crassa* and the synthesis of compound **3** has been completed by Otsuka et al²². In this paper we describe our efforts to prepare GSLs (1) and (2). We were particularly interested to develop a synthetic scheme which minimizes the number of protection and deprotection steps by exploiting the different reactivities of hydroxyl groups in galactose. This approach is expected to reduce the number of synthetic steps while at the same time improving the overall yield of the desired products.

2. Result and discussion

2.1 Total synthesis of glycosphingolipid 1

The tetrasaccharide glycosphingolipid 1 contains the neogala-core sequence $[\beta$ -D-Gal $p(1\rightarrow 6)$ - β -D-Gal $p(1\rightarrow 6)$ - β -D-Galp] was prepared by stepwise synthesis of galactosyl donors and acceptors (Scheme 1). Galactopyranosyl donor 4 and 5 were obtained from phenyl 4,6-O-benzylidene-1-thio- β -D-galactopyranoside (3)³⁹. Regioselective chloroacetylation and subsequently benzoylation of 3 provided donor 4^{40} . Benzoylation of the two free hydroxyl groups of **3** using standard condition provided 5^{41} . We envisaged to achieve regioselective 6-O-glycosylation of galactoside acceptor 6unprotected at both C-4 and C-6 by taking advantage of the greater steric hindrance between primary and secondary (axia) hydroxyl groups as well as using deactivating 2.3-O-benzoyl protecting groups. Disaccharide 7 was synthesized by selective glycosylation of diol-based glycosyl acceptor 6^{42} with thiogalactosyl donor 5 using *N*-iodosuccinimde (NIS)/ trifluoromethanesulfonic acid (TfOH) as promoter^{43,44}. The β -linkage in 7 was confirmed by ¹H-NMR spectroscopy. The nature of the new glycosidic linkage was determined by coupling constant of anomeric proton (H-1', $\delta =$ 4.94, J = 8.1 Hz). This reaction was achieved by using small amount of donor 5 (1.1 equv.) which reacted regioselectively with the 6-OH of acceptor 6. In addition, the glycosylation with the 6-OH was evidenced by HMBC correlation between the signal of C-6 at $\delta = 66.9$ and H-1' at $\delta = 4.94$. The 4-OH group in 7 was acetylated and the benzylidene was cleaved by treatment with 80% AcOH to produce diol 8 in 72% yield. Comparing the ¹H-NMR data of **7** with those of **8** showed that H-4 signal of Gal a residue was shifted downfield by 1.49 ppm. This also indicates that the Gal b was bound at the 6-position of Gal a. The same previously described regioselective glycosylation strategy was used to generate trisaccharide 9 from diol 8. NIS/TfOH promoted glycosylation of thiogalactoside donor 4 with acceptor 8 provided trisaccharide 9 in 78% yield as the only product. The anomeric proton of newly established anomeric

center appeared as a doublet at $\delta = 7.83$ (J = 8.2 Hz) consistent with the expected β -linkage. Trisaccharide acceptor **10** was synthesized by hydrolysis of the chloroacetyl group in **9** with aqueous pyridine in 92% yield.



Scheme 1. Reagents and conditions: (a) see ref. 40; (b) benzoyl chloride, pyridine, 87%; (c) NIS, TfOH, 4Å MS, CH₂Cl₂, **7** 86%, **9** 78%; (d) 1) Ac₂O, pyridine, 2) 80% AcOH, 72% (two steps); (e) pyridine, H₂O, 92%.

Regioselective glycosylation of thiomannosyl donor 11^{45} with the trisaccharide diol-acceptor 10 in the presence of NIS and TfOH gave desired disaccharide 12. As expected, the more reactive equatorial hydroxyl group at 3-position of acceptor 10 was successfully glycosylated. However, we were unable to isolate any other glycosylation product derived from glycosylation of the axial hydroxyl group indicating the highly regioselective nature of this glycosylation. The α -configuration of the new glycosidic linkage in 12 was indicated by the $J_{C-1',H-1'}$ value of 170 Hz. Furthermore, the binding to the 3-position was indicated by HMBC between the signal of C-3 of Gal c at $\delta = 74.8$ and H-1 of Man at $\delta = 5.10$. The successful selective introduction of mannose residue to reactive 3-OH makes this strategy very attractive and avoids time-consuming protection and deprotection steps. Removal of the benzylidene of 12 with 80% AcOH and acetylation gave protected tetrasaccharide 13 in 92% yield over two steps. Comparing the ¹H-NMR data of **12** with those of **13** showed that H-4 signal of Gal b residue was shifted downfield by 1.30 ppm, so it was confirmed that the mannose was bound to 3-position of the Gal c. Selective removal of 2-(trimethylsilyl)ethyl (TMS-ethyl) group in 13 with trifluoroacetic acid (TFA) in CH₂Cl₂, followed by treatment with CCl₃CN in

the presence of 1,8-diazabicyclo[5,4,0]-undeca-7-ene (DBU)⁴⁶ afforded corresponding α -trichloroacetimidate **14** in 90% yield over two steps. Glycosylation of phytoceramide acceptor (2*S*,3*R*,4*R*)-3,4-di-*O*-benzoyl-2-hexadecanamido-octadecane-1,3,4-triol **15**⁴⁷ with glycosyl donor **14** in the presence of trimethylsilyl trifluoromethanesulfonate (TMSOTf)⁴⁸ afford desired protected glycosphingolipid **16** in 56% yield. Finally, Zemplén-based deacylation of **16** and purification by column chromatography on Sephadex LH-20 produced glycosphingolipid **1** (Scheme 2) in 84% yield. The structure and purity of **1** were demonstrated by its ¹H NMR and HR-FABMS data.



Scheme 2. Reagents and Conditions: a) NIS, TfOH, 4Å MS, CH₂Cl₂, 75%; b) 1) 80%AcOH, 2) Ac₂O, pyridine, 92% (two steps); c) 1) TFA, CH₂Cl₂, 2) CCl₃CN, DBU, CH₂Cl₂, 90% (two steps); d) TMSOTf, 4Å MS, CH₂Cl₂, 56%; e) NaOMe, 1,4-dioxane/MeOH, 84%.

2.2 Total synthesis of glycosphingolipid 2

The pentasaccharide glycosphingolipid 2 contains differs from 1 by addition of a glucose residue to the unreactive axial hydroxyl group in tetrasaccharide 12. The pentasaccharide portion of glycolipid 2 was synthesized by glycosylation of reactive perbenzylated thioglucoside donor 17^{49} with tetrasaccharide acceptor 12. We studied

various reaction conditions varying solvent and temperature to optimize the desired α -selectivity in this reaction. Using a toluene/1,4-dioxane mixture⁵⁰ as solvent at -20°C resulted in complete α -selectivity in this reaction to produce **18** α in 77% yield. (Scheme 3. Table). The α -linkage was assigned on the homonuclear coupling constant (br. d) of newly anomeric proton signal at $\delta = 5.10$ and carbon signal at $\delta = 99.4$. The β -linkage in **18\beta** was assigned on proton signal $\delta = 4.36$ (d, J = 7.6 Hz) and carbon signal at $\delta = 103.8$.



Scheme 3.

Table α -glycosylation under various conditions

Ent.	Solvent	Temp.	Yield	α:β
1	CH ₂ Cl ₂	$-40^{\circ}C$	83%	1.2:1
2	$CH_2Cl_2/ether = 5/1$	$-20^{\circ}C$	79%	5:1
3	toluene/1,4-dioxane = $\frac{3}{1}$	$-20^{\circ}C$	77%	Only α

Removal of benzylidene and benzyl groups in 18α using catalytic hydrogenolysis over 10% Pd(OH)₂/C in THF/MeOH/AcOH followed by acetylation provided *O*-acetylated intermediate **19**. Compound **19** was exposed to TFA to produce the hemiacetal which was converted into α -trichloroacetimidate **20** in quantitative yield. TMSOTf-promoted glycosylation of phytoceramide acceptor **15** with glycosyl imidate donor **20** produced desired β -glycoside **21** in 42% yield. Finally standard deacylation of **21** and purification by column chromatography on Sephadex LH-20 afforded glycosphingolipid **2** (Scheme 4). The structure and purity of **2** were demonstrated by its ¹H NMR, ¹³C NMR and HR-FABMS data.



Scheme 4. Reagents and Conditions: a) 1) Pd(OH)₂-C/H₂, THF/MeOH/AcOH, 2) Ac₂O, Pyr, 86% (two steps); b) 1) TFA, CH₂Cl₂, 2) CCl₃CN, DBU, CH₂Cl₂, quant. (two steps); c) TMSOTf, 4Å MS, CH₂Cl₂, 42%; d) NaOMe, 1,4-dioxane/MeOH, 88%.

3. Conclusion

In summary, a systematic and integrated approach for the synthesis of two naturally occurring glycosphingolipids 1 and 2 from the fungus *Hirsutella rhossiliensis* has been accomplished. The synthetic strategy described may also be useful for the synthesis of related GSLs from other species. Biological activities of glycosphingolipids 1 and 2 is currently in progress, and the results will be reported in detail elsewhere.

4. Experimental

4.1 General

Optical rotations were measured with a Jasco P-1020 digital polarimeter. ¹H and ¹³C

NMR spectra were recorded with a JEOL 400 FT NMR spectrometer. Me₄Si or acetone were used as internal standards for CDCl₃, CD₃OD and D₂O. MALDI-TOFMS was recorded on an SHIMADZU AXIMA-CFR Plus mass spectrometer. High-resolution mass spectra were recorded on a JEOL T100LP under ESI conditions. TLC was performed on Silica Gel 60 F254 (E. Merck) with detection by quenching of UV fluorescence and by charring with 10% H₂SO₄ in EtOH. Column chromatography was carried out on Silica Gel 60N 100–210 μ m (KANTO KAGAKU). Ultra Pack A (ϕ 11 x 300, Silica gel 40 μ m, YAMAZEN), Ultra Pack B (ϕ 26 x 300, Silica gel 40 μ m, YAMAZEN), were used for flash column chromatography.

Phenyl 4,6-*O*-benzylidene-1-thio- β -D-galactopyranoside (**3**)³⁹, 2-(trimethylsilyl)ethyl 2,3-di-*O*-benzoyl- β -D-galactopyranoside (**6**)⁴², phenyl

2,3,4,6-tetra-*O*-acetyl-1-thio- α -D-mannopyranoside (**11**)⁴⁵, phenyl

2,3,4,6-tetra-*O*-benzyl-1-thio- β -D-glucopyranoside (**17**)⁴⁹ were prepared as reported. Benzoylceramide **15** was prepared from phytosphingosine, which was purchased from Degussa (The Netherlands) by the conventional four-step procedure.⁴⁷

4.2. Phenyl 2,3-di-O-benzoyl-4,6-O-benzylidene-β-D-galactopyranoside (5)

To a solution of **3** (600 mg, 1.67 mmol) in pyridine (10.0 mL) was added BzCl (0.58 mL, 5.00 mmol), and the mixture was stirred for 1.5 h at 0°C. The mixture diluted with CHCl₃, washed with 10% aq. HCl, sat. aq. NaHCO₃, and brine, dried (MgSO₄) and concentrated. The residue was purified by silica gel column chromatography (ϕ 50 x 130 mm) using 4:1 hexane-AcOEt to give **5** (824 mg, 87%) as a white solid.

[α]_D²⁵ -20.1 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.99–7.23 (m, 20H, 4 x Ph), 5.81 (t, 1H, $J_{1,2} = J_{2,3} = 9.9$ Hz, H-2), 5.51 (s, 1H, CHPh), 5.36 (dd, $J_{3,4} = 3.2$ Hz, 1H, H-3), 4.96 (d, 1H, $J_{1,2} = 9.8$ Hz, H-1), 4.59 (d, 1H, $J_{3,4} = 3.2$ Hz, H-4), 4.54 (dd, $J_{5,6a} = 1.6$ Hz, $J_{6a,6b} = 12.4$ Hz, 1H, H-6a), 4.09 (dd, 1H, $J_{5,6b} = 1.5$ Hz, $J_{6a,6b} = 12.5$ Hz, H-6b), 3.75 (br. d, 1H, J = 0.9 Hz, H-5); ¹³C NMR (100 MHz, CDCl₃): δ 166.1, 164.9, 137.5, 133.8, 133.3, 133.1, 131.0, 130.1, 129.9, 129.7, 129.6, 129.0, 128.7, 128.4, 128.3, 128.2, 128.1, 126.4, 100.9, 85.2 (C-1), 74.0, 73.6, 69.8, 69.1, 67.0.

HR-FABMS: calcd for C₃₃H₂₈O₇SNa: *m/z* 591.1453; found: *m/z* 591.1478 [M+Na]⁺.

4.3.

2-(Trimethylsilyl)ethyl

2,3-di-O-benzoyl-4,6-O-benzylidene- β -D-galactopyranosyl-(1 \rightarrow 6)-2,3-di-O-benzoyl- β -D-galactopyranoside (7)

A mixture of acceptor 2-(trimethylsilyl)ethyl 2,3-di-*O*-benzoyl- β -D-galactopyranoside (**6**) (2.30 g, 4.71 mmol), **5** (2.94 g, 5.18 mmol) and 4Å MS (4.50 g) in dry CH₂Cl₂ (25.0 mL) was stirred for 3 h at room temperature, then cooled to -60° C. NIS (1.75 g, 7.75 mmol) and TfOH (69.6 µL, 0.78 mmol) was added, and the mixture was stirred for 1 h at -60° C, then neutralized with Et₃N. The precipitates were filtrated off and washed with CHCl₃. The combined filtrate and washings were successively washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, brine, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography (ϕ 40 x 170 mm) using 3:1 hexane-AcOEt to give **7** (3.83 g, 86%) as an amorphous powder.

 $[\alpha]_D^{25}$ +103.9 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.01–7.27 (m, 25H, 5 x Ph), 5.92 (dd, 1H, $J_{1,2} = 8.1$ Hz, $J_{2,3} = 10.3$ Hz, H-2 of Gal b), 5.71 (dd, 1H, $J_{1,2} = 8.0$ Hz, $J_{2,3} = 10.3$ Hz, H-2 of Gal a), 5.56 (s, 1H, >CH–Ph), 5.42 (dd, 1H, $J_{2,3} = 10.2$ Hz, $J_{3,4} = 3.5$ Hz, H-3 of Gal b), 5.25 (dd, 1H, $J_{2,3} = 10.2$ Hz, $J_{3,4} = 3.2$ Hz, 1H, H-3of Gal a), 4.94 (d, 1H, $J_{1,2} = 8.1$ Hz, H-1 of Gal b), 4.62 (d, 1H, $J_{3,4} = 3.5$ Hz, H-4 of Gal b), 4.58 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal a), 4.42 (dd, 1H, $J_{5,6a} = 1.1$ Hz, $J_{6a,6b} = 12.6$ Hz, H-6a of Gal b), 4.29 (br. t, 1H, J = 3.4 Hz, H-4 of Gal a), 4.21 (dd, 1H, $J_{5,6a} = 6.5$ Hz, $J_{6a,6b} =$ 10.6 Hz, H-6a of Gal a), 4.15 (dd, 1H, $J_{5,6b} = 1.6$ Hz, $J_{6a,6b} = 12.4$ Hz, H-6b of Gal b), 4.03 (dd, 1H, $J_{5,6b} = 6.4$ Hz, $J_{6a,6b} = 10.7$ Hz, H-6b of Gal a), 3.92–3.86 (m, 2H, H-5 of Gal a, CH₂CH₂SiMe₃), 3.74 (s, 1H, H-5 of Gal b), 3.45–3.38 (m, 1H, CH₂CH₂SiMe₃), 2.78 (d, 1H, $J_{OH,4} = 5.3$ Hz, 4-OH of Gal a), 0.85–0.62 (m, 2H, CH₂CH₂SiMe₃), -0.12 (s, 9H, CH₂CH₂Si*Me*₃); ¹³C NMR (100 MHz, CDCl₃): δ 166.1, 165.6, 165.4, 165.2, 137.4, 133.3, 133.2, 133.1, 132.9, 129.9, 129.8, 129.7, 129.6, 129.4, 129.4, 128.99, 128.96, 128.9, 128.33, 128.32, 128.27, 128.13, 128.08, 126.2, 100.8 (C-1 of Gal b), 100.73, 100.65 (C-1 of Gal a), 74.1, 73.5, 73.4, 72.6, 69.8, 68.84, 68.81, 67.3, 67.0, 66.9, 66.5, 17.6, -1.50.

HR-FABMS: calcd for C₅₂H₅₄O₁₅SiNa: *m/z* 969.3130; found: *m/z* 969.3060 [M+Na]⁺.

4.4. 2-(Trimethylsilyl)ethyl 2,3-di-*O*-benzoyl-β-D-galactopyranosyl-(1→6)-4-*O*-acetyl-2,3-di-*O*-benzoyl-β-D-gal actopyranoside (8)

To a solution of 7 (3.83 g, 4.04 mmol) in pyridine (15.0 mL) was added Ac_2O (10 mL) at 0°C, and the mixture was stirred for 18 h at room temperature. The mixture diluted with CHCl₃, washed with 10% aq. HCl, sat. aq. NaHCO₃, and brine, dried

(MgSO₄) and concentrated. To a solution of the residue in AcOH (24 mL) was added H_2O (6.0 mL), and the mixture was stirred for 6 h at 70°C. The residue was purified by flash column chromatography (Hi-flash column L) using 1:1 hexane-AcOEt to give **8** (2.60 g, 72%) as an amorphous powder.

[α]_D²⁵ +74.3 (*c* = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.97–7.29 (m, 20H, 4 x Ph), 5.82 (dd, 1H, $J_{1,2}$ = 7.8 Hz, $J_{2,3}$ = 10.3 Hz, H-2 of Gal b), 5.78 (d, 1H, $J_{3,4}$ = 3.4 Hz, H-4 of Gal a), 5.63 (dd, 1H, $J_{1,2}$ = 8.0 Hz, $J_{2,3}$ = 10.3 Hz, H-2 of Gal a), 5.40 (dd, 1H, $J_{2,3}$ = 10.3 Hz, $J_{3,4}$ = 3.4 Hz, H-3 of Gal a), 5.30 (dd, 1H, $J_{3,4}$ = 3.2 Hz, $J_{2,3}$ = 10.3 Hz, H-3 of Gal b), 4.78 (d, 1H, $J_{1,2}$ = 7.8 Hz, H-1 of Gal b), 4.68 (d, 1H, $J_{1,2}$ = 8.0 Hz, H-1 of Gal a), 4.45 (br. s, 1H, H-4 of Gal b), 4.07 (t, 1H, $J_{5,6a}$ = $J_{5,6b}$ = 5.9 Hz, H-5 of Gal b), 3.97–3.85 (m, 6H, H-6 of Gal a, H-6 of Gal b), CH₂CH₂SiMe₃, 4-OH of Gal b), 3.74 (t, $J_{5,6a}$ = $J_{5,6b}$ = 5.9 Hz, 1H, H-5 of Gal b), 3.49–3.42 (m, 1H, CH₂CH₂SiMe₃), 3.18 (br. s, 1H, 6-OH of Gal b), 2.13 (s, 3H, COCH₃), 0.88–0.67 (m, 2H, CH₂CH₂SiMe₃), -0.14 (s, 9H, CH₂CH₂SiMe₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.78, 165.9, 165.6, 165.3, 165.2, 133.3, 133.2, 133.1, 129.8, 129.69, 129.66, 129.5, 129.1, 128.9, 128.4, 128.4, 128.3, 128.2, 101.2 (C-1 of Gal b), 100.8 (C-1 of Gal a), 74.4, 73.9, 72.3, 72.1, 69.7, 69.6, 68.4, 68.0, 67.7, 67.2, 62.6, 10.7, 17.7, -1.54.

HR-FABMS: calcd for C₄₇H₅₂O₁₆SiNa: *m/z* 923.2983; found: *m/z* 923.2922 [M+Na]⁺.

4.5.

2-(Trimethylsilyl)ethyl

2-*O*-benzoyl-4,6-*O*-benzylidene-3-*O*-chloroacetyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -2,3-di-*O*-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-*O*-acetyl-2,3-di-*O*-benzoyl- β -D-galactopyranoside (9)

A mixture of acceptor **8** (1.44 g, 1.60 mmol), donor **4** (0.950 g, 1.76 mmol) and 4Å MS (2.00 g) in dry CH₂Cl₂ (16.0 mL) was stirred for 1.5 h at room temperature, then cooled to -60° C. NIS (593 mg, 2.63 mmol) and TfOH (23.6 µL, 0.263 mmol) was added, and the mixture was stirred for 1 h at -60° C, then neutralized with Et₃N. The precipitates were filtrated off and washed with CHCl₃. The combined filtrate and washings were successively washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, brine, dried (MgSO₄), and concentrated. The product was purified by flash column chromatography (Ultra pack B) using 4:3 hexane-AcOEt to give **9** (1.65 g, 78%) as an amorphous powder.

 $[\alpha]_D^{25}$ +49.7 (*c* = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.99–7.15 (m, 30H, 6 x Ph), 5.70–5.63 (m, 2H, H-2 of Gal b, H-2 of Gal c), 5.60–5.56 (m, 2H, H-2, 4 of Gal a), 5.23 (s, 1H, >CH–Ph), 5.31–5.24 (m, 2H, H-3 of Gal a, H-3 of Gal c), 5.20 (dd, 1H, *J*_{2,3})

= 10.6 Hz, $J_{3,4}$ = 3.2 Hz, H-3 of Gal b), 4.83 (d, 1H, $J_{1,2}$ = 8.2 Hz, H-1 of Gal c), 4.67 (d, 1H, $J_{1,2}$ = 7.7 Hz, H-1 of Gal c), 4.39 (br. d, 1H, $J_{1,2}$ = 7.8 Hz, H-1 of Gal a), 4.47 (br. d, 1H, $J_{3,4}$ = 3.3 Hz, H-4 of Gal c), 4.39 (br. d, 1H, J = 11.5 Hz, H-6a of Gal c), 4.28 (br. s, 1H, H-4 of Gal b), 4.17–4.10 (m, 2H, H-6a of Gal b, H-6b of Gal c), 4.02 (d, 1H, J = 15.6 Hz, COC H_2 Cl), 3.99–3.91 (m, 3H, H-5, 6b of Gal b, COC H_2 Cl), 3.87–3.73 (m, 4H, H-5, 6 of Gal a, CH_2 CH₂SiMe₃), 3.67 (br. s, 1H, H-5 of Gal c), 3.40–3.33 (m, 1H, C H_2 CH₂SiMe₃), 2.85 (s, 1H, 4-OH of Gal b), 2.02 (s, 3H, COC H_3), 0.81–0.59 (m, 2H, CH₂CH₂SiMe₃), -0.16 (s, 9H, CH₂CH₂SiMe₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.0, 167.2, 165.6, 165.4, 165.3, 165.2, 165.1, 137.2, 133.5, 133.2, 133.03, 133.01, 129.8, 129.74, 129.67, 129.63, 129.58, 129.5, 129.19, 129.15, 129.0, 128.6, 128.33, 128.31, 128.29, 128.24, 128.22, 128.2, 126.3, 125.3, 101.1 (C-1 of Gal b), 101.0, 100.6 (C-1 of Gal a), 100.5 (C-1 of Gal c), 73.7, 73.5, 73.4, 73.1, 72.7, 71.9, 69.7, 69.5, 68.7, 68.6, 68.0, 67.5, 67.2, 66.8, 66.4, 66.0, 40.6, 21.4, 20.5, 17.6, -1.50.

HR-FABMS: calcd for C₆₉H₇₁ClO₂₃SiNa: m/z 1353.3661; found: m/z 1353.3653 [M+Na]⁺.

4.6.

2-(Trimethylsilyl)ethyl

$\label{eq:2-O-benzoyl-4,6-O-benzylidene-$\beta-D-galactopyranosyl-(1 \rightarrow 6)-2,3-di-$O-benzoyl-$\beta-D-galactopyranosyl-(1 \rightarrow 6)-4-$O-acetyl-2,3-di-$O-benzoyl-$\beta-D-galactopyranoside (10)$

To a solution of **9** (322 mg, 0.242 mmol) in pyridine (6.0 mL) was added H₂O (1.2 mL), and the mixture was stirred for 18 h at 70°C. The mixture diluted with CHCl₃, washed with 10% aq. HCl, sat. aq. NaHCO₃, and brine, dried (MgSO₄) and concentrated. The residue was purified by silica gel column chromatography (ϕ 30 x 150 mm) using 3:2 toluene-acetone to give **10** (280 mg, 91%) as an amorphous powder.

[α]_D²⁵ +29.1 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.06–7.16 (m, 30H, 6 x Ph), 5.70 (dd, 1H, $J_{1,2} = 7.8$ Hz, $J_{2,3} = 10.3$ Hz, H-2 of Gal b), 5.61–5.57 (m, 3H, H-2, 4 of Gal a, >CH–Ph), 5.36 (dd, 1H, $J_{1,2} = 8.0$ Hz, $J_{2,3} = 9.9$ Hz, H-2 of Gal c), 5.20 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} = 3.4$ Hz, H-3 of Gal a), 5.22 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} = 3.2$ Hz, H-3 of Gal b), 4.77 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal c), 4.67 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1 of Gal b), 4.57 (d, 1H, $J_{1,2} = 8.1$ Hz, H-1 of Gal a), 4.38 (br. d, 1H, J = 11.7 Hz, H-6a of Gal c), 4.28 (br. d, 2H, J = 3.0 Hz, H-4 of Gal b, H-4 of Gal c), 4.14–4.10 (m, 3H, H-6a of Gal b, H-6 of Gal c), 4.01–3.76 (m, 7H, H-5, 6 of Gal a, H-5, 6b of Gal b, H-3 of Gal c, $CH_2CH_2SiMe_3$), 3.61(br. s, 1H, H-5 of Gal b), 2.66 (d, 1H, $J_{OH,3} = 11.0$ Hz, 3-OH of Gal c), 2.03 (s, 3H, COCH₃), 0.81–0.59 (m, 2H, CH₂CH₂SiMe₃), -0.16 (s,

9H, CH₂CH₂Si*Me*₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 166.3, 165.6, 165.4, 165.17, 165.15, 137.2, 133.2, 133.04, 133.02, 129.82, 129.76, 129.7, 129.63, 129.58, 129.5, 129.3, 129.1, 128.99, 128.95, 128.5, 128.33, 128.28, 128.22, 128.18, 126.3, 101.5, 101.1 (C-1 of Gal b), 100.6 (C-1 of Gal a), 100.4 (C-1 of Gal c), 75.5, 73.7, 72.8, 72.5, 71.9, 71.5, 69.7, 69.6, 67.8, 68.1, 67.5, 67.4, 66.9, 66.8, 66.1, 20.5, 17.6, -1.49. HR-FABMS: calcd for C₆₇H₇₀O₂₂SiNa: *m*/*z* 1277.3873; found: *m*/*z* 1277.3826 [M+Na]⁺.

4.7.

4.8.

2-(Trimethylsilyl)ethyl

2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -2-*O*-benzoyl-4,6-*O*-benzylidene- β -D-galactopyranosyl- $(1\rightarrow 6)$ -2,3-di-*O*-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-*O*-ac etyl-2,3-di-*O*-benzoyl- β -D-galactopyranoside (12)

Compound **12** was prepared from **10** (1.19 g, 0.95 mmol) and **11** (459 mg, 1.04 mmol) as described for preparation of **7**. The product was purified by silica gel column chromatography (5:2 toluene-acetone) to give **12** (1.13 g, 75%).

 $[\alpha]_{D}^{25}$ +60.8 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.10–7.31 (m, 30H, 6 x Ph), 5.73 (dd, 1H, *J*_{1,2} = 8.0 Hz, *J*_{2,3} = 10.3 Hz, H-2 of Gal b), 5.64–5.58 (m, 4H, H-2, 4 of Gal a, H-2 of Gal c, >CH-Ph), 5.33 (dd, 1H, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 3.4$ Hz, H-3 of Gal a), 5.25 (dd, 1H, J_{2.3} = 10.2 Hz, J_{3.4} = 2.9 Hz, H-3 of Gal b), 5.21–5.14 (m, 3H, H-2, 3, 4 of Man), 5.10 (br. s, 1H, H-1 of Man), 4.85 (d, 1H, J_{1,2} = 8.0 Hz, H-1 of Gal c), 4.69 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal b), 4.61 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal a), 4.47–4.40 (m, 2H, H-4, 6a of Gal c), 4.33 (d, 1H, J_{3,4} = 2.9 Hz, H-4 of Gal b), 4.21–4.12 (m, 3H, H-6a of Gal b, H-3, 6b of Gal c), 4.01-3.76 (m, 8H, H-5, 6 of Gal a, H-5, 6b of Gal b, H-5, 6 of Man, 4-OH of Gal b, CH₂CH₂SiMe₃), 3.62(br. s, 1H, H-5 of Gal c), 3.44–3.37 (m, 1H, CH₂CH₂SiMe₃), 2.13, 2.08, 2.07, 1.94, 1.77 (each s, each 3H, COCH₃), 0.85–0.63 (m, 2H, CH₂CH₂SiMe₃), -0.13 (s, 9H, CH₂CH₂SiMe₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 170.03, 169.98, 169.5, 169.1, 165.6, 165.3, 165.2, 165.1, 164.9, 149.6, 137.2, 133.3, 133.2, 133.1, 133.0, 132.9, 129.8, 129.7, 129.63, 129.60, 129.55, 129.5, 129.2, 129.0, 128.9, 128.4, 128.3, 128.3, 128.2, 126.2, 126.1, 123.7, 101.0 (C-1 of Gal b), 100.8, 100.54 (C-1 of Gal a), 100.45 (C-1 of Gal c), 94.2 ($J_{C,H} = 170$ Hz, C-1 of Man), 74.8, 73.7, 73.2, 72.7, 71.9, 71.6, 69.9, 69.72, 69.66, 69.6, 68.9, 68.6, 68.3, 68.0, 67.4, 67.1, 66.7, 66.6, 65.8, 65.6, 61.7, 20.8, 20.6, 20.54, 20.50, 20.4, 17.5, -1.52.

HR-FABMS: calcd for C₈₁H₈₈O₃₁SiNa: *m*/*z* 1607.4977; found: *m*/*z* 1607.4806 [M+Na]⁺.

2-(Trimethylsilyl)ethyl

$2,3,4,6\text{-tetra-}\textit{O}\text{-}acetyl\text{-}\alpha\text{-}D\text{-}mannopyranosyl\text{-}(1 \rightarrow 3)\text{-}4,6\text{-}di\text{-}\textit{O}\text{-}acetyl\text{-}2\text{-}\textit{O}\text{-}benzoyl\text{-}\beta\text{-}D$

-galactopyranosyl- $(1\rightarrow 6)$ -6-*O*-acetyl-2,3-di-*O*-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-*O*-acetyl-2,3-di-*O*-benzoyl- α -D-galactopyranoside (13)

To a solution of **12** (156 mg, 98.1 μ mol) in AcOH (2.4 mL) was added H₂O (0.6 mL) at room temperature, and the mixture was stirred for 8.0 h at 70°C. After completion of the reaction, the mixture was concentrated. To a solution of the residue in pyridine (3.0 mL) was added Ac₂O (2.0 mL), and the mixture was stirred for 18 h at room temperature. The mixture diluted with CHCl₃, washed with 10% aq. HCl, sat. aq. NaHCO₃, and brine, dried (MgSO₄) and concentrated. The residue was purified by silica gel column chromatography (ϕ 22 x 180 mm) using 8:1 toluene-acetone to give **13** (147 mg, 92%) as an amorphous powder.

 $[\alpha]_D^{25}$ +55.2 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.05–7.31 (m, 25H, 5 x Ph), 5.63 (br. d, 1H, $J_{3,4} = 3.2$, H-4 of Gal b), 5.61–5.58 (m, 2H, H-2 of Gal a, H-2 of Gal b), 5.55 (br. d, 1H, *J*_{3,4} = 3.5 Hz, H-4 of Gal a), 5.45–5.39 (m, 2H, H-2, 4 of Gal c), 5.34 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} = 2.4$ Hz, H-3 of Gal b), 5.30 (dd, 1H, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 3.5$ Hz, H-3 of Gal a), 5.12–5.07 (m, 2H, H-2, 4 of Man), 5.01 (d, 1H, $J_{1,2} = 1.6$ Hz, H-1 of Man), 4.97 (dd, 1H, $J_{2,3} = 10.4$ Hz, $J_{3,4} = 3.4$ Hz, H-3 of Man), 4.75 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1 of Gal c), 4.69 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal b), 4.60 (d, 1H, $J_{1,2} =$ 8.0 Hz, H-1 of Gal a), 4.27 (dd, 1H, $J_{5,6a} = 6.6$ Hz, $J_{6a,6b} = 11.5$ Hz, H-6a of Gal c), 4.20 (dd, 1H, $J_{5,6b} = 6.4$ Hz, $J_{6a,6b} = 11.5$ Hz, H-6b of Gal c), 4.10 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4}$ = 6.4 Hz, H-3 of Gal c), 4.03–3.65 (m, 11H, H-5, 6 of Gal a, H-5, 6 of Gal b, H-5 of Gal c, H-5, 6 of Man, CH₂CH₂SiMe₃), 3.42–3.36 (m, 1H, CH₂CH₂SiMe₃), 2.23, 2.10, 2.083, 2.06, 1.90, 1.67 (each s, each 3H, 6 x COCH₃), 2.078 (s, 6H, 2 x COCH₃), 0.84–0.62 (m, 2H, CH₂CH₂SiMe₃), -0.15 (s, 9H, CH₂CH₂SiMe₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 170.4, 170.3, 169.9, 169.7, 169.31, 169.26, 165.34, 165.26, 165.11, 165.06, 164.4, 133.4, 133.2, 133.1, 133.0, 129.8, 129.62, 129.58, 129.5, 129.4, 129.3, 128.9, 128.8, 128.5, 128.3, 128.2, 128.1, 101.0 (C-1 of Gal c), 100.8 (C-1 of Gal b), 100.6 (C-1 of Gal a), 94.9 (C-1 of Man), 73.8, 72.6, 72.5, 71.9, 71.6, 70.8, 70.0, 69.5, 69.4, 68.6, 68.52, 68.45, 67.9, 67.7, 67.4, 67.0, 65.03, 65.00, 61.51, 61.47, 29.6, 20.7, 20.6, 20.5, 20.4, 20.3, 17.5, -1.55.

HR-FABMS: calcd for C₈₀H₉₀O₃₄SiNa: *m/z* 1645.4980; found: *m/z* 1645.4910 [M+Na]⁺.

4.9.

2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -4,6-di-*O*-acetyl-2-*O*-benzoyl- β -D -galactopyranosyl- $(1\rightarrow 6)$ -6-*O*-acetyl-2,3-di-*O*-benzoyl- β -D -galactopyranosyl- $(1\rightarrow 6)$ -4-*O*-acetyl-2,3-di-*O*-benzoyl- β -D -galactopyranosyl trichloroacetimidate (14)

To a solution of **13** (136 mg, 83.8 μ mol) in CH₂Cl₂ (2.0 mL) was added CF₃CO₂H (1.0 mL) at 0°C, and the mixture was stirred for 1.5 h at 0°C. After completion of the reaction, the mixture was concentrated. To a solution of the residue in CH₂Cl₂ (2.0 mL) was added CCl₃CN (69.0 μ L, 670 μ mol) and DBU (18.8 μ L, 126 μ mol) at 0°C. The reaction mixture was stirred for 3.0 h at rt. The mixture was concentrated and purified by silica gel column chromatography (ϕ 28 x 105 mm) using 20:1 chloroform-acetone to give **14** (126 mg, 90.2%) as an amorphous powder.

 $[\alpha]_D^{25}$ +79.7 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.23 (s, 1H, NH), 8.07– 7.31 (m, 25H, 5 x Ph), 6.61 (d, 1H, $J_{1,2} = 3.2$ Hz, H-1 of Gal a), 5.79 (dd, 1H, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 3.2$ Hz, H-3 of Gal a), 5.74 (dd, 1H, $J_{1,2} = 3.2$ Hz, $J_{2,3} = 10.5$ Hz, H-2 of Gal a), 5.70 (br. d, 1H, *J*_{4,3} = 3.2 Hz, H-4 of Gal a), 5.62 (br. d, 1H, *J*_{3,4} = 3.2 Hz, H-4 of Gal b), 5.57 (dd, 1H, $J_{1,2} = 8.0$ Hz, $J_{2,3} = 10.5$ Hz, H-2 of Gal b), 5.44 (br. d, 1H, $J_{3,4} = 3.5$ Hz, H-4 of Gal c), 5.40 (dd, 1H, $J_{1,2} = 8.0$ Hz, $J_{2,3} = 10.0$ Hz, H-2 of Gal c), 5.31 (dd, 1H, *J*_{2,3} = 10.5 Hz, *J*_{3,4} = 3.2 Hz, H-3 of Gal b), 5.13–5.08 (m, 2H, H-2, 4 of Man), 5.02 (d, 1H, $J_{1,2} = 1.4$ Hz, H-1 of Man), 4.98 (dd, 1H, $J_{2,3} = 3.2$ Hz, $J_{3,4} = 10.1$ Hz, H-3 of Man), 4.71 (d, 2H, $J_{1,2} = 8.0$ Hz, H-1 of Gal b, H-1 of Gal c), 4.47 (br. t, J = 5.5, 6.4 Hz 1H, H-6a of Gal a), 4.28–4.18 (m, 2H, H-6 of Gal c), 4.12 (dd, 1H, $J_{2,3} = 10.0$ Hz, $J_{3,4} =$ 3.5 Hz, H-3 of Gal c), 4.02–3.82 (m, 6H, H-6b of Gal a, H-5, 6 of Gal b, H-5 of Gal c, H-6a of Man), 3.75–3.68 (m, 3H, H-5 of Gal a, H-5, 6b of Man), 2.25, 2.14, 2.10, 2.074, 2.069, 2.06, 1.91, 1.68 (each s, each 3H, 8 x COCH₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 170.34, 170.29, 169.6, 169.5, 169.34, 169.28, 169.2, 165.5, 165.24, 165.18, 165.0, 164.4, 160.1, 133.41, 133.37, 1333.2, 133.0, 129.74, 129.66, 129.6, 129.48, 129.45, 129.2, 129.0, 128.9, 128.8, 128.6, 128.5, 128.31, 128.27, 128.2, 128.1, 101.0 (C-1 of Gal b), 100.6 (C-1 of Gal c), 94.9 (C-1 of Man), 93.3 (C-1 of Gal a), 90.5, 73.8, 72.7, 71.6, 70.7, 70.0, 69.1, 68.6, 68.5, 68.4, 68.2, 67.6, 67.5, 66.9, 65.0, 64.5, 61.4, 20.64, 20.60, 20.51, 20.48, 20.4, 20.3.

MALDI-TOFMS: calcd for C₇₇H₇₈Cl₃NO₃₄Na: *m/z* 1688.3; found: *m/z* 1688.7 [M+Na]⁺.

4.10. (2*S*, 3*S*, 4*R*)-3,4-dibenzyloxy-2-(hexadecanoylamino)-octadecyl 2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranosyl-(1 \rightarrow 3)-4,6-di-*O*-acetyl-2-*O*-benzoyl- β -D -galactopyranosyl-(1 \rightarrow 6)-6-*O*-acetyl-2,3-di-*O*-benzoyl- β -D-galactopyranosyl-(1 \rightarrow 6) -4-*O*-acetyl-2,3-di-*O*-benzoyl- β -D-galactopyranoside (16)

A mixture of solution of acceptor (2*S*,3*R*,4*R*)-3,4-di-*O*-benzoyl-2-hexadecanamido-octadecane-1,3,4-triol (**15**) (72.2 mg,

94.4 µmol), donor 14 (105 mg, 63.0 µmol) and 4Å MS (400 mg) in dry CH₂Cl₂ (0.80 mL) was stirred for 18 h at room temperature, then cooled to 0°C. TMSOTf (9.08 μ L, 50.4 μ mol) was added, and the mixture was stirred for 3.0 h at 0°C, then neutralized with Et₃N. The precipitates were filtrated off and washed with CHCl₃. The combined filtrate and washings were successively washed with brine, dried $(MgSO_4)$, and concentrated. The product was purified by flash column chromatography (Ultra Pack A) using 1:1 hexane-ethyl acetate to give 16 (80.0 mg, 56.0%) as an amorphous powder. $[\alpha]_{D}^{25}$ +31.4 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.06–7.30 (m, 35H, 7 x Ph), 6.05 (d, 1H J = 9.1 Hz, NH), 5.59 (d, 1H, J_{3,4} = 3.2 Hz, H-4 of Gal a), 5.55–5.25 (m, 9H, H-2, 3 of Gal a, H-2, 3, 4 of Gal b, H-2, 4 of Gal c, H-3, 4 of Cer), 5.12-5.06 (m, 2H, H-2, 4 of Man), 5.02 (br. s, 1H, H-1 of Man), 4.97 (dd, 1H, J_{2,3} = 3.4 Hz, J_{3,4} = 10.1 Hz, H-3 of Man), 4.67 (d, 1H, $J_{1,2} = 8.1$ Hz, H-1 of Gal c), 4.40 (d, 1H, $J_{1,2} = 7.3$ Hz, H-1 of Gal b), 4.32–4.18 (m, 3H, H-6a of Man, H-1a, 2 of Cer), 4.13–4.08 (m, 2H, J_{1,2} = 8.0 Hz, H-1 of Gal a, H-3 of Gal c), 3.95–3.65 (m, 9H, H-5 of Gal a, H-5, 6a, of Gal b, H-5, 6 of Gal c, H-5, 6b of Man, H-1b, 2 of Cer), 3.55 (dd, 1H, $J_{5,6a} = 4.1$ Hz, $J_{6a,6b} =$ 10.5 Hz, H-6a of Gal a), 3.09 (dd, 1H, $J_{5,6b} = 2.5$ Hz, $J_{6a,6b} = 9.4$ Hz, H-6b of Gal b), 3.01 (dd, 1H, $J_{5,6b} = 7.2$ Hz, $J_{6b,6a} = 10.5$ Hz, H-6b of Gal a), 2.21, 2.10, 2.09, 2.07, 1.91, 1.67 (each s, each 3H, 6 x COCH₃), 2.06 (s, 6H, 2 x COCH₃), 2.17-0.85 (m, 60H, alkyl-H); ¹³C NMR (100 MHz, CDCl₃): δ 172.7, 170.6, 170.43, 170.36, 170.0, 169.7, 169.4, 169.3, 166.1, 165.3, 164.9, 164.5, 164.4, 133.4, 133.34, 133.25, 133.3, 133.0, 132.8, 130.3, 130.1, 129.8, 129.6, 129.5, 129.3, 129.2, 129.1, 128.9, 128.8, 128.5, 128.3, 128.2, 100.9 (C-1 of Gal a), 100.8 (C-1 of Gal c), 100.3 (C-1 of Gal b), 95.0 (C-1 of Man), 73.8, 72.5, 72.0, 71.23, 71.15, 70.9, 70.2, 70.0, 69.4, 68.64, 68.57, 68.5, 67.8, 67.59, 67.55, 66.8, 66.6, 65.1, 61.53, 61.47, 47.6, 36.3, 31.8, 59.7, 69.6, 29.5, 29.4, 29.3, 29.1, 28.6, 25.6, 25.4, 22.6, 20.70, 20.66, 20.53, 20.46, 20.3, 14.1. MALDI-TOFMS: calcd for C₁₂₃H₁₅₃NO₃₉Na: *m/z* 2291; found: *m/z* 2291 [M+Na]⁺.

4.11. (2*S*, 3*S*, 4*R*)-2-(hexadecanoylamino)-3,4-dihydroxyoctadecyl α -D-mannopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 6)-4- β -D-galactopyranoside (1)

To a solution of **16** (68.3 mg, 30.1 μ mol) in MeOH (2.0 mL) was added 1,4-dioxane (2.0 mL) and NaOMe (200 mg) at room temperature, and the mixture was stirred for 18 h at room temperature then neutralized with IR-120B (H⁺). The precipitates were filtrated off and concentrated. The product was purified by gel filtration chromatography (Sephadex LH-20, ϕ 20 x 1000 mm) using 1:1 chloroform-MeOH to

give 1 (30.5 mg, 84%) as a white solid.

[α]_D²⁵ +12.8 (c = 0.2, 1:1 CHCl₃-MeOH). ¹H NMR (400 MHz, 1:1 CDCl₃-CD₃OD): δ 4.99 (d, 1H, J = 1.1 Hz, H-1 of Man), 4.36 (d, 1H, J = 7.3 Hz, H-1 of Gal c), 4.32 (d, 1H, J = 7.3 Hz, H-1 of Gal b), 4.27 (d, 1H, J = 7.3 Hz, H-1 of Gal a); ¹³C NMR (100 MHz, CDCl₃): δ 175.5, 105.0 (C-1 of Gal b, c), 104.9 (C-1 of Gal a), 97.4 (C-1 of Man), 77.7, 76.0, 74.90, 74.87, 74.5, 74.3, 74.1, 73.8, 72.7, 72.2, 72.1, 72.0, 71.7, 70.5, 70.3, 69.8, 69.7, 69.6, 69.5, 68.4, 65.6, 62.6, 62.2, 51.4, 37.1, 32.8, 30.5, 30.5, 30.4, 30.3, 30.2, 26.9, 26.8, 23.5, 14.4.

HR-FABMS: calcd for $C_{58}H_{109}NO_{24}Na$: m/z 1226.7237; found: m/z 1226.7118 $[M+Na]^+$.

4.12.

2-(Trimethylsilyl)ethyl

2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -2-O-benzoyl-4,6-O-benzylidene- β -D-galactopyranosyl- $(1\rightarrow 6)$ -[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl- $(1\rightarrow 4)$]-2, 3-di-O-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-O-acetyl-2,3-di-O-benzoyl- β -D-gluco pyranoside (18 α)

4.13.

2-(Trimethylsilyl)ethyl

2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -2-O-benzoyl-4,6-O-benzylidene- β -D-galactopyranosyl- $(1\rightarrow 6)$ -[2,3,4,6-tetra-O-benzyl- β -D-glucopyranosyl- $(1\rightarrow 4)$]-2, 3-di-O-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-O-acethyl-2,3-di-O-benzoyl- β -D-gluc opyranoside (18 β)

(Ent. 1) A mixture of acceptor **12** (207 mg, 0.130 mmol), donor **17** (247 mg, 0.390 mmol) and 4Å MS (250 mg) in dry CH₂Cl₂ (2.0 mL) was stirred for 2 h at room temperature, then cooled to -40° C. NIS (132 mg, 0.585 mmol) and TfOH (5.24 µL, 58.5 µmol) was added, and the mixture was stirred for 1 h at -40° C, then neutralized with Et₃N. The precipitates were filtrated off and washed with CHCl₃. The combined filtrate and washings were successively washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, brine, dried (MgSO₄), and concentrated. The product was purified by silica gel column chromatography (ϕ 30 x 170 mm) using 1:1 hexane-AcOEt to give **18** β (104 mg, 38%) and **18** α (123 mg, 45%) as amorphous powders.

(Ent. 2) A mixture of acceptor **12** (220 mg, 0.138 mmol), donor **17** (263 mg, 0.414 mmol) and 4Å MS (300 mg) in dry CH_2Cl_2 (2.0 mL) and dry diethyl ether (0.4 mL) was stirred for 4 h at room temperature, then cooled to $-20^{\circ}C$. NIS (140 mg, 0.621 mmol) and TfOH (5.56 μ L, 0.0621 mmol) was added, and the mixture was stirred for 1 h at $-20^{\circ}C$, then neutralized with Et₃N. The precipitates were filtrated off and washed with

CHCl₃. The combined filtrate and washings were successively washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, brine, dried (MgSO₄), and concentrated. The product was purified by flash column chromatography (Ultra Pack B) using 5:2 hexane-AcOEt to give **18** β (38.8 mg, 13%) and **18\alpha** (189 mg, 65%) as amorphous powders.

(Ent. 3) A mixture of acceptor **12** (270 mg, 0.170 mmol), donor **17** (323 mg, 0.511 mmol) and 4Å MS (300 mg) in dry toluene (1.8 mL) and dry 1,4-dioxane (0.6 mL) was stirred for 4 h at room temperature, then cooled to -20° C. NIS (173 mg, 0.767 mmol) and TfOH (6.87 µL, 0.0767 mmol) was added, and the mixture was stirred for 1 h at -20° C, then neutralized with Et₃N. The precipitates were filtrated off and washed with CHCl₃. The combined filtrate and washings were successively washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, brine, dried (MgSO₄), and concentrated. The product was purified by flash column chromatography (Ultra Pack B) using 5:2 hexane-AcOEt to give **18** α (277 mg, 77%) as amorphous powders.

18α: $[α]_D^{25}$ +78.8 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.26–7.28 (m, 50H, 10 x Ph), 5.79–5.70 (m, 5H, H-2, 4 of Gal a, H-2 of Gal b, H-2 of Gal c, >CH-Ph), 5.46 (dd, 1H, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 3.4$ Hz, H-3 of Gal a), 5.38–5.31 (m, 5H, H-3 of Gal b, H-2, 3 of Man, 2 x benzyl methylene), 5.24–5.21 (m, 2H, H-1, 3 of Man), 5.14–5.10 (m, 2H, H-1 of Glc, benzyl methylene), 5.03 (d, 1H, benzyl methylene), 4.95–4.89 (m, 4H, H-1 of Gal c, 3 x benzyl methylene), 4.74 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal b), 4.71 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1 of Gal a), 4.56–4.47 (m, 4H, H-4, 6a of Gal b, H-4 of Gal c, benzyl methylene), 4.26–4.19 (m, 4H, H-6b of Gal b, H-3, 5 of Gal c, H-3 of Glc), 4.09-3.85 (m, 8H, H-6 of Gal a, H-5 of Gal b, H-5, 6 of Man, H-4 of Glc, $CH_2CH_2SiMe_3$, 3.82 (br. s, 2H, H-6 of Gal c), 3.65 (dd, 1H, $J_{1,2} = 3.2$ Hz, $J_{2,3} = 9.8$ Hz, H-2 of Glc), 3.55-3.41 (m, 3H, H-5, 6a of Glc, $CH_2CH_2SiMe_3$), 3.07 (br. d, 1H, J = 9.8Hz, H-6b of Glc), 2.27, 2.21, 2.19, 2.08, 1.93 (each s, each 3H, COCH₃), 0.91–0.68 (m, 2H, CH₂CH₂SiMe₃), -0.03 (s, 9H, CH₂CH₂SiMe₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 170.1, 170.0, 169.6, 169.1, 165.8, 165.3, 165.2, 165.1, 164.8, 139.0, 138.6, 138.5, 138.0, 137.3, 133.2, 133.14, 133.09, 133.0, 132.9, 129.9, 129.8, 129.7, 129.60, 129.55, 129.3, 129.0, 128.9, 128.53, 128.47, 128.28, 128.26, 128.2, 127.9, 127.8, 127.7, 127.6, 127.5, 127.44, 127.42, 127.3, 127.2, 126.2, 100.9 (C-1 of Gal a), 100.7, 100.5 (C-1 of Gal b), 100.4 (C-1 of Gal c), 99.4 (C-1 of Glc), 94.4 (C-1 of Man), 81.5, 80.2, 77.5, 75.4, 75.2, 75.1, 74.7, 74.3, 73.5, 73.44, 73.35, 73.2, 72.7, 71.9, 71.8, 71.0, 70.5, 70.0, 69.9, 69.7, 68.6, 68.4, 68.1, 67.4, 66.5, 66.2, 65.6, 61.7, 29.7, 20.9, 20.6, 20.54, 20.46, 17.5, -1.49.

MALDI-TOFMS: calcd for C₁₁₅H₁₂₂O₃₆SiNa: m/z 2129.7; found: m/z 2130.1 [M+Na]⁺. **18β**: $[\alpha]_D^{25}$ +92.5 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.07–7.09 (m, 50H,

10 x Ph), 5.73 (dd, 1H, $J_{1,2} = 8.0$ Hz, $J_{2,3} = 10.3$ Hz, H-2 of Gal b), 5.59–5.55 (m, 2H, H-2 of Gal a, >CH-Ph), 5.50-5.46 (m, 2H, H-4 of Gal a, H-2 of Gal c), 5.38-5.32 (m, 2H, H-3 of Gal b, benzyl methylene), 5.26 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} = 3.4$ Hz, H-3 of Gal a), 5.22–5.11 (m, 3H, H-2, 3, 4 of Man), 5.04 (d, 1H, $J_{1,2} = 1.4$ Hz, H-1 of Man), 4.90–4.83 (m, 3H, H-1 of Gal c, 2 x benzyl methylene), 4.74–4.69 (m, 2H, 2 x benzyl methylene), 4.67 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1 of Gal b), 4.58–4.47 (m, 4H, H-1 of Gal a, 3 x benzyl methylene), 4.36 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1 of Glc), 4.34 (br. d, 1H, J = 11.9 Hz, H-6a of Gal c), 4.24-4.19 (m, 3H, H-4, 6a of Gal b, H-4 of Gal c), 4.09-4.00 (m, 3H, H-6b of Gal b, H-3, 6b of Gal c), 3.89-3.70 (m, 8H, H-6 of Gal a, H-5 of Gal b, H-5 of Gal c, H-5, 6 of Man, CH₂CH₂SiMe₃), 3.65 (br. s, 2H, H-6 of Glc), 3.57–3.52 (m, 2H, H-2, 4 of Glc), 3.44 (d, 1H, $J_{2,3} = J_{3,4} = 8.9$ Hz, H-3 of Glc), 3.34–3.28 (m, 2H, H-5 of Gal a, CH₂CH₂SiMe₃), 3.22–3.18 (m, 1H, H-5 of Glc), 2.08, 2.06, 2.00, 1.89, 1.76 (each s, each 3H, COCH₃), 0.76–0.52 (m, 2H, CH₂CH₂SiMe₃), -0.18 (s, 9H, CH₂CH₂SiMe₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 170.13, 170.07, 169.6, 169.1, 165.6, 165.4, 165.1, 164.84, 164.77, 138.8, 138.5, 138.3, 138.0, 137.4, 133.3, 133.2, 133.0, 132.9, 130.1, 129.8, 129.63, 129.56, 129.0, 128.8, 128.6, 128.5, 128.4, 128.30, 128.26, 128.2, 128.1, 127.9, 127.8, 127.7, 127.54, 127.47, 127.3, 126.2, 103.8 (C-1 of Glc), 100.9 (C-1 of Gal a), 100.7, 100.5 (C-1 of Gal c), 100.40 (C-1 of Gal b), 94.6 (C-1 of Man), 84.6, 82.3, 77.6, 75.6, 75.2, 75.1, 75.0, 74.9, 74.6, 73.9, 73.3, 72.9, 72.8, 72.1, 71.9, 70.9, 70.1, 70.0, 69.7, 69.1, 68.9, 68.6, 68.5, 68.4, 68.1, 67.5, 67.3, 66.3, 65.6, 61.6, 20.9, 20.61, 20.56, 20.53, 20.45, 17.5, -1.47.

MALDI-TOFMS: calcd for C₁₁₅H₁₂₂O₃₆SiNa: *m/z* 2129.7; found: *m/z* 2130.0 [M+Na]⁺.

4.14.

2-(Trimethylsilyl)ethyl

2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -4,6-di-*O*-acetyl-2-*O*-benzoyl- β -D -galactopyranosyl- $(1\rightarrow 6)$ -[2,3,4,6-tetra-*O*-acetyl- α -D-glucopyranosyl- $(1\rightarrow 4)$]-2,3-di-*O*-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-*O*-acethyl-2,3-di-*O*-benzoyl- β -D-glucopyr anoside (19)

Compound **18a** (373 mg, 0.177 mmol) in THF-MeOH-AcOH (1:1:1, 12 mL) was hydrogenolysis under hydrogen (0.11 MPa) in the presence of $Pd(OH)_2/C$ (300 mg) for 18 h at room temperature, and the mixture was then filtered and concentrated. The residue was acetylated with acetic anhydride (4.0 mL) in pyridine (6.0 mL). The reaction mixture was poured into ice H₂O and extracted with CHCl₃. The extract was washed sequentially with 5% HCl, aq NaHCO₃, and brine, dried (MgSO₄), and concentrated. The residue was purified by flash column chromatography (Ultra Pack A) using 8:1 toluene-acetone to give 19 (290 mg, 85.9%) as amorphous powders. $[\alpha]_{D}^{25}$ +54.3 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.04–7.31 (m, 25H, 5 x Ph), 5.61–5.50 (m, 4H, H-2, 4 of Gal a, H-2 of Gal b, H-2 of Glc), 5.45 (d, 1H, J_{3,4} = 3.0 Hz, H-4 of Gal c), 5.39–5.34 (m, 1H, H-2 of Gal c), 5.30 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} =$ 3.2 Hz, H-3 of Gal a), 5.18–4.95 (m, 6H, H-3 of Gal b, H-1, 3, 4 of Glc, H-1, 2, 3, 4 of Man), 4.73 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal c), 4.66 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1 of Gal b), 4.61 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal a), 4.34 (br. d, 1H, J = 10.1 Hz, H-5 of Glc), 4.25– 4.09 (m, 5H, H-5 of Gal a, H-4 of Gal b, H-3, 5, 6 of Gal c), 4.00-3.65 (m, 11H, H-5, 6 of Gal a, H-5, 6a of Gal b, H-6 of Glc, H-5, 6 of Man, CH₂CH₂SiMe₃), 3.57 (br. d, 1H, J = 11.2 Hz, H-6b of Gal b), 3.43–3.36 (m, 1H, CH₂CH₂SiMe), 2.24, 2.15, 2.10, 2.09, 2.07, 2.05, 2.01, 1.98, 1.94, 1.90, 1.66 (each s, each 3H, COCH₃), -0.17 (s, 9H, CH₂CH₂Si*Me*₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 170.5, 170.41, 170.37, 170.04, 169.98, 169.9, 169.5, 169.41, 169.38, 169.3, 165.6, 165.3, 165.2, 165.1, 164.5, 133.6, 133.3, 133.13, 133.08, 129.9, 129.74, 129.68, 129.6, 129.5, 129.3, 129.1, 129.0, 128.8, 128.6, 128.43, 128.37, 128.3, 128.2, 101.3 (C-1 of Gal b), 100.9 (C-1 of Gal c), 100.6 (C-1 of Gal a), 97.9 (C-1 of Glc), 95.1 (C-1 of Man), 75.7, 73.8, 72.9, 72.82, 72.76, 71.9, 70.7, 70.6, 70.2, 69.6, 69.5, 68.7, 68.6, 68.5, 68.1, 68.0, 67.8, 67.6, 66.3, 65.1, 65.0, 61.5, 61.2, 61.1, 60.4, 20.8, 20.7, 20.6, 20.4, 17.6, 14.2, -1.52

MALDI-TOFMS: calcd for $C_{92}H_{106}O_{42}SiNa: m/z$ 1933.6; found: m/z 1934.3 [M+Na]⁺.

4.15.

2,3,4,6-tetra-*O*-acetyl- β -D-mannopyranosyl- $(1\rightarrow 3)$ -4,6-di-*O*-acetyl-2-*O*-benzoyl- β -D -galactopyranosyl- $(1\rightarrow 6)$ -[2,3,4,6-tetra-*O*-acethyl- α -D-glucopyranosyl- $(1\rightarrow 4)$]-2,3-d i-*O*-benzoyl- β -D-galactopyranosyl- $(1\rightarrow 6)$ -4-*O*-acetyl-2,3-di-*O*-benzoyl- α -D-glucopy ranosyl trichloroacetimidate (20)

Compound **20** was prepared from **19** (271 mg, 142 µmol) as described for preparation of **14**. The product was purified by silica gel column chromatography (20:1 chloroform-acetone) to give **20** (283 mg, quant.) as amorphous powder. $[\alpha]_D^{25}$ +94.4 (c = 2.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.30 (s, 1H, NH), 8.06– 7.30 (m, 25H, 5 x Ph), 6.61 (d, 1H, $J_{1,2} = 4.4$ Hz, H-1 of Gal a), 5.81 (dd, 1H, $J_{2,3} = 10.8$ Hz, $J_{3,4} = 3.3$ Hz, H-3 of Gal a), 5.73–5.69 (m, 2H, H-2, 4 of Gal a), 5.54–5.48 (m, 2H, H-2 of Gal b, H-2 of Glc), 5.46 (d, 1H, $J_{3,4} = 3.0$ Hz, H-4 of Gal c), 5.39 (t, 1H, $J_{1,2} = J_{2,3} = 8.0$ Hz, H-2 of Gal c), 5.17–4.92 (m, 8H, H-3 of Gal b, H-1, 3, 4 of Glc, H-1, 2, 3, 4, of Man), 4.75 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1 of Gal c), 4.65 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1 of Gal b), 4.45 (br. d, 1H, $J_{5,6} = 6.2$ Hz, H-5 of Gal a), 4.35–4.32 (m, 1H, H-5 of Glc), 4.26–4.12 (m, 5H, H-6 of Gal b, H-3, 4, 5 of Gal c), 4.03–3.89 (m, 3H, H-5 of Gal b, H-6a of Gal c, H-6a of Glc), 3.82–3.64 (m, 6H, H-6 of Gal a, H-6b of Gal c, H-6b of Glc, H-5, 6 of Man), 2.25, 2.17, 2.13, 2.11, 2.00, 1.95, 1.93, 1.91, 1.67 (each s, each 3H, COC*H*₃), 2.08 (s, 6H, 2 x COC*H*₃); ¹³C NMR (100 MHz, CDCl₃): δ 170.54, 170.49, 170.4, 170.3, 170.0, 169.9, 169.5, 169.41, 169.36, 169.3, 165.63, 169.58, 165.1, 165.0, 164.5, 160.3, 133.6, 133.5, 133.4, 133.2, 133.0, 129.9, 129.7, 129.6, 129.5, 129.3, 129.1, 129.0, 128.7, 128.6, 128.4, 128.2, 100.9 (C-1 of Gal b, c), 97.8 (C-1 of Glc), 95.0 (C-1 of Man), 93.5 (C-1 of Gal a), 90.6, 76.0, 73.8, 73.1, 72.9, 70.8, 70.7, 70.6, 70.1, 69.3, 68.7, 68.6, 68.5, 68.2, 68.1, 67.7, 66.9, 66.6, 65.0, 64.9, 61.5, 64.2, 61.0, 20.7, 20.7, 20.6, 20.5, 20.5, 20.3, 14.0, 10.9.

MALDI-TOFMS: calcd for C₈₉H₉₄Cl₃NO₄₂Na: *m/z* 1976; found: *m/z* 1976 [M+Na]⁺.

4.16. (2*S*, 3*S*, 4*R*)-3,4-dibenzyloxy-2-(hexadecanoylamino)-octadecyl 2,3,4,6-tetra-*O*-acetyl- β -D-mannopyranosyl-(1 \rightarrow 3)-4,6-di-*O*-acetyl-2-*O*-benzoyl- β -D -galactopyranosyl-(1 \rightarrow 6)-[2,3,4,6-tetra-*O*-aceyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-6-*O*-ac etyl-2,3-di-*O*-benzoyl- β -D -galactopyranosyl-(1 \rightarrow 6)-4-*O*-acetyl-2,3-di-*O*-benzoyl- β -D -galactopyranoside (21)

Compound 21 was prepared from 15 (76.9 mg, 101 µmol) and 20 (131 mg, 67.1 µmol) as described for preparation of 16. The product was purified by silica gel column chromatography (1:1 hexane-ethyl acetate) to give 21 (72.1 mg, 42.0%). $[\alpha]_{D}^{25}$ +54.4 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ ; 8.04–7.31 (m, 35H, 5 x Ph), 5.99 (d, 1H, J = 9.6 Hz, NH of Cer), 5.52–5.23 (m, 9H, H-2, 3 of Gal a, H-2, 3 of Gal b, H-2, 4 of Gal c, H-2 of Glc, H-3, 4 of Cer), 5.14–4.95 (m, 8H, H-4 of Gal a, H-1, 3, 4 of Glc, H-1, 2, 3, 4 of Man), 4.70 (d, 1H, $J_{1,2} = 8.2$ Hz, H-1 of Gal c), 4.43 (d, 1H, $J_{1,2} = 7.5$ Hz, H-1 of Gal b), 4.31–4.29 (m, 2H, H-1a, 2 of Cer), 4.22–4.09 (m, 5H, H-4 of Gal b, H-3, 6a of Gal c, H-6 of Glc), 4.04 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1 of Gal a), 3.95– 3.49 (m, 11H, H-6 of Gal a, H-6 of Gal b, H-5, 6b of Gal c, H-5 of Glc, H-5, 6 of Man, H-1b of Cer), 3.15 (dd, 1H, J_{5,6a} = 2.8 Hz, J_{5,6b} = 9.4 Hz, H-5 of Gal a), 2.98 (dd, 1H, $J_{5,6a} = 6.4$ Hz, $J_{5,6b} = 10.8$ Hz, H-5 of Gal b), 2.22, 2.15, 2.11, 2.09, 2.063, 2.057, 2.00, 1.95, 1.91, 1.90, 1.66 (each s, each 3H, COCH₃), 1.78–0.85 (m, 60H, alkyl-H); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 170.50, 170.45, 170.4, 170.2, 169.9, 169.54, 169.45, 169.4, 169.3, 166.1, 165.6, 165.4, 165.2, 165.0, 164.5, 133.6, 133.5, 133.4, 133.3, 132.9, 130.3, 130.2, 129.9, 129.8, 129.7, 129.6, 129.5, 129.24, 129.17, 129.1, 129.0, 128.8, 128.6, 128.5, 128.4, 128.3, 101.4 (C-1 of Gal a), 101.0 (C-1 of Gal b), 100.3 (C-1 of Gal c), 97.7 (C-1 of Man), 95.1 (C-1 of Glc), 75.4, 73.8, 72.9, 72.7, 72.4, 72.2, 71.3,

70.8, 70.6, 70.3, 70.2, 69.6, 68.70, 68.66, 68.6, 68.1, 67.9, 67.7, 66.5, 66.0, 65.1, 65.0, 61.6, 61.2, 61.1, 47.8, 36.2, 31.9, 29.71, 29.68, 29.64, 29.59, 29.54, 29.47, 29.3, 29.2, 28.8, 25.6, 25.4, 22.7, 20.8, 20.7, 20.59, 20.55, 20.4, 14.1.

MALDI-TOFMS: calcd for C₁₃₅H₁₆₉NO₄₇Na: *m/z* 2579.1; found: *m/z* 2579.6 [M+Na]⁺.

4.17 (2*S*, 3*S*, 4*R*)-2-(hexadecanoylamino)-3,4-dihydroxyoctadecyl β -D-mannopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 6)-[α -D-glucopyranosyl-(1 \rightarrow 4)]- β -D-galactopyranosyl-(1 \rightarrow 6)-4- β -D-galactopyranoside (2)

Compound 2 was prepared from **21** (72.1 mg, 28.2 µmol) as described for preparation of **1**. The product was purified by gel filtration chromatography (Sephadex LH-20, ϕ 20 x 1000 mm) using 1:1 chloroform-MeOH to give **2** (33.9 mg, 88.0%). [α]_D²⁵ +50.3 (*c* = 0.2, 1:1 CHCl₃-MeOH). ¹H NMR (400 MHz, CDCl₃): δ 5.06 (d, 1H, *J*

[α]_D +50.3 (c = 0.2, 1:1 CHCl₃-MeOH). H NMR (400 MHz, CDCl₃): δ 5.06 (d, 1H, J = 3.7 Hz, H-1 of Glc), 4.98 (br. s, 1H, H-1 of Man), 4.40 (d, 1H, J = 6.9 Hz, H-1 of Gal c), 4.33 (d, 1H, J = 6.8 Hz, H-1 of Gal b), 4.28 (d, 1H, J = 7.5 Hz, H-1 of Gal a); ¹³C NMR (100 MHz, CDCl₃): δ 175.3, 104.8 (C-1 of Gal b), 104.6 (C-1 of Gal c), 104.1 (C-1 of Gal a), 101.0 (C-1 of Glc), 97.2 (C-1 of Man), 75.9, 74.1, 74.0, 73.9, 73.7, 73.5, 72.9, 71.9, 71.7, 71.4, 71.1, 70.4, 70.1, 69.8, 69.4, 68.0, 67.3, 65.7, 62.5, 62.2, 62.3, 62.1, 62.0, 50.9, 36.9, 32.5, 30.3, 30.2, 30.1, 30.0, 29.9, 26.7, 26.5, 24.0, 23.2, 14.3. HR-FABMS: calcd for C₆₄H₁₁₀NO₂₉Na: m/z 1388.7765; found: m/z 1388.7743 [M+Na]⁺.

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Stereocontrolled syntheses of two neutral glycosphingolipids from the fungus *Hirsutella rhossiliensis* have been achieved.

The synthetic methods may be useful for the synthesis of conjugation of the other glycan.