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Chuanzhou Tao^{ab}, Feng Liu^{ac}, Bin Xu^a, Zhiling Cao^{ab}, Huiyan Wang^a & Weiwei Liu^{ac}

^a School of Chemical Engineering, Huaihai Institute of Technology, Lianyungang, 222005, P.R. of China

^b Jiangsu Marine Resources Development Research Institute, Lianyungang, 222005, P.R. of China

^c School of Chemical Engineering, China University of Mining and Technology , XuZhou , 221116 , P.R. of China Published online: 11 Oct 2013.

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Copper-Catalyzed Synthesis of N-aryl-D-Glucosamines from Arylboronic Acids

Chuanzhou Tao,^{1,2} Feng Liu,^{1,3} Bin Xu,¹ Zhiling Cao,^{1,2} Huiyan Wang,¹ and Weiwei Liu^{1,3}

¹School of Chemical Engineering, Huaihai Institute of Technology, Lianyungang 222005, P.R. of China
²Jiangsu Marine Resources Development Research Institute, Lianyungang 222005, P.R. of China
³School of Chemical Engineering, China University of Mining and Technology, XuZhou 221116, P.R. of China

A convenient catalytic protocol was developed to synthesize N-aryl-D-glucosamines from the corresponding arylboronic acids. C–N cross-coupling between arylboronic acids and 1,3,4,6-tetra-O-benzyl- β -D-glucosamine was realized by copper catalyst under mild conditions. Subsequent deprotection of the benzyl ethers gave the N-arylation products, N-aryl-D-glucosamines.

Keywords N-aryl-D-glucosamine; Arylboronic acid; Copper catalysis; Cross-coupling

INTRODUCTION

D-Glucosamine and its *N*-substituted derivatives are found in numerous biologically active molecules such as cell surface *N*-glycoproteins, proteoglycans, glycosylphosphatidylinositol (GPI) anchors, glycosphingolipids, lipopolysaccharides, and chitin/chitosan.^[1] Synthetic chemists also used these molecules as ligands or organocatalysts to introduce chirality in catalytic asymmetric reactions.^[2] Recently, *N*-arylation derivatives from D-glucosamine (Fig. 1) were disclosed with many utilities such as a predictive method in ecotoxicology,^[3a,3b]

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Address correspondence to Chuanzhou Tao, School of Chemical Engineering, Huaihai Institute of Technology, Lianyungang 222005, P.R. of China, and Jiangsu Marine Resources Development Research Institute, Lianyungang 222005, P.R. of China E-mail: taocz@hhit.edu.cn.; and Weiwei Liu, School of Chemical Engineering, Huaihai Institute of Technology, Lianyungang 222005, P.R. of China, and School of Chemical Engineering, China University of Mining and Technology, XuZhou 221116, P.R. of China. E-mail: liuww323@yahoo.com.cn



Figure 1: N-aryl-D-glucosamine derivatives.

cancer cell detection,^[3c] a metabolic indicator for FACS analysis,^[3d] and measurement of glucose uptake in a single cell of living *E. coli*.^[3e,3f] However, chemical modifications of D-glucosamine at the *N*-position mainly rely on acylation,^[4] Schiff-base formation,^[5] azidation,^[6] and alkylation.^[7] *N*-Arylation of D-glucosamine has rarely been realized due to synthetic difficulty. Only in a few examples was *N*-arylation of D-glucosamine accomplished by the S_NAr reaction between D-glucosamine and ary halides containing a strong electronwithdrawing group.^[3,8]

Very recently, we developed a general protocol for synthesis of *N*-aryl-D-glucosamines by the copper-catalyzed cross-coupling between aryl halides and D-glucosamine.^[9,10] With aryl halides as arylation reagents, a number of functionalized *N*-aryl-1,3,4,6-tetra-*O*-benzyl- β -D-glucosamines were synthesized, which were subsequently deprotected to give *N*-aryl-D-glucosamines. The reaction represents a major advance in the *N*-aryl-D-glucosamines synthesis. However, aryl halides carrying an *ortho*-substituent failed to participate in the reaction. The cross-coupling had to be carried out at high temperature (130°C) and protected with inert gas. Besides, ligands were critical to the transformations.

Herein we report a convenient and mild protocol for synthesis of *N*-aryl-D-glucosamines via copper-catalyzed cross-coupling between arylboronic acids and D-glucosamine under ligand-free conditions, where aryl boronic acids were used as arylation reagents instead of aryl halides. Copper-catalyzed crosscouplings between arylboronic acids and N/O/S-nucleophiles (phenol, aniline, or thiophenol) are known as Chan-Evans-Lam-modified Ullmann condensation, and the advantage of this reaction is that much milder reaction conditions can be allowed such as lower temperature and aerobic environment.^[10g,11,12] Importantly, *N*-ortho-substituent-aryl-D-glucosamines could also be obtained through the present method. The fact that many aryl boronic acids are now commercially available makes this protocol more practically appealing.^[13]

RESULTS AND DISCUSSION

We commenced our study with phenylboronic acid (2a) as model substrate, Cu(OAc)₂ as catalyst, and CH₂Cl₂ as solvent (Scheme. 1). Initially, Dglucosamine (1a) was selected as substrate to directly couple with phenylboronic acid (2a). Nevertheless, we could not obtain the *N*-arylation product **3aa** from D-glucosamine (1a). We speculated that the hydroxyl groups, especially 1-hydroxyl, have influence on the cross-coupling reaction. Then, we turned to the substrates from *O*-protected-D-glucosamines.



Scheme 1: Copper-catalyzed arylation of D-glucosamines with arylboronic acids.

It is well known that acetyl is a versatile group for protecting hydroxyl in carbohydrate chemistry and can be removed under mild conditions.^[14] If 1,3,4,6-tetra-O-acetyl- β -D-glucosamine (1b) was used as a surrogate of D-glucosamine, it would probably cross-couple with phenyl boronic acid (2a) to produce N-aryl-1,3,4,6-tetra-O-acetyl- β -D-glucosamine, which would easily be converted into N-aryl-D-glucosamines with base. This hypothesis prompted us to examine a roundabout process for synthesis of N-aryl-Dglucosamines catalyzed by copper (Sch. 1). The surrogate 1,3,4,6-tetra-Oacetyl- β -D-glucosamine (1b) was synthesized according to the literature.^[15] With our standard conditions, however, the reaction was reluctant and only 9% of the product was obtained. On the other hand, as another protecting group for hydroxyl, benzyl may be more stable in the catalytic system (Sch. 1). So we turned to the benzyl-protected D-glucosamine and 1,3,4,6-tetra-O-benzyl- β -Dglucosamine (1c)^[16] was used as a model substrate. Gratifyingly, the designed product (3ca) was obtained with 48% yield. It is noteworthy that many classic methods^[14,17] are known to deprotect the benzyl group from alcohol. For

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instance, hydrogenolysis with H₂/Pd/C can readily remove benzyl groups to release the hydroxyl groups of glucosamine.

Subsequently, to optimize the reaction condition, 1,3,4,6-tetra-O-benzyl- β -D-glucosamine and phenyl boronic acid were selected as model substrates. It is reported that the undesired phenol and homocoupled biphenyl ether byproducts could be formed via copper-catalyzed C-O coupling with H₂O and phenol.^[18] So, the molecular sieves were introduced into the reaction mixture to reduce the amount of H₂O. As shown in Table 1, under a ligand-free and basefree condition, different solvents were screened including polar solvents DMF and MeOH and less polar solvents CHCl₃, ClCH₂CH₂Cl, PhCl, CH₃CN, and toluene (entries 1–8). However, lower yields were obtained even when the reaction temperature was raised to 130°C. To our satisfaction, 65% yield was achieved when a CH₂Cl₂ and CH₃CN (2/1) mixture was used as cosolvent (entry 9). Nonetheless, other cosolvents such as toluene and CH₃CN slightly increased the yield (entry 10). With CH₂Cl₂ and CH₃CN as the solvent, we screened the copper sources including cuprous and cupric salts (entries 11–14).

BnO OBn BnO NH ₂ OBr	ı ⁺ Ph-B(C	0H)₂ → BnO BnO	OBn OBn NH OBn Ph
1c	2a		3ca
Entry Catalys	t Solve	nt Temp/°C	2 Yield% ^b
1 Cu(OAc) 2 Cu(OAc) 3 Cu(OAc) 4 Cu(OAc) 5 Cu(OAc) 6 Cu(OAc) 7 Cu(OAc) 8 Cu(OAc) 9 Cu(OAc) 10 Cu(OAc) 11 CuSO4 12 CuO 13 Cul 14 CuBr	$\begin{array}{cccc} & & CH_2C\\ \hline \\ \hline$	$\begin{array}{cccc} L_2 & 65 \\ I_3 & 65 \\ H_2 CI & 80 \\ I & 130 \\ ne & 110 \\ N & 80 \\ I & 120 \\ N & 80 \\ I & 120 \\ N & 80 \\ I & 120 \\ H_3 CN & 65 \\ H_3 $	48 39 36 39 45 33 29 15 65 49 0 0 27 32

Table 1: Copper-catalyzed phenylation of 1,3,4,6-tetra-O-benzyl- β -D-glucosamine with phenylboronic acid^{α}

^oConditions: **1c** 0.5 mmol; **2a** 1.5 mmol; catalyst 20 mol%; 4ÅMS 400 mg; solvent = 3.0 mL (V_{sol} : $V_{MeCN} = 2:1$); O₂, 24 h.

The yields indicated that $Cu(OAc)_2$ was the best catalyst. Therefore, our optimal condition for the coupling of arylboronic acid and 1,3,4,6-tetra-*O*-benzyl- β -D-glucosamine was as follows: 20 mol% $Cu(OAc)_2$ as the catalyst relative to 1,3,4,6-tetra-*O*-benzyl- β -D-glucosamine, and CH_2Cl_2 and CH_3CN mixture as the cosolvent at 65°C.

With the optimized catalytic system $Cu(OAc)_2/CH_2Cl_2/CH_3CN$, we next examined whether the same catalytic system can be applied to the coupling of 1,3,4,6-tetra-O-benzyl- β -D-glucosamine with various arylboronic acids having different electronic and steric properties. It was found that both electron-rich and electron-deficient arylboronic acids can be smoothly converted to the desired products with satisfactory isolated yields (Sch. 2). It is noteworthy that



Scheme 2: Copper-catalyzed N-arylation of 1,3,4,6-tetra-O-benzyl- β -D-glucosamine (1c) with arylboronic acids (2).

arylboronic acids carrying an ortho-substituent were also found to readily participate in the reaction. For instance, 2-methylphenyl boronic acid could give an isolated yield of 54%, and the yield for 2-methoxylphenyl boronic acid is 51%.

Certainly, the above arylation products can now easily be converted to the target N-aryl-D-glucosamines. Hydrogenolysis catalyzed by Pd/C was employed to release the hydroxyl of N-aryl-1,3,4,6-tetra-O-benzyl- β -Dglucosamine. As shown in Scheme 3, under the standard conditions,^[17] Nphenyl-D-glucosamine (**3aa**) and N-(4-methoxyl phenyl)-D-glucosamine (**3ac**) were obtained with high yields (76% and 87%, respectively).



Scheme 3: Deprotection of N-aryl-1,3,4,6-tetra-O-benzyl-β-D-glucosamines.

CONCLUSIONS

To conclude, we have developed a mild and convenient catalytic protocol for the synthesis of *N*-aryl-*D*-glucosamines. 1,3,4,6-tetra-*O*-benzyl- β -D-glucosamine is used as D-glucosamine surrogate and Cu(OAc)₂ was used as catalyst to achieve the C-N coupling. The efficiency and functional-group tolerance of this procedure have been demonstrated by synthesis of various functionalized *N*-aryl-1,3,4,6-tetra-*O*-benzyl- β -D-glucosamines. Furthermore, *N*-ortho-substituent-aryl-1,3,4,6-tetra-*O*-benzyl- β -D-glucosamines were also synthesized with the standard conditions. Finally, free *N*-aryl-*D*-glucosamines were easily obtained by deprotection of the benzyl protecting group. Given the fact that D-glucosamine derivatives play an important role in biomedical research and chiral molecule design, we anticipate that the method described in the present report will find applications in a number of fields such as pharmaceutical research and organic material synthesis.

EXPERIMENTAL

General Experimental Procedures

The solvents were distilled from CaH_2 . All chemicals were obtained from commercial source and used without further purification. Flash column chromatography was performed on silica 230–400 mesh. Melting points were determined on a Yanaco melting point apparatus and were uncorrected. IR spectra were recorded on a Bruker Tensor 27 spectrometer with KBr pellets. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker Advance 400 spectrometer at ambient temperature in DMSO-d⁶ or D₂O. Chemical shifts are reported in ppm relative to TMS. HRMS analysis was performed on a Mariner ESI-TOF and Finnigan LCQ advantage Max Series MS System.

Copper-catalyzed Coupling of arylboronic (2) with 1,3,4,6tetra-O-benzyl-β-D-glucosamine (1c): General procedure

A 100-mL oven-dried round-bottom flask, containing a magnetic stirring bar, was charged with Cu(OAc)₂ (18 mg, 20 mol%), ArB(OH)₂ **2** (1.5 mmol, 3 eq.), and 4Å MS (400 mg). The solvent of CH₂Cl₂/CH₃CN (3.0 mL = 2.0 mL + 1.0 mL) was injected and the suspension was stirred for 10 min at rt. Then, 1,3,4,6-tetra-O-benzyl- β -D-glucosamine **1c** (270 mg, 0.5 mmol) was added. The reaction vessel was flushed with O₂, sealed, and stirred at 65°C for 24 h. The resulting mixture was cooled to rt and filtered through a pad of silica gel with the help of CH₂Cl₂ (30 mL). The filtrate was concentrated and the residue was purified by column chromatography (silica gel, EtOAc-PE) to afford the product **3**.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-Nphenyltetrahydro-2H-pyran-3-amine (3ca)

m.p. = 114–117°C. IR (KBr) v: 3548, 3478, 3414, 1636, 1620, 1111, 1072, 620. ¹H NMR (400 MHz, DMSO) δ 7.40–7.27 (m, 8H), 7.25–7.14 (m, 10H), 7.11–7.03 (m, 4H), 6.74 (d, J = 7.8 Hz, 2H), 6.54 (t, J = 7.0 Hz, 1H), 5.62 (d, J = 9.0 Hz, 1H), 4.81–4.71 (m, 3H), 4.65 (d, J = 10.8 Hz, 1H), 4.61–4.51 (m, 4H), 4.48 (d, J = 7.9 Hz, 1H), 3.76–3.63 (m, 3H), 3.59–3.51 (m, 2H), 3.47–3.37 (m, 1H). ¹³C NMR (100 MHz, DMSO) δ 149.08, 138.50, 138.32, 138.28, 137.84, 128.64, 128.23, 128.20, 127.98, 127.97, 127.73, 127.70, 127.58, 127.52, 127.42, 127.28, 127.16, 127.01, 115.70, 112.78, 102.56, 84.17, 77.99, 74.13, 74.09, 73.91, 72.36, 70.04, 68.92, 59.66. HRMS (ESI): m/z [M + Na]⁺ calcd for C₄₀H₄₁O₅NNa: 638.2877; found: 638.2872.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-N-ptolyltetrahydro-2H-pyran-3-amine (3cb)

m.p. = 115–116°C. IR (KBr) v: 3548, 3477, 3417, 1636, 1617, 1565, 1415, 1123, 1111, 1069, 617. ¹H NMR (400 MHz, DMSO) δ 7.39–7.27 (m, 8H), 7.25–7.16 (m, 10H), 7.14–7.07 (m, 2H), 6.89 (d, J = 8.2 Hz, 2H), 6.67 (d, J = 8.4 Hz, 2H), 4.84–4.70 (m, 3H), 4.66 (d, J = 10.9 Hz, 1H), 4.62–4.51

(m, 4H), 4.48 (d, J = 8.0 Hz, 1H), 3.79–3.60 (m, 3H), 3.60–3.50 (m, 2H), 3.41–3.36 (m, 1H), 2.17 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 146.85, 138.57, 138.33, 138.30, 137.87, 129.09, 128.23, 128.20, 127.97, 127.73, 127.69, 127.58, 127.51, 127.42, 127.28, 127.17, 127.03, 124.11, 124.09, 113.00, 102.66, 84.19, 78.01, 74.09, 74.07, 73.91, 72.37, 70.05, 68.95, 60.17, 20.08. HRMS (ESI): m/z [M + H]⁺ calcd for C₄₁H₄₄O₅N: 630.3214; found: 630.3212.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-N-(4methoxyphenyl)tetrahydro-2H-pyran-3-amine (3cc)

m.p. = 107–109°C. IR (KBr) v: 3548, 3474, 3417, 1637, 1618, 1562, 1516, 1451, 1244, 1116, 1070, 742, 694. ¹H NMR (400 MHz, DMSO) δ 7.40–7.15 (m, 18H), 7.15–7.09 (m, 2H), 6.70 (s, 4H), 4.83–4.64 (m, 4H), 4.62–4.50 (m, 4H), 4.47 (d, J = 8.0 Hz, 1H), 3.79–3.59 (m, 6H), 3.54 (d, J = 5.2 Hz, 2H), 3.29 (dd, J = 9.3, 8.2 Hz, 1H). ¹³C NMR (100 MHz, DMSO) δ 150.77, 143.36, 138.60, 138.33, 138.30, 137.88, 128.24, 128.22, 128.00, 127.99, 127.72, 127.71, 127.60, 127.53, 127.43, 127.30, 127.19, 127.04, 114.38, 113.93, 102.79, 84.23, 78.03, 74.07, 73.90, 72.37, 70.09, 68.95, 60.84, 55.36. HRMS (ESI): m/z [M + H]⁺ calcd for C₄₁H₄₄O₆N: 646.3163; found: 646.3165.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-N-(4chlorophenyl)tetrahydro-2H-pyran-3-amine (3cd)

m.p. = 118–120°C. IR (KBr) v: 3445, 3343, 3029, 2895, 2862, 1600, 1517, 1492, 1450, 1114, 1070, 995, 813, 749, 697. ¹H NMR (400 MHz, DMSO) δ 7.39–7.27 (m, 8H), 7.24–7.15 (m, 8H), 7.16 (dd, J = 7.0, 2.5 Hz, 2H), 7.12–7.05 (m, 4H), 6.74 (d, J = 8.9 Hz, 2H), 4.82–4.70 (m, 3H), 4.67 (d, J = 10.9 Hz, 1H), 4.60–4.52 (m, 4H), 4.47 (d, J = 8.1 Hz, 1H), 3.80–3.52 (m, 5H), 3.46–3.37 (m, 1H). ¹³C NMR (100 MHz, DMSO) δ 148.07, 138.46, 138.32, 138.25, 137.78, 128.30, 128.24, 128.22, 128.00, 127.73, 127.67, 127.60, 127.55, 127.43, 127.34, 127.24, 127.01, 118.85, 114.15, 102.47, 84.01, 78.01, 74.18, 74.10, 73.95, 72.38, 70.11, 68.88, 59.64. HRMS (ESI): m/z [M + H]⁺ calcd for C₄₀H₄₁O₅N³⁵Cl: 650.2667; found: 650.2668; calcd for C₄₀H₄₁O₅N³⁷Cl: 652.2638; found: 652.2639.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-N-(4bromophenyl)tetrahydro-2H-pyran-3-amine (3ce)

m.p. = 127–129°C. IR (KBr) v: 3345, 3061, 3029, 2895, 2866, 1578, 1485, 1450, 1069, 809, 745, 694. ¹H NMR (400 MHz, DMSO) δ 7.40–7.27 (m, 8H), 7.27–7.12 (m, 12H), 7.08 (dd, J = 6.6, 3.0 Hz, 2H), 6.70 (d, J = 8.9 Hz, 2H), 5.91 (d, J = 9.1 Hz, 1H), 4.83–4.63 (m, 4H), 4.62–4.50 (m, 4H), 4.46 (d, J = 8.0 Hz, 1H), 3.78–3.50 (m, 5H), 3.40 (dd, J = 17.6, 9.0 Hz, 1H). ¹³C NMR (100 MHz,

DMSO) δ 148.49, 138.44, 138.31, 138.24, 137.77, 131.12, 128.23, 128.21, 127.99, 127.73, 127.68, 127.67, 127.59, 127.54, 127.43, 127.33, 127.23, 127.01, 115.52, 114.76, 102.44, 84.00, 77.99, 74.17, 74.09, 73.93, 72.36, 70.09, 68.87, 59.55. HRMS (ESI): m/z [M + Na]⁺ calcd for C₄₀H₄₀O₅NNa⁷⁹Br: 716.1982; found: 716.1992; calcd for C₄₀H₄₀O₅NNa⁸¹Br: 718.1970; found: 718.1990.

Ethyl 4-((2R,3R,4R,5S,6R)-2,4,5-tris(benzyloxy)-6-(benzyloxymethyl)tetrahydro-2H-pyran-3-ylamino)benzoate (3cf)

m.p. = 115–116°C. IR (KBr) v: 3468, 3420, 3353, 3029, 2900, 2866, 1700, 1610, 1537, 1274, 1175, 1111, 1069, 752, 694. ¹H NMR (400 MHz, DMSO) δ 7.69 (d, J = 8.9 Hz, 2H), 7.40–7.26 (m, 8H), 7.22–7.14 (m, 10H), 7.06 (dd, J = 7.4, 2.1 Hz, 2H), 6.77 (d, J = 8.9 Hz, 2H), 4.84–4.63 (m, 4H), 4.62–4.47 (m, 5H), 4.22 (q, J = 7.1 Hz, 2H), 3.81–3.52 (m, 6H), 1.28 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, DMSO) δ 165.86, 153.15, 138.35, 138.31, 138.21, 137.72, 130.77, 128.25, 128.23, 128.01, 127.99, 127.76, 127.64, 127.61, 127.57, 127.44, 127.35, 127.26, 127.00, 116.33, 111.70, 102.04, 83.71, 77.96, 74.23, 74.14, 73.98, 72.39, 70.05, 68.84, 59.48, 58.69, 14.38. HRMS (ESI): m/z [M + H]⁺ calcd for C₄₃H₄₆O₇N: 688.3268; found: 688.3266.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-N-otolyltetrahydro-2H-pyran-3-amine (3cg)

Liquid, IR (KBr) ν : 3433, 3003, 2978, 2936, 2853, 1578, 1424, 1043, 1014, 649, 620. ¹H NMR (400 MHz, DMSO) δ 7.49 (dd, J = 5.1, 1.8 Hz, 1H), 7.43–7.35 (m, 6H), 7.33–7.28 (m, 3H), 7.26–7.18 (m, 7H), 7.17–7.12 (m, 4H), 7.09 (dd, J = 7.8, 1.7 Hz, 1H), 7.03 (dd, J = 7.2, 2.3 Hz, 1H), 6.84 (dd, J = 10.9, 4.4 Hz, 1H), 4.83 (m, 3H), 4.64–4.55 (m, 5H), 4.48 (dd, J = 11.0, 2.0 Hz, 1H), 3.89 (dd, J = 11.9, 9.3 Hz, 1H), 3.83–3.68 (m, 4H), 3.63 (t, J = 9.2 Hz, 1H), 2.08 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 146.24, 138.32, 138.25, 138.16, 137.78, 137.75, 129.20, 128.26, 128.24, 128.03, 128.01, 127.95, 127.73, 127.66, 127.62, 127.61, 127.12, 126.96, 126.36, 121.24, 115.93, 110.95, 102.54, 82.82, 79.18, 74.00, 73.65, 72.41, 70.36, 69.71, 68.77, 58.65, 30.62. HRMS (ESI): m/z [M + Na]⁺ calcd for C₄₁H₄₃O₆NNa: 652.3043; found: 652.3033.

(2R,3R,4R,5S,6R)-2,4,5-Tris(benzyloxy)-6-(benzyloxymethyl)-N-(2methoxyphenyl)tetrahydro-2H-pyran-3-amine (3ch)

Liquid, IR (KBr) ν : 3419, 3062, 3030, 2928, 2866, 1643, 1578, 1452, 1419, 1111, 1027, 734, 693. ¹H NMR (400 MHz, DMSO) δ 7.39–7.15 (m, 18H), 7.07 (dt, J = 4.6, 3.2 Hz, 2H), 6.91–6.80 (m, 2H), 6.74 (td, J = 7.7, 1.2 Hz, 1H), 6.56 (td, J = 7.7, 1.5 Hz, 1H), 4.85–4.51 (m, 9H), 3.85 (dd, J = 17.4, 8.2 Hz, 1H), 3.80–3.66 (m, 4H), 3.64–3.51 (m, 2H), 3.40 (dd, J = 17.7, 8.5 Hz, 1H), 3.29 (dd,

$$\begin{split} J &= 9.4, \, 8.0 \ \text{Hz}, \ 1\text{H}). \ ^{13}\text{C} \ \text{NMR} \ (100 \ \text{MHz}, \ \text{DMSO}) \ \delta \ 163.69, \ 146.31, \ 138.48, \\ 138.33, \ 138.29, \ 137.87, \ 128.24, \ 128.20, \ 128.00, \ 127.96, \ 127.70, \ 127.67, \ 127.61, \\ 127.58, \ 127.42, \ 127.27, \ 127.11, \ 126.98, \ 120.89, \ 115.70, \ 110.87, \ 109.92, \ 102.28, \\ 83.80, \ 78.06, \ 74.10, \ 74.08, \ 73.83, \ 72.37, \ 69.96, \ 68.95, \ 59.96, \ 55.41. \ \text{HRMS} \ (\text{ESI}): \\ m/z \ [\text{M} + \mbox{Na}]^+ \ \text{calcd for} \ C_{41} \mbox{H}_{43} \mbox{O}_6 \mbox{N} + \mbox{Na}: \ 668.2983; \ found: \ 668.2987. \end{split}$$

Deprotection of *N*-aryl-1,3,4,6-tetra-O-benzyl- β -D-glucosamine (3): General procedure

Under hydrogen atmosphere, a solution of **3** (0.5 mmol) in CH₃OH and EtOAc (4 mL + 2 mL) was stirred in the presence of Pd/C (10%, 100 mg) and trichloroacetic acid (80 mg, 0.5 mmol) at 40°C. After 40 h the reaction mixture was cooled to rt and filtered through a pad of silica gel with the help of CH₃OH. The filtrate was concentrated and the residue was purified by column chromatography (silica gel, CHCl₃-CH₃OH) to afford the deprotected product.

(2R,3R,4R,5S,6R)-6-(Hydroxymethyl)-3-(phenylamino)tetrahydro-2H-pyran-2,4,5-triol (3aa)

IR (KBr) v: 3383, 2975, 2927, 2898, 1644, 1450, 1412, 1382, 1088, 1049, 880, 637. ¹H NMR (400 MHz, D₂O) δ 7.68–7.38 (m, 5H), 5.05 (s, 0.81H), 4.90–4.84 (m, 0.35H), 4.11 (t, J = 9.7 Hz, 0.85H), 4.00–3.62 (m, 4.15H), 3.55 (t, J = 9.6 Hz, 1.04H), 3.51–3.32 (m, 0.77H). ¹³C NMR (100 MHz, D₂O) δ 136.70, 136.23, 132.91, 132.71, 131.82, 130.71, 125.17, 125.13, 94.95, 90.38, 78.43, 74.09, 73.17, 72.78, 72.48, 72.22, 66.59, 66.24, 63.04, 62.88. HRMS (ESI): m/z [M + Na]⁺ calcd for C₁₂H₁₇O₅NNa: 278.0999; found: 278.0993.

(2R,3R,4R,5S,6R)-6-(Hydroxymethyl)-3-(4-methoxyphenylamino) tetrahydro-2H-pyran-2,4,5-triol (3ac)

IR (KBr) ν : 3411, 2936, 2840, 1600, 1577, 1512, 1445, 1260, 1031, 759. ¹H NMR (400 MHz, D₂O) δ 7.45 (t, J = 9.4 Hz, 2H), 7.15 (d, J = 8.7 Hz, 2H), 5.01 (s, 0.83H), 4.08 (t, J = 9.8 Hz, 0.84H), 3.96–3.58 (m, 7.45H), 3.51 (t, J = 9.5 Hz, 1.1H), 3.35 (s, 1.8H). ¹³C NMR (100 MHz, D₂O) δ 162.23, 128.18, 127.89, 127.08, 118.08, 117.96, 94.17, 90.20, 78.42, 74.07, 72.78, 72.44, 72.21, 72.03, 66.99, 66.67, 62.98, 62.86, 58.30, 51.45. HRMS (ESI): m/z [M + H]⁺ calcd for C₁₃H₁₉O₆N: 286.1285; found: 286.1293.

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