## **Brief Communications**

## Phosphorylthioureas and phosphorylureas containing amino acid fragments

A. E. Shipov,\* G. K. Genkina, and P. V. Petrovskii

A. N. Nesmeyanov Institute of Organoelement Compounds, Russian Academy of Sciences, 28 ul. Vavilova, 119991 Moscow, Russian Federation. Fax: +7 (499) 135 5085. E-mail: shipov@ineos.ac.ru

A series of (dialkoxyphosphoryl)thioureas and their 1,3,2-oxazaphosphinane analogs containing fragments of glycine, alanine,  $\beta$ -alanine, L-aspartic and L-glutamic acids, as well as phosphorylureas derived from glycine and  $\beta$ -alanine were synthesized in the search for potential biologically active compounds (including possible inhibitors of aspartate transcarbamoylase).

**Key words:** phosphoryl isothiocyanates, phosphoryl isocyanates, amino acid esters, esters of phosphorylthiocarbamoylamino acid, esters of phosphorylcarbamoylamino acids, 1,3,2-oxazaphosphinanes.

Earlier,<sup>1</sup> we have described synthesis of structural analogs of PALA<sup>2,3</sup> (1), an active carcinolytic, *viz.*, phosphorylureas (2) that contain an L-aspartic acid fragment, including 2-oxo-1,3,2-oxazaphosphinane derivatives (2,  $RR' = HN(CH_2)_3O$ ) in which the oxazaphosphinane ring can play «transport» role delivering a pharmacophore group into a cell through cell membranes<sup>4</sup> (similarly to the antitumor agent cyclophosphamide).

It is also known that ThioPALA (3) is twice as active as PALA due to higher lipophilicity.<sup>5</sup> It was thus of interest to synthesize thiourea derivatives with L-aspartic acid fragment analogous to compounds 2 including compounds that contain 1,3,2-oxazaphosphinane ring. Phosphoryl-thiourea and phosphorylurea derivatives that contain other



$$X = O(1); S(3).$$

amino acid fragments can also be of definite interest as potential biologically active compounds.

The reactions of the known<sup>6</sup> phosphoryl isothiocyanates 4 and 5 with amino acid esters 6 afford phosphorylthioureas 7, 8 (Scheme 1, Table 1), the reaction proceeded smoothly at room temperature in benzene.

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Esters **6** react with 2-isothiocyano-2-oxo-1,3, $2\lambda^5$ oxazaphosphinane (**9**) under the same conditions resulting in phosphorylthioureas **10**, which contain a cyclic fragment (see Scheme 1, Table 1). It is worth noting that isothiocyanate **9**, as well as compounds **4**, **5**, was obtained by the reaction of the corresponding acid chloride, 2-chloro-2-oxo-1,3, $2\lambda^5$ -oxazaphosphinane<sup>7</sup> (**11**), with KSCN in MeCN, but in this case longer heating (15–18 h) at 45–50 °C and two equivalents of KSCN were required (Scheme 2).

Scheme 2



After removal of inorganic salts, compound **9**, as judged by the <sup>31</sup>P NMR spectral data, was free from any admixtures, it was used in further reactions without additional purification.

Besides phosphorylthioureas **7b,c**, phosphorylureas **12a,b** were obtained (analogous L-aspartic and L-glutamic acids derivatives have been described by us earlier<sup>1</sup>), which

Table 1. Yields, melting points and elemental analysis data for the obtained compounds

Com- pound	Yield (%)	M.p. /°C	Found (%) Calculated				Molecular formula
			С	Н	Ν	Р	
7 <b>a</b> <sup>a</sup>	70	Oil	$\frac{36.99}{37.10}$	<u>5.94</u> 5.94	<u>7.68</u> 7.86	<u>8.61</u> 8.69	$\mathrm{C}_{11}\mathrm{H}_{21}\mathrm{N}_{2}\mathrm{O}_{7}\mathrm{PS}$
7b	44	84—85	<u>33.88</u> 33.88	$\frac{6.09}{6.03}$	<u>9.84</u> 9.88	$\frac{10.88}{10.90}$	$\mathrm{C_8H_{17}N_2O_5PS}$
7c	66	Oil	$\frac{36.25}{36.24}$	$\frac{6.47}{6.42}$	<u>9.26</u> 9.39	—	$C_9H_{19}N_2O_5PS$
7d	60	Oil	<u>38.94</u> 38.92	$\frac{6.32}{6.26}$	$\frac{7.51}{7.56}$	—	$\mathrm{C}_{12}\mathrm{H}_{23}\mathrm{N}_{2}\mathrm{O}_{7}\mathrm{PS}$
<b>8</b> <sup>b</sup>	20	Oil	$\frac{43.66}{43.68}$	$\frac{7.19}{7.10}$	$\frac{6.57}{6.74}$	$\frac{7.31}{7.51}$	$\mathrm{C_{15}H_{29}N_2O_7PS}$
10a	71	с	$\frac{35.41}{35.40}$	$\frac{5.18}{5.35}$	$\frac{12.04}{12.38}$	—	$C_{10}H_{18}N_3O_6PS$
			$\frac{33.82}{33.73}$	<u>4.96</u> 5.05	<u>11.49</u> 11.57	—	$C_{10}H_{18}N_3O_6PS \cdot 0.2CHCl_3$
10b	78	123—124	<u>31.51</u> 31.46	$\frac{5.37}{5.28}$	<u>15.77</u> 15.72	—	$C_7H_{14}N_3O_4PS$
10c	76	с	<u>33.94</u> 34.16	<u>5.61</u> 5.73	<u>14.98</u> 14.94	<u>10.89</u> 11.01	$C_8H_{16}N_3O_4PS$
10d	81	С	$\frac{37.49}{37.39}$	$\frac{5.77}{5.70}$	<u>11.81</u> 11.89	—	$C_{11}H_{20}N_3O_6PS$
10e	70	С	<u>34.18</u> 34.16	<u>5.79</u> 5.73	<u>14.71</u> 14.94	—	$C_8H_{16}N_3O_4PS$
			$\frac{32.21}{32.28}$	$\frac{5.17}{5.35}$	<u>13.69</u> 13.77	—	$C_8H_{16}N_3O_4PS \boldsymbol{\cdot} 0.2CHCl_3$
12a	54	101-102	$\frac{35.84}{35.82}$	<u>6.51</u> 6.39	<u>10.49</u> 10.45	<u>11.51</u> 11.55	$C_8H_{17}N_2O_6P$
12b	70	98—99	_	_	<u>9.56</u> 9.90	$\frac{11.03}{11.00}$	$C_9H_{19}N_2O_6P$

<sup>a</sup> Found (%): S, 8.93, calculated (%): S, 9.00.

<sup>b</sup> Found (%): S, 7.71, calculated (%): S, 7.77.

<sup>*c*</sup> Amorphous compound without definite melting point.

allows a comparison of the biological activities of these classes of compounds. Synthesis of ureas 12a,b was carried out by the described procedure<sup>1</sup> using reaction of diethoxyphosphoryl isocyanate (13) with amino acids esters **6** (Scheme 3, see Table 1).

The composition and structures of the obtained compounds were confirmed by data from elemental analy-

sis (see Table 1), and the <sup>31</sup>P{<sup>1</sup>H} and <sup>1</sup>H NMR spectra (Table 2). Diastereomeric anisochronous effect was observed in the NMR spectra of compounds **10a,d,e**, which contain two chiral centres in the molecules. Compounds **10a,e** formed stable solvates with 0.2 molecule of CHCl<sub>3</sub>. Data on the biological evaluation of the synthesized compounds will be published elsewhere.

Table 2. <sup>31</sup>P{<sup>1</sup>H} and <sup>1</sup>H NMR spectra for the obtained compounds in CDCl<sub>3</sub>

Com- pound	<sup>31</sup> Ρ NMR, δ	<sup>1</sup> H NMR, δ, J/Hz
7a	-4.5	1.35 (t, 6 H, CH <sub>2</sub> CH <sub>3</sub> , ${}^{3}J_{H,H} = 7.0$ ); 3.03 (AB-system, 1 H, H <sub>B</sub> , NHCH <sub>2</sub> C(O), ${}^{3}J_{H,H_{B}} = 4.7$ , ${}^{2}J_{H_{A},H_{B}} = 17.2$ ); 3.09 (AB-system, 1 H, H <sub>A</sub> , NHCH <sub>2</sub> C(O), ${}^{3}J_{H,H_{A}} = 4.4$ , ${}^{2}J_{H_{A},H_{B}} = 17.2$ ); 3.68, 3.75 (both s, 3 H each, CH <sub>3</sub> O); 4.11-4.26 (m, 4 H, CH <sub>3</sub> CH <sub>2</sub> O); 5.38 (dt, 1 H, NHCHCH <sub>2</sub> ), ${}^{3}J_{CH} = 4.4$ , ${}^{3}J_{CH} = 5.3$ ); 7.23 (d, 1 H, NHC ${}^{2}J_{CH} = 9.8$ ); 9.38 (d, 1 H, NHCH ${}^{3}J_{CH} = 7.6$ )
7b	-4.04	$J_{\text{H},\text{H}} = 4.4, J_{\text{H},\text{H}} = 5.3, 7.25 \text{ (d, 1 H, N(\underline{H}), J_{\text{H},\text{P}} = 7.6)}, 7.58 \text{ (d, 1 H, N(\underline{H}), J_{\text{H},\text{H}} = 7.6)}$ 1.41 (t, 6 H, CH <sub>2</sub> CH <sub>3</sub> , ${}^{3}J_{\text{H},\text{H}} = 7.1$ ); 3.81 (s, 3 H, CH <sub>3</sub> O); 4.18–4.29 (m, 4 H, CH <sub>3</sub> CH <sub>2</sub> O); 4.43 (d, 2 H, NHCH <sub>2</sub> , ${}^{3}J_{\text{H},\text{H}} = 5.0$ ); 7.66 (d, 1 H, NHP, ${}^{2}J_{\text{H},\text{P}} = 6.6$ ); 9.34 (br.s. 1 H, NHCH <sub>2</sub> )
7c	-4.55	1.35 (t, 6 H, CH <sub>2</sub> CH <sub>2</sub> , ${}^{3}J_{H,H} = 6.6$ ); 2.70 (t, 2 H, CH <sub>2</sub> C(O), ${}^{3}J_{H,H} = 6.3$ ); 3.70 (s, 3 H, CH <sub>3</sub> O); 3.91 (dt, 2 H, NHCH <sub>2</sub> CH <sub>2</sub> , ${}^{3}J_{H,H} = 6.2$ , ${}^{3}J_{H,H} = 6.1$ ); 4.10–4.20 (m, 4 H, CH <sub>3</sub> CH <sub>2</sub> O); 6.57 (d, 1 H, NHP ${}^{2}J_{U,P} = 7.1$ ); 9.15 (br s. 1 H, NHCH <sub>3</sub> )
7d	-4.26	1.35, 1.36 (boht 1, 6 H, CH <sub>2</sub> C <u>H</u> <sub>3</sub> , ${}^{3}J_{\text{H,H}} = 6.7$ ); 2.10–2.36 (m, 2 H, C <u>H</u> <sub>2</sub> C(O)); 2.39–2.47 (m, 2 H, CHC <u>H</u> <sub>2</sub> ); 3.65, 3.74 (both s, 3 H each, C <u>H</u> <sub>3</sub> O); 4.13–4.23 (m, 4 H, C <u>H</u> <sub>2</sub> O); 5.04, 5.06 (both t, 1 H, CHC <u>H</u> <sub>2</sub> ); 3.65, 3.74 (both s, 2 H, NHP); 9.21 (d, 1 H, NHCH, ${}^{3}J_{\text{H,H}} = 7.2$ )
8	-2.11	$\begin{array}{l} 0.87, 0.88 \text{ (both t, 6H, CH2, 0, 3}, 1.10, (0.13, 1.14), (0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0.11, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,$
9	$-13.69^{a}$	
10a	-1.54, -1.57	1.81–1.89 <sup><i>b</i></sup> , 2.15–2.35 (both m, 1 H + 1 H, CH <sub>2</sub> CH <sub>2</sub> ); 2.94–3.09 (m, 2 H, CH <sub>2</sub> NHP); 3.20–3.40 (m, 2 H CH <sub>2</sub> OP); 3.67, 3.68 (both s, 3 H, CH <sub>3</sub> O); 3.73, 3.74 (both s, 3 H, CH <sub>3</sub> O); 4.03–4.15 (br., 1 H, CH <sub>2</sub> NHP); 4.33–4.43 (m, 2 H, CH <sub>2</sub> CH); 5.36 (dt, 1 H, NHCHCH, ${}^{3}L_{VV} = 4.9$ , ${}^{3}L_{VV} = 8.0$ ):
10b	-0.68	$\begin{array}{l} 7.79 (d, 1 H, N\underline{H}P, {}^{2}J_{H,P} = 9.1); 9.53 (br.s, 1 H, N\underline{H}CH) \\ 1.77-1.84, 2.02-2.13 (both m, 1 H each, CH_2C\underline{H}_2CH_2); 3.32-3.39 (m, 2 H, C\underline{H}_2NHP); 3.75 (s, 3 H, C\underline{H}_3O); 4.09 (br.s, 1 H, CH_2N\underline{H}P); 4.29 (AB-system, 1 H, HB, NHC\underline{H}_2C(O), {}^{3}J_{H,H_B} = 5.1, {}^{2}J_{H_A,H_B} = 18.0); 4.39 (AB-system, 1 H, HA, NHC\underline{H}_2C(O), {}^{3}J_{H,H_A} = 5.2, {}^{2}J_{H_A,H_B} = 18.0); \\ 4.26-4.46 (m, 2 H, C\underline{H}_2OP); 8.34 (d, 1 H, C(S)N\underline{H}P, {}^{2}J_{H,P} = 9.5); 8.92 (t, 1 H, C(S)N\underline{H}CH_2, \\ \end{array}$
10c	-1.06	${}^{3}J_{H,H} = 5.0$ ) 1.81–1.89, 2.00–2.08 (both m, 1 H each, ${}^{3}J_{H,H} = 4.6$ , CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> ); 2.64 (ABCD-system, 1 H, H <sub>B</sub> , CH <sub>2</sub> CH <sub>2</sub> C(O), ${}^{3}J_{H_{B},H_{C}} = 6.5$ , ${}^{3}J_{H_{B},H_{D}} = 2.5$ , ${}^{2}J_{H_{A},H_{B}} = 17.0$ ); 2.71 (ABCD-system, 1 H, H <sub>A</sub> , CH <sub>2</sub> CH <sub>2</sub> C(O), ${}^{3}J_{H_{A},H_{C}} = 6.4$ , ${}^{3}J_{H_{B},H_{D}} = 2.4$ , ${}^{2}J_{H_{A},H_{B}} = 17.0$ ); 3.68 (s, 3 H, CH <sub>3</sub> O); 3.28–3.42 (m, 2 H, CH <sub>2</sub> NHP); 3.83 (AB-system, 1 H, HB, C(S)NHCH <sub>2</sub> , ${}^{3}J_{H,H_{B}} = 2.1$ , ${}^{2}J_{H_{A},H_{B}} = 6.4$ ); 3.86 (AB-system, 1 H, HA, C(S)NHCH <sub>2</sub> , ${}^{3}J_{H,H_{B}} = 6.4$ ); 3.90 (br.s, 1 H, CH <sub>2</sub> NHP); 4.33–4.45 (m, 2 H, CH <sub>2</sub> OP); 8.12 (d, 1 H, PNHC(S), ${}^{2}J_{H,P} = 9.4$ ); 8.88 (t, 1 H, C(S)NHCH <sub>2</sub> ,
10d	-0.36, -0.47	$J_{H,H} = 5.6$ ) 1.78–1.90, 2.27–2.36 (both m, 1 H each, $CH_2C\underline{H}_2CH_2$ ); 2.00–2.15 (m, 2 H, $C\underline{H}_2C(O)$ ); 3.30–3.46 (m, 2 H, $C\underline{H}_2$ NHP); 3.65 (s, 3 H, $CH_2COOC\underline{H}_3$ ); 3.730, 3.736 (s, 3 H, $CHCOOC\underline{H}_3$ ); 4.02 (br.s, 1 H, $CH_2N\underline{H}P$ ); 4.35–4.48 (m, 2 H, $C\underline{H}_2OP$ ); 5.03 (dt, 1 H, $NHC\underline{H}CH_2$ ,
10e	0.21, 0.36	${}^{3}J_{\text{H},\text{H}} = 4.5, {}^{3}J_{\text{H},\text{H}} = 5.3$ ); 8.04 (br.s, 1 H, PN <u>H</u> C(S)); 9.00–9.05, 9.07–9.14 (both m, 1 H, N <u>H</u> CH) 1.48 (d, 3 H, CHC <u>H</u> <sub>3</sub> , ${}^{3}J_{\text{H},\text{H}} = 7.3$ ); 1.77–1.90, 1.97–2.12 (both m, 1 H each, CH <sub>2</sub> C <u>H</u> <sub>2</sub> CH <sub>2</sub> ); 3.23–3.48 (m, 2 H, C <u>H</u> <sub>2</sub> NHP); 3.72, 3.73 (both s, 3 H, OC <u>H</u> <sub>3</sub> ); 4.04 (br.d, 1 H, CH <sub>2</sub> N <u>H</u> P, ${}^{2}J_{\text{H},\text{P}} = 31.0$ ); 4.31–4.48 (m, 2 H, C <u>H</u> <sub>2</sub> OP); 4.90 (dq, 1 H, NHC <u>H</u> CH <sub>3</sub> , ${}^{3}J_{\text{H},\text{H}} = 7.3, {}^{3}J_{\text{H},\text{H}} = 6.0$ ); 7.75 (br.s. 1 H, PNHC(S)); 8.99 (dd, 1 H, NHCH, ${}^{3}J_{\text{H},\text{H}} = 7.0, {}^{2}J_{\text{H},\text{P}} = 54.0$ )
12a	-0.99	1.38 (t, 6 H, CH <sub>2</sub> CH <sub>3</sub> , ${}^{3}J_{H,H} = 7.0$ ); 3.76 (s, 3 H, CH <sub>3</sub> O); 4.01 (d, 2 H, NHCH <sub>2</sub> , ${}^{3}J_{H,H} = 5.7$ ); 4.15-4.26 (m, 4 H, CH <sub>2</sub> CH <sub>3</sub> O); 7.18 (s, 1 H, NHP); 7.90 (s, 1 H, NHCH <sub>2</sub> )
12b	-0.8	1.41 (t, 6 H, CH <sub>2</sub> C <u>H<sub>3</sub></u> , ${}^{3}J_{H,H} = 7.0$ ); 2.63 (t, 2 H, C <u>H<sub>2</sub></u> C(O), ${}^{3}J_{H,H} = 6.4$ ); 3.57 (dt, 2 H, NHC <u>H<sub>2</sub></u> CH <sub>2</sub> , ${}^{3}J_{H,H} = 6.4$ , ${}^{3}J_{H,H} = 6.2$ ); 3.76 (s, 3 H, C <u>H<sub>3</sub></u> O); 4.17–4.27 (m, 4 H, CH <sub>3</sub> C <u>H<sub>2</sub></u> O); 6.22 (d, 1 H, N <u>H</u> P, ${}^{2}J_{H,P} = 5.6$ ); 7.20 (br.s, 1 H, N <u>H</u> CH <sub>2</sub> )

<sup>a</sup> In MeCN.

<sup>b</sup> In CD<sub>3</sub>CN.



**12:**  $Y = CH_2(a), CH_2CH_2(b).$ 

## Experimental

NMR spectra were recorded on Bruker Avance-400 ( ${}^{1}$ H 400.13 MHz;  ${}^{31}$ P 161.98 MHz) and Bruker Avance-300 ( ${}^{1}$ H 300.13 MHz;  ${}^{31}$ P 121.50 MHz) spectrometers in CDCl<sub>3</sub>, the signals for the residual protons of the deuterated solvents were used as the internal standard ( ${}^{1}$ H), 85% H<sub>3</sub>PO<sub>4</sub> in CDCl<sub>3</sub> was used as the external standard ( ${}^{31}$ P).

Phosphoryl isothiocyanates **4**, **5** were synthesized by the described method,<sup>6</sup> 2-chloro-2-oxo-1,3,2 $\lambda^5$ -oxazaphosphinane **11** was obtained by the known method.<sup>7</sup> Phosphoryl isocyanate **13** (Aldrich) was distilled *in vacuo* before reaction. KSCN that was used for the reactions was dried azeotropically with C<sub>6</sub>H<sub>6</sub> and then *in vacuo* over P<sub>2</sub>O<sub>5</sub>. Amino acid esters were obtained by the known method,<sup>8</sup> they were isolated from hydrochlorides by treatment with a solution of NH<sub>3</sub> in CHCl<sub>3</sub> (see Ref. 9).

In some cases, the isolation of compounds was carried out using column chromatography on SiO<sub>2</sub> (130–270 mesh, Aldrich) at a ratio compound : SiO<sub>2</sub> 1 : 16 (w/w), gradient elution with a mixture CHCl<sub>3</sub>–MeOH from 100 : 1 to 10 : 1 or hexane–Me<sub>2</sub>CO from 100 : 1 to 3 : 2. Fractions were analyzed by TLC on SiO<sub>2</sub> in the solvent systems CHCl<sub>3</sub>–MeOH (10 : 1) or hexane–Me<sub>2</sub>CO (3 : 2).

**Dimethyl** *N*-[*N*-(diethoxyphosphoryl)thiocarbamoyl]-L-aspartate (7a). A solution of ester 6a (1.06 g, 6.7 mmol) in  $C_6H_6$  (3 mL) was added dropwise with stirring to a solution of isothiocyanate 4 (1.27 g, 6.5 mmol) in anhydrous  $C_6H_6$  (11 mL) (slight heating occurred). The course of the reaction was monitored by <sup>31</sup>P NMR spectroscopy by following disappearance of the signal of 4 ( $\delta$  –18.4). After 5 h at room temperature, the solution was concentrated *in vacuo*, and the residue (1.68 g) was chromatographed on SiO<sub>2</sub> in a hexane—acetone solvent system. Compound 7a was obtained in the form of a viscous yellowish oil (1.60 g, 70%) (see Tables 1 and 2).

*N*-[*N*-(Diethoxyphosphoryl)thiocarbamoyl]glycine methyl ester (7b) was obtained analogously by the reaction of 4 (1.04 g, 5.3 mmol) in  $C_6H_6$  (8 mL) with ester 6b (0.475 g, 5.4 mmol) in  $C_6H_6$  (1.5 mL). After 16 h, the product was filtered off and chromatographed in a hexane—acetone solvent system. Product 7b was isolated (0.55 g, 44%) as a white solid (see Tables 1 and 2).

*N*-[*N*-(Diethoxyphosphoryl)thiocarbamoyl]-β-alanine methyl ester (7c) was obtained under the same conditions from compound 4 (0.80 g, 4.0 mmol) in  $C_6H_6$  (4.5 mL) and ester 6c (0.43 g, 4.1 mmol) in  $C_6H_6$  (2.0 mL); removal of the solvent *in vacuo* and chromatography in a CHCl<sub>3</sub>-MeOH solvent system yielded 7c (0.79 g, 66%) (see Tables 1 and 2).

**Dimethyl** *N*-[*N*-(diethoxyphosphoryl)thiocarbamoyl]-L-glutamate (7d) was obtained similarly by the reaction of 4 (0.80 g, 4.0 mmol) in  $C_6H_6$  (4.5 mL) with ester 6d (0.72 g, 4.1 mmol); after chromatographic purification in a CHCl<sub>3</sub>-MeOH solvent system, product 7d was obtained (0.90 g, 60%) (see Tables 1 and 2).

**Dimethyl** *N*-[*N*-(dibutoxyphosphoryl)thiocarbamoyl]-L-aspartate (8) was obtained similarly from isothiocyanate 5 (1.15 g, 4.5 mmol) in acetone (5.5 mL) and ester 6a (0.75 g, 4.6 mmol) in acetone (1.5 mL); after chromatographic purification in a hexane—acetone solvent system, thiourea 8 was obtained (0.40 g, 20%) (see Tables 1 and 2).

**2-Isothiocyano-2-oxo-1,3,2\lambda^3-oxazaphosphinane (9).** Chloride **11** (<sup>31</sup>P NMR,  $\delta$  10.32) (0.23 g, 1.5 mmol) was added to a solution of KSCN (0.29 g, 3.0 mmol) in MeCN (5 mL), the reaction mixture was stirred at 45–50 °C for 17 h (<sup>31</sup>P NMR monitoring). The mixture was filtered, the precipitate was washed with MeCN; the filtrate was concentrated to dryness *in vacuo*, the residue was extracted with C<sub>6</sub>H<sub>6</sub> (5×5 mL), the extracts were concentrated *in vacuo* (1 Torr). Isothiocyanate **9** was obtained (0.23 g, 87%) (<sup>31</sup>P NMR,  $\delta$  –13.69). The product was used in the following reactions without additional purification.

**Dimethyl** *N*-[*N*-(2-oxo-1,3,2 $\lambda^3$ -oxazaphosphinan-2-yl)thiocarbamoyl]-L-aspartate (10a). Ester 6a (0.74 g, (4.6 mmol) in C<sub>6</sub>H<sub>6</sub> (3 mL) was added dropwise with stirring to a solution of isothiocyanate 9 (0.82 g, 4.6 mmol) in C<sub>6</sub>H<sub>6</sub> (10 mL) (slight self-heating from 22 to 27 °C occurred), and the reaction mixture was kept for 16 h. The mixture was concentrated *in vacuo*, the residue (1.47 g) was purified by chromatography in a CHCl<sub>3</sub>— MeOH solvent system. Compound 10a was isolated (1.11 g, 72%) (solid foam) in the form of a stable solvate with 0.2 molecule of CHCl<sub>3</sub> (See Table 1), the presence of the latter was confirmed by the <sup>1</sup>H NMR spectrum in CD<sub>3</sub>CN. To remove the solvate CHCl<sub>3</sub>, compound was dissolved in anhydrous MeOH, the solution was concentrated to dryness *in vacuo*, and the residue was dried over P<sub>2</sub>O<sub>5</sub> (see Tables 1 and 2).

*N*-[*N*-(2-Oxo-1,3,2 $\lambda^3$ -oxazaphosphinan-2-yl)thiocarbamoyl]glycine methyl ester (10b) was obtained similarly from isothiocyanate 9 (0.80 g, 4.5 mmol) and ester 6b (0.45 g, 5.0 mmol) in C<sub>6</sub>H<sub>6</sub> (13 mL). Fine crystals that precipitated were filtered off, the product was purified by precipitation with anhydrous ether from a solition in CHCl<sub>3</sub>. Compound 10b was isolated (0.94 g, 78%) (see Tables 1 and 2).

*N*-[*N*-(2-Oxo-1,3,2 $\lambda^3$ -oxazaphosphinane-2-yl)thiocarbamoyl]β-alanine methyl ester (10c) was obtained similarly from 9 (0.81 g, 4.55 mmol) and ester 6c (0.52 g, 5.0 mmol) in C<sub>6</sub>H<sub>6</sub> (14 mL). After removal of C<sub>6</sub>H<sub>6</sub> and chromatographic purification using a CHCl<sub>3</sub>-MeOH solvent system, compound 10c was isolated (0.97 g, 76%) as a solid foam (see Tables 1 and 2).

**Dimethyl** *N*-[*N*-(2-oxo-1,3,2 $\lambda^3$ -oxazaphosphinan-2-yl)thiocarbamoyl]-L-glutamate (10d) was obtained under the same conditions from 9 (0.60 g, 3.37 mmol) and ester 6d (0.61 g, 3.50 mmol) in C<sub>6</sub>H<sub>6</sub> (10 mL). After chromatographic purification in a CHCl<sub>3</sub>—MeOH solvent system, the isolated product was dissolved in anhydrous MeOH, the solution was concentrated *in vacuo*, the residue was dried over P<sub>2</sub>O<sub>5</sub>. Compound 10d was obtained (0.97 g, 81%) (see Tables 1 and 2) as solid foam.

*N*-[*N*-(**2**-Oxo-1,3,2 $\lambda^3$ -oxazaphosphinane-2-yl)thiocarbamoyl]alanine methyl ester (10e) was obtained similarly from 9 (0.87 g, 4.88 mmol) and ester 6e (0.57 g, 5.50 mmol) in C<sub>6</sub>H<sub>6</sub> (14 mL). The product was chromatographed in a CHCl<sub>3</sub>-MeOH solvent system and isolated in the form of a solvate (0.96 g, 70%) with 0.2 molecule of CHCl<sub>3</sub> (see Table 1). To remove the solvate CHCl<sub>3</sub>, compound was dissolved in anhydrous MeOH, the solution was concentrated to dryness *in vacuo*, and the residue was dried *in vacuo* over  $P_2O_5$  (see Tables 1 and 2).

*N*-[*N*-(Diethoxyphosphoryl)carbamoyl]glycine methyl ester (12a). Ester 6b (0.53 g, 5.8 mmol) was added dropwise with stirring to a solution of isocyanate 13 (1.00 g, 5.6 mmol) in  $C_6H_6$  (8 mL) (self-heating by 5 °C occurred). After 16 h at room temperature, the gel-like mixture was diluted with ether, the residue was filtered off and washed with ether. Solid product 12a was obtained (0.80 g, 54%) (see Tables 1 and 2).

*N*-[*N*-(Diethoxyphosphoryl)carbamoyl]-β-alanine methyl ester (12b) was obtained similarly from isocyanate 13 (0.59 g, 3.3 mmol) in C<sub>6</sub>H<sub>6</sub> (4.5 mL) and ester 6c (0.35 g, 3.4 mmol) in C<sub>6</sub>H<sub>6</sub> (1.5 mL), the precipitate was filtered off and washed with ether. Solid compound 12b was isolated (0.59 g, 70%) (see Tables 1 and 2).

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