# Enantioselective Friedel–Crafts Alkylation of Indoles with Aromatic α,β-Unsaturated Aldehydes Catalyzed by Diphenylprolinol Silyl Ether

Zhi-Jing Wang,<sup>a</sup> Jian-Guo Yang,<sup>b</sup> Jun Jin,<sup>a</sup> Xin Lv,<sup>a</sup> Weiliang Bao\*<sup>a</sup>

<sup>a</sup> Department of Chemistry, Zhejiang University, Xixi Campus, Hangzhou 310028, P. R. of China Fax +86(571)88273814; E-mail: wlbao@css.zju.edu.cn

<sup>b</sup> Office of Assets Administration, Taizhou University, Linhai 317000, P. R. of China

Received 15 June 2009; revised 30 July 2009

**Abstract:** A catalytic enantioselective Friedel–Crafts alkylation of indoles with aromatic  $\alpha$ , $\beta$ -unsaturated aldehydes has been developed. The process is promoted by (*S*)-diphenylprolinol trimethyl silyl ether to afford, after sodium borohydride mediated reduction, 3-indolyl-3-phenylpropanols in high enantioselectivities and good yields.

Key words: organocatalysis, enantioselectivity, Friedel–Crafts alkylation, indoles, aromatic  $\alpha$ , $\beta$ -unsaturated aldehydes

The Friedel-Crafts reaction of aromatic compounds is one of the most important reactions for the formation of C-C bonds and direct derivatization of aromatic compounds in organic synthesis.1 Various aromatic compounds, including benzenes with electron-donating substituents, furans, pyrroles and indoles, have been successfully applied in a number of Friedel-Crafts reactions with electrophiles. The indole framework represents a privileged structural motif of established value in biologically active natural products and pharmaceutical compounds.<sup>2</sup> In this regard, the enantioselective Friedel-Crafts reaction of indoles is one of the most extensive areas of research. Catalytic enantioselective versions of this fundamental transformation have been reported, most of which use metal-based chiral complex catalysts.<sup>3</sup> Recently, asymmetric organocatalysis has attracted considerable attention in terms of its unique properties,<sup>4</sup> and there have been several reports of asymmetric Friedel-Crafts reactions of indole derivatives with organocatalysts. In 2002, MacMillan and coworkers first reported the asymmetric Friedel-Crafts reaction of indoles and N-alkyl indoles with alkyl  $\alpha$ ,  $\beta$ -unsaturated aldehydes using chiral imidazolidinones as catalysts, and demonstrated the feasibility of organocatalytic strategies based on LUMO-lowering activation of  $\alpha$ , $\beta$ -unsaturated aldehydes via the reversible formation of an iminium ion.<sup>5</sup> Since then, a variety of catalytic asymmetric reactions of indoles with electrophiles including alkyl  $\alpha,\beta$ -unsaturated aldehydes,<sup>6</sup> nitroalkenes,<sup>7</sup>  $\alpha,\beta$ -unsaturated ketones,8 imines,9 trifluoropyruvate,10 and electron-rich alkenes<sup>11</sup> have been reported.

However, the catalytic enantioselective Friedel–Crafts alkylation of indoles with aromatic  $\alpha$ , $\beta$ -unsaturated alde-

hydes is not well documented. There is, so far, only one example of the Friedel-Crafts reaction of N-methyl indole with cinnamaldehyde, 5,6c and no reaction of 1*H*-indole with cinnamaldehyde has been reported. The NH moiety on the indole has significant influence on its reactivity and 1H-indoles and N-alkyl indoles show different reactivities in many cases. In some reactions, 1H-indoles were more reactive than N-alkyl indoles,<sup>8b,10</sup> while in others, N-alkyl indoles were superior to 1H-indoles. In the enantioselective addition of indole to N-acyl imines reported by Antilla and co-workers, 1H-indole provided lower yields and enantioselectivities than N-alkyl indole.9c Due to the difference between 1H-indoles and N-alkyl indoles, the enantioselective Friedel-Crafts alkylation of 1H-indoles with aromatic  $\alpha,\beta$ -unsaturated aldehydes is of significance.

The products from  $\alpha$ , $\beta$ -unsaturated aldehydes are more versatile than those from nitroalkenes and  $\alpha$ , $\beta$ -unsaturated ketones, and the 1*H*-indole derivatives could easily undergo further derivatization on the nitrogen atom. Therefore, the enantioselective Friedel–Crafts reactions of 1*H*-indoles and  $\alpha$ , $\beta$ -unsaturated aldehydes would afford potentially useful 3-substituted indole derivatives.

Diarylprolinol silyl ethers, which were first utilized as efficient asymmetric catalysts by Jørgensen in 2005,<sup>12a</sup> have now been shown to be excellent organocatalysts for a wide variety of asymmetric reactions.<sup>12–16</sup> However, to the best of our knowledge, its application to catalytic enantioselective Friedel–Crafts alkylation has not been reported. Herein, we would like to report the catalytic enantioselective Friedel–Crafts reaction of indoles with aromatic  $\alpha$ , $\beta$ -unsaturated aldehyde derivatives catalyzed by diphenylprolinol silyl ether (Scheme 1).

The proposed enantioselective Friedel–Crafts reaction was first examined using indole (2a), cinnamaldehyde (3a), and 15 mol% of (S)-1 at room temperature in toluene. The desired Friedel–Crafts reaction product 4a was obtained, which was reduced by sodium borohydride in situ to the corresponding  $\gamma$ -alcohol 5a with moderate yield and enantioselectivity (50%, 57% ee). Addition of benzoic acid did not improve the result (Table 1, entry 2). Various solvents were then screened and, as shown in Table 1, it is noteworthy that not only the catalytic yield but also the asymmetric induction were highly dependent on the solvents employed. Among the solvents tested, polar protic methanol was found to be the best with respect to both

SYNTHESIS 2009, No. 23, pp 3994–4000 Advanced online publication: 12.10.2009 DOI: 10.1055/s-0029-1217038; Art ID: F12009SS © Georg Thieme Verlag Stuttgart · New York



Scheme 1 Enantioselective Friedel–Crafts reaction of indole with cinnamaldehyde

the catalytic activity and the asymmetric induction (Table 1, entry 5). As expected, enantiomeric excess increased with a decrease in reaction temperature, reaching 83% ee at -10 °C, and good yield was attained after 96 h (Table 1, entry 14). After screening of the loading of the catalyst, 20 mol% was selected as the optimal amount. Thus, **5a** was formed in the presence of 20 mol% of the catalyst (*S*)-**1**, in methanol at -10 °C, with 87% yield and 86% ee, respectively (Table 1, entry 15).

Under the above optimized reaction conditions (Table 1, entry 15), the generality of the reaction was investigated; the results are summarized in Table 2. All substituted cinnamaldehydes tested were converted with very high levels of stereocontrol into the expected products 5a-j (Table 2

entries 1–10). Cinnamaldehydes with electron-withdrawing groups (4-Br, 4-Cl, 4-F, 4-CF<sub>3</sub>, 3-CF<sub>3</sub>, 3-Cl, and 2-Cl) reacted efficiently with indole **2a** and gave good yields as well as excellent enantioselectivities (Table 2, entries 2– 8). The electron-donating groups (4-Me, 4-MeO, and 2-MeO) on the aromatic ring of cinnamaldehyde were tolerated, but had a negative influence on the the yield (Table 2, entries 9–10). The Friedel–Crafts alkylation of electron-rich indoles (**2b–e**) afforded the corresponding products **5k–n** with good results (Table 2, entries 11–14). Electron-deficient indoles (5-NO<sub>2</sub>) caused a decrease in the yield (Table 2, entry 15). As a limitation of the approach, it is worth mentioning that substitution in the  $\alpha$ -position ( $\alpha$ -methyl) of the cinnamaldehyde and the

Table 1 Optimization of the Reaction Conditions<sup>a</sup>

	+ H Ph	1) (S)-1 (cat.), solvent 2) excess NaBH <sub>4</sub> , MeOH 0 °C, 20 min						
Entry	3a Solvent	Time (h)	Temn (°C)	5a Cat (mol%)	Vield (%) <sup>b</sup>	ee (%)°		
1	toluene	36	r.t.	15	50	57		
2	toluene	36	r.t.	15	40	50 <sup>d</sup>		
3	CHCl <sub>3</sub>	36	r.t.	15	44	70		
4	CH <sub>2</sub> Cl <sub>2</sub>	36	r.t.	15	42	60		
5	MeOH	36	r.t.	15	65	73		
6	EtOH	36	r.t.	15	42	49		
7	MeCN	36	r.t.	15	46	72		
8	THF	36	r.t.	15	33	63		
9	DMF	36	r.t.	15	trace	n.d. <sup>e</sup>		
10	MeOH	72	0	10	67	77		
11	MeOH	72	0	15	70	80		
12	MeOH	72	0	20	76	82		
13	MeOH	96	-10	10	72	81		
14	MeOH	96	-10	15	75	83		
15	МеОН	96	-10	20	87	86		

Ph

<sup>a</sup> Reaction conditions: indole (1.0 mmol), cinnamaldehyde (0.5 mmol), catalyst (S)-1 (15 mol%) and solvent (1.0 mL).

<sup>b</sup> Isolated yield.

<sup>c</sup> Enantiomeric excess was determined by chiral HPLC analysis.

 $^{\rm d}$  Benzoic acid (15 mol%) was added as a co-catalyst.

<sup>e</sup> Not determined.

Table 2Ca	atalytic Asymmetric Frie	del–Crafts Reaction of Substituted	Indoles and Cinnamal	dehydes <sup>a</sup>		
R <sup>1</sup> IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	$+$ $H$ $R^2$ 3a-j	1) ( <i>S</i> )-1 (20 mol%), MeOH -10 °C, 96 h 2) excess NaBH₄, MeOH 0 °C, 20 min		НС		
Entry	<b>R</b> <sup>1</sup>	R <sup>2</sup>	Product	Yield (%) <sup>b</sup>	ee (%) <sup>e</sup>	
1	H ( <b>2a</b> )	Ph ( <b>3a</b> )	5a	87	86	
2	H ( <b>2a</b> )	$4-BrC_{6}H_{4}(\mathbf{3b})$	5b	79	91	
3	H ( <b>2a</b> )	$4\text{-}\mathrm{ClC}_{6}\mathrm{H}_{4}\left(\mathbf{3c}\right)$	5c	74	86	
4	H ( <b>2a</b> )	$4-FC_{6}H_{4}(\mathbf{3d})$	5d	79	87	
5	H ( <b>2a</b> )	$4-F_{3}CC_{6}H_{4}$ ( <b>3e</b> )	5e	72	86	
6	H ( <b>2a</b> )	$3-F_{3}CC_{6}H_{4}$ ( <b>3f</b> )	5f	75	93	
7	H ( <b>2a</b> )	$3\text{-ClC}_{6}\text{H}_{4}(\mathbf{3g})$	5g	70	89	
8	H ( <b>2a</b> )	$2\text{-ClC}_{6}\text{H}_{4}(\mathbf{3h})$	5h	79	95	
9	H ( <b>2a</b> )	$4-MeC_{6}H_{4}(3i)$	5i	56	86	
10	H ( <b>2a</b> )	$4\text{-}\text{MeOC}_{6}\text{H}_{4}\left(\mathbf{3j}\right)$	5j	63	87	
11	5-MeO ( <b>2b</b> )	Ph ( <b>3a</b> )	5k	70	88	

Table 2	Catalytic .	Asymmetric	Friedel-	Crafts	Reaction	of	Substituted	Indoles	and	Cinnamaldel	hydes <sup>a</sup>
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<sup>a</sup> Reaction conditions: 2 (1.0 mmol), 3 (0.5 mmol), (S)-1 (0.10 mmol), MeOH (1.0 mL).

Ph (3a)

Ph (3a)

Ph (3a)

Ph (3a)

51

5m

5n

50

<sup>b</sup> Isolated yield.

12

13

14

15

<sup>c</sup> Enantiomeric excess was determined by chiral HPLC analysis.

6-MeO (2c)

5-Me (2d)

7-Me (2e)

 $5 - O_2 N (2f)$ 

4-position (4-Me and 4-COOMe) of the indole ring had a detrimental effect on reactivity. The NH-moiety on the indole ring could not be blocked. For example, N-methyl indole did not react with cinnamaldehyde. This feature has already been observed in other organocatalytic Friedel-Crafts reactions of indoles believed to proceed via a dual activation mechanism.<sup>7a,10</sup> Alkyl  $\alpha,\beta$ -unsaturated aldehydes such as crotonaldehyde seemed unreactive under our catalytic conditions.



Scheme 2 The absolute configuration of the product 5a was determined as S by correlation to 8a

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For the determination of the absolute configuration of the catalytic product 5a, we designed a route as shown in Scheme 2. From the comparison of its optical rotation with that of (S)-8a, which was synthesized by the reaction of cinnamaldehyde with 1-methylindole catalyzed by chiral imidazolidinones,<sup>5</sup> the absolute configuration of **5a** was determined to be 3S.

74

75

77

60

87

87

94

86

In summary, we have developed a highly enantioselective Friedel–Crafts reaction of indoles with aromatic  $\alpha,\beta$ -unsaturated aldehydes using diphenylprolinol silyl ether as the organocatalyst. It could be a good complementary approach to MacMillan's protocol. The reaction is applicable to a variety of aromatic  $\alpha,\beta$ -unsaturated aldehydes and indoles, and high enantioselectivities are attained. The approach avoids the use of any additional acid or base, and the products are potentially useful. Further investigations into synthetic applications of the method are underway.<sup>17</sup>

Commercial reagents were used as received, unless otherwise stated. The organocatalyst (S)-1 was prepared following the procedure described in the literature. Racemates of 1 were prepared using racemic diphenylprolinol TMS-ether as the catalyst. All <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were measured in CDCl<sub>3</sub> on a Bruker Avance 400 MHz instrument with TMS as the internal standard. Chemical shifts are expressed in ppm and *J* values are given in Hz. Data for <sup>1</sup>H NMR are reported as follows: chemical shift (ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet) and integral. Data for <sup>13</sup>C NMR are reported as ppm. MS was obtained using EI ionization on an HP6989B mass spectrometer. HRMS was obtained using ESI ionization on an Apex III (7.0 tesla) Fourier transform ion cyclotron resonance (FTICR) mass spectrometer. Melting points are uncorrected. Optical rotations were measured using a 1 mL cell with a 1 dm path length on a Perkin–Elmer 341 digital polarimeter and are reported as follows:  $[\alpha]_D^{20}$  (*c* in g per 100 mL of solvent). HPLC analysis was performed using Daicel Chiralpak AD-H and OD-H columns.

# Indole Addition to α,β-Unsaturated Aldehydes (Table 2, Entry 1); Typical Procedure

To catalyst (*S*)-1 (32 mg, 0.10 mmol, 20 mol%) dissolved in MeOH (1 mL) in a 12 mL vial at -10 °C, was added cinnamaldehyde (66 mg, 0.5 mmol). The mixture was stirred for 30 min, and indole (117 mg, 1.0 mmol) was added to the mixture. The vial was capped and the mixture was stirred at -10 °C for 96 h. Excess NaBH<sub>4</sub> (57 mg, 1.5 mmol) was added, followed by the addition of MeOH (1 mL). The -10 °C cooling bath was replaced by an ice bath, and the mixture was stirred for a further 20 min. The mixture was slowly added to sat. NH<sub>4</sub>Cl (5 mL) at 0 °C and extracted with Et<sub>2</sub>O (10 × 3 mL). The organic layer was collected, washed with H<sub>2</sub>O (5 mL) and brine (5 mL), then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc–hexane, 1:1) to afford **5a**.

#### (S)-3-(1*H*-indol-3-yl)-3-phenylpropan-1-ol (5a)

Yield: 109 mg (87%); colorless oil;  $[\alpha]_D^{20}$  +4.0 (*c* 0.8, CHCl<sub>3</sub>); 86% ee by HPLC (Daicel Chiralpak OD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major}$  = 44.6 min;  $\tau_{minor}$  = 57.8 min).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.00 (br s, 1 H), 7.43 (d, *J* = 7.6 Hz, 1 H), 7.22–7.31 (m, 5 H), 7.14 (dd, *J* = 8.8, 7.6 Hz, 2 H), 6.98–7.02 (m, 2 H), 4.37 (t, *J* = 7.8 Hz, 1 H), 3.60–3.67 (m, 2 H), 2.40–2.49 (m, 1 H), 2.21–2.30 (m, 1 H), 1.44 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 38.6, 39.2, 61.3, 110.1, 119.3, 119.5, 119.6, 121.1, 122.0, 126.1, 126.8, 127.8, 128.4, 136.5, 144.7.

EI-MS:  $m/z = 251 [M^+]$ .

Anal. Calcd for  $C_{17}H_{17}NO$ : C, 81.24; H, 6.82; N, 5.77. Found: C, 81.13; H, 6.86; N, 5.47.

#### 5b

Pale-red oil;  $[\alpha]_D^{20}$  –12.0 (*c* 1.0, CHCl<sub>3</sub>); 91% ee by HPLC (Daicel Chiralpak OD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 43.8 \text{ min}; \tau_{minor} = 66.0 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.05 (br s, 1 H), 7.35–7.40 (m, 3 H), 7.31 (d, *J* = 8.0 Hz, 1 H), 7.13–7.18 (m, 3 H), 6.99–7.03 (m, 2 H), 4.35 (t, *J* = 7.8 Hz, 1 H), 3.58–3.69 (m, 2 H), 2.38–2.47 (m, 1 H), 2.16–2.24 (m, 1 H), 1.44 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 38.4, 38.5, 61.0, 111.1, 119.0, 119.3, 121.1, 122.2, 126.7, 129.6, 131.4, 136.6, 143.8.

EI-MS:  $m/z = 329 [M^+; {}^{79}Br], 331 [M^+; {}^{81}Br].$ 

Anal. Calcd for  $C_{17}H_{16}BrNO$ : C, 61.83; H, 4.88; N, 4.24. Found: C, 61.85; H, 4.94; N, 4.31.

#### 5c

Pale-yellow oil;  $[\alpha]_D^{20}$  –9.8 (*c* 2.0, CHCl<sub>3</sub>); 86% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 27.2 \text{ min}; \tau_{minor} = 19.1 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.04 (br s, 1 H), 7.40 (d, *J* = 7.6 Hz, 1 H), 7.33 (d, *J* = 7.8 Hz, 1 H), 7.21–7.25 (m, 4 H), 7.15 (t,

J = 7.6 Hz, 1 H), 7.00-7.05 (m, 2 H), 4.38 (t, J = 7.6 Hz, 1 H), 3.60-3.71 (m, 2 H), 2.40-2.47 (m, 1 H), 2.17-2.26 (m, 1 H), 1.37 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 38.5 (2 × C), 61.0, 111.1, 119.2, 119.3, 119.4, 121.0, 122.2, 126.7, 128.5, 129.2, 131.7, 136.6, 143.3. EI-MS: *m*/*z* = 285 [M<sup>+</sup>; <sup>35</sup>Cl], 287 [M<sup>+</sup>; <sup>37</sup>Cl].

Anal. Calcd for C<sub>17</sub>H<sub>16</sub>ClNO: C, 71.45; H, 5.64; N, 4.90. Found: C, 71.36; H, 5.69; N, 4.82.

#### 5d

Pale-yellow oil;  $[\alpha]_D^{20}$  –3.8 (c = 1.27, CHCl<sub>3</sub>); 87% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 24.8 \text{ min}$ ;  $\tau_{minor} = 18.4 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.05$  (br s, 1 H), 7.38 (d, J = 8.0 Hz, 1 H), 7.28 (d, J = 8.4 Hz, 1 H), 7.20–7.23 (m, 2 H), 7.13 (t, J = 7.6 Hz, 1 H), 7.00 (t, J = 7.4 Hz, 1 H), 6.89–6.95 (m, 3 H), 4.34 (t, J = 7.6 Hz, 1 H), 3.56–3.66 (m, 2 H), 2.36–2.44 (m, 1 H), 2.13–2.20 (m, 1 H), 1.68 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 38.3, 38.6, 61.0, 111.1, 115.0, 115.2, 119.3, 119.4, 121.0, 122.1, 126.7, 129.2 (d, *J* = 7.4 Hz), 136.5, 140.4 (d, *J* = 2.3 Hz), 161.3 (d, *J* = 242.2 Hz).

HRMS (ESI): m/z [M + Na]<sup>+</sup> calcd for C<sub>17</sub>H<sub>16</sub>FNONa: 292.1108; found: 292.1104.

EI-MS:  $m/z = 269 [M^+]$ .

#### 5e

Pale-yellow oil;  $[a]_D^{20}$  +5.6 (*c* 0.8, CHCl<sub>3</sub>); 86% ee by HPLC (Daicel Chiralpak OD-H column; *n*-hexane-*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 36.5 \text{ min}; \tau_{minor} = 56.0 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.15 (br s, 1 H), 7.53 (d, *J* = 8.0 Hz, 2 H), 7.43 (d, *J* = 8.4 Hz, 3 H), 7.35 (d, *J* = 8.0 Hz, 1 H), 7.19 (t, *J* = 7.6 Hz, 1 H), 7.02–7.07 (m, 2 H), 4.49 (t, *J* = 7.6 Hz, 1 H), 3.61–3.72 (m, 2 H), 2.44–2.52 (m, 1 H), 2.22–2.31 (m, 1 H), 1.65 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 38.3, 38.8, 60.8, 111.2, 118.5, 119.2, 119.5, 121.2, 122.3, 125.3 (q, *J* = 4.0 Hz), 125.6 (t, *J* = 280.5 Hz), 128.1, 128.2, 136.5, 149.0.

HRMS (ESI): m/z [M + Na]<sup>+</sup> calcd for C<sub>18</sub>H<sub>16</sub>F<sub>3</sub>NONa: 342.1076; found: 342.1079.

EI-MS:  $m/z = 319 [M^+]$ .

#### 5f

Pale-yellow oil;  $[\alpha]_D^{20}$  +7.2 (*c* 2.2, CHCl<sub>3</sub>); 93% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 11.3 \text{ min}; \tau_{minor} = 10.8 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.15$  (br s, 1 H), 7.62 (s, 2 H), 7.50 (d, J = 7.6 Hz, 1 H), 7.45 (d, J = 7.6 Hz, 1 H), 7.33–7.38 (m, 2 H), 7.19 (t, J = 7.6 Hz, 1 H), 7.04–7.08 (m, 2 H), 4.91 (t, J = 7.8 Hz, 1 H), 3.60–3.71 (m, 2 H), 2.43–2.52 (m, 1 H), 2.22–2.32 (m, 1 H), 1.79 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 38.4, 38.8, 60.8, 111.2, 118.5, 119.1, 119.4, 121.2, 122.2, 123.0 (q, *J* = 3.8 Hz), 124.2 (q, *J* = 270.8 Hz), 124.4 (q, *J* = 3.8 Hz), 126.6, 128.3, 131.3 (2 × C), 136.5, 145.9.

HRMS (ESI): m/z [M + Na]<sup>+</sup> calcd for C<sub>18</sub>H<sub>16</sub>F<sub>3</sub>NONa: 342.1076; found: 342.1072.

EI-MS:  $m/z = 319 [M^+]$ .

# 5g

Colorless oil;  $[\alpha]_D^{20}$ –4.6 (*c* 1.33, CHCl<sub>3</sub>); 89% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 18.0 \text{ min}; \tau_{minor} = 16.0 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.06 (br s, 1 H), 7.42 (d, *J* = 8.0 Hz, 1 H), 7.31 (d, *J* = 8.4 Hz, 1 H), 7.27 (s, 1 H), 7.12–7.20 (m, 4 H), 7.00–7.03 (m, 2 H), 4.36 (t, *J* = 7.8 Hz, 1 H), 3.58–3.69 (m, 2 H), 2.37–2.45 (m, 1 H), 2.17–2.27 (m, 1 H), 1.76 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 38.4, 38.7, 60.9, 111.2, 118.8, 119.3, 119.4, 121.1, 122.2, 126.1, 126.4, 126.7, 127.9, 129.6, 134.2, 136.5, 147.0.

EI-MS: *m*/*z* = 285 [M<sup>+</sup>; <sup>35</sup>Cl], 287 [M<sup>+</sup>; <sup>37</sup>Cl].

Anal. Calcd for C<sub>17</sub>H<sub>16</sub>ClNO: C, 71.45; H, 5.64; N, 4.90. Found: C, 71.55; H, 5.60; N, 4.88.

#### 5h

Colorless oil;  $[a]_D^{20}$  +73.1 (*c* 2.33, CHCl<sub>3</sub>); 95% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 26.7 \text{ min}; \tau_{minor} = 17.9 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.10 (br s, 1 H), 7.44 (d, *J* = 8.0 Hz, 1 H), 7.34–7.36 (m, 1 H), 7.29 (d, *J* = 8.4 Hz, 1 H), 7.18–7.22 (m, 1 H), 6.99–7.15 (m, 5 H), 4.93 (t, *J* = 7.0 Hz, 1 H), 3.56–3.79 (m, 2 H), 2.41–2.45 (m, 1 H), 2.19–2.27 (m, 1 H), 1.98 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 34.9, 38.4, 61.2, 111.1, 118.5, 119.3, 119.4, 121.5, 122.1, 126.9, 127.1, 127.3, 129.1, 129.4, 133.5, 136.4, 142.2.

EI-MS:  $m/z = 285 [M^+; {}^{35}Cl], 287 [M^+; {}^{37}Cl].$ 

Anal. Calcd for  $C_{17}H_{16}$ ClNO: C, 71.45; H, 5.64; N, 4.90. Found: C, 71.42; H, 5.57; N, 4.90.

#### 5i

Yellow oil;  $[\alpha]_D^{20}$  –9.0 (*c* 1.13, CHCl<sub>3</sub>); 86% ee by HPLC (Daicel Chiralpak OD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 38.0 \text{ min}; \tau_{minor} = 52.8 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.02$  (br s, 1 H), 7.48 (d, J = 8.0 Hz, 1 H), 7.32 (d, J = 8.0 Hz, 1 H), 7.22 (d, J = 8.0 Hz, 2 H), 7.16 (t, J = 7.6 Hz, 1 H), 7.09 (d, J = 7.6 Hz, 1 H), 7.02–7.05 (m, 2 H), 4.36 (t, J = 7.6 Hz, 1 H), 3.66–3.71 (m, 2 H), 2.42–2.50 (m, 1 H), 2.31 (s, 3 H), 2.23–2.28 (m, 1 H), 1.45 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 20.9$ , 38.7, 38.8, 61.4, 111.0, 119.2, 119.5, 119.8, 121.0, 122.0, 126.9, 127.6, 129.1, 135.6, 136.5, 141.7.

EI-MS:  $m/z = 265 [M^+]$ .

Anal. Calcd for  $C_{18}H_{19}NO$ : C, 81.47; H, 7.22; N, 5.28. Found: C, 81.47; H, 7.19; N, 5.27.

#### 5j

Pale-yellow oil;  $[\alpha]_D^{20}$  –8.8 (*c* 0.8, CHCl<sub>3</sub>); 87% ee by HPLC (Daicel Chiralpak OD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 63.7 \text{ min}; \tau_{minor} = 82.1 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 8.01 (br s, 1 H), 7.44 (d, *J* = 7.6 Hz, 1 H), 7.31 (d, *J* = 8.0 Hz, 1 H), 7.22 (d, *J* = 8.0 Hz, 2 H), 7.14 (t, *J* = 7.6 Hz, 1 H), 6.99–7.02 (m, 2 H), 6.80 (d, *J* = 8.8 Hz, 2 H), 4.33 (t, *J* = 7.8 Hz, 1 H), 3.75 (s, 3 H), 3.66–3.77 (m, 2 H), 2.40–2.48 (m, 1 H), 2.19–2.27 (m, 1 H), 1.36 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 38.4, 38.8, 55.2, 61.4, 111.0, 113.8, 119.3, 119.5, 120.0, 120.9, 122.0, 126.8, 128.7, 136.6, 136.8, 157.9.

#### EI-MS: m/z = 281 [M<sup>+</sup>].

Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO<sub>2</sub>: C, 76.84; H, 6.81; N, 4.98. Found: C, 76.84; H, 6.76; N, 4.90.

#### 5k

Yellow oil;  $[\alpha]_D^{20}$ –42.5 (*c* 0.53, CHCl<sub>3</sub>); 88% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 28.9 \text{ min}; \tau_{minor} = 98.7 \text{ min}$ ).

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<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.99 (br s, 1 H), 7.22–7.30 (m, 4 H), 7.14–7.16 (m, 2 H), 6.95 (d, *J* = 1.6 Hz, 1 H), 6.84 (d, *J* = 2.4 Hz, 1 H), 6.78 (dd, *J* = 6.4, 2.4 Hz, 1 H), 4.30 (t, *J* = 7.6 Hz, 1 H), 3.71 (s, 1 H), 3.60–3.66 (m, 2 H), 2.36–2.44 (m, 1 H), 2.18–2.27 (m, 1 H), 1.70 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 38.5, 39.1, 55.8, 61.2, 101.5, 111.7, 111.9, 119.2, 121.9, 126.1, 127.2, 127.8, 128.4, 131.7, 144.7, 153.6.

EI-MS: m/z = 281 [M<sup>+</sup>].

Anal. Calcd for  $C_{18}H_{19}NO_2$ : C, 76.84; H, 6.81; N, 4.98. Found: C, 76.87; H, 6.85; N, 4.86.

### 51

Pale-yellow oil;  $[\alpha]_D^{20}$  +5.3 (*c* 1.13, CHCl<sub>3</sub>); 87% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 47.5$  min;  $\tau_{minor} = 41.6$  min).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.94 (br s, 1 H), 7.22–7.29 (m, 5 H), 7.14 (t, *J* = 7.0 Hz, 1 H), 6.85 (d, *J* = 2.0 Hz, 1 H), 6.75 (d, *J* = 2.0 Hz, 1 H), 6.66 (dd, *J* = 6.4, 2.2 Hz, 1 H), 4.29 (t, *J* = 8.0 Hz, 1 H), 3.75 (s, 1 H), 3.57–3.65 (m, 2 H), 2.36–2.44 (m, 1 H), 2.17–2.27 (m, 1 H), 1.70 (br s, 1 H).

 ${}^{13}C NMR (100 MHz, CDCl_3): \delta = 38.5, 39.2, 55.6, 61.2, 94.6, 109.1, 119.4, 119.9, 120.0, 121.3, 126.1, 127.8, 128.4, 137.2, 144.8, 156.3.$ 

EI-MS:  $m/z = 281 [M^+]$ .

Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO<sub>2</sub>: C, 76.84; H, 6.81; N, 4.98. Found: C, 76.80; H, 6.77; N, 4.95.

#### 5m

Colorless oil;  $[\alpha]_D^{20}$  –26.8 (*c* 0.93, CHCl<sub>3</sub>); 87% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 20.4$  min;  $\tau_{minor} = 17.0$  min).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.91 (br s, 1 H), 7.22–7.30 (m, 5 H), 7.13–7.16 (m, 2 H), 6.95 (d, *J* = 8.4 Hz, 1 H), 6.88 (s, 1 H), 4.31 (t, *J* = 7.8 Hz, 1 H), 3.56–3.65 (m, 2 H), 2.37–2.42 (m, 1 H), 2.36 (s, 3 H), 2.19–2.26 (m, 1 H), 1.70 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 21.5$ , 38.7, 39.1, 61.3, 110.8, 118.8, 118.9 (2 × C), 121.3, 123.6, 126.0, 127.0, 127.8, 128.4, 134.8, 144.8.

EI-MS:  $m/z = 265 [M^+]$ .

Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO: C, 81.47; H, 7.22; N, 5.28. Found: C, 81.47; H, 7.23; N, 5.30.

#### 5n

Colorless oil;  $[\alpha]_D^{20}$  +21.3 (*c* 1.2, CHCl<sub>3</sub>); 94% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major} = 15.9 \text{ min}; \tau_{minor} = 17.8 \text{ min}$ ).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 8.02 (br s, 1 H), 7.28–7.36 (m, 3 H), 7.19–7.22 (m, 2 H), 7.03 (d, J = 2.4 Hz, 1 H), 6.97–7.00 (m, 2 H), 4.40 (t, J = 7.6 Hz, 1 H), 3.66–3.71 (m, 2 H), 2.45–2.53 (m, 1 H), 2.46 (s, 3 H), 2.26–2.34 (m, 1 H), 1.68 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 16.5, 38.6, 39.3, 61.3, 117.2, 119.4, 120.0, 120.2, 120.8, 122.5, 126.0, 126.3, 127.8, 128.3, 136.0, 144.8.

EI-MS:  $m/z = 265 [M^+]$ .

Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO: C, 81.47; H, 7.22; N, 5.28. Found: C, 81.35; H, 7.28; N, 5.21.

#### 50

Yellow solid; mp 157–159 °C;  $[\alpha]_D^{20}$ –169.9 (*c* 0.67, CHCl<sub>3</sub>); 86% ee by HPLC (Daicel Chiralpak AD-H column; *n*-hexane–*i*-PrOH, 90:10; flow rate: 1.0 mL/min;  $\tau_{major}$  = 23.2 min;  $\tau_{minor}$  = 38.8 min).

<sup>1</sup>H NMR (400 MHz,  $CDCl_3$ ):  $\delta = 8.54$  (br s, 1 H), 8.41 (s, 1 H), 8.05 (d, J = 9.2 Hz, 1 H), 7.19–7.36 (m, 7 H), 4.44 (t, J = 7.8 Hz, 1 H), 3.64–3.71 (m, 2 H), 2.42–2.48 (m, 1 H), 2.26–2.34 (m, 1 H), 1.64 (br s, 1 H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 38.5, 38.8, 60.8, 111.0, 116.8, 117.9, 122.6, 123.9, 126.3, 126.7, 127.7, 128.7, 139.5, 141.5, 143.6.

EI-MS:  $m/z = 296 [M^+]$ .

Anal. Calcd for  $C_{17}H_{16}N_2O_3$ : C, 68.91; H, 5.44; N, 9.45. Found: C, 68.82; H, 5.41; N, 9.46.

# Acknowledgment

This work was financially supported by the Specialized Research Fund for the Doctoral Program of Higher Education of China (20060335036) and Zhejiang Provincial Natural Science Foundation of China under Grant No. Y406049.

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