

Article

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What is the Structure of the Antitubercular Natural Product Eucapsitrione?

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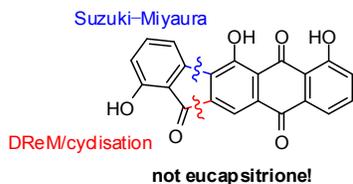
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ABSTRACT

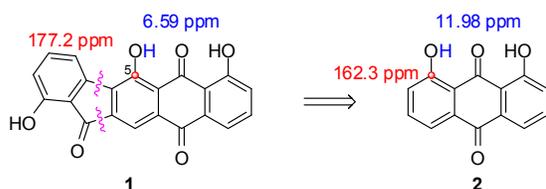
1,5,7-Trihydroxy-6*H*-indeno[1,2-*b*]anthracene-6,11,13-trione (**1**), proposed to be the antitubercular natural product eucapsitrione, has been synthesised in 43% overall yield, and six steps, including a key Suzuki–Miyaura biaryl coupling and a directed remote metalation (DReM)-initiated cyclisation. The physical and spectroscopic properties of **1** do not match the data reported for the natural product. At this time there is insufficient information available to enable a structure reassignment. During the optimization of the Suzuki–Miyaura

coupling an unprecedented biaryl coupling *ortho* to the borono group was observed. The scope of this unusual reaction has been investigated.

INTRODUCTION

In 2010, an investigation of a cyanobacterium of the previously unexplored genus *Eucapsis* was conducted with the aim of isolating and identifying new antitubercular agents.¹ The study led to the isolation of a novel natural product, which was named eucapsitrione and assigned structure **1** (Scheme 1) on the basis of spectroscopic and mass spectrometric data.¹

Scheme 1. The proposed structure **1** of eucapsitrione and anomalous NMR data compared to chrysazin (**2**), a potential precursor for total synthesis.



Eucapsitrione exhibits potent activity against rapidly growing *M. tuberculosis* with a minimum inhibitory concentration of 3.1 μM . Importantly, the natural product was also active at a similar concentration in the low-oxygen-recovery assay (LORA),² which has been developed to mimic the non-replicating persistent (NRP) state that makes tuberculosis (TB) difficult to treat, and contributes to antimicrobial resistance in *M. tuberculosis*.³ Conversely, eucapsitrione did not affect the viability of *Staphylococcus aureus*, *Escherichia coli*, *Candida albicans*, or *Mycobacterium smegmatis* at 55 μM , and had an $\text{IC}_{50} > 28 \mu\text{M}$ against the mammalian Vero cell line, suggesting a mode of action that is selective for *M. tuberculosis*. These properties make eucapsitrione a promising lead for the discovery of novel drugs for the treatment of TB infection, as noted in several reviews.⁴⁻⁸ In addition, no fluorenones, and only a few anthraquinones,⁹ have ever been isolated from a cyanobacterium. Indeed, the

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3 fused pentacyclic 6*H*-indeno[1,2-*b*]anthracene-6,11,13-trione skeleton of **1** is unique amongst
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5 natural products. Thus, **1** was an attractive candidate for synthesis.
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10 Our interest in eucapsitrione was further piqued because several of the natural product's
11 spectroscopic data did not seem to fit the proposed structure. Most noticeable amongst these
12 was the assignment of a signal at 6.59 ppm in the ¹H NMR spectrum (in *d*₆-DMSO) to the C5
13 phenolic proton. Such protons, *peri* to carbonyl groups, are typically strongly hydrogen-
14 bonded and thus resonate considerably downfield of non-hydrogen bonded phenols. For
15 example, in chrysazin (**2**) (Scheme 1), the analogous protons resonate at 11.98 ppm in *d*₆-
16 DMSO. Furthermore, a signal in the ¹³C NMR spectrum of eucapsitrione at 177.2 ppm was
17 assigned to C5. Such carbons generally resonate in the range 150–160 ppm.¹⁰ Indeed, the
18 corresponding carbons of chrysazin (**2**) give rise to a signal at 162.3 ppm (Scheme 1).¹¹
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32 The report that **1** produces only one carbonyl absorption band in its IR spectrum (1616 cm⁻¹)
33 also seemed incongruous with the proposed structure. The absorption frequency of a carbonyl
34 group is typically lowered when hydrogen-bonded to a *peri* phenolic proton, as evident in the
35 infrared spectrum of chrysazin (**2**, 1678 and 1621 cm⁻¹),¹² and related α-
36 hydroxyanthraquinones.^{12,13} As such it would be reasonable to expect the structure **1** to give
37 rise to at least two distinct carbonyl absorption bands. It is also worth noting that the exact
38 mass determined for eucapsitrione [M-H]⁻ (357.04291)¹ is 6.7 ppm out from the calculated
39 mass (357.0405) of the C₂₁H₉O₆⁻ ion, raising some doubt about the molecular formula.
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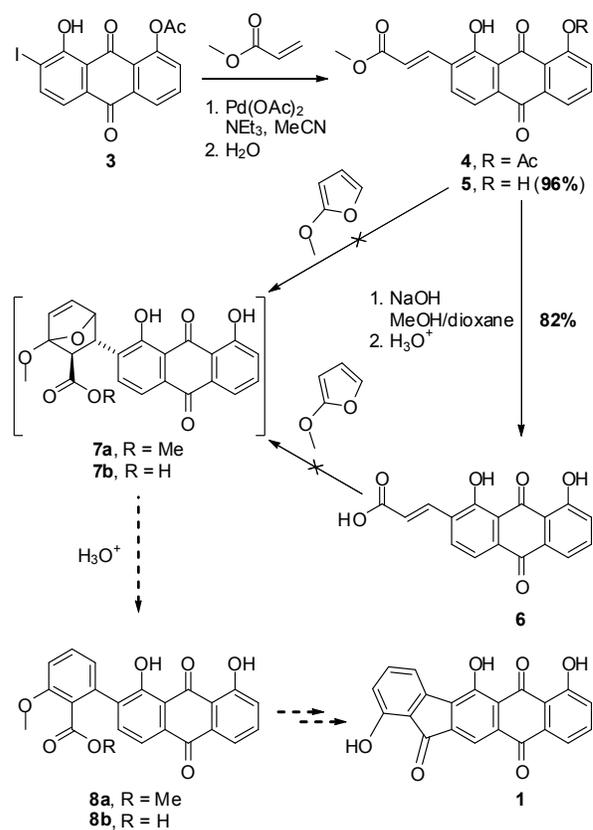
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52 The irregularities in the mass spectrometric and spectroscopic data discussed above suggested
53 that the proposed structure **1** required validation.
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RESULTS AND DISCUSSION

Synthesis of the ring system represented by **1** has only been reported a handful of times, often without substitution;^{14,15} otherwise, substituted derivatives have been produced in low yield¹⁶ and as reaction byproducts.¹⁷ With no existing practical routes upon which to base the synthesis of the structure **1**, two novel pathways were devised beginning from the cheap and readily available chrysazin (**2**) (Scheme 1).

The first approach is outlined in Scheme 2. The known iodide **3**, prepared simply in two steps from chrysazin (**2**),^{18,19} was subjected to a Heck reaction with methyl acrylate, under standard conditions,²⁰ initially providing a mixture of the expected coupling product **4** and the corresponding phenol **5** resulting from deacetylation. Deprotection of the phenol was actually desired, and was forced to completion by addition of water to the reaction mixture once the Heck coupling was complete, affording **5** exclusively, in excellent yield. Our intention was to effect a regioselective Diels–Alder cycloaddition of **5** with 2-methoxyfuran to provide adduct **7a**, which we assumed would aromatise upon acidic workup to give the biaryl **8a**. Subsequent electrophilic ring closure could then provide access to **1**.

Scheme 2. An unsuccessful approach to eucapsitrione involving key Heck and Diels–Alder reactions.



Although 2-methoxyfuran reacts readily with very good dienophiles,²¹⁻²⁴ and with some less electron-deficient dienophiles in the presence of Lewis acid catalysts,²⁵ our attempts to induce a Diels–Alder reaction with **5** were unsuccessful. Heating the pair under reflux in toluene led to partial decomposition of 2-methoxyfuran and no trace of a Diels–Alder adduct by ^1H NMR spectroscopy.

It was reasoned that addition of a Lewis acid catalyst may be able to activate the dienophile through coordination to both the ester carbonyl group and chelation with the anthraquinone carbonyl/*peri* hydroxy moiety in **5**, the latter mitigating the electron-donating effects of the phenol. With multiple potential coordination sites in the dienophile **5**, experiments were

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3 therefore conducted in the presence of both sub- and super-stoichiometric quantities of Lewis
4 acids, and a range of temperatures. The attempted Diels–Alder reaction of **5** in the presence
5 of Yb(OTf)₃;²⁶⁻²⁹ a silica-supported TiCl₄-based catalyst that has been used previously with
6 2-methoxyfuran;²⁵ and phenylboronic acid, which was expected to form a chelate borate
7 complex with α -hydroxyquinone moiety;³⁰ were all unsuccessful, leading only to
8 decomposition of the diene.
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18 Arylboronic acids have successfully catalysed a number of Diels–Alder cycloadditions of
19 furan, though where the dienophile is an α,β -unsaturated carboxylic acid.^{31,32} In these cases,
20 *ortho*-bromo- and -iodobenzeneboronic acids are noted as being amongst the most effective
21 catalysts,³¹⁻³³ and the carboxylic acid group of the dienophile is key to their mode of
22 activation.^{31,34,35} Thus the acrylic acid **6** was prepared and, the cycloaddition with 2-
23 methoxyfuran was attempted in the presence of *ortho*-bromobenzeneboronic acid. The
24 reaction was first tried in 1,2-dichloroethane (DCE); however, **6** is poorly soluble in DCE and
25 other solvents compatible with arylboronic acid catalysis,³² so, lastly, solvent-free
26 experiments with a ten-fold excess of diene were attempted. In all cases, only decomposition
27 of 2-methoxyfuran was observed with no evidence for a Diels–Alder reaction.
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43 With further experimentation, and judicious choice of catalyst, and/or phenol protecting
44 groups that render the dienophile more electron-deficient, it may have been possible to
45 achieve the desired cycloaddition. However, with no indication of any cycloadduct formation
46 across all attempts, an alternative synthetic pathway was pursued.
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53 In the second, and ultimately successful synthetic route to **1**, a Suzuki–Miyaura cross-
54 coupling of iodide **3** was used to construct the key biaryl bond (Scheme 3 and Table 1).
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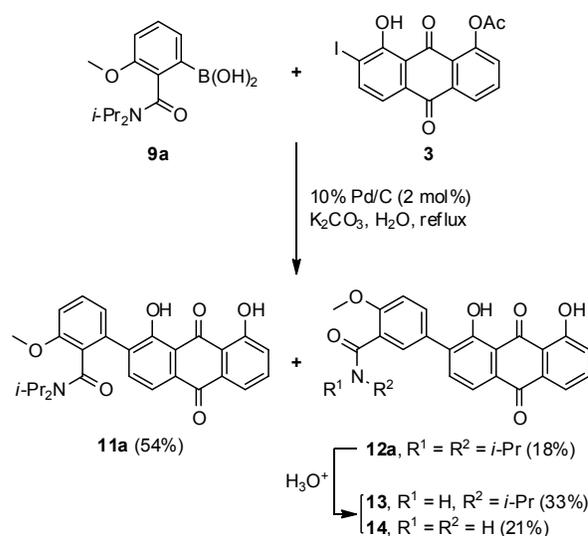
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3 Initially the boronic acid **9a**,³⁶ derived via directed *ortho*-lithiation/borylation of the
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5 corresponding diisopropylamide, was investigated for this purpose.
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10 Suzuki–Miyaura coupling of halophenols can be carried out under very convenient
11 conditions (aqueous K₂CO₃, Pd/C),^{37,38} and we thought these should be applicable to the
12 current synthesis. Following work-up and chromatography, the reaction of boronic acid **9a**
13 with iodide **3** under these conditions yielded a product that displayed twice the number of ¹H
14 NMR resonances expected of biaryl **11a**. Initially, this observation was tentatively attributed
15 to atropisomerism about the biaryl and aryl-carboxamide bonds, giving rise to a mixture of
16 diastereomers. A simplification of the ¹H NMR spectrum of this material at high temperature
17 would have confirmed our hypothesis; however, there was little change in the spectrum on
18 heating. Similarly, cleavage of the very bulky diisopropylamide should lead to a simplified
19 ¹H NMR spectrum in the resulting carboxylic acid if atropisomerism was at play, but the
20 amide was remarkably resistant to hydrolysis under basic conditions, with only starting
21 material recovered after nine days of heating under reflux in 1 M NaOH/dioxane.
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38 An attempt to effect hydrolysis under strongly acidic conditions instead resulted in a mixture
39 of products arising from *N*-dealkylation.³⁹ Analysis of this mixture made it clear that our
40 atropisomerism hypothesis was incorrect and, rather, the Suzuki–Miyaura coupling had
41 produced two constitutional isomers. Two of the products isolated from this reaction
42 appeared, by ¹H NMR spectroscopy, to possess a 1,2,4-trisubstituted benzene moiety.
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48 Ultimately, NOESY and 2D NMR spectroscopic experiments confirmed the unexpected
49 substitution pattern in compounds **13** and **14** (Scheme 3).
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This revelation prompted a reinvestigation of the Suzuki–Miyaura coupling of **3** and **9a**, and careful preparative TLC allowed the separation of the two isomeric products of this reaction. Thus, in addition to the expected biaryl **11a**, this reaction gave the minor regioisomer **12a**, which must arise from direct arylation of the aromatic methine *para* to the methoxy group – a process which must either be preceded or followed by a deboronation step in order to furnish **12a**.

Scheme 3. Suzuki coupling of iodide **3** and boronic acid **9a** produces two regioisomers **11a** and **12a**, discovered after isolation of the *N*-dealkylation products **13** and **14**. Yields of **11a** and **12a** are NMR yields.

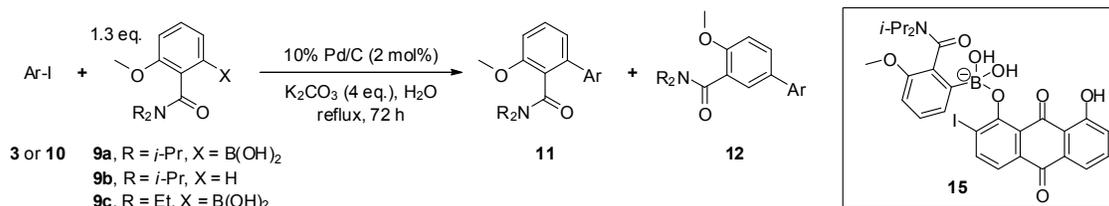


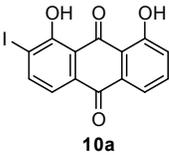
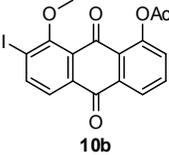
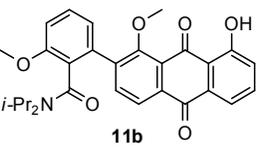
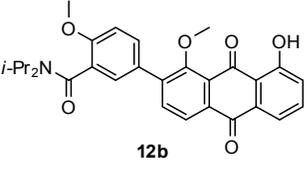
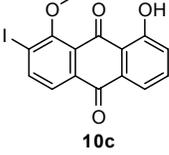
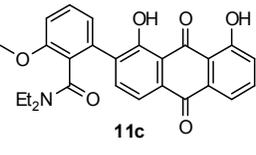
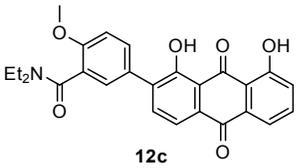
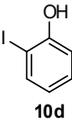
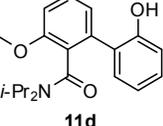
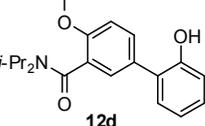
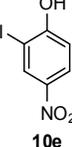
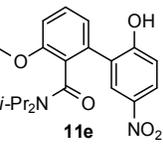
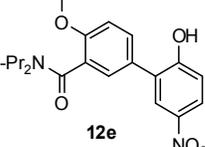
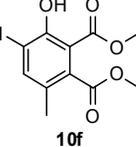
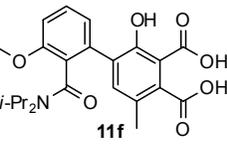
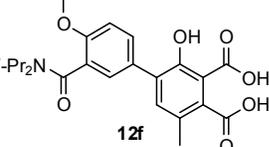
Reports of Pd/C-catalysed C–H activations,^{40–49} whilst still relatively uncommon, have become more prevalent in recent years, as palladium on carbon is being applied to transformations previously reserved for more complicated homogeneous catalytic systems.^{50–52} Amongst these, direct arylations remain limited,^{53–58} and are rarely reported for non-heteroaromatic systems.⁵⁹ The formation of a significant proportion of **12a** from the reaction

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3 of **3** and **9a** under simple conditions therefore presented as an interesting side reaction worth
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5 investigating further (Table 1).
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10 The Suzuki–Miyaura reaction conditions were applied to 2-iodochryszin (**10a**), which
11 confirmed that the direct arylation to give **12a** was not an aberration, and that the acetyl
12 group of **3** likely has no role in the reaction (entry 2). These initial experiments revealed that
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14 **9a** is readily deboronated under the reaction conditions,⁶⁰⁻⁶² as a significant quantity of amide
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16 **9b** was observed in the crude reaction product in both cases. Therefore, to assess the
17
18 importance of the borono group, the arylation with **9b** was attempted (entry 3). This reaction
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20 gave only a trace of biaryl **12a**. Whilst this result is informative, hinting at a mechanism
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22 involving electrophilic attack at the most activated position *para*-to the methoxy substituent in
23
24 both **9a** and **9b**, the increased yield of **12a** in entry 1 suggests that the borono group directs
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26 *ortho*-arylation of **9a**, prior to deboronation. To the best of our knowledge, this
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28 transformation is unique amongst metal-catalysed arylation reactions.*
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59 * However, a similar transformation has been achieved with a hypervalent iodine.⁶³
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Table 1. Scope and mechanism investigation of the direct arylation of arylboronic acid **9a** and related compounds.

Entry	Iodide	9	11	Yield % ^a	12	Yield % ^a
1	3^b	9a	11a	54	12a	18
2		9a	11a	54	12a	13
3	3 	9a	11b 	—	12a 	—
4		9a	11b	—	12b	—
5	3	9c	11c 	79 86 (82) ^c	12c 	2 1 ^c
6		9a	11d 	56 (51)	12d 	—
7		9a	11e 	7	12e 	—
8		9a	11f 	60	12f 	—
9	10d^d	9a	11d	21	12d	—
10	10d^e	9a	11d	51	12d	—

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3 ^a Yields determined by ¹H NMR spectroscopy using 1,3,5-trimethoxybenzene as an internal
4 standard. ^b Reaction time 7 d instead of 72 h. ^c 3 mol % of Pd/C used, reaction time 36 h
5 instead of 72 h. ^d BQ (1 eq.) added to the reaction mixture. ^e Chrysazin (1 eq.) added to the
6 reaction mixture.
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14 The regioselectivity of the reaction, and prevalence of intramolecular direct arylation
15 reactions in the literature,⁶⁴⁻⁶⁹ suggest that the borono substituent is acting as a directing
16 group, perhaps via an intermediate phenyl boronate such as **15**. Protection of the free
17 hydroxyl of iodide **3** should preclude formation of such an intermediate, and so the methyl
18 ethers **10b** and **10c** were prepared (entries 3 and 4). Unfortunately, neither iodide **10b**⁷⁰ nor
19 **10c** underwent any reaction with **9a**, (i.e. the normal Suzuki–Miyaura coupling was also
20 completely suppressed), providing no insight into the mechanism by which the borono
21 substituent activates the *ortho* position.
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34 Before conducting more intensive investigations regarding the mechanism of this direct
35 arylation, the scope of the reaction was probed. It was apparent from the reaction of the less
36 hindered *N,N*-diethylamidoboronic acid **9c** that the extremely bulky substituent *ortho* to the
37 boronic acid is required for the direct arylation to compete with conventional cross-coupling
38 as only a trace of the direct arylation product **12c** was observed (entry 5). The hindered
39 boronic acid **9a** was therefore retained in subsequent reactions with selected alternative aryl
40 iodides **10d–f**. The reaction of 2-iodophenol (**10d**) gave only the expected Suzuki–Miyaura
41 coupling product **11d** in modest yield (entry 6). This suggested that the anthraquinone moiety
42 in the iodochrysazins **3** and **10a** played a role in the direct arylation reaction. The carbonyl
43 groups in the anthraquinones enhance the acidity of the phenolic hydroxyl, decrease the
44 electron density of the aryl iodide and provide some steric encumbrance. We thus chose other
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3 iodides that mimicked these properties. 4-Nitro-2-iodophenol (**10e**) gave only a low yield of
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5 the normal Suzuki–Miyaura product **11e** (entry 7), indicating that enhanced phenol acidity is
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7 not sufficient to promote direct arylation. The Suzuki–Miyaura coupling of dimethyl
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9 iodophthalate **10f**, which should quite closely mimic the stereoelectronic properties of the
10
11 iodochryszins, was accompanied by saponification to give the phthalic acid **11f**, but again,
12
13 no direct arylation product **12f** was observed (entry 8). The importance of oxidative
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15 properties of the chryszin anthraquinone moiety^{71,72} of **3** were also considered;
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17 benzoquinone (BQ) promotes a variety of C–H activations,⁷³⁻⁷⁵ and so was also tested as an
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19 additive in the reaction with 2-iodophenol (entry 9), but this only led to a reduction in yield of
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21 the expected Suzuki–Miyaura product **11d**. And, finally, addition of an equivalent of
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23 chryszin (**2**) to the experiment with 2-iodophenol (**10d**) had no significant effect on the
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25 outcome of that reaction.
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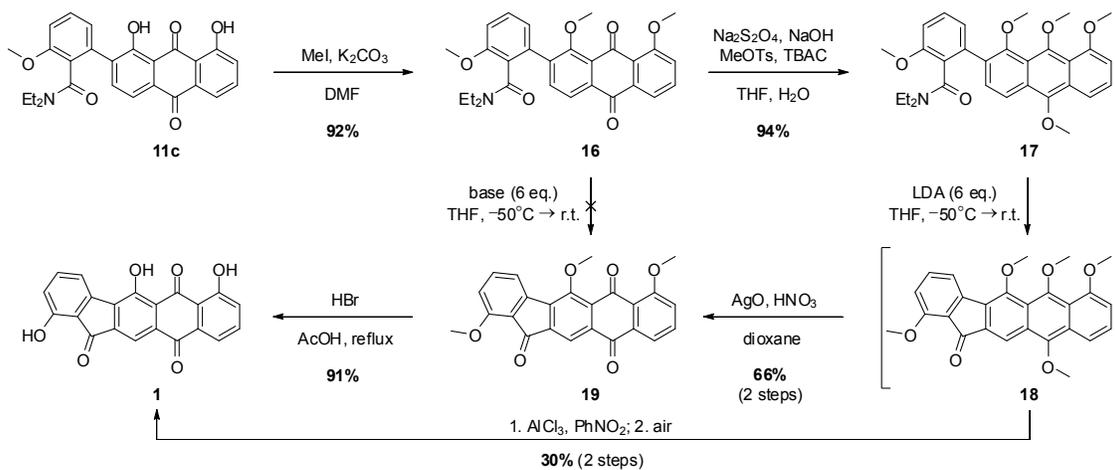
32 Thus, while an interesting diversion, the scope of this direct arylation appears to be limited.
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34 Moreover, the side-reaction detracted from the efficiency of the desired synthesis of **1**.
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36 Fortunately, the Suzuki–Miyaura coupling of iodide **3** with the less hindered *N,N*-
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38 diethylamide **9c** produced biaryl **11c** in good yield with only traces of the undesired direct
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40 arylation product. The yield of **11c** was improved simply by increasing the loading of Pd/C to
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42 3 mol%, which furnished the biaryl in 82% isolated yield.
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47 With a good yield of the biaryl intermediate **11c** in hand, our attention turned to the end game
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49 (Scheme 4). Cyclisation of **11c** via an electrophilic mechanism (e.g., Friedel–Crafts
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51 acylation) was considered, but predicted to be troublesome for several reasons. The amide is
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53 not reactive enough, and hydrolysis was likely to be very difficult. In addition, acylation
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55 would be required at the deactivated 3-position on the pendant anthraquinone moiety.
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5 Therefore, we set out to employ the powerful directing ability of the diethylamido group of
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7 **11c** in a directed remote metalation (DReM).⁷⁶⁻⁷⁹ Literature precedents suggested that both
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9 the phenolic hydroxyls and quinone carbonyl groups of **11c** required protection for a
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11 successful DReM-initiated cyclisation.^{76,78} Indeed, when **11c** was converted to the dimethyl
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13 ether **16**, and treated with excess LDA a drastic colour change resulted, but returned only
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15 starting material on work-up. A subsequent D₂O quench experiment revealed that no
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17 lithiation of **16** had occurred. It was reasoned that LDA was reducing the quinone through α -
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19 hydride transfer,⁸⁰ and the hydroquinone dianion was responsible for the intense colour.
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21 Lithium hexamethyldisilazide (LHMDS) or lithium 2,2,6,6-tetramethylpiperidide (LTMP)
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23 cannot effect reduction through this mechanism; however, treatment of **16** with these bases
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25 delivered the same results as treatment with LDA. We surmise that the lithium amide bases
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27 form an electron-transfer complex with **16** in a similar manner to alkyllithiums with
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29 quinones,⁸¹ resulting in a colour change but no apparent reaction post work-up.
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36 Relenting, we prepared the anthracene **17** in excellent yield by reductive methylation of **16**
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38 (Scheme 4). Methyl tosylate⁸² proved to be as effective as dimethyl sulfate for this reaction,
39
40 which was fortunate given the scheduling of the latter. DReM-cyclisation of **17** proceeded
41
42 smoothly, though attempts to purify fluorenone **18** resulted in significant degradation – likely
43
44 via photooxidation to its corresponding endoperoxide^{83,84} – and as a result it was not isolated.
45
46 Instead, freshly prepared, crude **18** was subjected to global demethylation,⁸⁵ and aerial
47
48 oxidation⁸⁶ furnished the target structure **1** in modest yield across two steps from **17**. The
49
50 remarkably poor solubility of **1** complicated purification, contributing to the low yield.
51
52 Therefore, oxidative demethylation of crude **18**, followed by demethylation of **19** proved a
53
54 more convenient and higher yielding route to **1**.
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Scheme 4. Completion of the total synthesis of the proposed structure **1** of eucapsitrione, by DReM/cyclisation and deprotection.



The first empirical evidence indicating that **1** does not represent the structure of the natural product eucapsitrione relates to solubility. The spectroscopic data for the natural product were obtained in *d*₆-DMSO, a solvent in which the synthetic material was practically insoluble. Even at elevated temperature, a ¹H NMR spectrum of **1** in DMSO could not be obtained, thus a direct comparison of synthetic **1** with the natural product could not be achieved. The recalcitrant insolubility of **1** forced us to acquire NMR spectra in *d*₅-pyridine.

¹³C resonances are typically less solvent dependent than those for protons, and occur over a wide spectral range;^{87,88} thus, comparison of the ¹³C NMR spectrum of **1** with that of the natural product (Figure 1) is more useful than comparing ¹H NMR resonances, though the latter is included for completeness (Table 2). To estimate the effect of the solvent on the NMR chemical shifts of **1**, chryszin (**2**) was used as a comparator. Indeed the ¹³C resonances

of chryszazin in d_5 -pyridine and in d_6 -DMSO¹¹ were very similar, having a mean absolute difference of 0.3 ppm, and a maximum difference of 0.7 ppm (Table S1).

The ¹³C NMR spectroscopic data obtained for **1** vary significantly from those reported for eucapsitrione, with the majority of the resonances differing by > 0.7 ppm (the maximum solvent-dependent difference observed with chryszazin) (Table S1), and a number even deviating by > 5 ppm (Figure 1). As predicted, **1** does not give rise to a resonance close to 177.2 ppm, the signal in the ¹³C NMR spectrum of eucapsitrione that raised our suspicions about the assigned structure of the natural product.

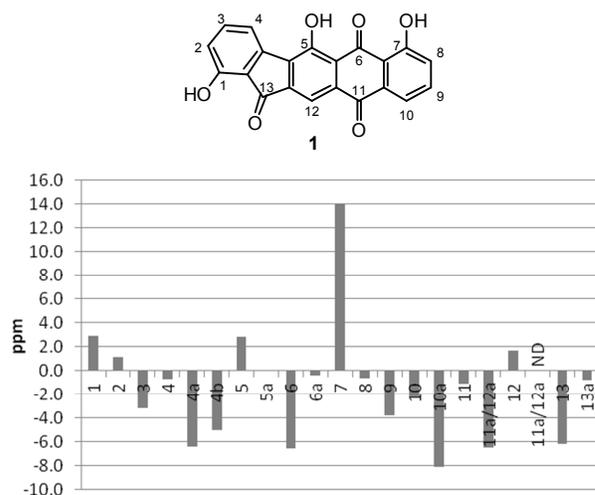


Figure 1. ¹³C NMR chemical shift differences between eucapsitrione (d_6 -DMSO)¹ and synthetic **1** (d_5 -pyridine). The mean $|\Delta\delta|$ is 3.7 ppm and the maximum is 14.0 ppm. A ¹³C NMR resonance of C11a or C12a in **1** could not be experimentally observed (ND = No Data); it is likely obscured by the solvent peak at 136.5–135.5 ppm based upon ¹³C NMR chemical shift predictions (see Table S2 and experimental for **1**), and so was excluded from these shift difference calculations.

Although less meaningful, comparison of the ¹H NMR data (Table 2) did reveal a particularly significant discrepancy. The distinctive singlet at 8.34 ppm, arising from H12, *peri* to two

carbonyl groups in **1**, is reported to appear much further upfield at 7.15 ppm in eucapsitrione.¹

Table 2. ¹H NMR and IR spectroscopic data for **1** versus those reported for eucapsitrione.¹

1 (d ₅ -pyr) ^{b, c}	¹ H NMR (ppm)		IR (ν _{max} cm ⁻¹) ^a	
	1 (CDCl ₃) ^b	eucapsitrione ¹ (d ₆ -DMSO) ^d	1	eucapsitrione ¹
	12.48 (s, OH) ^e	13.99 (s, OH)	3380	
	11.95 (s, OH) ^e	13.28 (s, OH)	1696	
	8.39 (s, OH) ^e		1669	
8.34 (s)	8.16 (s)	7.60 (t)	1628	1616
7.91 (dd)	7.89 (dd)	7.56 (m)		1559
7.78 (d)	7.75 (dd)	7.56 (m)		1457
7.67 (app. t)	7.60 (dd)	7.52 (dd)		1372
7.51 (dd)	7.49 (dd)	7.15 (s)		1331
7.39 (d)	7.35 (dd)	7.14 (dd)		1273
7.13 (d)	6.88 (dd)	7.12 (dd)		1210
		6.59 (s, OH)		1152

^a Neat, ATR ^b 500 MHz. ^c Exchangeable protons not observed due to exchange with residual D₂O. ^d 600 MHz. ^e Confirmed by deuterium exchange experiment.

In contrast the NMR spectroscopic data, the infrared spectra are directly comparable, and the spectrum of synthetic **1** shows considerable disparity with that of eucapsitrione (Table 2). As predicted, **1** gives rise to three distinct carbonyl absorption bands. The absorptions at 1669 and 1628 cm⁻¹ are, respectively, characteristic of the free and hydrogen-bonded carbonyl groups of an α -hydroxyanthraquinone.^{12,13} The remaining peak at 1696 cm⁻¹ is therefore attributed to the fluorenone carbonyl group. The lack of an absorption band at this frequency in the IR spectrum of eucapsitrione suggests that it is not a fluorenone.

Though the discrepancies between their physical and spectroscopic properties are already strong evidence that eucapsitrione does not possess structure **1**, the inability to directly

compare the NMR spectroscopic data is rather unsatisfying. Accordingly, the ^{13}C NMR frequencies for **1** in DMSO solution were calculated.

To determine a suitable computational methodology to apply to **1**, chrysazin (**2**) reprised its role as a standard, as the experimental ^{13}C NMR spectrum could be compared with calculated frequencies (Table 3). Several methodology and basis set combinations were employed for these calculations, of which HF 6-31G* proved to be the most accurate for chrysazin (**2**), giving a mean absolute difference between calculated and experimental ^{13}C shifts of 2.9 ppm, and a maximum difference of 7.0 ppm.

Table 3. Comparison of experimental and calculated ^{13}C NMR chemical shifts for chrysazin (**2**).

δ_{exp} 2 ¹¹ (d_6 -DMSO)	δ_{calc} 2 HF/6-31G* (DMSO)	δ_{calc} 2 B3LYP/6-31G* //HF/6-311+G(2d,p) (DMSO)	δ_{calc} 2 B3LYP/ 6-31G* (DMSO)
192.9	192.7	196.2	186.4
182.2	178.8	194.0	177.4
162.3	160.2	166.8	157.1
162.3	160.2	166.8	157.1
138.3	137.0	144.9	131.9
138.3	137.0	144.9	131.9
134.1	133.3	142.0	128.3
134.1	133.3	142.0	128.3
125.3	120.3	128.9	119.5
125.3	120.3	128.9	119.5
120.2	118.1	126.1	116.3
120.2	118.1	126.1	116.3
116.8	109.8	124.5	112.3
116.8	109.8	124.5	112.3
Mean $\Delta\delta$	2.9	6.2	5.3
Maximum $\Delta\delta$	7.0	11.8	6.5

All values in ppm.

Using this basis set, the mean and maximum differences between the ^{13}C NMR chemical shifts calculated for **1** and those reported for eucapsitrione were 5.3 and 18.9 ppm, respectively (Figure 2; see also Table S2 for calculations carried out using alternative methodology and basis set combinations). Notably, HF 6–31G* most poorly predicted the shift of the most upfield ^{13}C -resonances in chrysozin, whilst for eucapsitrione, the significantly larger maximum shift difference again arose from the carbon resonating at 177.2 ppm.

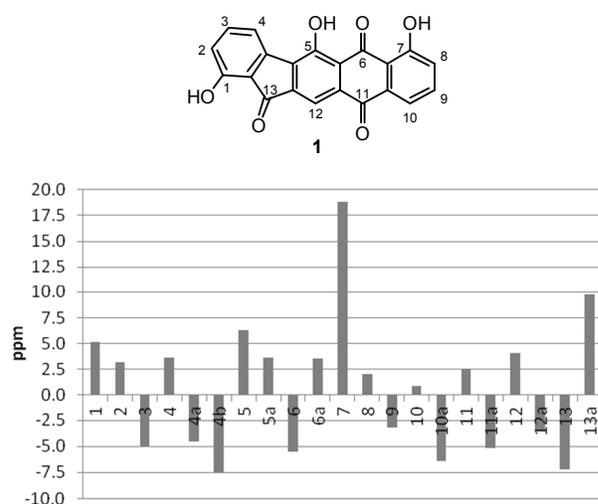


Figure 2. ^{13}C NMR chemical shift differences between eucapsitrione (d_6 -DMSO)¹ and the calculated shifts for **1** in DMSO (HF 6–31G*). The mean $|\Delta\delta|$ is 5.3 ppm and the maximum is 18.9 ppm.

CONCLUSION

In conclusion, 1,5,7-trihydroxy-6*H*-indeno[1,2-*b*]anthracene-6,11,13-trione (**1**) has been synthesised in 43% overall yield, and six steps, from the known boronic acid **9c** and iodide **3** (eight steps from commercially available starting materials). The physical characteristics and spectroscopic data for **1** do not match those reported for the natural product eucapsitrione. Unfortunately, we believe the existing, published spectra of eucapsitrione do not permit a

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3 reassignment of structure, partly because of uncertainty about whether spurious peaks in the
4 NMR spectra arise from the natural product, or significant impurities. Given the promising
5 biological activity of the eucapsitrione, another isolation and reinvestigation of its structure is
6 warranted.
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14 During the course of the synthesis of **1**, an unprecedented direct arylation reaction was
15 observed, in which a borono group appears to act as a traceless *ortho*-activator. At this stage
16 the reaction appears to have rather specific substrate requirements, but with optimization and
17 broadening of scope, this reaction could prove valuable.
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24 25 **EXPERIMENTAL SECTION**

26 27 **Materials and Methods**

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29 All solvents were distilled prior to use. Anhydrous THF was obtained from a Pure Solv 5-
30 Mid Solvent Purification System (Innovative Technology Inc.). ‘Dry DMF’ and ‘dry MeCN’
31 refers to solvent that was stored over activated 3A molecular sieves for at least 24 h.⁹⁰ ‘Dry
32 acetone’ refers to acetone that was stirred over anhydrous CaSO₄ for 4 h before being
33 distilled under N₂. Tetramethylethylenediamine (TMEDA) and diisopropylamine were dried
34 over and distilled from CaH₂ under N₂ onto KOH pellets and stored as such under N₂.^{77,90}
35
36 Triisopropyl borate was distilled and stored under N₂. The concentrations of solutions of *n*-
37 and *sec*-BuLi were determined by titration with *N*-benzylbenzamide.⁹¹ 1-Hydroxy-8-acetoxy-
38 9,10-athraquinone was prepared from chrysazin (**2**) by a known method.¹⁸ *N,N*-diethyl-2-
39 methoxybenzamide⁹² and *N,N*-diisopropyl-2-methoxybenzamide (**9b**)⁹³ were prepared by
40 known methods from *o*-anisic acid. Dimethyl 3-hydroxy-6-methylphthalate⁹⁴ was prepared
41 according to a known method. All other reagents and materials were purchased from
42 commercial suppliers and used as received.
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5 All reactions, excepting the preparations of 1-hydroxy-8-acetoxy-9,10-anthraquinone and 1-
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7 hydroxy-2-iodo-8-acetoxy-9,10-anthraquinone (**3**) were conducted in flame-dried glassware
8
9 under an atmosphere of N₂ with the use of syringe and septum-cap techniques. Where
10
11 indicated, reaction temperatures refer to the temperature of the heating or cooling bath. All
12
13 organic extracts were dried over MgSO₄, filtered and evaporated under reduced pressure at
14
15 40–45°C. Trace residual solvent was removed under a stream of N₂ or, where indicated,
16
17 using high vacuum.
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19

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23 Reaction progress was monitored by thin layer chromatography (TLC) using Merck
24
25 aluminium-backed TLC silica gel 60 F₂₅₄ plates, which were also used for preparative TLC.
26
27 Spots were visualised directly (coloured compounds) or using ultraviolet light. Flash column
28
29 chromatography was performed using Davisil chromatographic silica media LC60A 40–63
30
31 μm.
32
33

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36 ¹H and ¹³C nuclear magnetic resonance (NMR) spectra were acquired using Bruker Avance
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38 IIIHD (600 MHz for ¹H and 150 MHz for ¹³C), Bruker Avance IIIHD (500MHz for ¹H and
39
40 125 MHz for ¹³C), and Varian (400 MHz for ¹H and 100 MHz for ¹³C) spectrometers, as
41
42 indicated. Deuteriochloroform (CDCl₃) was used as the solvent for NMR samples unless
43
44 otherwise indicated. Spectra were calibrated against CHCl₃ (for ¹H spectra; δ 7.26 ppm) or
45
46 CDCl₃ (for ¹³C spectra; δ 77.16 ppm) peaks. Where *d*₅-pyridine was used as a solvent, spectra
47
48 were calibrated against the most upfield peaks of C₅D₄HN (for ¹H spectra; δ 7.22 ppm) and
49
50 C₅D₅N (for ¹³C spectra; δ 123.90). Where *d*₆-DMSO was used as a solvent, spectra were
51
52 calibrated against CD₃SOCD₂H (for ¹H spectra; δ 2.50) or (CD₃)₂SO (for ¹³C spectra; δ
53
54 39.52). ¹H and ¹³C NMR assignments were made based upon 2D and NOE NMR
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3 experiments, as noted for each assigned compound. Complete atom-numbered structures of
4
5 each compound synthesised can be found in the Supporting Information.
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9
10 High resolution mass spectra were recorded on a Waters Liquid Chromatograph Premier
11
12 mass spectrometer with time-of-flight mass analyser, using atmospheric pressure chemical
13
14 ionisation (APCI) in positive or negative mode as indicated. Infrared spectra were recorded
15
16 on a Perkin-Elmer Spectrum One FT-IR spectrometer with Attenuated Total Reflectance
17
18 (ATR) using neat samples. Melting points were determined using a Reichert hot stage
19
20 melting point apparatus.
21
22

23 24 25 **Synthesis**

26
27 *1-Hydroxy-2-iodo-8-acetoxy-9,10-anthraquinone (3)*.¹⁹ Iodic acid (4.38 g, 24.9 mmol) was
28
29 added to a stirred mixture of 1-hydroxy-8-acetoxy-9,10-anthraquinone¹⁸ (7.06 g, 25.0 mmol)
30
31 in water (60 mL) and dioxane (190 mL). Iodine (3.17 g, 12.5 mmol) was added, and the
32
33 resulting mixture was heated at reflux for 18 h before being cooled to room temperature. The
34
35 mixture was then poured into water (2 L) and extracted with CHCl₃ (3 × 900 mL). The
36
37 organic extract was washed with 1M aqueous Na₂S₂O₃ solution (500 mL), water (500 mL)
38
39 and brine (500 mL), dried and evaporated. The crude solid residue was subjected to flash
40
41 chromatography. Elution with PhMe gave **3** (2.38 g) as an orange solid. Additionally, impure
42
43 fractions were recrystallised from PhMe to give **3** as orange needles (2.06 g, total yield 44%),
44
45 mp 224–226°C [lit.¹⁹ 222–224°C]. R_f(PhMe): 0.25; IR (ATR) ν_{max} cm⁻¹: 1765 (OC=O), 1667
46
47 (C10=O), 1639 (C9=O); ¹H NMR (400 MHz, CDCl₃) δ 13.50 (s, 1H), 8.28 (dd, *J* = 7.6, 1.2
48
49 Hz, 1H), 8.20 (d, *J* = 8.0 Hz, 1H), 7.84 (dd [app. t], *J*₁ = *J*₂ = 8.0 Hz, 1H), 7.56 (d, *J* = 8.0
50
51 Hz, 1H), 7.45 (dd, *J* = 8.0, 1.2 Hz, 1H), 2.47 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 187.8,
52
53 181.5, 169.6, 161.4, 150.9, 146.5, 136.1, 135.3, 132.8, 130.6, 126.2, 124.3, 120.5, 116.0,
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95.7, 21.3; HRMS (APCI-) m/z: $[M]^{*-}$ Calcd for $C_{16}H_9IO_5$ 407.9495; Found 407.9510. This compound has been reported previously, but only with 1H NMR data provided.¹⁹ The 1H NMR data obtained match those in literature.

(E)-Methyl 3'-(8-acetoxy-1-hydroxy-9,10-anthraquinon-2-yl)acrylate (**4**). A mixture of iodide **3** (0.410 g, 1.00 mmol) in MeCN (45 mL) was sparged with N_2 before methyl acrylate (0.29 mL, 3.2 mmol), NEt_3 (0.43 mL, 3.1 mmol) and $Pd(OAc)_2$ (45 mg, 0.20 mmol, 20 mol%) were added. The reaction mixture was then stirred at 70°C under N_2 for 14 h before being cooled to room temperature. The mixture was diluted with CH_2Cl_2 (200 mL), and the solution was washed with brine (3 × 30 mL), vacuum filtered through Celite, dried and evaporated. The residue was subjected to flash chromatography. Elution with CH_2Cl_2 afforded **5** (77 mg, 24%) identical with the material described below. Further elution with CH_2Cl_2 gave **4** (52 mg, 14%), which crystallised from EtOAc as red prisms, mp 226–230°C. $R_f(CH_2Cl_2)$: 0.25; IR (ATR) ν_{max} cm^{-1} : 1765 (OC=O), 1716 (C1'=O), 1667 (C10=O), 1633 (C9=O); 1H NMR (400 MHz, $CDCl_3$) δ 13.39 (s, 1H), 8.28 (dd, $J = 7.6, 1.2$ Hz, 1H), 8.00 (d, $J = 16.4$ Hz, 1H), 7.87 (d, $J = 8.0$ Hz, 1H), 7.84 (dd [app. t], $J_1 = J_2 = 8.0$ Hz, 1H), 7.81 (d, $J = 8.0$ Hz, 1H), 7.44 (dd, $J = 8.0, 1.2$ Hz, 1H), 6.77 (d, $J = 16.4$ Hz, 1H), 3.84 (s, 3H), 2.49 (s, 3H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 188.4, 181.4, 169.7, 167.3, 161.7, 150.8, 137.7, 136.0, 135.6, 135.4, 133.2, 130.6, 130.1, 126.2, 124.7, 122.4, 119.0, 116.9, 52.1, 21.3; HRMS (APCI-) m/z: $[M]^{*-}$ Calcd for $C_{20}H_{14}O_7$ 366.0740; Found 366.0735.

(E)-Methyl 3'-(1,8-dihydroxy-9,10-anthraquinon-2-yl)acrylate (**5**). A mixture of iodide **3** (0.404 g, 0.990 mmol) in dry MeCN (60 mL) was sparged with N_2 before NEt_3 (0.43 mL, 3.1 mmol), methyl acrylate (0.29 mL, 3.2 mmol) and $Pd(OAc)_2$ (42 mg, 0.19 mmol, 19 mol%) were added. The resulting mixture was stirred at 70°C under N_2 for 5 h then water (3 mL)

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3 was added and stirring was continued at 70°C 64 h. The reaction mixture was cooled to room
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5 temperature, diluted with CH₂Cl₂ (300 mL) and washed with brine (3 × 50 mL). The organic
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7 phase was vacuum filtered through Celite, washed with saturated aqueous citric acid (2 × 100
8
9 mL), brine (100 mL) and then dried and evaporated. The residue was dissolved in CH₂Cl₂,
10
11 filtered through a plug of silica gel and washed through with CH₂Cl₂, affording **5** (0.308 g,
12
13 96%) as an orange solid, mp 215–218°C. R_f (CH₂Cl₂): 0.45; IR (ATR) ν_{max} cm⁻¹: 1716
14
15 (C1'=O), 1668 (C10=O), 1621 (C9=O); ¹H NMR (400 MHz, CDCl₃) δ 12.86 (s, 1H), 11.96
16
17 (s, 1H), 8.00 (d, *J* = 16.4 Hz, 1H), 7.89 (d, *J* = 8.0 Hz, 1H), 7.844 (dd, *J* = 7.2, 1.2 Hz, 1H),
18
19 7.842 (d, *J* = 8.0 Hz, 1H), 7.71 (dd, *J* = 8.4, 7.6 Hz, 1H), 7.32 (dd, *J* = 8.6, 1.0 Hz, 1H), 6.77
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21 (d, *J* = 16.0 Hz, 1H), 3.84 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 193.4, 181.3, 167.2, 162.8,
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23 161.5, 137.7, 137.5, 136.0, 134.2, 133.6, 130.0, 125.0, 122.6, 120.4, 119.7, 116.3, 115.9,
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25 52.1; HRMS (APCI-) *m/z*: [M]⁻ Calcd for C₁₈H₁₂O₆ 324.0634; Found 324.0635.
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32 (*E*)-3'-(1,8-Dihydroxy-9,10-anthraquinon-2-yl)acrylic acid (**6**). 4 M NaOH (0.29 mL) was
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34 added to a suspension of acrylate **5** (91 mg, 0.28 mmol) in 3:1 dioxane:MeOH (4 mL), and
35
36 the resulting purple mixture was heated under reflux for 24 h. The reaction mixture was
37
38 acidified with 1 M HCl (5 mL), forming an orange precipitate, which was collected by
39
40 vacuum filtration, washed with water (*ca.* 300 mL) and dried. The aqueous washes were
41
42 extracted with EtOAc (3 × 60 mL). The extract was dried and evaporated, giving an orange
43
44 solid, which was combined with the collected precipitate and triturated with boiling CH₂Cl₂
45
46 to give **6** (72 mg, 83%) as a dark orange solid, mp 291–293°C. R_f (1:49 AcOH:CH₂Cl₂) 0.25;
47
48 IR (ATR) ν_{max} cm⁻¹: 3200–2400 (COOH), 1684 (C1'=O), 1662 (C10=O), 1623 (C9=O); ¹H
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50 NMR (500 MHz, *d*₆-DMSO) δ 12.43 (br s, 3H, 3 × OH), 8.23 (d, *J* = 8.0 Hz, 1H, H3), 7.86
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52 (d, *J* = 16.2 Hz, 1H, H3'), 7.83 (dd, *J* = 8.2, 7.7 Hz, 1H, H6), 7.72 (dd, *J* = 7.5, 1.0 Hz, 1H,
53
54 H5), 7.69 (d, *J* = 8.0 Hz, 1H, H4), 7.40 (dd, *J* = 8.4, 1.0 Hz, 1H, H7), 6.79 (d, *J* = 16.2 Hz,
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3 1H, H2'); ¹³C NMR (125 MHz, *d*₆-DMSO) δ 192.1 (C9), 181.1 (C10), 167.3 (C1'), 161.4
4 (C8), 160.1 (C1), 137.6 (C6), 136.0 (C3'), 135.5 (C3), 133.7 (C4a), 133.3 (C10a), 129.0 (C2),
5
6 124.6 (C7), 123.1 (C2'), 119.4 (C5), 118.6 (C4), 116.4 (C9a), 116.0 (C8a); HRMS (APCI-)
7
8 m/z: [M]⁻ Calcd for C₁₇H₁₀O₆ 310.0477; Found 310.0480. NMR assignments were made
9
10 with the assistance of COSY, HQSC and HMBC experiments.
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16 (2-(Diisopropylcarbamoyl)-3-methoxyphenyl)boronic acid (**9a**).⁹⁵ A stirred solution of
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18 TMEDA (1.30 mL, 8.67 mmol) in anhydrous THF (30 mL) at -78°C under N₂ was treated
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20 with a 0.96 M solution of *sec*-BuLi in cyclohexane (8.60 mL, 8.26 mmol). To this was added
21
22 a solution of *N,N*-diisopropyl-2-methoxybenzamide⁹³ (1.76 g, 7.50 mmol) in anhydrous THF
23
24 (15 mL) dropwise over 10 min. The resulting mixture was stirred at -78°C for 2 h before
25
26 being treated with B(Oi-Pr)₃ (5.20 mL, 22.5 mmol) and allowed to warm to room temperature
27
28 overnight. The mixture was cooled and neutralised with saturated NH₄Cl (10 mL), then most
29
30 of the THF was evaporated. The residue was diluted with water (30 mL), acidified to pH 3–4
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32 with 1 M HCl (ca. 20 mL) and extracted with CH₂Cl₂ (3 × 50 mL). The extract was
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34 evaporated and the residue was dissolved in Et₂O (150 mL) and extracted with 1 M NaOH (3
35
36 × 50 mL). The basic extracts were back-extracted with Et₂O (50 mL) then cooled to 0°C and
37
38 acidified to pH 3–4 with 5 M HCl (ca. 35 mL), forming a white suspension, which was
39
40 extracted with CH₂Cl₂ (5 × 50 mL). The extract was dried and evaporated, affording **9a** (1.49
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42 g, 71%) as a white solid, which did not require further purification, mp 233–234°C [lit.⁹⁶
43
44 149–150°C]. ¹H NMR (500 MHz, CDCl₃) δ 7.46 (dd, *J* = 7.3, 0.8 Hz, 1H), 7.34 (dd, *J* = 8.0,
45
46 7.5 Hz, 1H), 6.97 (dd, *J* = 8.3, 0.8 Hz, 1H), 5.91 (s, 2H), 3.81 (s, 3H), 3.61 (sept., *J* = 6.7 Hz,
47
48 1H), 3.52 (sept., *J* = 6.8 Hz, 1H), 1.57 (d, *J* = 6.5 Hz, 6H), 1.08 (d, *J* = 7.0 Hz, 6H). The ¹H
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50 NMR data match those in literature.⁹⁶
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3 *General procedure for the Suzuki–Miyaura Cross-couplings.* The selected iodide **3** or **10**
4 (1 equiv.), boronic acid **9** (or amide in the case of **9b**) (1.3 equiv.), K₂CO₃ (4 equiv.) and 10
5 wt% Pd/C (2 mol%) were suspended in water (10 mL/mmol of iodide). The suspension was
6 stirred with heating under reflux under N₂ for 72 h before being cooled to room temperature,
7 acidified with 1 M HCl, diluted with water and extracted with CH₂Cl₂. The extract was
8 filtered through a pad of Celite, washed with water and brine, dried and evaporated to give
9 the crude product. Table 1 yields were determined via ¹H NMR spectroscopy of the crude
10 product with use of 1,3,5-trimethoxybenzene as the internal standard.
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23 *Suzuki–Miyaura Cross-coupling of 1-hydroxy-2-iodo-8-acetoxy-9,10-anthraquinone (3)*
24 *with (2-(Diisopropylcarbamoyl)-3-methoxyphenyl)boronic acid (9a).* The general procedure
25 was used with iodide **3**¹⁹ (1.63 g, 4.00 mmol) and boronic acid **9a**⁹⁵ (1.43 g, 5.12 mmol),
26 except that the reaction time was 7 d instead of 72 h. The crude product was subjected to
27 flash chromatography. Elution with 1:199 MeOH:CH₂Cl₂ gave a tertiary mixture of **11a**, **12a**
28 and *N,N*-diisopropyl-2-methoxybenzamide (1.69 g, 72% yield of **11a** and **12a** with a 3:1 ratio
29 of **11a**:**12a** by ¹H NMR) as an orange solid. A sample of the mixture was subjected to
30 preparative thin-layer chromatography. Development with 2:3 EtOAc:hexanes gave a pure
31 sample of **12a**; the sample of **11a** still contained *N,N*-diisopropyl-2-methoxybenzamide,
32 which was removed by crystallisation from 1:1 EtOAc:EtOH.
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45 *2'-(1,8-Dihydroxy-9,10-anthraquinon-2-yl)-N,N-diisopropyl-6'-methoxybenzamide (11a).*
46 Yellow-orange solid, mp 263–264°C. R_f (2:3 EtOAc:hexanes): 0.5; IR (ATR) ν_{max} cm⁻¹:
47 1671 (C10=O), 1623 (C9=O, NC=O); ¹H NMR (500 MHz, CDCl₃) δ 12.55 (s, 1H), 12.04 (s,
48 1H), 8.01 (d, *J* = 8.0 Hz, 1H), 7.87 (dd, *J* = 7.5, 1.0 Hz, 1H), 7.86 (d, *J* = 7.5 Hz, 1H), 7.71
49 (dd [app. t], *J*₁ = *J*₂ = 8.0 Hz, 1H), 7.39 (dd [app. t], *J*₁ = *J*₂ = 8.0 Hz, 1H), 7.32 (dd, *J* = 8.5,
50 1.0 Hz, 1H), 7.07 (dd, *J* = 7.8, 0.8 Hz, 1H), 6.97 (dd, *J* = 8.3, 0.8 Hz, 1H), 3.87 (s, 3H), 3.70
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3 (sept., $J = 6.7$ Hz, 1H), 3.22 (sept., $J = 6.8$ Hz, 1H), 1.48 (d, $J = 7.0$ Hz, 3H), 1.03 (d, $J = 7.0$
4 Hz, 3H), 1.02 (d, $J = 7.0$ Hz, 3H), 0.70 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ
5 193.4, 181.6, 166.6, 162.8, 160.3, 156.0, 140.3, 137.5, 135.1, 133.9, 133.0, 132.9, 128.4,
6 128.2, 124.8, 123.0, 120.3, 119.6, 116.1, 115.6, 110.9, 55.9, 51.0, 45.7, 21.0, 20.6, 20.3;
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12 HRMS (APCI $^-$) m/z : $[\text{M}]^{\bullet-}$ Calcd for $\text{C}_{28}\text{H}_{27}\text{NO}_6$ 473.1838; Found 473.1848.

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14 *5'-(1,8-Dihydroxy-9,10-anthraquinon-2-yl)-N,N-diisopropyl-2'-methoxybenzamide (12a).*

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16 Red-orange solid, mp 222–223°C. R_f (2:3 EtOAc:hexanes): 0.4; IR (ATR) ν_{max} cm^{-1} : 1731,
17 1666 (C10=O), 1621 (C9=O), 1606 (NC=O); ^1H NMR (500 MHz, CDCl_3) δ 12.79 (s, 1H),
18 12.07 (s, 1H), 7.90 (d, $J = 8.0$ Hz, 1H), 7.86 (dd, $J = 7.5, 1.0$ Hz, 1H), 7.76 (d, $J = 7.5$ Hz,
19 1H), 7.72 (dd, $J = 8.5, 7.5$ Hz, 1H), 7.67 (dd, $J = 8.5, 2.5$ Hz, 1H), 7.46 (d, $J = 2.5$ Hz, 1H),
20 7.33 (dd, $J = 8.5, 1.0$ Hz, 1H), 7.00 (d, $J = 8.5$ Hz, 1H), 3.89 (s, 3H), 3.80 (sept., $J = 6.7$ Hz,
21 1H), 3.53 (sept., $J = 6.7$ Hz, 1H), 1.58 (d, $J = 6.5$ Hz, 3H), 1.56 (d, $J = 7.0$ Hz, 3H), 1.19 (d, J
22 = 6.5 Hz, 3H), 1.09 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 193.7, 181.6, 168.1,
23 162.8, 160.2, 155.5, 137.6, 137.5, 136.6, 133.9, 132.3, 130.6, 128.8, 128.4, 128.1, 124.8,
24 120.4, 120.2, 116.2, 116.1, 110.8, 55.8, 51.2, 46.0, 20.93, 20.90, 20.7, 20.6; HRMS (APCI $^-$)
25 m/z : $[\text{M}]^{\bullet-}$ Calcd for $\text{C}_{28}\text{H}_{27}\text{NO}_6$ 473.1838; Found 473.1841.

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41 *5'-(1,8-Dihydroxy-9,10-anthraquinon-2-yl)-N-isopropyl-2'-methoxybenzamide (13).* A
42 tertiary mixture of biaryl amides **11**, **12** and *N,N*-diisopropyl-2-methoxybenzamide was finely
43 divided and then washed with cold 1:1 EtOH:EtOAc, giving a binary mixture of **11** and **12**.
44 This mixture (92 mg, 0.19 mmol, containing 0.078 mmol of **12** by ^1H NMR spectroscopy)
45 was treated with conc. H_2SO_4 (2 mL), forming a dark purple mixture. After stirring at 80°C
46 for 24 h, the resulting mixture was cooled to room temperature, poured into ice/water (40
47 mL) and extracted with CH_2Cl_2 (5 \times 25 mL). The combined extracts were washed with water
48 (30 mL), dried and evaporated to give an orange solid, which was subjected to flash
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3 chromatography. Elution with 3:7 EtOAc:hexanes afforded **13** (11 mg, 33%) as a red-orange
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5 solid, mp 232–234°C. R_f (3:7 EtOAc:hexanes): 0.3; IR (ATR) ν_{\max} cm⁻¹: 3331 (N–H), 1664
6
7 (C10=O), 1619 (C9=O, NC=O); ¹H NMR (600 MHz, CDCl₃) δ 12.76 (s, 1H, OH1), 12.06 (s,
8
9 1H, OH8), 8.46 (d, *J* = 2.4 Hz, 1H, H6'), 7.90 (d, *J* = 7.8 Hz, 1H, H4), 7.86 (dd, *J* = 7.2, 1.2
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11 Hz, 1H, H5), 7.83 (dd, *J* = 8.4, 2.4 Hz, 1H, H4'), 7.80 (d, *J* = 7.8 Hz, 1H, H3), 7.70 (dd, *J* =
12
13 8.4, 7.8 Hz, 1H, H6), 7.69 (br d, *J* = 7.8 Hz, 1H, N–H), 7.31 (dd, *J* = 8.4, 0.6 Hz, 1H, H7),
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15 7.08 (d, *J* = 9.0 Hz, 1H, H3'), 4.31 (m [pseudo oct], *J* = 6.8 Hz, 1H, NCH), 4.03 (s, 3H,
16
17 OCH₃), 1.28 (d, *J* = 6.6 Hz, 6H, 2 × CH₃); ¹³C NMR (150 MHz, CDCl₃) δ 193.6 (C9), 181.6
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19 (C10), 164.0 (NC=O), 162.7 (C8), 160.2 (C1), 157.6 (C2'), 137.7 (C3), 137.5 (C6), 136.2
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21 (C2), 133.9 (C10a), 133.6 (C4'), 133.3 (C6'), 132.5 (C4a), 129.0 (C5'), 124.7 (C7), 122.2
22
23 (C1'), 120.3 (C4), 120.2 (C5), 116.2 (C8a or C9a), 116.1 (C8a or C9a), 111.5 (C3'), 56.3
24
25 (OCH₃), 41.8 (NCH), 23.0 (2 × CH₃); HRMS (APCI⁻) *m/z*: [M]^{•-} Calcd for C₂₅H₂₁NO₆
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27 431.1369; Found 431.1379. NMR assignments were made with the assistance of COSY,
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29 HSQC, HMBC and NOESY experiments.
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34 Further elution with 1:99 MeOH:CH₂Cl₂ afforded 5'-(1,8-Dihydroxy-9,10-anthraquinon-2-
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36 yl)-2'-methoxybenzamide (**14**) (6.5 mg, 21%) as a red-orange solid, mp 288–290°C. R_f (1:49
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38 MeOH:CH₂Cl₂): 0.2; IR (ATR) ν_{\max} cm⁻¹: 3442 (N–H), 3168, 1686 (C=O), 1674 (C=O),
39
40 1620 (C9=O); ¹H NMR (600 MHz, CDCl₃) δ 12.79 (s, 1H, OH), 12.07 (s, 1H, OH), 8.49 (d, *J*
41
42 = 2.4 Hz, 1H, H6'), 7.92 (d, *J* = 7.8 Hz, 1H, H3 or H4), 7.90 (dd, *J* = 8.4, 2.4 Hz, 1H, H4'),
43
44 7.88 (dd, *J* = 7.5, 0.9 Hz, 1H, H5), 7.81 (d, *J* = 7.8 Hz, 1H, H3 or H4), 7.74 (br s, 1H, NH),
45
46 7.71 (dd, *J* = 8.4, 7.8 Hz, 1H, H6), 7.32 (dd, *J* = 8.4, 0.6 Hz, 1H, H7), 7.13 (d, *J* = 9.0 Hz, 1H,
47
48 H3'), 5.79 (br s, 1H, NH), 4.06 (s, 3H, OCH₃); ¹H NMR (600 MHz, *d*₅-pyridine) δ 9.03 (d, *J*
49
50 = 2.4 Hz, 1H, H6'), 8.62 (br s, 1H, NH), 8.32 (br s, 1H, NH), 7.99 (d, *J* = 7.8 Hz, 1H, H4),
51
52 7.97 (dd, *J* = 9.0, 2.4 Hz, 1H, H4'), 7.95 (d, *J* = 7.2 Hz, 1H, H5), 7.80 (d, *J* = 7.8 Hz, 1H, H3),
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54 7.66 (dd, *J* = 8.4, 7.8 Hz, 1H, H6), 7.39 (d, *J* = 8.4 Hz, 1H, H7), 7.19 (d, *J* = 9.0 Hz, 1H, H3'),
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3 3.85(s, 3H, OCH₃); ¹³C NMR (150 MHz, *d*₅-pyridine) δ 193.8 (C9), 181.9 (C10), 167.4
4 (NC=O), 163.1 (C8), 160.6 (C1), 158.7 (C1'), 138.1 (C3 or C6), 138.0 (C3 or C6), 136.6
5 (C2), 134.6 (C10a), 134.5 (C4'), 134.1 (C6'), 133.1 (C4a), 129.4 (C5'), 125.1 (C7), 123.7[†]
6 (C1'), 120.4 (C4), 120.3 (C5), 117.0 (C8a or C9a), 116.9 (C8a or C9a), 112.6 (C3'), 56.5
7 (OCH₃); HRMS (APCI⁻) *m/z*: [M]⁻ Calcd for C₂₂H₁₅NO₆ 389.0899; Found 389.0898. NMR
8 assignments were made with the assistance of COSY, HSQC and HMBC experiments.
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19 *1,8-Dihydroxy-2-iodo-9,10-anthraquinone (10a)*. Conc. H₂SO₄ (80 mL) was added to 1-
20 hydroxy-8-acetoxy-2-iodo-9,10-anthraquinone (**3**)¹⁹ (2.05 g, 5.02 mmol) and the resulting
21 mixture was stirred for 45 min. The mixture was poured into ice-water (400 mL) and
22 extracted with CHCl₃ (3 × 250 mL). The extract was dried and evaporated affording **10a** as
23 an orange solid (1.82 g, 99 %), which crystallised from EtOAc as orange needles, mp 200–
24 202°C. R_f (PhMe): 0.65; IR (ATR) ν_{max} cm⁻¹: 1660 (C10 C=O), 1620 (C9 C=O); ¹H NMR
25 (400 MHz, CDCl₃) δ 12.97 (s, 1H, OH), 11.91 (s, 1H, OH), 8.22 (d, *J* = 8.0 Hz, 1H), 7.84
26 (dd, *J* = 7.6, 1.0 Hz, 1H), 7.71 (dd, *J* = 8.4, 7.6 Hz, 1H), 7.58 (d, *J* = 8.0 Hz, 1H, H3), 7.33
27 (dd, *J* = 8.4, 1.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 192.8, 181.4, 162.9, 161.2, 147.0,
28 137.9, 133.7, 133.6, 125.1, 121.1, 120.4, 115.6, 115.4, 95.3; HRMS (APCI⁻) *m/z*: [M]⁻
29 Calcd for C₁₄H₇IO₄ 365.9389; Found 365.9376.
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45 *1-Methoxy-2-iodo-8-acetoxy-9,10-anthraquinone (10b)*. A mixture of 1-hydroxy-2-iodo-8-
46 acetoxy-9,10-anthraquinone (**3**)¹⁹ (0.15 g, 0.37 mmol) and K₂CO₃ were suspended in dry
47 acetone (8 mL). MeOTs (0.24 mL, 1.6 mmol) was then added, and the mixture was heated
48 under reflux under N₂ with stirring for 12 h. Additional dry acetone (2 mL), and MeOTs
49 (0.24 mL, 1.6 mmol) were added, and the mixture was stirred with heating under reflux for a
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57 [†] This chemical shift is reported based upon the observed correlation to H3' in the HMBC spectrum, as the
58 signal was obscured by solvent in the ¹³C NMR spectrum.
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3 further 24 h. The reaction mixture was cooled to room temperature, filtered, and concentrated
4
5 under reduced pressure, and the residue was dissolved in CH₂Cl₂ (20 mL). The organic phase
6
7 was washed with 1 M HCl (3 × 5 mL), water (5 mL), and brine (5 mL), dried and evaporated.
8
9 The residue was subjected to flash chromatography. Elution with CH₂Cl₂ gave **10b** (0.11 g,
10
11 69%) as a yellow solid, mp 170–173°C. R_f (CH₂Cl₂): 0.4; IR (ATR) ν_{max} cm⁻¹: 1752
12
13 (OC=O), 1668 (C9=O, C10=O); ¹H NMR (500 MHz, CDCl₃) δ 8.21 (d, *J* = 8.0 Hz, 1H, H3),
14
15 8.19 (dd, *J* = 8.0, 1.2 Hz, 1H, H5), 7.79 (d, *J* = 8.0 Hz, 1H, H4), 7.76 (dd [app. t], *J* = 8.0 Hz,
16
17 1H, H6), 7.43 (dd, *J* = 8.0, 1.2 Hz, 1H, H7), 3.92 (s, 3H, OCH₃), 2.49 (s, 3H, CH₃); ¹³C NMR
18
19 (125 MHz, CDCl₃) δ 182.3 (C10), 181.2 (C9), 169.9 (OC=O), 159.8 (C1), 149.9 (C8), 144.7
20
21 (C3), 135.1 (C4a), 134.6 (C6), 134.4 (C10a), 130.2 (C7), 126.8 (C9a), 126.2 (C8a), 125.4
22
23 (C5), 124.5 (C4), 104.5 (C2), 62.3 (OCH₃), 21.4 (CH₃); HRMS (APCI⁻) *m/z*: [M]⁻ Calcd for
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25 C₁₇H₁₁IO₅ 421.9651; Found 421.9643. NMR assignments made with the assistance of COSY,
26
27 HSQC and HMBC experiments.
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34 *1-Methoxy-2-iodo-8-hydroxy-9,10-anthraquinone (10c)*. Conc. H₂SO₄ (1.5 mL) was added
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36 to acetate **10b** (63 mg, 0.15 mmol) and the mixture was stirred for 45 min then poured into
37
38 ice/water (15 mL) and extracted with CH₂Cl₂ (3 × 20 mL). The extract was dried and
39
40 evaporated, giving a yellow-green solid, which was subjected to flash chromatography.
41
42 Elution with 7:3 PhMe:hexanes afforded **10c** (43 mg, 75%) as a yellow solid, mp 164–166°C.
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44 R_f (7:3 PhMe:hexanes): 0.25; IR (ATR) ν_{max} cm⁻¹: 1672 (C10=O), 1636 (C9=O); ¹H NMR
45
46 (500 MHz, CDCl₃) δ 12.77 (s, 1H, OH), 8.28 (d, *J* = 8.0 Hz, 1H, H3), 7.86 (d, *J* = 8.0 Hz, 1H,
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48 H4), 7.80 (dd, *J* = 7.5, 1.0 Hz, 1H, H5), 7.67 (dd, *J* = 8.5, 7.5 Hz, 1H, H6), 7.33 (dd, *J* = 8.5,
49
50 1.0 Hz, 1H, H7), 3.98 (s, 3H, OCH₃); ¹³C NMR (125 MHz, CDCl₃) δ 187.9 (C9), 182.1
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52 (C10), 162.8 (C8), 160.3 (C1), 145.7 (C3), 136.7 (C6), 136.0 (C4a), 132.7 (C10a), 125.6
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54 (C9a), 125.3 (C4), 125.2 (C7), 119.3 (C5), 116.7 (C8a), 105.0 (C2), 62.0 (OCH₃); HRMS
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(APCI⁻) m/z: [M]⁻ Calcd for C₁₅H₉IO₄ 379.9546; Found 379.9556. NMR assignments were made with the assistance of COSY, HSQC and HMBC experiments.

(2-(Diethylcarbamoyl)-3-methoxyphenyl)boronic acid (**9c**).³⁶ A 0.96 M solution of *sec*-BuLi in cyclohexane (17.0 mL, 16.3 mmol) was added to a solution of TMEDA (2.50 mL, 16.7 mmol) in anhydrous THF (40 mL) at -78°C under N₂. To this a solution of *N,N*-diethyl-2-methoxybenzamide⁹² (3.08 g, 14.8 mmol) in anhydrous THF (20 mL) was added dropwise over 20 min with stirring. The resulting mixture was stirred at -78°C for 1 h then treated with B(*Oi*-Pr)₃ (11.5 mL, 49.8 mmol). The reaction mixture was allowed to warm to room temperature and stirring was continued overnight. The reaction mixture was cooled and neutralised with saturated NH₄Cl (15 mL), and most of the THF was evaporated. The residue was diluted with water (60 mL), acidified to pH 3–4 with 1 M HCl, and extracted with CHCl₃ (3 × 100 mL). The extract was evaporated and the residue was dissolved in Et₂O and extracted with 1 M NaOH (2 × 50 mL). The basic extract was back-extracted with Et₂O (50 mL), then cooled to 0°C and acidified with 1 M HCl to pH 3–4. The resulting mixture was extracted with CHCl₃ (3 × 100 mL) and the combined extracts were dried and evaporated, affording **9c** as a white solid (1.85 g, 50%), which was used as such without further purification. A sample crystallised from EtOAc/hexanes as colourless microneedles, mp 106–108°C. IR (ATR) ν_{\max}^{-1} : 3355 (OH), 1606 (NC=O); ¹H NMR (500 MHz, CDCl₃) δ 7.49 (dd, *J* = 7.0, 1.0 Hz, 1H), 7.37 (dd [app. t], *J* = 8.5, 7.5 Hz, 1H), 6.98 (dd, *J* = 8.0, 1.0 Hz, 1H), 5.97 (s, 2H), 3.82 (s, 3H), 3.61 (q, *J* = 7.0 Hz, 2H), 3.12 (q, *J* = 7.0 Hz, 2H), 1.26 (t, *J* = 7.0 Hz, 3H), 1.02 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.5, 154.6, 130.5, 129.8, 127.7, 113.0, 55.6, 43.4, 39.6, 13.8, 12.8. HRMS (APCI⁻) m/z: [M-H]⁻ Calcd for C₁₂H₁₇BNO₄ 250.1251; Found 250.1247. This compound has been reported previously,³⁶ but the free boronic acid has not been characterised.

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5 *2'-(1,8-Dihydroxy-9,10-anthraquinon-2-yl)-N,N-diethyl-6'-methoxybenzamide (11c)*. The
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7 general procedure was used with iodide **3**¹⁹ (1.63 g, 4.00 mmol), boronic acid **9c**³⁶ (1.31 g,
8 5.22 mmol), and 10% Pd/C (125 mg, 3 mol%). The crude product was subjected to flash
9
10 chromatography. Elution with CH₂Cl₂ gave a binary mixture of **11c** and the by-product *N,N*-
11
12 diethyl-2-methoxybenzamide as a red gum. The residue was heated under high vacuum to
13
14 give **11c** (1.46 g, 82%) as an orange solid, mp 212–214°C. R_f (CHCl₃): 0.1; IR (ATR) ν_{max}
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16 cm⁻¹: 1669 (C10=O), 1629 (C9=O), 1615 (NC=O); ¹H NMR (400 MHz, CDCl₃) δ 12.50 (s,
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18 1H), 12.03 (s, 1H), 7.86 (dd, *J* = 7.8, 1.4 Hz, 1H), 7.84 (AB, *J* = 8.0 Hz, 2H), 7.71 (dd, *J* =
19
20 8.4, 7.6 Hz, 1H), 7.42 (dd, *J* = 8.4, 8.0 Hz, 1H), 7.31 (dd, *J* = 8.4, 1.2 Hz, 1H), 7.05 (dd, *J* =
21
22 7.6, 0.8 Hz, 1H), 7.00 (dd, *J* = 8.4, 0.8 Hz, 1H), 3.88 (s, 3H), 3.73 (m [pseudo sextet], *J* = 6.8
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24 Hz, 1H), 3.25 (m [pseudo sextet], *J* = 7.0 Hz, 1H), 2.96 (m [pseudo sextet], *J* = 7.0 Hz, 1H),
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26 2.92 (m [pseudo sextet], *J* = 7.1 Hz, 1H), 0.96 (dd [app. t], *J*₁ = *J*₂ = 7.2 Hz, 3H), 0.75 (dd
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28 [app. t], *J*₁ = *J*₂ = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 193.5, 181.6, 167.0, 162.8,
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30 160.2, 156.0, 139.6, 137.6, 135.3, 134.1, 133.9, 133.0, 129.1, 126.7, 124.8, 122.9, 120.3,
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32 119.7, 116.1, 115.7, 110.9, 55.8, 42.7, 38.1, 13.7, 12.2; HRMS (APCI–) *m/z*: [M]⁺ Calcd for
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34 C₂₆H₂₃NO₆ 445.1525; Found 445.1514.
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43 *2'-Hydroxy-N,N-diisopropyl-3-methoxy-[1,1'-biphenyl]-2-carboxamide (11d)*. The general
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45 procedure was used with iodide **10d** (0.113 g, 0.514 mmol) and boronic acid **9a**⁹⁵ (0.185 g,
46 0.663 mmol). The crude product was subjected to flash chromatography. Elution with 1:4
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48 EtOAc:hexanes gave **11d** (85 mg, 51%) as a white powder, mp 189–191°C. R_f (1:4
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50 EtOAc:hexanes): 0.30; IR (ATR) ν_{max} cm⁻¹: 3400–2900 (OH), 1610 (NC=O); ¹H NMR (500
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52 MHz, CDCl₃) δ 8.34 (s, 1H), 7.37 (dd, *J* = 8.3, 7.8 Hz, 1H), 7.24 (ddd, *J* = 8.1, 7.4, 1.9 Hz,
53
54 1H), 7.06 (br d, *J* = 7.5 Hz), 7.02 (dd, *J* = 8.0, 1.0 Hz, 1H), 6.92 (dd, *J* = 8.3, 0.8 Hz, 1H),
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3 6.92 (ddd [app. td], $J = 7.4, 1.3$ Hz, 1H), 6.86 (dd, $J = 7.8, 0.8$ Hz), 3.86 (s, 3H), 3.59 (sept, J
4 = 6.7 Hz, 1H), 3.28 (sept, $J = 6.8$ Hz, 1H), 1.49 (d, $J = 6.5$ Hz, 3H), 1.09 (d, $J = 7.0$ Hz, 3H),
5 1.03 (d, $J = 7.0$ Hz, 3H), 0.78 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.2,
6 154.7, 154.3, 137.3, 130.8, 129.9, 129.8, 129.7, 127.3, 123.7, 120.6, 120.2, 109.8, 55.8, 51.5,
7 46.2, 20.6, 20.5, 20.4, 19.7; HRMS (APCI+) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{20}\text{H}_{26}\text{NO}_3^+$ 328.1913;
8 Found 328.1917.
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18 *2-Iodo-4-nitrophenol (10e)*.⁹⁷ I_2 (5.08 g, 20.0 mmol) was dissolved in saturated aqueous KI
19 (200 mL) and the resulting solution was added dropwise to a solution of 4-nitrophenol (2.78
20 g, 20.0 mmol) in 25% aqueous NH_3 (150 mL) at 0°C . The reaction mixture was warmed to
21 room temperature and stirred for 4 d. Additional I_2 (1.67 g, 6.58 mmol) in saturated aqueous
22 KI (150 mL) was added dropwise and the reaction mixture was stirred at room temperature
23 for a further 2 d. A third portion of I_2 (1.02 g, 4.02 mmol) in saturated aqueous KI (40 mL)
24 was added dropwise, followed by fresh 25% aqueous NH_3 (150 mL) and the reaction mixture
25 was stirred for a further 17 d, after which time thin-layer chromatography still showed
26 incomplete consumption of 4-nitrophenol. The reaction mixture was acidified with 6 M HCl
27 to $\sim\text{pH}$ 3 and the resulting suspension was extracted with Et_2O (3×300 mL). The extract was
28 washed with 0.5 M $\text{Na}_2\text{S}_2\text{O}_3$ (2×200 mL), water (200 mL), and brine and then dried and
29 evaporated. The crude residue was subjected to flash chromatography. Elution with CH_2Cl_2
30 afforded **10e** (3.86 g, 73%) as a yellow solid, mp $89\text{--}91^\circ\text{C}$ [lit.⁹⁸ $85\text{--}87$]. ^1H NMR (400
31 MHz, CDCl_3) δ 8.60 (d, $J = 2.4$ Hz, 1H), 8.18 (dd, $J = 9.0, 2.4$ Hz, 1H), 7.07 (d, $J = 9.2$ Hz,
32 1H), 6.06 (s, 1H). The ^1H NMR data match those in literature.⁹⁸
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54 *Dimethyl 3-hydroxy-4-iodo-6-methylphthalate (10f)*. A mixture of dimethyl 3-hydroxy-6-
55 methylphthalate⁹⁴ (1.11 g, 4.97 mmol), NaIO_4 (1.09 g, 5.08 mmol), and NaCl (0.59 g, 10
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mmol) was dissolved in 9:1 AcOH:water (20 mL) and KI (0.84 g, 5.06 mmol) was added portion-wise. The mixture was stirred at room temperature for 15 min, then sealed and stirred at 50°C for 21 h before being cooled to room temperature and poured into water (150 mL). The resulting suspension was extracted with CH₂Cl₂ (3 × 100 mL), and the extract was washed with 0.2 M Na₂S₂O₃ (2 × 50 mL), water (2 × 50 mL) and brine (50 mL). The organic phase was dried and evaporated to give **10f** as an off-white solid (1.68 g, 97%) which required no further purification, mp 89–92°C. R_f (CH₂Cl₂) 0.6; IR (ATR) ν_{max} cm⁻¹: 3136 (OH), 1721 (C8=O), 1678 (C10=O); ¹H NMR (500 MHz, CDCl₃) δ 11.63 (s, 1H, OH), 7.84 (s, 1H, H5), 3.94 (s, 3H, 3 × H11), 3.89 (s, 3H, 3 × H9), 2.19 (s, 3H, 3 × H7); ¹³C NMR (125 MHz, CDCl₃) δ 169.1 (C10), 168.9 (C8), 158.6 (C3), 146.6 (C5), 135.3 (C1), 127.7 (C6), 109.1 (C2), 87.1 (C4), 53.5 (C11), 52.6 (C9), 18.2 (C7); HRMS (APCI-): [M-H]⁻ Calcd for C₁₁H₁₀IO₅⁻ 348.9578; Found 348.9578. NMR assignments were made with the assistance of HSQC and HMBC experiments.

2'-(Diisopropylcarbamoyl)-2-hydroxy-3'-methoxy-5-methyl-[1,1'-biphenyl]-3,4-dicarboxylic acid (11f). The general procedure was used with iodide **10f** (0.175 g, 0.50 mmol) and boronic acid **9a**⁹⁵ (0.183 g, 0.66 mmol). Repeated crystallisation from Et₂O/CH₂Cl₂/hexanes afforded a pure sample of **11f** as white granules, mp >300°C. R_f (1:4:45 TFA:MeOH:CH₂Cl₂): 0.25; IR (ATR) ν_{max} cm⁻¹: 3000–2700 (OH), 2700–2250 (OH), 2100–1850, 1672 (C8=O), 1648 (C7=O), 1616 (NC=O); ¹H NMR (500 MHz, MeOD) δ 7.46 (d, *J* = 0.4 Hz, 1H, H6), 7.37 (dd, *J* = 8.3, 7.8 Hz, 1H, H5'), 7.06 (dd, *J* = 8.3, 0.8 Hz, 1H,), 7.04 (dd, *J* = 7.8, 0.8 Hz, 1H,), 3.87 (s, 3H, OCH₃), 3.68 (sept, *J* = 6.6 Hz, 1H, NCH), 3.34 (sept, *J* = 6.8 Hz, 1H, NCH), 2.23 (d, *J* = 0.4 Hz, 3H, 3 × H9), 1.47 (d, *J* = 6.8 Hz, 3H, CH₃), 1.07 (d, *J* = 6.8 Hz, 3H, CH₃), 1.02 (d, *J* = 6.6 Hz, 3H, CH₃), 0.69 (d, *J* = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, MeOD), δ 173.0 (C8), 172.7 (C7), 169.7 (C7'), 158.7 (C2), 157.3 (C3'),

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3 139.8 (C6), 137.9 (C4), 135.3 (C1'), 129.7 (C5'), 129.2 (C1 or C3), 128.5 (C2'), 124.9 (C5),
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5 124.7 (C6'), 111.4 (C4'), 110.8 (C1 or C3), 56.2 (C8'), 52.4, 46.9, 21.0, 20.8, 20.3, 20.2, 18.5
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7 (C9); HRMS (APCI-): $[M-H]^-$ Calcd for $C_{23}H_{26}NO_7^-$ 428.1715; Found 428.1725. NMR
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9 assignments were made with the assistance of COSY, HSQC and HMBC experiments.
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14 *2'-(1,8-Dimethoxy-9,10-anthraquinon-2-yl)-N,N-diethyl-6'-methoxybenzamide (16)*. MeI
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16 (3.2 mL, 51 mmol) was added to a mixture of amide **11c** (1.12 g, 2.51 mmol) and K_2CO_3
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18 (6.22 g, 45.0 mmol) in dry DMF (50 mL). The resulting mixture was flushed with N_2 , sealed,
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20 and stirred at 60°C for 48 h. The mixture was poured into water (200 mL) and extracted with
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22 EtOAc (6 × 100 mL) until the extracts were colourless. The extract was washed with water (8
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24 × 200 mL) and brine (6 × 200 mL), dried and evaporated. The crude residue was filtered
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26 through a plug of silica, alternating washes with hexanes and CH_2Cl_2 until the washes were
27
28 colourless. Elution with 9:1 CH_2Cl_2 :MeOH afforded **16** as a yellow solid (1.09 g, 92%), mp
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30 183–185°C. R_f (1:49 MeOH: CH_2Cl_2): 0.4; IR (ATR) ν_{max} cm^{-1} : 1673 (C9=O, C10=O), 1624
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32 (NC=O); 1H NMR (400 MHz, $CDCl_3$) δ 7.97 (d, J = 8.0 Hz, 1H, H4), 7.86 (dd, J = 7.6, 1.2
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34 Hz, 1H, H5), 7.66 (dd, J = 8.6, 7.8 Hz, 1H, H6), 7.59 (br s, 1H, H3), 7.38 (dd, J = 8.2, 7.8 Hz,
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36 1H, H4'), 7.31 (dd, J = 8.6, 1.0 Hz, 1H, H7), 7.02 (dd, J = 7.8, 0.6 Hz, 1H, H3'), 6.96 (dd, J =
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38 8.6, 1.0 Hz, 1H, H5'), 4.01 (s, 3H, 3 × H3''), 3.86 (s, 3H, 3 × H1''), 3.75 (s, 3H, 3 × H2''), 3.71
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40 (m [pseudo sextet], J = 7.0 Hz, 1H, NCH₂), 3.35 (m [pseudo sextet], J = 7.0 Hz, 1H, NCH₂),
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42 3.10 (br m, 1H, NCH₂), 3.00 (br m, 1H, NCH₂), 1.08 (dd [app. t], $J_1 = J_2 = 6.8$ Hz, 3H, CH₃),
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44 0.80 (br s, 3H, CH₃); ^{13}C (100 MHz, $CDCl_3$) δ 183.5, 183.0, 167.0, 159.6, 157.9, 155.7,
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46 141.5, 136.0, 135.7 (br, C3), 135.2, 134.3 (C6), 134.2, 129.0 (C4'), 128.4, 126.7, 124.1, 122.9
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48 (C3'), 121.9 (C4), 119.2 (C5), 118.1 (C7), 110.3 (C5'), 62.6 (C2''), 56.7 (C3''), 55.6 (C1''),
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50 42.8, 37.8, 13.6, 12.1; HRMS (APCI-) m/z : $[M]^+$ 473.1841, ; $C_{28}H_{27}NO_6^+$ Calcd 473.1838.
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NMR assignments made with the assistance of COSY, HSQC and NOESY experiments.

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5 2'-(1,8,9,10-Tetramethoxyanthracen-2-yl)-N,N-diethyl-6'-methoxy-benzamide (**17**). Tetra-
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8 *n*-butylammonium chloride hydrate (0.320 g, 1.15 mmol) and anthraquinone **16** (1.12 g, 2.36
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10 mmol) were dissolved in THF (110 mL). The orange solution was treated with a solution of
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12 Na₂S₂O₄ (2.47 g, 14.2 mmol) in water (20 mL) and stirred for 1 h. A solution of NaOH (2.83
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14 g, 70.8 mmol) in water (20 mL) was added, and the resulting dark red mixture was stirred for
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16 30 min. MeOTs (14.5 mL, 96.1 mmol) was added and the reaction mixture was stirred under
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18 N₂ for 24 h. The resulting yellow mixture was poured into water (300 mL) and extracted with
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20 CH₂Cl₂ (3 × 200 mL). The extract was dried and evaporated, and the residue was filtered
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22 through a plug of silica, washing with CH₂Cl₂ to remove excess MeOTs, then eluting with
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24 EtOAc. The filtrate was evaporated, and the resulting orange film was subjected to high
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26 vacuum, affording **17** (1.12 g, 94%) as a yellow foam, mp 88–92°C. R_f (EtOAc): 0.6; IR
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28 (ATR) ν_{max} cm⁻¹: 1621 (NC=O); ¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, *J* = 9.2 Hz, 1H), 7.86
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30 (dd, *J* = 8.8, 1.2 Hz, 1H), 7.41–7.36 (br m, 1H), 7.39 (dd, *J* = 8.4, 7.6 Hz, 1H), 7.38 (dd, *J* =
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32 8.8, 7.6 Hz, 1H), 7.11 (br d, *J* = 7.2 Hz, 1H), 6.96 (dd, *J* = 8.4, 0.8 Hz, 1H), 6.80 (d, *J* = 7.2
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34 Hz, 1H), 4.07 (s, 3H), 4.04 (s, 3H), 3.92 (s, 3H), 3.88 (s, 3H), 3.76 (s, 3H), 3.70 (m [pseudo
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36 sextet], *J* = 6.8 Hz, 1H), 3.49 (br s, 1H), 2.97 (br s, 1H), 2.88 (br m, 1H), 1.06 (dd [app. t], *J*₁
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38 = *J*₂ = 6.4 Hz, 3H) 0.56 (br s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 167.5, 157.4, 155.7 (br),
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40 153.6, 149.9, 148.3, 138.1, 129.6, 129.1 (br), 128.7, 127.6, 127.5, 127.0, 125.8, 123.3, 120.7,
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42 119.6, 117.6, 114.9, 109.7, 104.3, 64.0, 63.2 (br), 62.9, 56.6, 55.6, 42.8, 37.7, 13.8, 12.1;
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44 HRMS (APCI+) *m/z*: [M+H]⁺ Calcd for C₃₀H₃₄NO₆⁺ 504.2386; Found 504.2381.
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52 1,5,6,7,11-Pentamethoxy-13H-indeno[1,2-*b*]anthracen-13-one (**18**). A solution of
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54 anhydrous *i*-Pr₂NH (1.03 mL, 7.35 mmol) in anhydrous THF (20 mL) under N₂ was cooled to
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56 –50°C and treated with a 1.2 M solution of *n*-BuLi in hexanes (5.4 mL, 6.5 mmol). The
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3 resulting mixture was stirred for 30 min before a solution of amide **17** (0.531 g, 1.05 mmol)
4 in THF (12 mL) was added dropwise over 10 min, whereupon a deep orange colour
5 developed. The reaction mixture was stirred at -50°C for 2 h, then allowed to slowly warm to
6 room temperature, and stirring was continued for 60 h, before being quenched with saturated
7 NH_4Cl (90 mL) and water (30 mL). The mixture was extracted with CH_2Cl_2 (3×50 mL), and
8 the extract was dried and evaporated to give **18** as a red gum, which formed a red foam
9 (0.506 g) under high vacuum. Attempts at purification led to decomposition of **18**, so the ^1H
10 NMR data reported are for the crude product. ^1H NMR (600MHz, CDCl_3) δ 8.49 (s, 1H),
11 7.86 (dd, $J = 8.7, 0.9$ Hz, 1H), 7.84 (dd, $J = 7.2, 0.6$ Hz, 1H), 7.58 (dd, $J = 8.4, 7.8$ Hz, 1H),
12 7.44 (dd, $J = 8.7, 7.5$ Hz, 1H), 6.90 (d, $J = 7.8$ Hz, 1H), 6.88 (d, $J = 8.4$ Hz, 1H), 4.091 (s,
13 3H), 4.089 (s, 3H), 4.04 (s, 3H), 4.03 (s, 3H), 3.98 (s, 3H).
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30 *1,5,7-Trimethoxy-6H-indeno[1,2-b]anthracene-6,11,13-trione (19)*. Crude fluorenone **18**
31 synthesised as described above from amide **17** (143 mg, 0.283 mmol) was dissolved in 1,4-
32 dioxane (10 mL) and treated with AgO (0.175 g, 1.41 mmol). The resulting mixture was
33 stirred under N_2 for 5 min, then 4 M HNO_3 (3 mL) was added dropwise over 5 min. The
34 resulting dark orange solution was stirred under N_2 for 30 min before being diluted with
35 water (20 mL) and extracted with CH_2Cl_2 (3×30 mL). The extract was dried and evaporated
36 and the residue was subjected to flash chromatography. Elution with 1:199 MeOH: CH_2Cl_2
37 gave **19** (74 mg, 66% over 2 steps) as an orange solid, mp $277\text{--}279^{\circ}\text{C}$. R_f (1:199
38 MeOH: CH_2Cl_2): 0.25; IR (ATR) ν_{max} cm^{-1} : 1704 (C13=O), 1671 (C6=O, C11=O); ^1H NMR
39 (500 MHz, CDCl_3) δ 8.32 (s, 1H), 7.87 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.70 (d, $J = 7.5$ Hz, 1H),
40 7.69 (dd, $J = 8.0, 7.0$ Hz, 1H), 7.57 (dd, $J = 8.5, 7.5$ Hz, 1H), 7.34 (dd, $J = 8.5, 1.0$ Hz, 1H),
41 6.96 (d, $J = 8.0$ Hz, 1H), 4.11 (s, 3H), 4.04 (s, 3H), 4.02 (s, 3H); ^{13}C NMR (125 MHz,
42 CDCl_3) δ 189.6, 182.9, 182.7, 159.7, 158.7, 155.9, 144.0, 140.7, 139.2, 138.0, 136.4, 135.0,
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3 134.8, 133.6, 123.6, 120.6, 119.5, 118.3, 118.2, 118.1, 114.3, 62.4, 56.8, 56.2; HRMS
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5 (APCI⁻) m/z: [M]⁻ Calcd for C₂₄H₁₆O₆ 400.0947; Found 400.0935.
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10 *1,5,7-Trihydroxy-6H-indeno[1,2-b]anthracene-6,11,13-trione (1)*. *Method 1*: Crude
11 fluorenone **18** synthesised as described above from amide **17** (125 mg, 0.248 mmol) was
12 treated with AlCl₃ (1.68 g, 12.6 mmol) and nitrobenzene (15 mL). The resulting dark green
13 mixture was stirred at 60°C under N₂ for 6.5 days before being poured into a mixture of ice
14 (75 g), water (30 mL) and 10 M HCl (45 mL). The resulting mixture was stirred for 3 d
15 before being extracted with Et₂O (3 × 60 mL). The extract was diluted with hexanes (~200
16 mL) resulting in a dark red precipitate, which was collected by vacuum filtration, washed
17 with hexanes and air dried. The aqueous phase was allowed to stand for 3 d and the resulting
18 precipitate was collected by vacuum filtration, washed with water, and air dried. The
19 precipitates were combined and crystallised from pyridine to give **1** as burgundy
20 microcrystals (27 mg, 30%), identical with the product described below.
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36 *Method 2*: A stirred suspension of fluorenone **19** (73 mg, 0.18 mmol) in glacial AcOH (20
37 mL) was treated with 48% aq. HBr (15 mL) and then heated under reflux under N₂ for 4.5
38 days. More glacial AcOH (10 mL) and 48% aq. HBr (7.5 mL) were added and the mixture
39 was refluxed under N₂ for a further 2.5 days before being cooled to room temperature and
40 poured into water (100 mL). The resulting precipitate was collected by vacuum filtration,
41 washed with water and air dried, giving **1** as a dark red solid (59 mg, 91%), mp >295°C. R_f
42 (1:99 AcOH:CH₂Cl₂): 0.45; IR (ATR) ν_{max} cm⁻¹: 3380 (OH), 1696 (C13=O), 1669 (C11=O),
43 1628 (C6=O); ¹H NMR (500 MHz, CDCl₃) δ 12.48 (s, 1H, OH), 11.95 (s, 1H, OH), 8.39 (br
44 s, 1H, OH1), 8.16 (s, 1H, H12), 7.89 (dd, *J* = 7.5, 1.5 Hz, 1H, H10), 7.75 (dd, *J* = 8.5, 7.5 Hz,
45 1H, H9), 7.60 (dd, *J* = 7.0, 0.5 Hz, 1H, H4), 7.49 (dd, *J* = 8.5, 7.0 Hz, 1H, H3), 7.35 (dd, *J* =
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3 8.3, 1.3 Hz, 1H, H8), 6.88 (dd, $J = 8.5, 0.5$ Hz, 1H, H2); ^1H NMR (500 MHz, d_5 -pyridine) δ
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5 8.34 (s, 1H, H12), 7.91 (dd, $J = 7.5, 0.5$ Hz, 1H, H10), 7.78 (d, $J = 7.0$ Hz, 1H, H4), 7.67 (dd
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7 [app. t], $J_1 = J_2 = 8.0$ Hz, 1H, H9), 7.51 (dd, $J = 8.3, 7.3$ Hz, 1H, H3), 7.39 (d, $J = 8.0$ Hz, 1H,
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9 H8), 7.13 (d, $J = 8.5$ Hz, 1H, H2); ^{13}C NMR (125 MHz, d_5 -pyridine) δ 193.7 (C6), 191.0
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11 (C13), 181.3 (C11), 163.2 (C7), 159.0 (C1), 158.3 (C5), 143.6 (C4a), 141.7 (C11a or C12a),[‡]
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13 138.3 (C3 and C9), 136.0[§] (C4b), 134.4 (C10a), 125.2 (C8), 121.7 (C2), 121.6 (C5a), 120.5
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15 (C10), 119.0 (C13a), 118.0 (C4), 116.8 (C6a), 114.5 (C12); HRMS (APCI⁻) m/z : $[\text{M}]^-$
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17 Calcd for $\text{C}_{21}\text{H}_{10}\text{O}_6$ 358.0477; Found 358.0488. NMR assignments were made with the
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19 assistance of COSY, HSQC and HMBC experiments (S53, S54 and S55, respectively).
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26 ^{13}C NMR Chemical Shift Calculations

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28 Calculations conducted at the HF 6-31G* level of theory were carried out using the Spartan
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30 08 software package.⁹⁹ Those performed at the HF/6-311+G(2d,p), B3LYP/6-31G* and
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32 B3LYP/6-311+G(2d,p) levels of theory were conducted using the Gaussian 09 software
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34 package.¹⁰⁰ Where two different levels of theory are noted, the ^{13}C NMR shifts were
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36 predicted at the first level of theory, and the structure was optimised at the second. Where
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38 only one level of theory is noted, both the structure optimisation and ^{13}C NMR chemical shift
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40 prediction were conducted using that level of theory. All calculations were conducted with
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42 solvation modelled in DMSO. Shifts are reported based on a tetramethylsilane reference
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44 calculated at the same level of theory as the structure optimisation.
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52 [‡] A signal corresponding to C11a or C12a could not be observed in the ^{13}C NMR spectrum, or any related NMR
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54 experiments. This carbon exhibited no correlations in the HMBC spectrum, which is not unexpected given that
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56 the analogous resonance at 141.7 ppm corresponds to either C11a or C12a and also exhibits a lack of carbon-
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58 proton correlations. It is therefore concluded that 'missing' signal corresponding to either C11a or C12a is also
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60 obscured by a solvent peak. Based upon ^{13}C NMR calculations (see Tables 1 and 3 and Table S2), it is expected
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62 that this resonance is obscured by the solvent signal at 136.3–135.5 ppm.

[§] This signal is obscured by a solvent peak in the ^{13}C NMR, and so the chemical shift is approximated from
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64 correlations observed in the HMBC spectrum.

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3 The calculated shifts were sorted in numerical order and then directly compared to the
4 experimentally determined shifts (also sorted in numerical order) for the molecule in question
5 (chrysazin in d_6 -DMSO¹¹ for Table 2, and **1** in d_5 -pyridine for Table S2). The mean of the
6 absolute value of the shift differences is reported, along with the absolute value of the largest
7 shift difference at the foot of Table 2, Figure 2, and Table S2. For Table S2, the ¹³C NMR
8 resonance of C11a or C12a in **1** could not be experimentally observed (see experimental for
9 **1**), and so was excluded from these shift difference calculations.
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20 ASSOCIATED CONTENT

21 Supporting Information

22
23 Additional ¹³C and ¹H NMR data, ¹³C NMR chemical shift predictions and ¹H and ¹³C NMR
24 spectra of new and known compounds. This material is available free of charge via the
25 Internet at <http://pubs.acs.org>.
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32 AUTHOR INFORMATION

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37
38 Author Contributions

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40 The manuscript was written through contributions of all authors. All authors have given
41 approval to the final version of the manuscript.
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