## SYNTHESIS OF AN ETHYLENE GLYCOL CROSS-LINKED AMINO ACID<sup>1</sup>

## Mengfei Ho\*, Weiheng Wang, Maria Douvlos, Thuha Pham, Timothy Klock Department of Medicinal Chemistry, School of Pharmacy State University of New York at Buffalo, Buffalo, NY 14260

**Abstract:** Synthesis of an ethylene glycol-containing amino acid,  $(2R,9S)-N^2-Cbz-N^9-Boc-2,9$ diamino-4,7-dioxadecanedioic acid 1-phenacyl 10-methyl diester 1, was studied through direct alkylation of N-Boc-L-serine and through BF3-catalyzed ring opening of N-acylaziridinecarboxylates.

In our modeling studies on conformationally constrained peptide analogs, we found that the ethylene glycol unit could provide certain structural features currently unavailable in peptide models containing disulfide cross-links. The favorable gauche placement for the O-C-C-O bond and anti placement for the C-O-C-C bond in oligoethylene glycols and crown ethers<sup>2,3</sup> would allow the ethylene glycol unit to serve as a covalent link which is longer than a disulfide bond, but which is less flexible and more hydrophilic than other carbon chain linkers. In order to test the effectiveness of such a cross-link in stabilizing peptide conformations, we have designed an ethylene glycol cross-linked amino acid 1. We have included four different protecting groups in this design to allow the possibility of manipulating two chiral centers when this cross-linked amino acid is incorporated into peptide analogs. These stereochemical and multifunctional considerations make the synthesis of 1 quite challenging. We report here our studies toward the synthesis of 1.



Ether-containing amino acids such as 4-oxalysine<sup>4</sup> and 5-oxahomolysine<sup>5</sup> have been prepared from alkylation of phthalimidomalonate. This method is incompatible with our objective to incorporate an ether-cross-linked amino acid into peptide analogs through conventional peptide synthesis, wherein the integrity of chiral centers and the application of orthogonally labile protecting groups are frequently emphasized. Direct O-alkylation of N-Boc-serine dianion prepared with NaH in DMF has been reported for the preparation of N-Boc-O-benzyl-L-serine (57%).<sup>6</sup> The alkylation appeared to be specific for ether formation and virtually no racemization was detected in the product. This procedure has also been reported for the preparation of O-allylserine derivatives.<sup>7</sup> We have similarly found that O- methylserine derivatives can easily be obtained with methyl iodide. Even though we observed no O-alkylation products with ethylene dibromide or ethylene diiodide under similar conditions, we were, however, able to obtain N-Boc-O-(2-benzyloxyethyl)-L-serine through direct alkylation with 2-benzyloxyethyl mesylate (15%) at room temperature.<sup>8</sup>

When benzyloxyethyl iodide (BEI) was used for O-alkylation (19 hr, r.t.) and was followed by methyl iodide workup, the ester of the desired product 2 (21.5%), the ester of recovered N-Boc-L-serine 4 (26%), methyl ether 3 (1.5%),  $\beta$ -elimination products 5 (< 2%) and 6 (yield undetermined) were obtained (Scheme 1).<sup>9</sup> The occurrence of 3 indicated that the alkylating agent was prematurely consumed. Further investigation showed that the yield of the alkylation product 2 could be increased substantially (up to 41%) with excess NaH and BEI (See table below). It is important to note that BEI apparently caused very little esterification even when 3 equivalents were used.



Scheme 2



Attempts to carry out the synthesis of 1 via direct alkylation of Boc-serine is outlined in Scheme 2. Mesylate 10 was prepared from chirogenic glycerol acetonide through the series of transformations shown, including an  $S_N 2$  step to introduce the nitrogen at the  $\alpha$ -position. Most of these steps can be carried out in excellent yield (close to or greater than 90%) with step vii (61%) as the only exception. This critical step was carried out by heating alcohol 9 with ClCH<sub>2</sub>CH<sub>2</sub>OTHP (2.5 eq.) and KOH (2.5 eq.) at 60° (Caution: Heating a mixture of 9 and KOH had resulted in an explosion in our laboratory). Ethylene glycol ether 11 was

obtained from 10 in 22% yield with two-fold excess of Boc-serine dianion and in 12% yield with one equivalent of the dianion. Longer reaction times (up to 3 days) with one equivalent of Boc-serine dianion did not improve the yield. Although most unreacted 10 can be recovered and the efficiency of this cross-linking alkylation may be improved with a large excess of Boc-serine dianion, the lengthy synthesis of 11 made this approach less appealing. Thus, we turned to a more practical alternative.

Nakajima et al.<sup>10</sup> have reported a convenient synthesis of optically active  $\beta$ -alkoxy- $\alpha$ amino acids via Lewis acid catalyzed ring opening of 2-aziridinecarboxylates (Azy). N-Acyl-Azy derivatives have also been shown to undergo ring opening by attack with other nucleophiles such as carboxylates, thiols and enolates.<sup>11</sup> Lanthionine and other thioether cross-linked amino acid derivatives have also been prepared through this route.<sup>14</sup> Although >90% yields could be achieved for alcoholysis of N-acyl-Azy derivatives using excess alcohol as co-solvent,<sup>10</sup> the synthesis of thioether cross-linked amino acids using only an equivalent of thiol nucleophile gave poor yields (up to 37%) and required unusually long reaction times (up to 5 days).<sup>14</sup> Despite the apparent need for a large excess of alcohol, we were able to obtain 1 from a 4:3 ratio of 14 and 16 in 64% yield.



As outlined in Scheme 3, Boc-L-Azy-OMe 13 and Cbz-D-Azy-OPac 15 were prepared from L-serine and D-serine via their N-trityl derivatives following published procedures.<sup>12,13</sup> Alcoholysis of 13 by 2-benzyloxyethanol gave 2 and subsequent hydrogenolysis yielded 14. The final coupling step was carried out as follows:

To a stirred solution of alcohol 14 (4.4 mmol) and aziridinecarboxylate 16 (3.3 mmol) in 5 mL of dry chloroform (distilled over P<sub>2</sub>O<sub>5</sub>) under N<sub>2</sub> at r.t., 0.5 mL of 10% BF<sub>3</sub> etherate (prepared in dry chloroform) was added. Three hours later, the reaction mixture was loaded on a silica gel column and purified by chromatography. Ethylene glycol cross-linked product (2*R*,9*S*)-*N*<sup>2</sup> -Cbz-*N*<sup>9</sup> -Boc-2,9-diamino-4,7-dioxadecanedioic acid 1-phenacyl 10-methyl diester 1 was obtained as an oil (1.28 g, 2.12 mmol; 64%): <sup>1</sup>H NMR (90 MHz, CDCl<sub>3</sub>)  $\delta$  1.40(9H, s), 3.40(4H, s), 3.68 (3H, s), 3.7-4.0(4H, m), 4.42(1H, m), 4.68(1H, m),

5.15(2H, s), 5.20, 5.23(2H, ABq, J=16.8 Hz), 5.64(1H, d, J=8.4 Hz), 6.0(1H, d, J=7.8 Hz), 7.35(5H, s) 7.4-7.6(3H, m), 7.92(2H, m); IR  $v_{max}$ (neat) 965, 1062, 1167, 1367, 1450, 1511, 1709, 1749 2850-3050, 3300-3400 cm<sup>-1</sup>; [ $\alpha$ ]<sup>D</sup> = +4.9 (c=7; EtOAc).<sup>15</sup>

In summary, direct alkylation of Boc-serine dianion with excess alkylating agent is an effective method for introducing an ethylene glycol unit into an amino acid side chain, but is less satisfactory for cross-linking two amino acid moieties. Lewis acid catalyzed ring opening of N-acyl aziridinecarboxylates with ethylene glycol-containing amino acid derivatives is a more practical method for preparation of ethylene-glycol cross-linked amino acids.

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## **References and Notes**

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- 8. The yield was based on the free acid purified by flash chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>:MeOH/4:1). If the reaction mixture was treated with methyl iodide before workup, the ester product was obtained after chromatography (EtOAc:Hexanes/1:4) in 20% yield.
- Methyl N Boc-N (2-Benzyloxyethyl)-2-aminoacrylate 5: <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.41(9H, s),
  3.67 (4H, s), 3.75(3H, s), 4.52(2H, s), 5.63(1H, s), 5.94(1H, s), 7.32(5H, s). Benzyl vinyl ether 6: <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 4.03(1H, dd, J=6.7 Hz, 2.0 Hz), 4.25(1H, dd, J=14.1 Hz, 2.0 Hz), 4.69(2H, s), 6.50(1H, dd, J=14.1 Hz, 6.7 Hz), 7.27(5H, s).
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- Compound 1 is not distinguishable from its (2S,9S)-diastereomer by TLC or NMR. However, derivatives of D- and L-Azy-OMe with Gerlach reagent (Gerlach, H.; Zagalak, B. J.C.S. Chem. Comm. 1973, 274.) showed that in our preparations both were enatiomerically pure within the limit of 400 MHz NMR instrument.

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