Diboron-Catalyzed Regio- and 1,2-*cis*- α -Stereoselective Glycosylation of *trans*-1,2-Diols

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arbohydrates play important roles in many biological processes, including cell proliferation, tumor metastasis, and immune recognition.¹ Methodologies for the efficient synthesis of homogeneous and structurally well-defined carbohydrates have thus attracted attention in various research fields to aid in understanding the precise biological roles and structure-activity relationships of these carbohydrates. Although there are many methods for the chemical synthesis of complex natural glycosides,² protecting groups must be carefully selected to control the glycosylation positions and anomeric configuration. The development of an efficient glycosylation method requiring minimal or no use of protecting groups is thus highly desirable. This has led to the development of regio- and stereoselective glycosylation methods using organoboron reagents, which can reversibly bind to *cis*-1,2- and 1,3-diols under mild conditions.³

In 1999, Aoyama et al.⁴ reported the regio- and Koenigs-Knorr-type glycosylation of a tetra-coordinate boronate ester, formed using a stoichiometric amount of an arylboronic acid and L-fucoside. Recently, Taylor et al. reported an efficient borinic-acid-catalyzed regio- and 1,2-trans-stereoselective glycosylation reaction.⁵ In a different approach, Kaji et al. reported a regio- and 1,2-trans-stereoselective glycosylation using an arylboronic acid as a transient masking reagent, which decreased the nucleophilicity of undesired hydroxyl groups by the formation of the corresponding boronic ester.⁶ These glycosylation methods require neighboring group participation from the 2-O-acyl functionality of the glycosyl donor to produce the corresponding 1,2-trans-glycoside with high stereoselectivity. On the other hand, the efficient synthesis of 1,2-cis-glycosides is difficult due to the unavailability of neighboring group participation. To address this, our group

has recently developed a boronic-acid-catalyzed regio- and 1,2cis-stereoselective glycosylation approach (Scheme 1A),7a-f namely boron-mediated aglycon delivery (BMAD).⁷ Cyclic boronic ester 4 or 5, derived from arylboronic acid 3 and cis-1,2-diol 1 or 1,3-diol 2, respectively, activated 1,2-anhydro donor 6 and induced glycosylation to provide the corresponding 1,2-cis glycoside 7 or 8 with high regio- and stereoselectivities. However, the binding affinity between an arylboronic acid and a trans-1,2-diol on a pyranose ring is quite low, preventing the effective application of BMAD to trans-1,2-diols on pyranosides. In this context, Morken et al. reported that tetrahydroxydiboron (10) could reversibly bind to trans-1,2-diols on a pyran ring to form 1,2-bonded diboron species.⁸ We, therefore, hypothesized that 1,2-bonded diboron species 11, derived from trans-1,2-diol 9 and diboron 10, can activate 6 and induce glycosylation to form 1,2-cis glycoside 12 with high stereoselectivity (Scheme 1B). Herein, we report the diboron-catalyzed regio- and 1,2-cis- α -stereoselective glycosylation of trans-1,2-diol sugar acceptors and its application to the synthesis of α -1,3-glucan pentasaccharide.

We selected 3,4,6-tri-O-benzyl-1,2-anhydroglucose $(14)^9$ and diboron 10 as the glycosyl donor and organoboron catalyst, respectively, and phenylboronic acid (15) as a negative control catalyst. First, the glycosylation of methyl

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Scheme 1. (A) Boronic-Acid-Catalyzed Regio- and 1,2-*cis*-Stereoselective Glycosylations of *cis*-1,2- and 1,3-Diol Sugar Acceptors; (B) Diboron-Catalyzed Regio- and 1,2-*cis*-Stereoselective Glycosylations of *trans*-1,2-Diol Sugar Acceptors



4,6-O-benzilidene- α -D-glucoside (13) with 14 was attempted using catalytic amounts of 15 in MeCN at room temperature for 4 h. The reaction proceeded poorly, providing α -glycosides 16 α and 17 α in 9% total yield with low regioselectivity but complete α -stereoselectivity, along with recovered 13 in 75% yield (Table 1, entry 1). Although we have not obtained clear

Table 1. Glycosylation of 1,2-Anhydro Donor 14 and trans-1,2-Diol 13



evidence at this stage, this result may suggest the formation of the relatively unstable seven-membered ring diarylpyroboronic ester 18^{10} (Figure 1A) and glycosylation catalysis with 14. In sharp contrast, the use of diboron 10 under the same reaction conditions uniquely resulted in smooth glycosylation to provide 16α and 17α in 93% total yield with modest regioselectivity ($16\alpha/17\alpha = 71:29$) and complete α -stereoselectivity, suggesting that relatively stable 1,2-bonded diboron



Figure 1. Proposed chemical equilibrium for the formation of (A) diarylpyroboronic ester 18 and (B) 1,2-bonded diboron species 19.

species 19 was formed as an active species (Figure 1B). Next, to confirm the generation of 19 under these reaction conditions, we conducted ¹H NMR studies using 13 (1.0 equiv) and 10 (0.2 equiv) in CD₃CN at room temperature. The results showed a 1:1 complex formation consistent with the proposed 19, as shown in Figure 2. We thus propose that



Figure 2. (A) ¹H NMR spectrum of 13 (100 mM) in CD_3CN_5 (B) ¹H NMR spectrum of 13 (100 mM) and 10 (20 mM) in CD_3CN .

the reaction mechanism comprises the formation of **19**, which activates glycosyl donor **14** without the need for additives. Sequential rearrangement of the B–O moiety in **20** affords **21**. Finally, a diol exchange reaction between **21** and **13** regenerates **19** and provides 1,2-*cis*- α -glycosides effectively (Scheme 2).

Next, we investigated regioselectivity using the present glycosylation method and several trans-1,2-diol sugar acceptors (22-27) under the same reaction conditions, as shown in Table 2. Using 22^{11} with the cyclic protecting group DTBS (di-tert-butylsilylene) as an acceptor resulted in smooth glycosylation with 14 to provide the corresponding α glycosides 22A and 22B in high yields with complete α stereoselectivity and high $\alpha(1,3)$ -regioselectivity (22A/22B = 92:8). In sharp contrast, when 23,¹² possessing TBS groups at the 4 and 6 positions, was used, the reaction provided $\alpha(1,2)$ glycoside 23B in moderate yield as a single isomer (entry 2, Table 2), suggesting that steric hindrance by the protecting groups at the 4 and 6 positions significantly affects regioselectivity in this glycosylation reaction. We thus focused on the size of the protecting groups and investigated the glycosylation of acceptors 24-27 possessing Ac,¹³ Bn,¹⁴ Bz, and Piv groups, respectively. In all cases, glycosylation proceeded smoothly to provide the corresponding 1,2-cis- α glycoside in a moderate to high yield, and $\alpha(1,2)$ -



^{*a*}Only the pathway for the generation of disaccharide 16α is shown.

Table 2. Glycosylations Using Several trans-1,2-Diols



regioselectivity increased as the size of the protecting group increased (entries 3–6, Table 2). These results indicate that the 1,2-*cis*- α -stereoselectivity was consistently very high, and the regioselectivity could be controlled by the size of the protecting group of the acceptor.

Next, we applied the present method to the synthesis of α -1,3-glucan (Scheme 3), a major cell wall component of various fungal pathogens, including *Aspergillus fumigatus*.¹⁵ α -1,3-Glucans were recently synthesized by Nifantiev et al.¹⁶ and Codée et al.¹⁷ In our synthetic strategy, we selected 3-*O*-Bn-4,6-*O*-DTBS-1,2-anhydroglucose **30**¹⁸ and *trans*-1,2-diol **22** as the glycosyl donor and initial glycosyl acceptor, respectively, to achieve high α (1,3)-selectivity. The Bn group at the 3 position of **30** would be selectively removed under mild conditions after the glycosylation of **22** with **30** to generate the *trans*-1,2-diol

Scheme 3. Efficient Synthesis of α -1,3-Glucan Pentasaccharide Using Diboron-Catalyzed Regio- and 1,2*cis-* α -Stereoselective Glycosylation Reactions^{*a*}



^aReagents and conditions: (a) NaOMe, MeOH, rt, 40 min, 98%; (b) ^bBu₂Si(OTf)₂, 2,6-lutidine, DMF, 0 °C, 30 min, 78%; (c) NaH, BnBr, TBAI, 0 °C to rt, 16 h, 82%; (d) dimethyldioxirane, CH₂Cl₂, 0 °C, 30 min, 98%.; (e) **30**, **10**, MeCN, 40 °C, 6 h, 84% (α (1,3) only) (92% BRSM); (f) Pd(OH)₂/C, H₂, EtOAc, rt, 30 min, 95%; (g) **30**, **10**, MeCN, 60 °C, 24 h, 73% (α (1,3) only) (90% BRSM); (h) Pd(OH)₂/ C, H₂, EtOAc, rt, 30 min, 81%; (i) **30**, **10**, MeCN, 50 °C, 24 h, 63% (α (1,3) only) (98% BRSM); (j) Pd(OH)₂/C, H₂, EtOAc, rt, 30 min, 83%; (k) **14**, **10**, MeCN, rt, 24 h, 81% (α (1,3) glycoside **37A**/ α (1,2) glycoside **37B** = 83:17); (l) Pd(OH)₂/C, H₂, EtOAc, rt, 30 min; (m) TBAF, THF, 0 °C to rt, 19 h, 53% in 2 steps.

sugar acceptor used in the next glycosylation. First, **29** was prepared from 3,4,6-tri-O-Ac-glycal (**28**) in three steps.¹⁹ Treatment of **29** with DMDO provided **30** in 98% yield with high diastereoselectivity ($\alpha/\beta = 92$:8). Next, we investigated the diboron-catalyzed glycosylation of **22** with **30** in the presence of **10**. After several attempts to optimize the reaction conditions, we found that the glycosylation of **22** (1.0 equiv) and **30** (3.0 equiv) using **10** (0.2 equiv) in MeCN at 40 °C for 6 h provided the best result, producing $\alpha(1,3)$ -glycoside **31** in 84% yield (92% BRSM) with excellent regio- and stereo-

selectivities. The deprotection of the Bn group of 31 under hydrogenolysis conditions gave 32. Next, glycosylation of triol sugar acceptor 32 with 30 provided the desired $\alpha(1,3)$ glycoside 33 as a single isomer. Deprotection of the Bn group of 33 gave trisaccharide 34, which possesses four free hydroxyl groups. Repetition of the glycosylation of 34 with 30 and the deprotection cycle generated tetrasaccharide 36, which possesses five free hydroxyl groups. The next glycosylation of 36 with 14 at room temperature proceeded smoothly to provide $\alpha(1,3)$ -glycoside 37A in 67% yield with good regioand excellent stereoselectivities. The anomeric configurations and glycosylated positions were confirmed by the coupling constants of the anomeric (H1) protons and the correlations between OH and H2 protons in the ¹H-¹H COSY NMR spectrum (see Supporting Information). Finally, deprotection of all protecting groups afforded α -1,3-glucan 38.

In conclusion, we developed the regio- and highly 1,2-*cis*- α stereoselective glycosylation of *trans*-1,2-diol sugar acceptors to extend the utility of our boron-mediated aglycon delivery (BMAD) method. NMR studies suggested the formation of the proposed 1,2-bonded diboron species **19** intermediate, and reactions proceeded smoothly to provide the corresponding 1,2-*cis*- α -glycosides. The stereoselectivity of the reaction products was consistently very high, and their regioselectivity was controlled by the protecting groups of the acceptor. Furthermore, this glycosylation method was applied successfully to the efficient synthesis of α -1,3-glucan pentasaccharide **38**. Detailed mechanistic studies of this method, its application to other types of *trans*-1,2-diol acceptors, and synthetic studies of other biologically active compounds using BMAD are now in progress.

EXPERIMENTAL SECTION

General Experimental Methods. NMR spectra were recorded on a JEOL ECA-500 (500 MHz for ¹H, 125 MHz for ¹³C) or JEOL ECZ-400 (400 MHz for ¹H, 100 MHz for ¹³C) spectrometer. ¹H NMR data are reported as follows: chemical shift in parts per million (ppm) downfield or upfield form CDCl₃ (δ 7.26), D₂O (δ 4.79), CD₃CN (δ 1.94), or tetrametry share (δ c.e., ..., multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = 1 constants (Hz). ¹³C quartet, and m = multiplet), and coupling constants (Hz). chemical shifts are reported in ppm downfield or upfield from CDCl₃ (δ 77.0). Using D₂O as an NMR solvent, ¹³C chemical shifts are reported in ppm downfield or upfield from acetone (δ 29.8) as an external reference. ESI-TOF mass spectra were measured on a Waters LCT Premier XE. Melting points were determined on a micro hotstage (Yanako MP-S3). Optical rotations were measured on a JASCO P-2200 polarimeter. Silica gel TLC and column chromatography were performed using Merck TLC 60F-254 (0.25 mm) and Silica Gel 60N (spherical, neutral, 63–210 $\mu{\rm m}$ or 40–50 $\mu{\rm m})$ (Kanto Chemical Co., Inc.), respectively. Reverse-phase column chromatography separation was performed using a Sep-Pak C18 reversed-phase cartridge (Waters). Air- and/or moisture-sensitive reactions were carried out under an argon atmosphere using oven-dried glassware.

Procedures for the Synthesis of Compound 26. To a solution of methyl 2,3-di-*O*-Bn-*α*-D-glucoside²⁰ (727 mg, 1.94 mmol) in pyridine (0.3 M, 6.47 mL) was added benzoyl chloride (541 μL, 4.66 mmol) at 0 °C under an Ar atmosphere. After the reaction mixture was stirred for 17 h at room temperature, the reaction was quenched by the addition of H₂O (10 mL). The aqueous layer was extracted with EtOAc (20 mL × 3), and then the combined extracts were washed with brine (20 mL), dried over anhydrous sodium sulfate, filtered, and concentrated *in vacuo*. Purification of the residue by silica gel column chromatography (2:1 *n*-hexane/EtOAc) gave methyl 2,3-di-*O*-Bn-4,6-di-*O*-Bz-*α*-D-glucoside (39) (1.09 g, 1.88 mmol, 97% yield).

Compound 39: white solid; mp 96–97 °C; R_f 0.43 (2:1 *n*-hexane/EtOAc); $[\alpha]_D^{26}$ +14.8° (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.01–7.97 (4H, m), 7.64–7.30 (12H, m), 7.14–7.06 (4H, m), 5.39 (1H, dd, *J* = 9.6 Hz, *J* = 10.0 Hz), 4.85 and 4.65 (2H, ABq, *J* = 11.2 Hz), 4.83 and 4.68 (2H, ABq, *J* = 12.0 Hz), 4.65 (1H, d, *J* = 3.6 Hz), 4.50 (1H, dd, *J* = 2.8, 12.0 Hz), 4.33 (1H, dd, *J* = 5.6, 12.0 Hz), 4.16 (1H, m), 4.11 (1H, dd, *J* = 9.2, 9.6 Hz), 3.68 (1H, dd, *J* = 3.6, 9.2 Hz), 3.43 (3H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 166.1, 165.2, 137.9, 137.8, 133.5, 133.2, 132.9, 130.0, 129.7, 129.6 × 2, 129.4, 129.3, 128.4, 128.3 × 2, 128.2, 128.1, 128.0 × 2, 127.9, 127.4, 98.1, 79.6, 78.9, 75.5, 73.5, 70.8, 67.5, 63.2, 55.4; HRMS (ESITOF) m/z [M + Na]⁺ calcd for C₃₅H₃₄O₈Na 605.2151, found 605.2180.

A solution of methyl 2,3-di-O-Bn-4,6-di-O-Bz- α -D-glucoside (39) (304 mg, 0.522 mmol) in dry THF (0.3 M, 1.74 mL) was stirred under H₂ atmosphere in the presence of 20% Pd(OH)₂/C (30.4 mg) for 2 h at room temperature. After changing the atmosphere to Ar, the mixture was filtered through a Celite pad (washed with EtOAc), and then the filtrate was concentrated *in vacuo*. Purification of the residue by silica gel column chromatography (2:1 toluene/acetone) gave 26 (199 mg, 0.496 mmol, 95% yield).

Compound 26: colorless foam; $R_f 0.23$ (2:1 toluene/acetone); $[\alpha]_D^{27}$ +166.3° (*c* 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.07–8.01 (4H, m), 7.60–7.53 (2H, m), 7.46–7.38 (4H, m), 5.25 (1H, dd, J = 9.6, 10.0 Hz), 4.88 (1H, d, J = 4.0 Hz), 4.58 (1H, dd, J = 2.8, 12.4 Hz), 4.40 (1H, dd, J = 5.6, 12.4 Hz), 4.22 (1H, m), 4.04 (1H, dd, J = 9.2, 9.6 Hz), 3.72 (1H, dd, J = 4.0, 9.2 Hz), 3.49 (3H, s), 2.76 (1H, br-s), 2.34 (1H, br-s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 166.1, 165.9, 133.3, 133.0, 129.8 × 2, 129.6 × 3, 129.2, 128.3 × 2, 128.2 × 2, 99.1, 72.4 × 2, 71.6, 67.6, 63.4, 55.4; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₂₁H₂₂O₈Na 425.1192, found 425.1192.

Procedures for the Synthesis of Compound 27. To a solution of methyl 2,3-di-O-Bn- α -D-glucoside²⁰ (183 mg, 0.489 mmol) in pyridine (0.3 M, 1.63 mL) was added pivaloyl chloride (147 μ L, 1.17 mmol) at 0 °C under an Ar atmosphere. After the reaction mixture was stirred for 22.5 h at 80 °C, the reaction was guenched by the addition of H_2O (5 mL). The aqueous layer was extracted with EtOAc (10 mL \times 3), and then the combined extracts were washed with brine (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated in vacuo. Purification of the residue by silica gel column chromatography (4:1 n-hexane/EtOAc) gave 2,3-di-O-Bn-4,6-di-O-Piv- α -D-glucoside (40) (249 mg, 0.460 mmol, 94% yield). Compound **40**: white solid; mp 117–118 °C; R_f 0.57 (2:1 *n*-hexane/EtOAc); $[\alpha]_{D}^{26}$ +25.6° (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.34– 7.24 (10H, m), 5.01 (1H, dd, J = 9.6, 10.0 Hz), 4.90 and 4.63 (2H, ABq, J = 11.2 Hz), 4.77 and 4.61 (2H, ABq, J = 12.0 Hz), 4.60 (1H, d, I = 3.6 Hz, 4.13 (1H, dd, I = 2.0, 12.0 Hz), 4.02 (1H, dd, I = 6.0, 12.0 Hz), 3.93 (1H, dd, J = 9.6, 10.0 Hz), 3.90 (1H, m), 3.59 (1H, dd, J = 3.6, 9.6 Hz, 3.40 (3H, s), 1.21 (9H, s), 1.17 (9H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 178.1, 176.8, 138.3, 137.9, 128.5 \times 2, 128.2×2 , 128.1×2 , 128.0, 127.4, 127.2×2 , 98.1, 79.6, 79.4, 75.4, 73.5, 69.6, 67.9, 62.4, 55.3, 38.8, 38.7, 27.1 × 3, 27.0 × 3; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₃₁H₄₂O₈Na 565.2777, found 565.2786.

A solution of methyl 2,3-di-O-Bn-4,6-di-O-Piv- α -D-glucoside (40) (174 mg, 0.321 mmol) in dry THF (0.3 M, 1.07 mL) was stirred under H₂ atmosphere in the presence of 20% Pd(OH)₂/C (17.4 mg) for 3 h at room temperature. After changing the atmosphere to Ar, the mixture was filtered through a Celite pad (washed with EtOAc), and then the filtrate was concentrated *in vacuo*. Purification of the residue by silica gel column chromatography (2:1 *n*-hexane/acetone) gave 27 (110 mg, 0.305 mmol, 95% yield).

Compound 27: colorless syrup; $R_f 0.37$ (1:5 *n*-hexane/EtOAc); $[\alpha]_{D}^{27}$ +105.6° (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 4.83 (1H, dd, J = 9.2, 10.0 Hz), 4.81 (1H, d, J = 4.0 Hz), 4.20 (1H, dd, J = 2.4, 12.0 Hz), 4.12 (1H, dd, J = 5.6, 12.0 Hz), 3.94 (1H, m), 3.83 (1H, ddd, J = 4.0, 9.2, 9.6 Hz), 3.59 (1H, ddd, J = 4.0, 8.4, 9.6 Hz), 3.45 (3H, s), 2.55 (1H, d, J = 4.0 Hz), 2.21 (1H, d, J = 8.4 Hz), 1.23 (9H, s), 1.22 (9H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 178.1 × 2, 99.1, 72.8, 72.7, 70.3, 67.9, 62.4, 55.4, 38.9, 38.8, 27.1 × 3, 27.0 ×

3; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₁₇H₃₁O₈ 363.2019, found 363.2015.

Compound 16\alpha and 17\alpha. To a solution of tetrahydroxydiboron (10) (0.80 mg, 8.92 μ mol) and 13²¹ (12.6 mg, 44.6 μ mol) in dry MeCN (0.2 M, 446 μ L) was added a solution of 14⁹ (38.6 mg, 89.2 μ mol) in dry MeCN (0.2 M, 446 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 9.81 μ mol). The aqueous layer was extracted with EtOAc (3 mL \times 5), and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. Purification of the residue by silica gel column chromatography (1:1 n-hexane/EtOAc) gave a mixture of **16** α and **17** α (29.7 mg, 41.5 μ mol, 93% yield, $\alpha(1,3)/\alpha(1,2) = 2.5:1$) as an inseparable mixture.²² Compound **16** α : white solid; mp 164– 165 °C; R_f 0.35 (1:1 *n*-hexane/EtOAc); $[\alpha]_D^{27}$ +76.4° (c 0.76, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.47–7.45 (2H, m), 7.39– 7.24 (16H, m), 7.16-7.13 (2H, m), 5.56 (1H, s), 5.12 (1H, s, J = 2.8 Hz), 4.93 and 4.78 (2H, ABq, J = 11.6 Hz), 4.80 (1H, d, J = 4.0 Hz), 4.82 and 4.47 (2H, ABq, J = 10.8 Hz), 4.57 and 4.51 (2H, ABq, J = 12.0 Hz), 4.31 (1H, dd, J = 4.8, 10.0 Hz), 4.19 (1H, m), 3.99 (1H, dd, J = 9.2, 9.6 Hz, 3.83 (1H, m), 3.77–3.72 (3H, m), 3.68–3.55 (4H, m), 3.46 (1H, dd, J = 8.8, 10.0 Hz), 3.42 (3H, s), 3.14 (1H, d, J = 7.6 Hz), 2.41 (1H, d, J = 9.2 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 138.7, 138.0, 137.7, 136.8, 129.0, 128.3, 128.2, 127.8, 127.6, 127.5, 125.8, 101.4, 100.2, 100.1, 83.4, 80.4, 80.1, 77.3, 75.1, 74.9, 73.4, 71.7, 70.9, 68.9, 68.7, 62.2, 55.4; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₄₁H₄₆O₁₁Na 737.2938, found 737.2928.

Compound 17*a***:** white solid; mp 166–167 °C; R_f 0.35 (1:1 *n*-hexane/EtOAc); $[\alpha]_D^{27}$ +109.6° (*c* 0.98, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.52–7.48 (2H, m), 7.40–7.24 (16H, m), 7.15–7.12 (2H, m), 5.53 (1H, s), 4.98 (1H, d, *J* = 3.6 Hz), 4.97 and 4.84 (2H, ABq, *J* = 11.6 Hz), 4.85 (1H, d, *J* = 5.6 Hz), 4.81 and 4.49 (2H, ABq, *J* = 10.8 Hz), 4.63 and 4.48 (2H, ABq, *J* = 12.0 Hz), 4.29 (1H, m), 4.12 (1H, m), 4.05 (1H, dd, *J* = 8.0, 8.8 Hz), 3.85–3.61 (8H, m), 3.48 (1H, dd, *J* = 8.8, 9.6 Hz), 3.43 (3H, s), 2.89 (1H, d, *J* = 2.4 Hz), 2.41 (1H, d, *J* = 9.6 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 138.7, 138.0, 137.8, 137.0, 129.2, 128.3 × 2, 127.9 × 3, 127.7 × 2, 127.6, 126.3, 102.0, 97.8, 97.7, 83.2, 81.2, 78.3, 77.2, 75.3, 75.1, 73.4, 72.8, 70.9, 69.3, 68.9, 68.3, 62.0, 55.3; HRMS (ESI-TOF) *m*/*z* [M + H]⁺ calcd for C₄₁H₄₇O₁₁ 715.3118, found 715.3107.

Compound 22A and 22B. To a solution of 10 (0.56 mg, 6.25 μ mol) and 22¹¹ (10.4 mg, 31.2 μ mol) in dry MeCN (0.2 M, 313 μ L) was added a solution of 14 (27.0 mg, 62.5 μ mol) in dry MeCN (0.2 M, 313 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 6.86 μ mol). The aqueous layer was extracted with EtOAc (3 mL × 5), and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC (4:1 toluene/EtOAc) to afford **22A** (18.4 mg, 24.0 μ mol, 77% yield) and **22B** (1.6 mg, 2.18 μ mol, 7% yield).

Compound 22A: colorless syrup; $R_f 0.57$ (2:1 *n*-hexane/EtOAc); $[\alpha]_{D}^{25}$ +61.6° (*c* 1.20, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.38 (2H, m), 7.34–7.25 (11H, m), 7.14–7.12 (2H, m), 5.04 (1H, d, *J* = 3.5 Hz), 5.03 and 4.76 (2H, ABq, *J* = 11.0 Hz), 4.83 and 4.45 (2H, ABq, *J* = 11.0 Hz), 4.75 (1H, d, *J* = 5.5 Hz), 4.56 and 4.52 (2H, ABq, *J* = 12.5 Hz), 4.15–4.12 (2H, m), 3.88 (1H, dd, *J* = 10.0, 10.0 Hz), 3.81–3.69 (5H, m), 3.66 (1H, dd, *J* = 2.0, 10.0 Hz), 3.58–3.53 (2H, m), 3.45–3.41 (4H, m), 3.30 (1H, d, *J* = 5.5 Hz), 3.03 (1H, d, *J* = 10.0 Hz), 1.07 (9H, s), 1.01 (9H, s); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 138.8, 138.0, 137.7, 128.4, 128.3 × 2, 128.0, 127.9, 127.8, 127.7 × 2, 127.5, 101.5, 99.8, 85.1, 83.9, 76.2, 75.3, 75.1, 74.2, 73.4, 71.5, 71.2, 68.7, 66.6, 66.0, 55.6, 27.5, 26.9, 22.7, 19.9; HRMS (ESITOF) m/z [M + Na]⁺ calcd for C₄₂H₅₈O₁₁SiNa 789.3646, found 789.3622.

Compound 22B: colorless syrup; R_f 0.49 (2:1 *n*-hexane/acetone); $[\alpha]_{D}^{25}$ +37.2° (*c* 0.46, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.38 (2H, m), 7.35–7.24 (11H, m), 7.16–7.14 (2H, m), 4.98 and

4.82 (2H, ABq, J = 11.5 Hz), 4.96 (1H, d, J = 4.0 Hz), 4.83 and 4.50 (2H, ABq, J = 11.0 Hz), 4.78 (1H, d, J = 4.0 Hz), 4.65 and 4.47 (2H, ABq, J = 12.0 Hz), 4.19 (1H, m), 4.11 (1H, dd, J = 5.0, 10.0 Hz), 3.86 (1H, dd, J = 10.0, 10.0 Hz), 3.82–3.77 (3H, m), 3.74–3.63 (6H, m), 3.42 (3H, s), 2.64 (1H, br-s), 2.46 (1H, d, J = 10.0 Hz), 1.05 (9H, s), 0.99 (9H, s); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 138.8, 138.3, 137.9, 128.3 × 2, 127.9 × 2, 127.8, 127.6 × 2, 127.5, 97.3, 83.6, 77.4, 76.4, 75.3, 74.9, 73.4, 73.0, 72.1, 70.6, 68.3, 66.4, 65.6, 55.4, 27.4, 26.9, 22.7, 19.9; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₄₂H₅₉O₁₁Si 767.3827, found 767.3803.

Compound 23B. To a solution of **10** (0.57 mg, 6.36 μ mol) and 23^{12} (13.4 mg, 31.8 μ mol) in dry MeCN (0.2 M, 318 μ L) was added a solution of 14 (27.5 mg, 63.6 μ mol) in dry MeCN (0.2 M, 318 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 7.00 μ mol). The aqueous layer was extracted with EtOAc $(3 \text{ mL} \times 5)$, and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC (5:1 n-hexane/acetone) to afford 23B (12.3 mg, 14.3 µmol, 45% yield): colorless syrup; R_f 0.31 (5:1 nhexane/acetone); $[\alpha]_{D}^{28}$ +102.9° (c 0.73, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.40-7.39 (2H, m), 7.34-7.26 (11H, m), 7.13-7.11 (2H, m), 4.99 and 4.82 (2H, ABq, J = 11.0 Hz), 4.96 (1H, d, J = 4.0 Hz), 4.82 and 4.47 (2H, ABq, J = 11.0 Hz), 4.78 (1H, d, J = 3.5 Hz), 4.61 and 4.51 (2H, ABq, J = 12.0 Hz), 4.04 (1H, m), 3.85 (1H, br-d, J = 11.0 Hz), 3.77-3.67 (6H, m), 3.60-3.55 (2H, m), 3.51-3.49 (2H, m), 3.38 (3H, s), 2,57 (1H, d, J = 3.5 Hz), 2.50 (1H, d, J = 10.0 Hz), 0.89 (9H, s), 0.88 (9H, s), 0.12 (3H, s), 0.09 (3H, s), 0.06 (6H, s); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 138.7, 138.0, 137.8, 128.4 × 2, 128.0, 127.9, 127.7 × 2, 127.6, 97.8, 96.7, 83.5, 78.7, 75.3, 75.1, 73.4, 73.1, 72.9, 72.3, 71.1, 71.0, 68.3, 62.4, 54.7, 25.9 × 2, 18.4, 18.3, -3.9, -4.9, -5.1, -5.4; HRMS (ESI-TOF) m/z [M + K]⁺ calcd for C46H70O11Si2K 893.4094, found 893.4112.

Compound 24A and 24B. To a solution of 10 (0.64 mg, 7.14 μ mol) and 24¹³ (10.2 mg, 35.7 μ mol) in dry MeCN (0.2 M, 357 μ L) was added a solution of 14 (30.9 mg, 71.4 μ mol) in dry MeCN (0.2 M, 357 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 7.85 μ mol). The aqueous layer was extracted with EtOAc (3 mL × 5), and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacua*. The residue was purified by preparative TLC (4:1 toluene/EtOAc) to afford 24A (9.3 mg, 13.0 μ mol, 36% yield) and 24B (8.9 mg, 12.9 μ mol, 36% yield).

Compound 24A: colorless syrup; $R_f 0.42$ (4:1 toluene/acetone); $[\alpha]_{D}^{25}$ +89.3° (*c* 0.72, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.37–7.27 (13H, m), 7.15–7.13 (2H, m), 5.05 (1H, dd, J = 9.6, 10.0 Hz), 4.91 (1H, d, J = 4.0 Hz), 4.90 and 4.80 (2H, ABq, J = 11.2 Hz), 4.80 (1H, d, J = 5.2 Hz), 4.80 and 4.46 (2H, ABq, J = 11.2 Hz), 4.56 and 4.49 (2H, ABq, J = 13.2 Hz), 4.28–4.21 (2H, m), 4.07 (1H, br-d, J = 12.4 Hz), 3.90–3.84 (2H, m), 3.72–3.55 (5H, m), 3.45 (1H, dd, J = 9.2, 9.6 Hz), 3.40 (3H, s), 3.24 (1H, d, J = 8.0 Hz), 2.10 (3H, s), 2.07 (3H, s), 1.93 (1H, d, J = 9.2 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.7 × 2, 138.5, 138.0, 137.6, 128.4 × 2, 127.9 × 2, 127.8, 127.7, 127.6, 100.4, 99.6, 83.1, 81.8, 77.5, 75.4, 74.9, 73.5, 72.9, 71.1, 70.9, 69.3, 68.8, 67.3, 62.1, 55.5, 20.9, 20.7; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₃₈H₄₆O₁₃Na 733.2836, found 733.2801.

Compound 24B: white solid; mp 125–126 °C; R_f 0.36 (4:1 toluene/acetone); $[\alpha]_D^{25}$ +103.8° (*c* 0.95, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.39–7.27 (13H, m), 7.15–7.13 (2H, m), 4.96–4.88 (3H, m), 4.84–4.79 (3H, m), 4.59 and 4.48 (2H, ABq, *J* = 12.4 Hz), 4.48 (1H, ABq, *J* = 12.8 Hz), 4.25 (1H, dd, *J* = 4.8, 12.4 Hz), 4.10–4.06 (2H, m), 3.94 (1H, dd, *J* = 9.2, 9.2 Hz), 3.90 (1H, m), 3.78–3.66 (5H, m), 3.58 (1H, dd, *J* = 9.2, 9.2 Hz), 3.41 (3H, s), 3.03 (1H, br-s), 2.35 (1H, br-s), 2.08 (3H, s), 2.07 (3H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.7, 170.3, 138.6, 138.0, 137.7, 128.4, 127.9 × 2, 127.7, 127.6, 97.9, 97.3, 83.2, 78.6, 77.2, 75.3, 75.0, 73.5, 72.7, 71.0,

70.5, 70.3, 68.5, 67.2, 62.2, 55.3, 20.8, 20.7; HRMS (ESI-TOF) m/z [M + K]⁺ calcd for C₃₈H₄₆O₁₃K 749.2576, found 749.2556.

Compound 25A and 25B. To a solution of **10** (0.51 mg, 5.69 μ mol) and **25**¹⁴ (10.6 mg, 28.4 μ mol) in dry MeCN (0.2 M, 285 μ L) was added a solution of **14** (24.6 mg, 56.9 μ mol) in dry MeCN (0.2 M, 285 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 6.25 μ mol). The aqueous layer was extracted with EtOAc (3 mL × 5), and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC (4:1 CHCl₃/EtOAc) to afford **25A** (4.7 mg, 5.82 μ mol, 21% yield) and **25B** (8.1 mg, 10.0 μ mol, 35% yield).

Compound 25A: colorless syrup; $R_f 0.47$ (4:1 CHCl₃/EtOAc); $[\alpha]_D^{27}$ +131.7° (*c* 0.19, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.37– 7.16 (25H, m), 5.19 (1H, d, *J* = 3.5 Hz), 4.86–4.77 (5H, m), 4.67 (1H, ABq, *J* = 12.0 Hz), 4.59–4.45 (5H, m), 4.27 (1H, m), 3.92 (1H, dd, *J* = 9.0, 9.0 Hz), 3.79–3.56 (9H, m), 3.48 (1H, dd, *J* = 8.5 Hz, *J* = 8.0 Hz), 3.42 (1H, d, *J* = 10.0 Hz), 3.37 (3H, s), 2.22 (1H, d, *J* = 8.0 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 138.6, 138.0, 137.8 × 2, 128.4, 128.0, 127.9 × 2, 127.8 × 2, 127.7, 99.6, 83.1 × 2, 77.7, 77.2, 75.3, 74.9, 74.6, 73.6, 73.5, 72.9, 71.5, 71.0, 69.9, 68.8, 68.4, 55.2; HRMS (ESI-TOF) *m/z* [M + Na]⁺ calcd for C₄₈H₅₄O₁₁Na 829.3564, found 829.3571.

Compound 25B: colorless syrup; $R_f 0.33$ (4:1 CHCl₃/EtOAc); $[\alpha]_D^{25}$ +94.7° (*c* 0.69, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.14 (25H, m), 4.98–4.96 (2H, m), 4.84–4.79 (4H, m), 4.64–4.60 (2H, m), 4.52–4.48 (4H, m), 4.07 (1H, m), 3.99 (1H, dd, *J* = 9.0, 9.0 Hz), 3.77–3.52 (10H, m), 3.38 (3H, s), 2.76 (1H, d, *J* = 3.0 Hz), 2.37 (1H, d, *J* = 9.5 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 138.7, 138.4, 138.1, 128.0, 137.8, 128.4, 127.9, 127.7, 127.6, 97.9, 97.2, 83.4, 78.7, 77.6, 77.2, 77.1, 75.3, 75.0, 74.8, 73.5 × 2, 73.0, 72.9, 71.0, 69.8, 68.5, 55.0; HRMS (ESI-TOF) *m*/*z* [M + Na]⁺ calcd for C₄₈H₅₄O₁₁Na 829.3564, found 829.3527.

Compound 26A and 26B. To a solution of 10 (0.68 mg, 7.59 μ mol) and 26 (15.3 mg, 37.9 μ mol) in dry MeCN (0.2 M, 380 μ L) was added a solution of 14 (32.8 mg, 75.9 μ mol) in dry MeCN (0.2 M, 380 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 8.34 μ mol). The aqueous layer was extracted with EtOAc (3 mL × 5), and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC (5:1 toluene/acetone) to afford 26A (10.3 mg, 12.3 μ mol, 32% yield) and 26B (20.2 mg, 24.1 μ mol, 64% yield).

Compound 26A: colorless syrup; R_f 0.52 (5:1 toluene/acetone); $[\alpha]_D^{25}$ +84.2° (*c* 0.61, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.02–8.01 (4H, m), 7.58–7.53 (2H, m), 7.44–7.39 (4H, m), 7.34–7.24 (13H, m), 7.13–7.12 (2H, m), 5.42 (1H, dd, *J* = 9.5, 9.5 Hz), 4.94 (1H, d, *J* = 4.0 Hz), 4.83 (1H, d, *J* = 4.0 Hz), 4.83 and 4.73 (2H, ABq, *J* = 11.5 Hz), 4.78 and 4.43 (2H, ABq, *J* = 10.5 Hz), 4.58–4.48 (3H, m), 4.38 (1H, dd, *J* = 5.5, 12.0 Hz), 4.26–4.19 (2H, m), 4.08 (1H, dd, *J* = 9.0, 9.5 Hz), 3.75–3.64 (3H, m), 3.56–3.49 (2H, m), 3.45 (3H, s), 3.43 (1H, d, *J* = 8.0 Hz), 3.39 (1H, dd, *J* = 9.5, 10.0 Hz), 1.82 (1H, d, *J* = 9.5 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 166.2, 166.0, 138.6, 138.0, 137.6, 133.6, 133.1, 129.8, 129.7, 129.1, 128.6, 128.4, 127.9, 127.8, 127.7, 127.6, 100.2, 99.5, 83.0, 81.8, 77.2, 75.4, 74.9, 73.5, 72.9, 71.3, 70.9, 70.5, 68.8, 67.4, 63.1, 55.5; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₄₈H₅₀O₁₃Na 857.3149, found 857.3174.

Compound 26B: colorless syrup; $R_f 0.45$ (5:1 toluene/acetone); $[\alpha]_D^{25} + 51.9^{\circ}$ (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.06–8.01 (4H, m), 7.59–7.52 (2H, m), 7.44–7.37 (6H, m), 7.34–7.20 (11H, m), 7.14–7.11 (2H, m), 5.28 (1H, dd, *J* = 9.0, 9.0 Hz), 5.01 (1H, d, *J* = 4.0 Hz), 4.95 and 4.83 (2H, ABq, *J* = 11.0 Hz), 4.90 (1H, d, *J* = 3.5 Hz), 4.80 and 4.45 (2H, ABq, *J* = 11.5 Hz), 4.59–4.55 (2H, m), 4.48 (1H, ABq, *J* = 12.5 Hz), 4.38 (1H, dd, *J* = 5.5, 12.5 Hz), 4.21 (1H, m), 4.18–4.07 (2H, m), 3.82–3.70 (3H, m), 3.66 (2H, d), 3.59 (1H, dd, *J* = 9.0, 10.0 Hz), 3.46 (3H, s), 3.01 (1H, d, *J* = 4.5 Hz), 2,30

(1H, d, J = 9.5 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 166.2, 165.9, 138.6, 138.0, 137.6, 133.4, 133.1, 129.9, 129.7, 129.4, 128.4 × 2, 127.9 × 2, 127.8, 127.7 × 2, 98.0, 97.3, 83.2, 78.6, 77.2, 75.3, 75.0, 73.5, 72.8, 71.5, 71.0, 70.7, 68.4, 67.4, 63.2, 55.4; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₄₈H₅₀O₁₃Na 857.3149, found 857.3113.

Compound 27A and 27B. To a solution of 10 (0.45 mg, 5.02 μ mol) and 27 (9.10 mg, 25.1 μ mol) in dry MeCN (0.2 M, 251 μ L) was added a solution of 14 (21.7 mg, 50.2 μ mol) in dry MeCN (0.2 M, 251 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 4 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 5.52 μ mol). The aqueous layer was extracted with EtOAc (3 mL × 5), and then the combined extracts were washed with sat. NH₄Cl (aq, 2 mL), sat. NaHCO₃ (aq, 2 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC (3:1 toluene/EtOAc) to afford 27A (3.0 mg, 3.77 μ mol, 15% yield) and 27B (8.9 mg, 11.3 μ mol, 45% yield).

Compound 27A: colorless syrup; R_f 0.48 (3:1 toluene/EtOAc); $[\alpha]_D^{25} + 93.8^{\circ}$ (*c* 0.31, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.38–7.37 (2H, m), 7.32–7.25 (11H, m), 7.13–7.11 (2H, m), 5.05 (1H, dd, *J* = 9.0, 9.5 Hz), 4.97 and 4.78 (2H, ABq, *J* = 11.5 Hz), 4.90 (1H, d, *J* = 4.0 Hz), 4.81 and 4.44 (2H, ABq, *J* = 10.5 Hz), 4.76 (1H, d, *J* = 4.0 Hz), 4.56 and 4.49 (2H, ABq, *J* = 12.5 Hz), 4.27 (1H, m), 4.17 (1H, br-d, 12.0 Hz), 4.04 (1H, dd, *J* = 5.5, 12.0 Hz), 3.92 (1H, m), 3.86 (1H, dd, *J* = 9.0, 9.5 Hz), 3.77–3.74 (1H, dd, *J* = 6.5, 6.5 Hz), 3.70–3.56 (5H, m), 3.44 (1H, dd, *J* = 8.5, 8.5 Hz), 3.39 (3H, s), 3.08 (1H, d, *J* = 9.0 Hz), 1.90 (1H, d, *J* = 10.5 Hz), 1.22 (18H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 178.1, 138.7, 138.1, 137.7, 128.4, 128.3, 128.0, 127.9 × 2, 127.7 × 2, 127.5, 100.4, 99.5, 83.3, 81.5, 77.2, 75.4, 74.9, 73.6, 73.1, 71.2, 70.7, 69.3, 68.9, 67.6, 62.1, 55.5, 39.1, 38.9, 27.1, 27.0; HRMS (ESI-TOF) *m*/*z* [M + Na]⁺ calcd for C₄₄H₅₈O₁₃Na 817.3775, found 817.3813.

Compound 27B: colorless syrup; R_f 0.34 (3:1 toluene/EtOAc); $[\alpha]_{D}^{25}$ +89.0° (*c* 0.86, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.37 (2H, m), 7.34–7.26 (11H, m), 7.13–7.11 (2H, m), 4.98–4.94 (2H, m), 4.89–4.81 (3H, m), 4.80 and 4.47 (2H, ABq, *J* = 10.5 Hz), 4.60 and 4.48 (2H, ABq, *J* = 12.0 Hz), 4.18 (1H, dd, *J* = 1.5, 12.0 Hz), 4.13–4.06 (2H, m), 3.98–3.92 (2H, m), 3.76–3.62 (5H, m), 3.59 (1H, dd, *J* = 8.5, 9.5 Hz), 3.42 (3H, s), 2.78 (1H, d, *J* = 5.0 Hz), 2.34 (1H, d, *J* = 9.5 Hz), 1.22 (9H, s), 1.21 (9H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 178.1, 177.8, 138.7, 138.1, 137.7, 128.4 × 2, 128.3, 128.0, 127.9, 127.8 × 2, 127.7, 127.6, 97.7, 97.1, 83.3, 78.3, 77.2, 75.3, 75.0, 73.4, 72.8, 70.9, 70.8, 70.2, 68.3, 67.5, 62.3, 55.3, 38.9, 27.1, 27.0; HRMS (ESI-TOF) *m*/*z* [M + Na]⁺ calcd for C₄₄H₅₈O₁₃Na 817.3775, found 817.3813.

Compound 31. To a solution of 10 (34.2 mg, 0.381 mmol) and 22 (637.7 mg, 1.90 mmol) in dry MeCN (28.6 mL) was added a solution of $3 \breve{0}^{18}$ (2.24 g, 5.72 mmol) in dry MeCN (28.6 mL) at 40 °C under an Ar atmosphere. After the reaction mixture was stirred for 6 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 0.419 mmol, 7.6 mL). The aqueous layer was extracted with EtOAc (50 mL \times 3), and then the combined extracts were washed with sat. $\rm NH_4Cl$ (aq, 50 mL), sat. $\rm NaHCO_3$ (aq, 50 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. Purification of the residue by flash silica gel column chromatography (5:1 toluene/ EtOAc) gave 31 (1.16 g, 1.596 mmol, 84% yield) and unreacted substrate 22 (58.2 mg, 174 µmol): colorless foam; Rf 0.74 (2:1 toluene/EtOAc); $[\alpha]_{D}^{26}$ +69.1° (*c* 0.97, CHCl₃); ¹H NMR (400 MHz, $CDCl_3$) δ 7.46–7.44 (2H, m), 7.34–7.24 (3H, m), 5.11 (1H, d, J = 4.4 Hz), 5.01 and 4.80 (2H, ABq, J = 11.6 Hz), 4.77 (1H, d, J = 4.0 Hz), 4.18-4.08 (3H, m), 3.90-3.83 (4H, m), 3.74-3.57 (5H, m), 3.45 (3H, s), 2.75 (1H, d, J = 8.8 Hz), 2.43 (1H, d, J = 8.4 Hz), 1.08 (9H, s), 1.07 (9H, s), 1.01 (9H, s), 1.00 (9H, s); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 139.1, 128.1, 127.7, 127.3, 101.1, 96.6, 83.0, 82.1, 77.7, 76.7, 74.8, 72.5, 71.2, 67.0, 66.6, 66.5, 66.1, 55.6, 27.4, 27.3, 22.6 \times 2, 19.9, 19.8; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₃₆H₆₂O₁₁NaSi₂ 749.3728, found 749.3735.

Compound 32. To a solution of **31** (704.9 mg, 0.970 mmol) in dry EtOAc (19.4 mL) was added 20% $Pd(OH)_2/C$ (704.9 mg) at room temperature, and the mixture was stirred under a H_2 atmosphere. After the reaction mixture was stirred for 30 min at the same temperature, the reaction was filtered through a Celite pad, and the filtrate was concentrated *in vacuo*. Purification of the residue by silica gel column chromatography (2:1 toluene/EtOAc) gave **32** (586.6 mg, 0.921 mmol, 95% yield): colorless foam; R_f 0.30 (2:1 toluene/EtOAc); $[\alpha]_D^{22} + 72.4^{\circ}$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 5.08 (1H, d, *J* = 4.0 Hz), 4.77 (1H, d, *J* = 4.0 Hz), 4.15–4.08 (3H, m), 3.91–3.81 (3H, m), 3.77–3.56 (6H, m), 3.45 (3H, s), 2.75 (1H, d, *J* = 10.0 Hz), 2.69 (1H, br-s), 2.47 (1H, d, *J* = 8.4 Hz), 1.07 (9H, s), 1.05 (9H, s), 1.00 (9H, s), 0.99 (9H, s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 100.9, 99.7, 83.4, 77.2, 76.8, 74.7, 72.6, 71.3, 66.8, 66.4 × 2, 66.2, 55.6, 27.4 × 2, 27.0, 26.9, 22.6 × 2, 19.9, 19.8; HRMS (ESI-TOF) *m*/*z* [M + H]⁺ calcd for C₂₉H₅₇O₁₁Si₂ 637.3439, found 637.3427.

Compound 33. To a solution of 10 (3.68 mg, 0.041 mmol) and 32 (130.8 mg, 0.205 mmol) in dry MeCN (616 μ L) was added a solution of 30 (241.9 mg, 0.616 mmol) in dry MeCN (616 μ L) at 60 °C under an Ar atmosphere. After the reaction mixture was stirred for 24 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 0.045 mmol, 820 μ L). The aqueous layer was extracted with EtOAc (50 mL \times 3), and then the combined extracts were washed with sat. NH₄Cl (aq, 50 mL) and sat. NaHCO₃ (aq, 50 mL), and brine, dried over anhydrous Na2SO4, and concentrated in vacuo. Purification of the residue by flash silica gel column chromatography (4:1 toluene/EtOAc) gave 33 (153.4 mg, 0.149 mmol, 73% yield) and unreacted substrate 32 (24.8 mg, 38.9 μ mol): colorless foam; R_d 0.71 (2:1 toluene/EtOAc); $[\alpha]_{D}^{24}$ +85.5° (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.46–7.26 (5H, m), 5.18 (1H, d, J = 2.4 Hz), 5.10 (1H, d, J = 4.0 Hz), 5.03 and 4.87 (2H, ABq, J = 12.0 Hz), 4.76 (1H, d, J = 3.6 Hz), 4.31-4.07 (5H, m), 3.91-3.56 (13H, m), 3.45(3H, s), 2.60 (1H, d, J = 8.4 Hz), 2.54 (1H, d, J = 9.6 Hz), 2.34 (1H, d, J = 8.8 Hz), 1.07–0.99 (54H, 6s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 139.3, 128.1, 127.7, 127.3, 101.2, 100.6, 99.7, 83.2, 82.3, 82.0, 77.9, 77.3, 74.8, 72.5, 71.3, 71.1, 66.7, 66.5, 66.4, 66.2, 55.7, 27.4 × 2, 27.0, 26.9, 22.6, 19.9, 19.8; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C43H83O16Si3 939.4989, found 939.4984.

Compound 34. To a solution of 33 (533.5 mg, 0.518 mmol) in dry EtOAc (5.18 mL) was added 20% Pd(OH)₂/C (533.5 mg) at room temperature, and the mixture was stirred under a H₂ atmosphere. After the reaction mixture was stirred for 30 min at the same temperature, the reaction was filtered through a Celite pad, and the filtrate was concentrated in vacuo. Purification of the residue by silica gel column chromatography (2:1 toluene/EtOAc) gave 34 (394.1 mg, 0.420 mmol, 81% yield): colorless foam; R_f 0.31 (2:1 toluene/EtOAc); $[\alpha]_D^{26}$ +101.6° (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, $CDCl_3$) δ 5.09 (1H, d, J = 3.6 Hz), 5.03 (1H, d, J = 4.4 Hz), 4.69 (1H, d, I = 4.0 Hz), 4.21-4.00 (5H, m), 3.83-3.48 (13H, m), 3.37(3H, s), 2.74 (1H, br-s), 2.59 (1H, d, J = 9.6 Hz), 2.50 (1H, d, J = 9.6 Hz), 2.35 (1H, d, J = 8.8 Hz), 1.00–0.91 (54H, 6s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 101.2, 100.4, 99.7, 83.2, 82.2, 77.2, 76.8, 74.7, 72.6, 71.3, 71.0, 66.5, 66.4, 66.3, 66.2 × 2, 55.6, 27.4 × 2, 27.3, 26.9 × 3, 22.6, 22.5, 19.9, 19.8; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C36H62O11NaSi2 749.3728, found 749.3735.

Compound 35. To a solution of 10 (0.38 mg, 4.28 μ mol) and 34 (20.1 mg, 21.4 μ mol) in dry MeCN (64 μ L) was added a solution of **30** (25.2 mg, 64.2 μ mol) in dry MeCN (64 μ L) at 50 °C under an Ar atmosphere. After the reaction mixture was stirred for 24 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 4.71 μ mol, 85.6 μ L). The aqueous layer was extracted with EtOAc (50 mL × 3), and then the combined extracts were washed with sat. NH₄Cl (aq, 50 mL) and sat. NaHCO₃ (aq, 50 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. Purification of the residue by flash silica gel column chromatography (4:1 *n*-hexane/ EtOAc) gave **35** (17.9 mg, 13.4 μ mol). 63% yield) and unreacted substrate **34** (7.3 mg, 7.77 μ mol): colorless foam; R_f 0.74 (2:1 toluene/EtOAc); $[\alpha]_{22}^{22}$ +102.8° (*c* 0.86, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.46–7.24 (5H, m), 5.19 (1H, d, *J* = 2.8 Hz), 5.17 (1H, d, *J* = 4.0 Hz), 5.09 (1H, d, *J* = 4.0 Hz), 5.03 and 4.87 (2H, ABq, *J* = 11.2 Hz), 4.76 (1H, d, *J* = 3.6 Hz), 4.33–4.27 (2H, m), 4.20–4.06 (5H, m), 3.90–3.56 (17H, m), 3.45 (3H, s), 2.64 (1H, d, *J* = 8.0 Hz),

2.53 (1H, d, *J* = 9.6 Hz), 2.43 (1H, d, *J* = 9.6 Hz), 2.33 (1H, d, *J* = 8.8 Hz), 1.07–0.96 (72H, 8s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 139.4, 128.1, 127.7, 127.3, 101.3, 100.7, 100.6, 99.8, 83.4, 82.4, 82.3, 82.0, 77.9, 77.2, 77.1, 76.8, 74.9, 72.6, 71.4, 71.2, 71.1, 66.8, 66.5, 66.4, 66.3, 66.2, 66.1, 55.7, 27.4 × 2, 27.1, 27.0, 26.9, 22.6 × 3, 20.0 × 2, 19.9, 19.8; HRMS (ESI-TOF) *m*/*z* [M + H]⁺ calcd for C₆₄H₁₁₅O₂₁Si₄ 1331.7008, found 1331.7008.

Compound 36. To a solution of **35** (46.0 mg, 34.5 μ mol) in dry EtOAc (1.73 mL) was added 20% $Pd(OH)_2/C$ (23.0 mg) at room temperature, and the mixture was stirred under a H₂ atmosphere. After the reaction mixture was stirred for 30 min at the same temperature, the reaction was filtered through a Celite pad, and the filtrate was concentrated in vacuo. Purification of the residue by silica gel column chromatography (2:1 toluene/EtOAc) gave 36 (35.6 mg, 28.7 μ mol, 83% yield): colorless foam; R_f 0.48 (2:1 toluene/EtOAc); $[\alpha]_{D}^{22}$ +113.0° (c 0.8, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 5.16 (2H, dd, I = 3.6 Hz, I = 4.0 Hz), 5.09 (1H, d, I = 3.6 Hz), 4.76 (1Hd, J = 3.6 Hz), 4.33-4.23 (2H, m), 4.20-4.06 (5H, m), 3.90-3.54 (17H, m), 3,45 (3H, s), 2.72 (1H, br-s), 2.66 (1H, d, J = 10.4 Hz), 2.53 (1H, d, J = 10.0 Hz), 2.46 (1H, d, J = 9.2 Hz), 2.35 (1H, d, J = 8.8 Hz), 1.05–0.98 (72H, 8s); $^{13}C{^1H}$ NMR (100 MHz, CDCl₃) δ 101.3, 100.8, 100.4, 99.7, 83.4, 82.4, 77.2, 76.8, 74.8, 72.7, 71.5, 71.2, 71.1, 66.5, 66.4, 66.3, 66.2, 66.1, 55.7, 27.4 × 2, 27.3, 27.0, 26.9 × 2, 22.6 × 2, 22.5, 20.0, 19.9, 19.8; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C57H109O21Si4 1241.6539, found 1241.6504.

Compound 37A and 37B. To a solution of **10** (0.25 mg, 2.74 μ mol) and **36** (17.0 mg, 13.7 μ mol) in dry MeCN (51 μ L) was added a solution of **14** (17.8 mg, 41.1 μ mol) in dry MeCN (51 μ L) at room temperature under an Ar atmosphere. After the reaction mixture was stirred for 24 h, the reaction was quenched by the addition of 50 mM NaBO₃ (aq, 3.01 μ mol, 54.8 μ L). The aqueous layer was extracted with EtOAc (50 mL × 3), and then the combined extracts were washed with sat. NH₄Cl (aq, 50 mL) and sat. NaHCO₃ (aq, 50 mL), and brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC (2:1 *n*-hexane/EtOAc) to afford **37A** (15.3 mg, 9.14 μ mol, 67% yield) and **37B** (3.1 mg, 1.85 μ mol, 14% yield).

Compound 37A: colorless foam; $R_f 0.70$ (2:1 toluene/EtOAc); $[\alpha]_{D}^{22}$ +105.4° (c 0.99, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.40– 7.11 (15H, m), 5.26 (1H, d, J = 4.0 Hz), 5.14 (1H, d, J = 2.8 Hz), 5.14 (1H, d, J = 4.0 Hz), 5.10 (1H, d, J = 4.0 Hz), 5.04 and 4.79 (2H, ABq, J = 11.2 Hz), 4.83 and 4.45 (2H, ABq, J = 10.8 Hz), 4.76 (1H, d, J = 4.0 Hz), 4.62 and 4.48 (2H, ABq, J = 12.4 Hz), 4.40 (1H, m), 4.31-4.05 (7H, m), 3.90-3.51 (22H, m), 3.45 (3H, s), 2.95 (1H, d, J = 9.6 Hz), 2.82 (1H, d, J = 8.4 Hz), 2.53 (1H, d, J = 9.6 Hz), 2.37 (1H, d, J = 9.6 Hz), 2.32 (1H, d, J = 8.4 Hz), 1.06-0.94 (72H, 8s); $^{13}C{^{1}H}$ NMR (100 MHz, CDCl₃) δ 139.1, 138.4, 138.0, 128.4, 128.3, 128.2, 128.0, 127.9, 127.8, 127.6, 127.5, 127.4, 101.3 \times 2, 100.9, 100.2, 99.8, 84.0, 83.9, 83.4, 82.5, 81.2, 77.2, 76.9, 76.8, 75.2, 74.9, 74.2, 73.3, 71.7, 71.2, 71.1, 70.7, 68.5, 66.5 × 2, 66.4, 66.3 × 2, 66.0, 65.9, 55.7, 27.4 × 2, 27.3, 27.0, 26.9, 22.6 × 3, 22.5, 20.0 × 2, 19.9; HRMS (ESI-TOF) m/z [M + H]⁺ calcd for C₈₄H₁₃₇O₂₆Si₄ 1673.8475, found 1673.8419.

Compound 37B: colorless foam; R_f 0.80 (2:1 toluene/EtOAc); $[\alpha]_{D}^{22}$ +102.5° (*c* 0.7, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.42–7.16 (15H, m), 5.76 (1H, d, *J* = 3.6 Hz), 5.24 (1H, d, *J* = 3.6 Hz), 5.11 (1H, d, *J* = 3.6 Hz), 5.08 (1H, d, *J* = 3.6 Hz), 5.04 and 4.53 (2H, ABq, *J* = 10.8 Hz), 4.83 and 4.79 (2H, ABq, *J* = 11.6 Hz), 4.76 (1H, d, *J* = 3.6 Hz), 4.65 and 4.53 (2H, ABq, *J* = 11.0 Hz), 4.37–4.28 (2H, m), 4.23–3.54 (28H, m), 3.45 (3H, s), 2.67 (1H, d, *J* = 9.2 Hz), 2.14 (1H, d, *J* = 9.6 Hz), 1.07–0.95 (72H, 8s); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 139.0, 138.5 138.2, 128.3, 127.9 × 2, 127.8, 127.5 × 2, 127.4, 101.3, 99.8, 99.7, 95.7, 94.5, 84.0, 83.6, 80.9, 78.5, 77.5, 77.2, 76.8, 76.0, 75.4, 74.9, 74.6, 73.2, 73.0, 72.4, 71.0, 70.9, 70.5, 68.6, 66.3, 65.9, 65.8, 64.9, 55.7, 27.5, 27.4 × 2, 27.2, 27.1, 27.0, 26.9, 23.0, 22.6, 22.4, 20.0 × 2, 19.9; HRMS (ESI-TOF) m/z [M + Na]⁺ calcd for C₈₄H₁₃₆O₂₆NaSi₄ 1695.8295, found 1695.8324.

Compound 38. To a solution of 37 (41.8 mg, 25.0 μ mol) in dry EtOAc (2.5 mL) was added 20% Pd(OH)₂/C (41.8 mg) at room

temperature, and the mixture was stirred under H₂ atmosphere. After the reaction mixture was stirred for 30 min at the same temperature, the reaction was filtered through a Celite pad, and the filtrate was concentrated in vacuo. To a solution of the residue in THF (1.25 mL) was added tetrabutyl-ammonium fluoride (43.4 μ L, 0.150 mmol) at 0 °C under an Ar atmosphere. After the reaction mixture was stirred for 22 h at room temperature, the reaction was quenched by $H_2O(1 \text{ mL})$ and EtOAc (1 mL). The aqueous layer was concentrated in vacuo. Purification of the residue by reverse phase column chromatography (H₂O) gave 38 (11.1 mg, 13.2 μ mol, 53% yield): colorless foam; R_{i} $0.23 (10:10:3 \text{ CHCl}_3/\text{MeOH}/\text{H}_2\text{O}); [\alpha]_D^{25} + 55.1^\circ (c \ 1.11, \text{H}_2\text{O}); {}^1\text{H}$ NMR (400 MHz, D₂O) δ 5.38-5.34 (4H, m), 4.83 (1H, br-s), 4.05-3.99 (4H, m), 3.92–3.63 (25H, m), 3.55 (1H, dd, J = 4.0, 10.0 Hz), 3.41 (3H, s); ¹³C{¹H} NMR(100 MHz, D₂O) δ 99.7, 99.6, 99.5, 99.4, 80.2, 80.1, 79.9, 73.2, 72.1, 72.0, 71.9, 71.8, 71.6, 70.7, 70.6, 70.3, 70.2, 70.1 × 2, 69.8, 60.8, 60.6 × 2, 60.4, 55.3; HRMS (ESI-TOF) m/ $z [M + Na]^+$ calcd for $C_{31}H_{54}O_{26}Na$ 865.2801, found 865.2817.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.0c02093.

¹H, ¹³C, and ¹H-¹H COSY NMR spectrum charts for all new compounds (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) (a) Carbohydrates in Chemistry and Biology; Ernst, B., Hart, G. W., Sinaÿ, P., Eds; Wiley-VCH: Weinheim, 2000; Vols. 1–4.
 (b) Glycoscience, Chemistry and Chemical Biology; Fraser-Reid, B. O., Tatsuta, K., Thiem, J., Eds.; Springer: Berlin, 2001.

(2) Handbook of Chemical Glycosylation: Advances in Stereoselectivity and Therapeutic Relevance; Demchenko, A. V., Ed; Wiley-VCH: Weinheim, 2008.

(3) (a) Lorand, J. P.; Edwards, J. O. Polyol Complexes and Structure of the Benzeneboronate Ion. J. Org. Chem. **1959**, 24, 769–774. (b) Springsteen, G.; Wang, B. A detailed examination of boronic acid-diol complexation. Tetrahedron **2002**, 58, 5291–5300. (c) Boronic Acids in Organic Synthesis and Chemical Biology; Hall, D. G., Ed.; Wiley-VCH: Weinheim, 2005.

(4) Oshima, K.; Aoyama, Y. Regiospecific Glycosidation of Unprotected Sugars via Arylboronic Activation. J. Am. Chem. Soc. **1999**, 121, 2315–2316.

(5) (a) Gouliaras, C.; Lee, D.; Chan, L.; Taylor, M. S. Regioselective Activation of Glycosyl Acceptors by a Diarylborinic Acid-Derived Catalyst. J. Am. Chem. Soc. 2011, 133, 13926–13929. (b) Beale, T. M.; Taylor, M. S. Synthesis of Cardiac Glycoside Analogs by Catalyst-Controlled, Regioselective Glycosylation of Digitoxin. Org. Lett. 2013, 15, 1358–1361.

(6) (a) Kaji, E.; Nishino, T.; Ishige, K.; Ohya, Y.; Shirai, Y. Regioselective glycosylation of fully unprotected methyl hexopyranosides by means of transient masking of hydroxy groups with arylboronic acids. *Tetrahedron Lett.* **2010**, *51*, 1570–1573. (b) Kaji, E.; Yamamoto, D.; Shirai, Y.; Ishige, K.; Arai, Y.; Shirahata, T.; Makino, K.; Nishino, T. Thermodynamically Controlled Regioselective Glycosylation of Fully Unprotected Sugars through Bis(boronate) Intermediates. *Eur. J. Org. Chem.* **2014**, *2014*, 3536–3539.

(7) (a) Nakagawa, A.; Tanaka, M.; Hanamura, S.; Takahashi, D.; Toshima, K. Regioselective and 1,2-cis- α -Stereoselective Glycosylation Utilizing Glycosyl-Acceptor-Derived Boronic Ester Catalyst. Angew. Chem., Int. Ed. 2015, 54, 10935-10939. (b) Nishi, N.; Nashida, J.; Kaji, E.; Takahashi, D.; Toshima, K. Regio- and stereoselective β mannosylation using a boronic acid catalyst and its application in the synthesis of a tetrasaccharide repeating unit of lipopolysaccharide derived from E. Coli O75. Chem. Commun. 2017, 53, 3018-3021. (c) Tanaka, M.; Nakagawa, A.; Nishi, N.; Iijima, K.; Sawa, R.; Takahashi, D.; Toshima, K. Boronic-Acid-Catalyzed Regioselective and 1,2-cis-Stereoselective Glycosylation of Unprotected Sugar Acceptors via S_Ni-Type Mechanism. J. Am. Chem. Soc. 2018, 140, 3644-3651. (d) Nishi, N.; Sueoka, K.; Iijima, K.; Sawa, R.; Takahashi, D.; Toshima, K. Stereospecific β -L-Rhamnopyranosylation through an S_Ni-Type Mechanism by Using Organoboron Reagents. Angew. Chem., Int. Ed. 2018, 57, 13858-13862. (e) Tanaka, M.; Sato, K.; Yoshida, R.; Nishi, N.; Oyamada, R.; Inaba, K.; Takahashi, D.; Toshima, K. Diastereoselective desymmetric 1,2-cis-glycosylation of meso-diols via chirality transferfrom a glycosyl donor. Nat. Commun. 2020, 11, 2431. (f) Inaba, K.; Endo, M.; Iibuchi, N.; Takahashi, D.; Toshima, K. Total Synthesis of Terpioside B. Chem. - Eur. J. 2020, 26, 10222-10225. (g) Tanaka, M.; Nashida, J.; Takahashi, D.; Toshima, K. Glycosyl-Acceptor-Derived Borinic Ester-Promoted Direct and β -Sterreoselective Mannosylation with a 1,2-Anhydromannose Donor. Org. Lett. 2016, 18, 2288-2291. (h) Tanaka, M.; Takahashi, D.; Toshima, K. 1,2-cis- α -Stereoselective Glycosylation Utilizing a Glycosyl-Acceptor-Derived Borinic Ester and Its Application to the Total Synthesis of Natural Glycosphingolipids. Org. Lett. 2016, 18, 5030-5033. (i) Nashida, J.; Nishi, N.; Takahashi, Y.; Igarashi, M.; Hayashi, C.; Takahashi, D.; Toshima, K. Systematic and Stereoselective Total Synthesis of Mannosylerythritol Lipids and Evaluation of Their Antibacterial Activity. J. Org. Chem. 2018, 83, 7281-7289. (8) Yan, L.; Meng, Y.; Haeffner, F.; Leon, R. M.; Crockett, M. P.; Morken, J. P. On the Carbohydrate/DBU Co-Catalyzed Alkene Diboration: Mechanistic Insight Provides Enhanced Catalytic

Efficiency and Substrate Scope. J. Am. Chem. Soc. 2018, 140, 3663–3773. (9) Padungros, P.; Alberch, L.; Wei, A. Glycal Assembly by the *in situ* Generation of Glycosyl Dithiocarbamates. Org. Lett. 2012, 14,

3380–3383.
(10) Meiland, M.; Heinze, T.; Guenther, W.; Liebert, T. Sevenmemberd ring boronates at *trans*-diol moieties of carbohydrates.

Tetrahedron Lett. 2009, 50, 469-472.

(11) Ohtawa, M.; Tomoda, H.; Nagamitsu, T. Regioselective Mono-Deprotection of Di-*tert*-butylsilylene Acetal Derived from 1,3-Diol with Ammonium Fluoride. *Bull. Chem. Soc. Jpn.* **2014**, *87*, 113–118. (12) (a) Hiruma, K.; Tamura, J.-i.; Horito, S.; Yoshimura, J.; Hashimoto, H. Convenient Synthesis of Pyruvate Acetals of Carbohydrates by Coupling of Trialkylsilylated Diols and Pyruvates. *Tetrahedron* **1994**, *50*, 12143–12158. (b) Chung, M.-K.; Orlova, G.; Goddard, J. G.; Schlaf, M.; Harris, R.; Beveridge; White, G.; Hallett, F. R. Regioselective Silylation of Sugars through Palladium Nanoparticle-Catalyzed Silane Alcoholysis. *J. Am. Chem. Soc.* **2002**, *124*, 10508–10518.

(13) Hanessian, S.; Kagotani, M. Novel methods for the preparation of partially acetylated carbohydrates. *Carbohydr. Res.* **1990**, 202, 67–79.

(14) Attouche, A.; Urban, D.; Beau, J.-M. A Tin-Free Regioselective Radical De-O-benzylation by an Intramolecular Hydrogen Atom Transfer on Carbohydrate Templates. *Angew. Chem., Int. Ed.* **2013**, *52*, 9572–9575.

(15) Johnston, I. R. The Partial Acid Hydrolysis of a Highly Dextrorotatory Fragment of the Cell Wall of *Aspergillus niger*. *Biochem. J.* **1965**, *96*, 659–664.

(16) (a) Komarova, B. S.; Orekhova, M. V.; Tsvetkov, Y. E.; Beau, R.; Aimanianda, V.; Latge, J. P.; Nifantiev, N. E. Synthesis of a Pentasaccharide and Neoglycoconjugated Related to Fungal α -(1 \rightarrow 3)-Glucan and Their Use in the Generation of Antibodies to Trace *Aspergillus fumigatus* Cell Wall. *Chem. - Eur. J.* **2015**, 21, 1029–1035. (b) Komarova, B. S.; Wong, S. S. W.; Orekhova, M. V.; Tsvetkov, Y. E.; Krylov, V. B.; Beauvais, A.; Bouchara, J.-P.; Kearney, J. F.; Aimanianda, V.; Latgé, J.-P.; Nifantiev, N. E. Chemical Synthesis and Application of Biotinylated Oligo- α -(1 \rightarrow 3)-D-Glucosides To Study the Antibody and Cytokine Response against the Cell Wall α -(1 \rightarrow 3)-D-Glucan of *Aspergillus fumigatus. J. Org. Chem.* **2018**, 83, 12965–12976.

(17) Wang, L.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Reagent Controlled Stereoselective Assembly of α -(1,3)-Glucans. *Eur. J. Org. Chem.* **2019**, 2019, 1994–2003.

(18) Hassan, H. H. A. M. Preparation of a set of selectively protected disaccharides for modular synthesis of heparan sulfate fragments: towards the synthesis of several *O*-sulfonated [β -D-GlcUA-(1 \rightarrow 4)- β -D-GlcNAc] OPr types. *Cent. Eur. J. Chem.* **2005**, *3*, 803–829.

(19) Bucher, C.; Gilmour, R. Fluorine-Directed Glycosylation. Angew. Chem., Int. Ed. 2010, 49, 8724-8728.

(20) Matwiejuk, M.; Thiem, J. Defining oxyanion reactivities in basepromoted glycosylations. *Chem. Commun.* **2011**, *47*, 8379–8381.

(21) Hollingsworth, R. I.; Hrabak, E. M.; Dazzo, F. B. Synthesis of 3,6-dideoxy-3-(methylamino)hexoses for G.L.C.-M.S. identification of *Rhizobium* lipopolysaccharide components. *Carbohydr. Res.* **1986**, 154, 103–113.

(22) Analytically pure compounds 16α and 17α were obtained after acetylation of the mixture, chromatographic purification on silica gel, followed by deprotection of Ac groups.