#### Letter

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# Stereoselective Synthesis of Coreoside D and Determination of Its Absolute Configuration

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**Abstract** We report the stereoselective synthesis of (35)- and (3*R*)-coreoside D. The conjugated diyne in the C1–C14 moiety was synthesized through two types of palladium-catalyzed cross-coupling reaction. The introduction of the glucopyranose was achieved by a glycosylation reaction using an imidate derivative in the presence of a Lewis acid. The asymmetric center at the C3-position was constructed by the chiralpool method using D-malic acid. The stereochemistry at the C3-position of the natural product was determined to be *R* by comparing the  $[\alpha]_D$  values of the synthetic stereoisomers with that reported for the natural product.

**Key words** asymmetric synthesis, coreoside D, polyacetylene glycoside, palladium catalysis, coupling reaction, glycosylation

Plains coreopsis (*Coreopsis tinctoria*) is an annual plant of the asteraceae family, and contains such chemical components as polyacetylene glycosides, flavonoids, and phenolic compounds.<sup>1</sup> In China, it is consumed as 'snow tea' or 'snow chrysanthemum' as a medicinal drink for treating hypertension and hyperlipidemia.<sup>1a</sup> In addition, it has been reported that a polyacetylene glycoside component of *C. tinctoria*, exhibits various physiological activities, such as antiallergic, antiinflammatory, and anti-HIV activities.<sup>1e,2</sup> Consequently, polyacetylene glycosides are attracting attention as pharmaceuticals and health foods. However, although there have been many reports on the biological activity of *C. tinctoria*, the absolute stereostructure of its bioactive compounds remained unclear.

In 2013, Zhang et al. isolated coreosides A–D (**1–4**, respectively) from the capitula of plains coreopsis (Figure 1).<sup>1a</sup> Coreosides E (**5**) and F (**6**) were subsequently isolated from the plains coreopsis by Guo et al. in 2017.<sup>1b</sup> These coreosides are polyacetylene glycosides that combine an aglycone containing a diyne skeleton with glucopyranose. The

coreosides have been reported to exhibit potent cyclooxygenase-2 (COX-2) inhibitory activity. Zhang et al. reported that only trace amounts of these compounds can be obtained from their natural sources, and that the stereochemistry of the C3-position was yet to be determined. Here, we report the first stereoselective syntheses of (3S)- and (3R)coreoside D in an attempt to determine the absolute conformations of natural coreosides.



Figure 1 The structures of coreosides A-F

Scheme 1 outlines our proposed synthesis of (3S)-coreoside D (**4a**). We surmised that (3S)-coreoside D (**4a**) might be obtainable by glycosylation by imidate **7** of the C1–C14 aglycone **8**, synthesized by a Sonogashira coupling reaction between iodoolefin **9** and acetylene **10**. Iodoolefin **9**, in turn, might be synthesized from D-malic acid by a Takai reaction to give the *trans*-iodoalkene moiety. On the other hand, diyne **10** might be prepared from propargyl alcohol by two types of palladium-catalyzed coupling reaction.

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Diethyl ester **12** was prepared from D-malic acid (**11**) by esterification with AcCl/EtOH (Scheme 2). Reduction of 12 with  $BH_3$ ·SMe<sub>2</sub>/NaBH<sub>4</sub> gave diol **13** as the major product in a 93:7 mixture, as observed by <sup>1</sup>H NMR spectroscopy; the minor product was formed by reduction of the other ester moiety (the structure of the minor product is not shown).<sup>3</sup> After protection of alcohol 13 using TBDPSCl, reduction of the ester group afforded 14.<sup>4</sup> Diol 14 was protected by treatment with cyclohexanone in the presence of PPTS to give acetal 15. At this stage, the desired acetal 15 was cleanly isolated as the sole isomer by silica gel column chromatography. Subsequently, deprotection of acetal 15 with TBAF afforded alcohol 16 in a good yield. Parikh-Doering oxidation of 16 followed by a Horner-Wadsworth-Emmons reaction gave the  $\alpha$ . $\beta$ -unsaturated ester **17** in 90% yield. Ester 17 was converted into alcohol 18 upon reduction of the C=C double bond and ester moiety. Parikh-Doering oxidation of **18** gave aldehvde **19**, which was subjected to a Takai reaction in 1,4-dioxane-THF by a modified version of the reported procedure.<sup>5</sup> The resulting iodoolefin was treated with TsOH·H<sub>2</sub>O to afford diol **20** (E/Z = 91:9). The E- and Zolefins could not be separated by purification, and were used as a mixture in the next reaction. Finally, the hydroxy group in 20 was protected by using TBDPSCI to give iodoalkene 9 in 87% yield.

The results of the two types of palladium-catalyzed coupling reaction used to prepare diyne **10** are summarized in Scheme 3. Propargyl alcohol (**21**) was converted into iodoolefin **22** in 82% yield.<sup>6</sup> Sonogashira coupling of iodoolefin **22** with (trimethylsilyl)acetylene afforded enyne **23**, which was then subjected to a deprotection step to remove the trimethylsilyl group to produce **24** in good yield. A palladium-catalyzed C(sp)–C(sp) cross-coupling reaction was



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then carried out to afford diyne **25**.<sup>7</sup> Finally, the trimethylsilyl group in **25** was removed by using  $K_2CO_3$ /MeOH to give the desired diyne **10**.

In the last stage, we completed the synthesis and characterization of (3S)-coreoside D (4a), as shown in Scheme 4. Construction of the C1–C14 moiety of (3S)-coreoside D (4a) was achieved by Sonogashira coupling of 9(E/Z = 91:9) and 10 to give alcohol 8.8 In this coupling reaction, up to 5% of the the E-olefin at the C12-C13 position isomerized to the Z-isomer upon conducting the reaction for more than two hours. In addition, (Z)-9 did not react in the coupling reaction. Subsequently, the glycosylation reaction with imidate 7<sup>9</sup> in the presence of TMSOTf and 4Å MS proceeded smoothly to afford acetal 26.<sup>10</sup> Finally, deprotection and hydrolvsis of **26** afforded (3S)-coreoside D (**4a**) in 71% vield from 8. The <sup>1</sup>H NMR, <sup>13</sup>C NMR, and UV spectral data for the synthetic (3S)-coreoside D (4a) were in good agreement with those reported for the natural product. However, the optical rotation of the synthetic compound was inconsistent with the literature data:  $[\alpha]_D^{21}$  – 31 (*c* 0.06, MeOH); Lit.<sup>1</sup>  $[\alpha]_{D}^{21}$  –13 (c 0.04, MeOH). This result suggested that the stereochemistry at the C3-position was incorrect, and prompted us to synthesize the corresponding (3R)-isomer 4b.



The synthesis of the (3R)-isomer (4b) is summarized in Scheme 5. Iodoolefin **9** was converted into *ent*-**9** in 68% yield by a Mitsunobu reaction. Thereafter, (3R)-coreoside D (4b) was synthesized in the same manner as shown in Scheme 4. The <sup>1</sup>H NMR, <sup>13</sup>C NMR, and UV spectral data obtained for (3*R*)-coreoside D (**4b**) were in good agreement with those reported for the natural product. The optical rotation was also in agreement with the literature data:  $[\alpha]_D^{22}$  –13 (*c* 0.05, MeOH); Lit.<sup>1</sup>  $[\alpha]_D^{22}$  –13 (*c* 0.04, MeOH).



**Scheme 5** Synthesis of (3*R*)-coreoside D (4b)

In conclusion, we have accomplished stereoselective syntheses of (3S)-coreoside D (4a) and its (3R)-isomer 4b. The key intermediate, iodoolefin 9 was synthesized by Horner-Wadsworth-Emmons and Takai reactions from Dmalic acid (11) as a starting material. On the other hand, divne **10** was derived from propargyl alcohol (**21**) by two types of palladium-catalyzed coupling reaction. The C1-C14 carbon chain was constructed by a Sonogashira coupling reaction of iodoolefin 9 and divne 10, after which the glucose moiety was introduced to synthesize (3S)-coreoside (4a) over 18 steps in a total yield of 7.6% from D-malic acid (11). The chiral carbon in alcohol 9 was inverted through a Mitsunobu reaction to afford ent-9, which was transformed into (3R)-coreoside D (4b) in the same manner. A comparison of the  $[\alpha]_{D}$  values of **4a** and **4b** with that reported for natural coreoside D was decisive in determining the R configuration of the C3 stereocenter in coreoside D and thus, the reported chirality was revised.

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## **Supporting Information**

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#### (11) (3S)-Coreoside (4a)

A 1.02 M solution of NaOMe in MeOH (0.25 mL, 0.255 mmol) was added to a solution of diol (3*S*)-**27** (0.126 mmol) in 1:1 MeOH–THF (4 mL), and the mixture was stirred at r.t. for 1 h, then neutralized with Amberlite IR-120 (hydrogen form). The resulting mixture was filtered through paper and concentrated. The residue was purified by chromatography [silica gel, CH<sub>2</sub>Cl<sub>2</sub>–MeOH (4:1)] to give a white solid; yield: 35.6 mg [71% from alcohol (*S*)-**8**]; mp 40 °C;  $R_f = 0.25$  (CH<sub>2</sub>Cl<sub>2</sub>–MeOH, 4:1);  $[\alpha]^{21}_{D} - 31$  (*c* 0.06, MeOH).

<sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD): δ = 1.56–1.83 (m, 4 H), 2.30 (q, *J* = 7.1 Hz, 2 H), 3.16 (t, *J* = 8.3 Hz, 1 H), 3.21–3.40 (m, 3 H), 3.62–3.72 (m, 2 H), 3.72–3.81 (m, 1 H), 3.81–3.92 (m, 2 H), 4.14 (dd, *J* = 4.7, 2.0 Hz, 2 H), 4.34 (d, *J* = 8.3 Hz, 1 H), 5.66 (d, *J* = 16.0 Hz, 1 H), 5.83 (d, *J* = 15.6 Hz, 1 H), 6.35 (dt, *J* = 15.6, 7.1 Hz, 1 H), 6.37 (dt, *J* = 16.0, 4.7 Hz, 1 H). <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD): δ = 30.1, 35.4, 38.2, 59.5, 62.68, 62.75, 71.6, 73.1, 74.9, 75.3, 77.8, 77.9, 78.2, 79.7, 81.2, 103.9, 109.0, 109.7, 147.4, 150.0. HRMS (FD): *m/z* [M]<sup>+</sup> calcd for C<sub>20</sub>H<sub>28</sub>O<sub>8</sub>: 396.17842; found: 396.17676. UV (MeOH):  $\lambda_{max}$  262, 276, 293, 313 nm.