



A common and stereoselective strategy for the synthesis of *N,O,O,O*-tetra-acetyl *D*-ribo-(2*S*,3*S*,4*R*)-phytosphingosine and 2-*epi*-jaspine B

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ABSTRACT

A common and stereoselective strategy for the synthesis of *N,O,O,O*-tetra-acetyl *D*-ribo-(2*S*,3*S*,4*R*)-phytosphingosine and 2-*epi*-jaspine B was achieved by using Grignard addition on *N*-benzyl sugar lactamine and Wittig olefination as key steps.

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Sphingoids are long-chain amino-diol and -triol bases that form the backbone and the characteristic structural unit of sphingolipids, which are important membrane constituents and play a vital role in cell regulation as well as signal transduction.¹ Furthermore, glycosphingolipids show important biological activities, such as antitumor,^{2a} antiviral,^{2b} antifungal,^{2c} and cytotoxic properties.^{2d} Phytosphingosines, one of the major classes of sphingoids, have been isolated and identified either separately or as parts of sphingolipids from plants,^{3a} marine organisms,^{3b,c} fungi,^{3d} yeasts^{3e} and even mammalian tissues^{3f-k} of kidney,^{3g} liver,^{3h} uterus,³ⁱ intestine,^{3j} and skin.^{3k} In addition to being base components of sphingolipids in membranes, phytosphingosines themselves are found to be bioactive lipids. For example, *ribo*-phytosphingosine **1** is a potential heat stress signal in yeast cells.⁴ It has also been established that various diastereomers of sphingosines exhibit different activities and metabolism.⁵ Some of its derivatives exhibit important physiological activities.⁶ For instance KRN 7000 (Fig. 1) which is a glucosphingolipid exhibits natural killer activity and strongly inhibits tumor metastasis in mice.^{6b} Since it is costly to isolate these lipids from the natural sources, and also not possible to obtain homogeneous material for biological studies, a great work has been carried out for the synthesis of these compounds. Among all the eight possible stereoisomers of phytosphingosine (2*S*,3*S*,4*R*)-2-aminooctadecane-1,3,4-triol **1** with *D*-ribo-stereochemistry is the most common one. Owing to their interesting biological activities many synthetic approaches to enantiomerically pure *D*-ribo-phytosphingosine **1** have been reported in the literature.⁷

Jaspine B, also known as pachastrissamine (**2a**, Fig. 1), is a cyclic anhydro-phytosphingosine derivative, which was initially isolated from a marine sponge *Pachastrissa* sp. (family Calthropellidae) in 2002 by Higa and co-workers,⁸ and found to have cytotoxicity at a level of IC₅₀ 0.01 µg/mL against P388, A549, HT29, and MEL28

cell lines. Later, Debitus and co-workers⁹ isolated the same compound from a Vanuatuan marine sponge genus *Jaspis* and named it as jaspine B **2a**. This compound displayed remarkable bioactivity (IC₅₀ 0.24 µM) against the A549 human lung carcinoma cell line using the ATPlite assay and represented as the most potent anti-cancer agent on this cell line. Delgado and co-workers reported that the potency of cytotoxicity is dependent on the stereochemistry of the tetrahydrofuran moiety.¹⁰

The novel structural features and interesting biological activity of pachastrissamine have prompted chemists to develop several syntheses for jaspine B **2a** and its isomers.¹¹ Our group published the first total synthesis of jaspine B **2a** and its C2 epimer **2b**.^{12a} We also published the synthesis of the C2 and C3 epimer **2c** and an enantiomer of jaspine B **2d** and their biological activity in comparison to jaspine B **2a**^{12b} and 3-*epi* jaspine **2e**.^{12c}

In continuation of our efforts in this area, we envisaged a concise, efficient, and common method for the synthesis of *N,O,O,O*-tetra-acetyl *ribo*-(2*S*,3*S*,4*R*)-phytosphingosine **3** and 2-*epi*-jaspine B **2b**. The starting material for our approach is *D*-ribose, which is an inexpensive sugar and possesses desired chirality. The retrosynthetic analysis of *N,O,O,O*-tetra-acetyl *D*-ribo-(2*S*,3*S*,4*R*)-phytosphingosine **3** and 2-*epi*-jaspine **2b** is depicted in Scheme 1. Compound **4** is a common intermediate for the synthesis of **3** and **2b** and it can be obtained from compound **5** after appropriate manipulations. The amino group can be introduced by nucleophilic addition on ribosylamine **6** and the required lipid chain can be introduced by Wittig olefination.

Earlier we explored stereoselective Grignard addition on chiral imines for the synthesis of polyhydroxylated pyrrolidines¹³ and carbasugars.¹⁴ In this report we expanded the utility of this sugar imine Grignard addition, for the synthesis of *N,O,O,O*-tetra-acetyl *ribo*-phytosphingosine **3** and 2-*epi*-jaspine **2b**.

The starting material 5-*O*-*tert*-butyl dimethylsilyl-2,3-*O*-isopropylidene-*D*-ribofuranose **7**, required for our synthesis was prepared from *D*-ribose using our earlier procedure (Scheme 2).¹⁵ Reaction on **7** with benzylamine gave ribosylamine **6**. Treatment of the crude ribosylamine **6** with vinylmagnesium bromide at –78 °C

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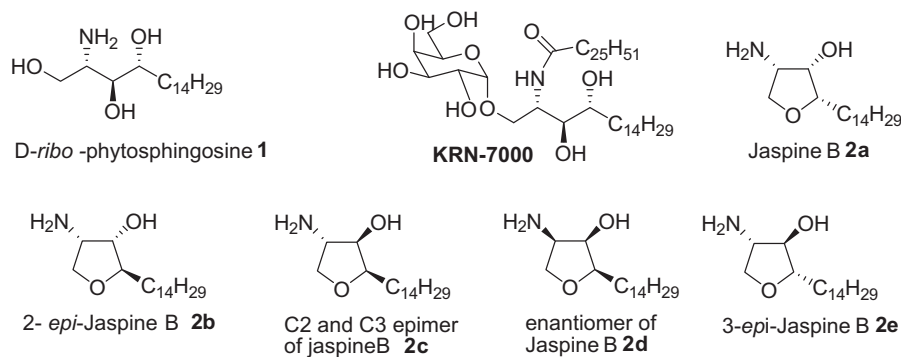
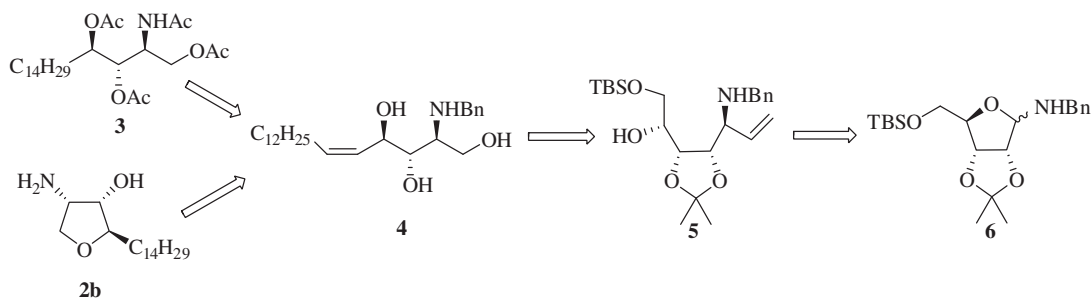
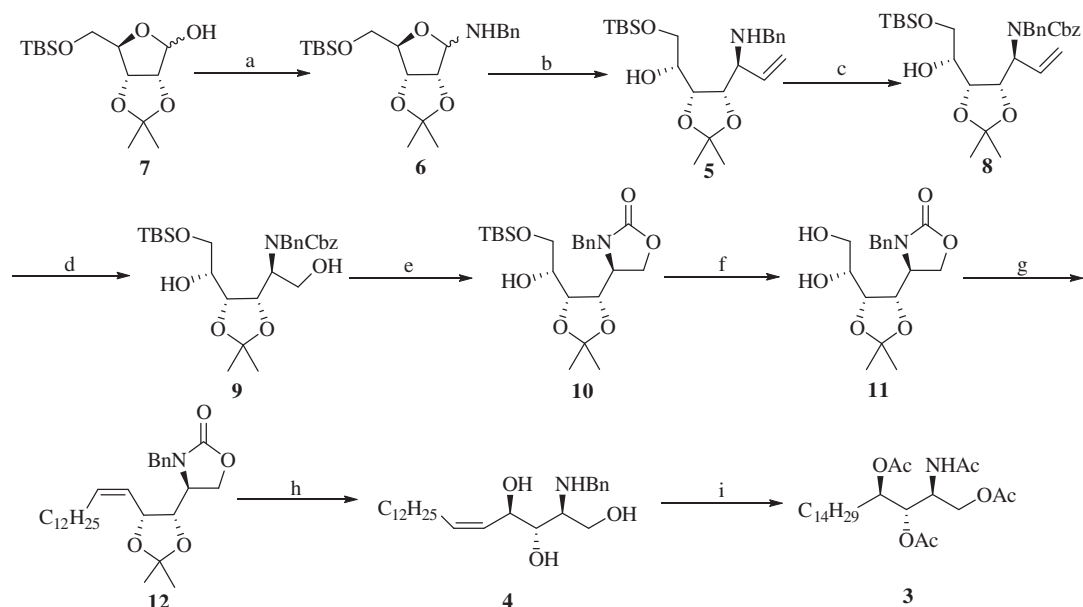


Figure 1. Structures of D-ribo-(2S,3S,4R)-phytosphingosine, KRN 7000, jaspine B, and its isomers.



Scheme 1. Retrosynthetic analysis of N,O,O,O-tetra-acetyl D-ribo-phytosphingosine **3** and 2-epi-jaspine B **2b**.



Scheme 2. Synthesis of N,O,O,O-tetra-acetyl D-ribo-phytosphingosine **3**. Reagents and conditions: (a) BnNH₂, MeOH, reflux; (b) vinylmagnesium bromide, THF, −78 °C, 2 h, 72%; (c) Cbz-Cl, NaHCO₃, MeOH, 1 h, 98%; (d) (i) O₃, CH₂Cl₂, −78 °C, 30 min and DMS, (ii) NaBH₄, MeOH, 1 h (85% for two steps); (e) NaH, THF, 30 min, 92%; (f) TBAF, THF, 2 h, 96%; (g) (i) NaIO₄, CH₂Cl₂, H₂O, rt, 4 h, (ii) C₁₃H₂₇Ph₃P⁺Br[−], *n*-BuLi, THF, 1 h (90% for two steps); (h) 6 N HCl, EtOH, reflux, 6 h, 94%; (i) (i) H₂, Pd/C, MeOH, 6 h, (ii) (Ac)₂O, N(Et)₃, CH₂Cl₂, 6 h (85% for two steps).

gave exclusively **5** as a single isomer. The absolute configuration of the newly generated stereo center was not confirmed at this stage, but as per our earlier observation it should give the *erythro*-isomer.¹⁴ The formation of *erythro*-isomer can be explained via seven membered transition state **A**¹⁶ or Felkin-Anh model **B** (Fig. 2). In fact the formation of *erythro*-isomer shows that chelation between sugarimine and isopropylidene group has not taken place due to steric strain.

The amino functionality in **5** was converted into carbamate with CbzCl to give **8**. Oxidative degradation of the alkene functionality in **8** with ozone afforded the aldehyde, which on treatment with NaBH₄ in MeOH gave alcohol **9**. In order to protect the primary hydroxyl group, compound **9** was treated with sodium hydride to give the cyclic carbamate **10**. Deprotection of silylether in **10** with TBAF gave diol **11**. The diol functionality of compound **11** was oxidatively cleaved to aldehyde using NaIO₄ in CH₂Cl₂/

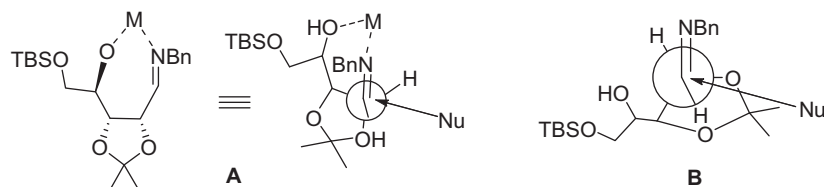
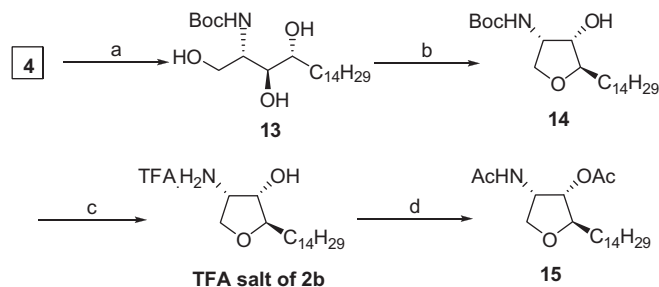


Figure 2. Seven-membered transition state (A) and Felkin Anh Model (B).



Scheme 3. Synthesis of TFA salt of 2-epi-jaspine B **2b**. Reagents and conditions: (a) (i) H₂, Pd/C, MeOH, 6 h, (ii) (Boc)₂O, Et₃N, CH₂Cl₂, 2 h (95% over two steps); (b) (i) TsCl, Et₃N, DMAP, CH₂Cl₂, 86%; (c) TFA, CH₂Cl₂, 2 h; 89%; (d) (Ac)₂O, Et₃N, CH₂Cl₂, 4 h, 94%.

H₂O, which was carried to the next step without any purification. Wittig reaction on crude aldehyde with C₁₃H₂₇PPh⁺Br[−] gave **12**. Global deprotection of carbamate and acetone in **12** with 6 N HCl in EtOH under reflux conditions gave **4**. Hydrogenation of **4** gave the D-ribo-phytosphingosine **1**. For the sake of characterization, it was converted as such into N,O,O,O-tetra-acetyl D-ribo-phytosphingosine **3**,¹⁷ by treating with acetic anhydride and Et₃N, whose physical properties are in good agreement with the reported values.¹⁸

To convert the compound **4** into 2-epi-jaspine B **2b** the following reactions were carried out (Scheme 3). Hydrogenation of compound **4** using Pd/C in MeOH followed by treatment of the resultant amino functionality with (Boc)₂O afforded carbamate **13**. Regioselective tosylation of the primary hydroxy group of **13** prompted spontaneous cyclization to give the tetrahydrofuran derivative **14**.^{11b} Deprotection of the Boc group in **14** with TFA/CH₂Cl₂ provided the desired 2-epi-jaspine B **2b** as TFA salt^{19a} and TFA salt of **2b** on acetylation with acetic anhydride in the presence of excess Et₃N gave the acetyl derivative **15**.^{19b} The physical properties of the TFA salt of 2-epi-jaspine B **2b**^{13a} and its diacetate derivative **15**^{12b} were in good agreement with the reported values.

In conclusion we have successfully demonstrated a general strategy for the synthesis of N,O,O,O-tetra-acetyl D-ribo-phytosphingosine **3** and 2-epi-jaspine B **2b** and its diacetate derivative **15** by using stereoselective vinyl Grignard on ribosylamine. Application of this strategy to make some more analogs of phytosphingosine and jaspine B are in progress.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2011.07.032.

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 17. Spectral data of compound **3**: $[\alpha]_D^{26} + 23.1$ (c 0.62, CHCl₃); [lit.¹⁸ $[\alpha]_D^{20} + 21.9$ (c 1.1, CHCl₃)]; IR $\nu_{\max}$: 2924, 2854, 1741, 1659, 1543, 1370, 1221, 1044 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 6.00 (d, 1H, *J* = 9.4 Hz), 5.04 (dd, 1H, *J* = 3.0 and 8.3 Hz), 4.83–4.91 (m, 1H), 4.40 (m, 1H), 4.22 (dd, 1H, *J* = 4.7 and 11.5 Hz), 3.94 (dd, 1H, *J* = 3.0 and 11.5 Hz), 2.01 (s, 3H), 1.99 (s, 6H), 1.96 (s, 3H), 1.76 (m, 1H), 1.51 (m, 1H), 1.05–1.31 (m, 24H), 0.80 (t, 3H, *J* = 6.6 Hz); ¹³C NMR (CDCl₃, 75 MHz): δ 171.1, 170.8, 170.0, 169.7, 72.9, 71.9, 62.8, 47.6, 31.9, 29.6, 29.5, 29.4, 29.3, 28.1, 25.5, 23.3, 22.6, 21.0, 20.7, 14.1; ESIMS *m/z*: 486 [M+H]⁺; HRMS (ESI): calcd for C₂₆H₄₈NO₇ [M+H]⁺ 486.3430, found 486.3441.
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 19. (a) Spectral data of compound of **2a** as TFA salt: $[\alpha]_D^{26} + 16.2$ (c 1.1, EtOH), [lit.^{12a} $[\alpha]_D^{25} + 14.46$ (c 1.5, EtOH)]; IR $\nu_{\max}$: 2920, 2851, 1669, 1209, 1147 cm⁻¹; ¹H NMR (CD₃OD, 300 MHz): δ 4.14 (m, 1H), 4.03 (t, 3H, *J* = 5.7 Hz), 3.65–3.80 (m, 3H), 1.17–1.70 (m, 26H), 0.89 (t, 3H, *J* = 6.8 Hz); ¹³C NMR (CD₃OD, 75 MHz): δ 85.3, 74.4, 69.4, 53.7, 47.9, 34.1, 33.1, 30.8, 30.7, 30.6, 30.5, 26.9, 23.7, 14.4; ESIMS *m/z*: 300 (M⁺+1–CF₃COOH); HRMS (ESI): calcd for C₁₈H₃₈NO₂ [M+H]⁺ 300.2902, found 300.2902.
 - (b) Spectral data of compound **15**: $[\alpha]_D^{26} - 15.1$ (c 1.2, CHCl₃), [lit.^{11b} $[\alpha]_D^{22} - 15.4$ (c 1.0, CHCl₃)]; IR $\nu_{\max}$: 3298, 2920, 2851, 1740, 1658, 1557, 1376, 1232, 1072 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 5.68 (br d, 1H, *J* = 7.7 Hz), 4.66 (m, 1H), 4.18 (t, 1H, *J* = 7.7 Hz), 3.84–3.90 (m, 1H), 3.52 (t, 1H, *J* = 8.5 Hz), 2.14 (s, 3H), 2.02 (s, 3H), 1.49–1.68 (m, 2H), 1.15–1.47 (m, 26H), 0.88 ((t, 1H, *J* = 6.6 Hz); ¹³C NMR (CDCl₃, 75 MHz): δ 170.1, 169.9, 84.0, 76.7, 69.7, 49.9, 33.5, 31.9, 29.6, 29.5, 29.5, 29.4, 29.3, 25.5, 23.2, 22.7, 21.0, 14.1; ESIMS *m/z*: 406 (M+Na)⁺; HRMS (ESI): calcd for C₂₂H₄₁NO₄Na [M+Na]⁺ 406.2933, found 406.2933.