

## Asymmetric Synthesis of 1,3-Dithiolane Nucleoside Analogues

Romualdo Caputo,<sup>[a]</sup> Annalisa Guaragna,<sup>\*[a]</sup> Giovanni Palumbo,<sup>[a]</sup> and Silvana Pedatella<sup>[a]</sup>*Dedicated to Prof. L. Mangoni on his 70th birthday***Keywords:** Nucleosides / Sulfur heterocycles / Chirality / Sulfoxides / Antiviral agents

We report the ready asymmetric synthesis of nucleoside analogues containing a 1,3-dithiolane ring that mimics the sugar moiety of natural nucleosides. The synthesis is accomplished in three main steps from benzoyloxyethanal 1,3-dithiolane, the key step being its conversion into a chiral monosulfoxide

by a modified Sharpless sulfo-oxygenation reaction. The nucleoside analogues were obtained in good overall yields and with excellent enantiomeric excesses.

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2003)

**Introduction**

The discovery of modified nucleosides that are effective in the treatment of Acquired Immunodeficiency Syndrome<sup>[1]</sup> (AIDS) has aroused a great interest in the synthesis of such molecules during the last decade. Only a few nucleoside analogues, which act as reverse transcriptase inhibitors<sup>[2]</sup> (NRTI), have been approved so far for the treatment of HIV-infected individuals, including AZT<sup>[3]</sup> (Zidovudine), ddC<sup>[4]</sup> (Zalcitabine), ddI<sup>[5]</sup> (Didanosine), d4T<sup>[6]</sup> (Stavudine), 3TC<sup>[7]</sup> (Lamivudine), and the carbocyclic analogue ABC<sup>[8]</sup> (Abacavir).

In general, nucleoside analogues having one or more heteroatoms replacing the ring atoms of the sugar moiety turn out to be more potent,<sup>[9]</sup> possess improved selectivity toward mutant viruses, and show enhanced cellular uptake. The sugar configuration also seems to affect the biological activity of the NRTIs;<sup>[10]</sup> in fact,  $\beta$ -L(-)-2'-deoxy-3'-thiacytidine (lamivudine), which is the sole L-nucleoside analogue in use, is much less cytotoxic<sup>[11]</sup> than its D-enantiomer (BCH-189), although both of them are roughly equipotent against the replication of HIV-1. All these considerations have spurred chemical research towards the synthesis of new L-sugar-based nucleoside analogues as potential reverse transcriptase inhibitors.

As a consequence, several methods are available in the recent literature for the synthesis of various nucleoside analogues,<sup>[12]</sup> although the real challenge is the difficulty in preparing such compounds in optically pure forms.

In this paper, we report a new asymmetric synthesis of 1,3-dithiolane nucleoside analogues, a class of thioribonucleosides having a second sulfur atom replacing the 3' ribose carbon atom. Synthetic paths leading to such compounds are not very common and, to the best of our knowledge, so far only two synthetic routes have been reported,<sup>[13,14]</sup> although both of them lead to racemic products that need resolution by HPLC on chiral columns to get enantiomerically pure compounds.

**Results and Discussion**

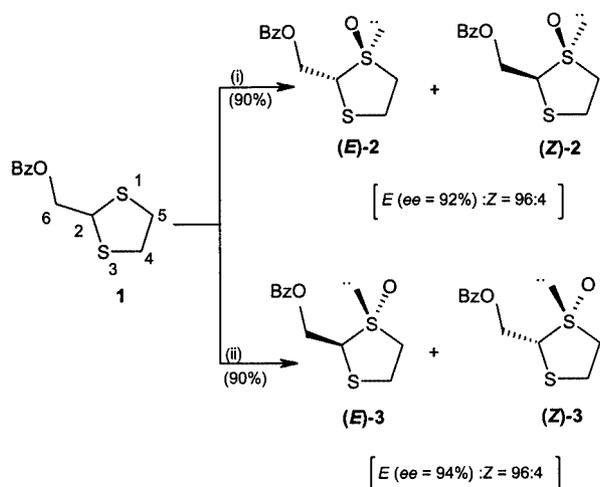
As a part of our current interest in the synthesis of modified sugars<sup>[15]</sup> for their incorporation into nucleoside analogues, we have devised a new general approach to the asymmetric synthesis of chiral 1,3-dithiolanyl nucleosides (2,4-disubstituted 1,3-dithiolanes) with excellent enantiomeric excesses, in three main steps starting from benzoyloxyethanal 1,3-dithiolane.<sup>[16]</sup>

The key step of our approach is the preparation of the chiral sulfoxides **2** (Scheme 1). 2-[(Phenylcarbonyloxy)methyl]-1,3-dithiolane (**1**) was obtained in very high yield (98%) by treatment of benzoyloxyethanal<sup>[17]</sup> (from monobenzoylglycerol) with ethanedithiol in the presence of polystyryl diphenylphosphane-iodine complex, which acts as both a Lewis acid and a dehydrating agent, according to a procedure formerly developed in our laboratory<sup>[18]</sup>. The conversion of achiral **1** into the chiral sulfoxides **2** was then performed by a Di Furia-Modena oxidation<sup>[19]</sup> with *tert*-butyl hydroperoxide and diethyl D-tartrate in the presence of Ti<sup>IV</sup> isopropoxide as catalyst.

In order to avoid the double oxidation of **1** at both the sulfur atoms (and overoxidation to the sulfone as well) we used a 5:1 molar ratio of **1** and the oxidizing system (*t*BuOOH/D-DET/Ti<sup>IV</sup>). Under such conditions, the oxida-

<sup>[a]</sup> Dipartimento di Chimica Organica e Biochimica, Università di Napoli Federico II  
Via Cynthia, 4, 80126 Napoli, Italy  
E-mail: guaragna@unina.it

<sup>[b]</sup> Consorzio Interuniversitario Nazionale, La Chimica per l'Ambiente  
Via della Libertà, 5/12, 30175 Marghera (VE), Italy



Scheme 1. Conversion of 1,3-dithiolane **1** into chiral sulfoxides **2** and **3**; i. D-DET/Ti(OiPr)<sub>4</sub>/tBuOOH in CH<sub>2</sub>Cl<sub>2</sub>, -20 °C, 16 h; ii. same as i., but with L-DET

tion of **1** led in high yield (90%) to a mixture of the diastereoisomeric *E* and *Z* pairs (**2**) in a 96:4 ratio (Scheme 1). After chromatographic separation, the more-abundant *E* pair exhibited an *ee* of 92%, as determined by <sup>1</sup>H NMR spectroscopic analysis using Eu(hfc)<sub>3</sub> (Aldrich) as the chiral shift reagent.

The replacement of diethyl D-tartrate with its L-enantiomer led also to a diastereoisomeric (*E/Z* = 96:4) mixture (**3**) in which the more-abundant *E* pair exhibited an *ee* of 94% (by <sup>1</sup>H NMR spectroscopy). The (2*R*,*S**R*) stereochemistry assigned to the sulfoxide (*E*)-**3** was supported by X-ray crystallographic analysis (85% confidence). The final step of the synthesis consisted of the coupling of the more-abundant chiral sulfoxides (*E*)-**2** and (*E*)-**3** with both *N*<sup>4</sup>-acetylcytosine and 6-Cl-purine under modified Pummerer rearrangement conditions;<sup>[20]</sup> i.e., by direct treatment of the heterocyclic base silylated in situ with trimethylsilyl triflate in the presence of triethylamine. The results obtained are displayed in Figure 1. The *cis/trans* configurations were assigned by NOE difference experiments (see Exp. Sect.).

Treatment of the protected dithiolanyl nucleosides with sodium methoxide in methanol afforded the corresponding free cytosinyl nucleosides **5a**, **5b**, **9a**, and **9b**, and purinyl nucleosides **7a** and **7b** in high yields.

This procedure for the synthesis of thiosugar-based nucleoside analogues, via chiral sulfoxides, is an appealing one for the preparation of such compounds belonging to both D and L series, with excellent enantiomeric excesses, merely by judicious choice of a suitable ligand to use as the chirality inductor in the asymmetric sulfo-oxygenation step.

While this work was in progress, we encountered a paper<sup>[14]</sup> that describes the synthesis and separation on a chiral HPLC column of two enantiomeric compounds matching our products **5b** and **9b**. In our opinion, this report is rather questionable and should be reconsidered since the physical data of those products are not those expected for enantiomers (e.g., their <sup>1</sup>H NMR spectra are quite dif-

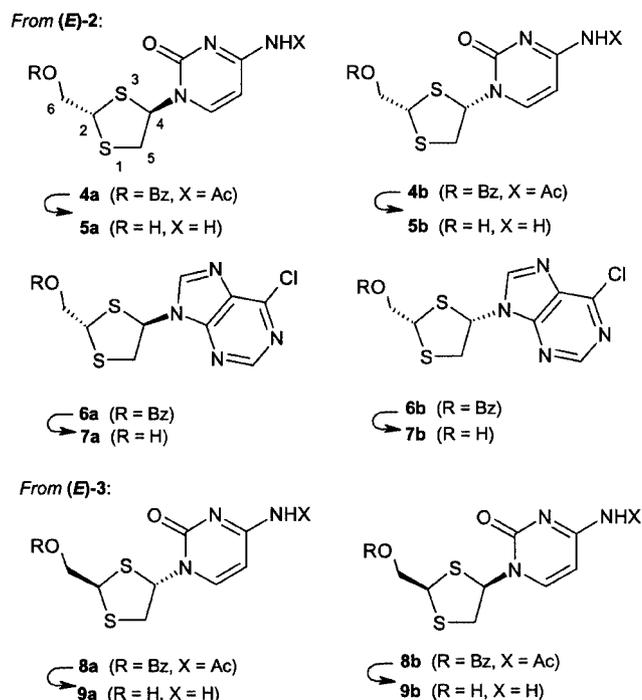


Figure 1. 1,3-Dithiolane nucleoside analogues from chiral sulfoxides **2** and **3**

ferent, and their melting points differ by more than 90 °C) nor do they match our data for the same compounds.

Our new approach to the asymmetric synthesis of hetero-substituted nucleoside analogues may represent a rapid entry to optically pure materials and potentially is amenable to the preparation of multi-gram quantities of these important compounds.

## Experimental Section

**General:** <sup>1</sup>H and <sup>13</sup>C NMR spectra: Varian Inova 500, Bruker DRX-400, Varian Gemini 200 spectrometers, CDCl<sub>3</sub> unless otherwise specified, TMS internal standard. Optical rotations: Jasco P-1010 (1.0 dm cell). Combustion analyses: Perkin–Elmer Series II 2400, CHNS analyzer. HPLC analyses: Gynkotek M480 chromatograph equipped with UVD 160S detector. Chiral columns: Cyclodextrin I 2000 RSP 4.6 mm ID × 250 mm (Astec) and DIACEL-chiralcel-OD-R 4.6 mm ID × 250 mm (JT Baker). TLC analyses: silica gel Merck 60 F<sub>254</sub> plates (0.2 mm layer thickness). Column chromatography: Merck Kieselgel 60 (70–230 mesh). Dry solvents were distilled immediately before use.

**Compound 1:** A solution of benzoyloxyethanal (3.7 g, 22.6 mmol) in dry MeCN (25 mL) was added with a syringe in one portion to a magnetically stirred suspension of polystyryl diphenylphosphane–iodine complex (22.6 mmol, prepared in situ) in dry MeCN (150 mL), at room temperature and under an atmosphere of dry N<sub>2</sub>. After 10 min, 1 M ethanedithiol in the same solvent (23 mL) was added in one portion. Benzoyloxyethanal was fully consumed (TLC monitoring) within 2 h. Solid K<sub>2</sub>CO<sub>3</sub> (excess) was added, the suspension stirred for a couple of minutes, and then filtered. The residual solid was washed with chloroform (3 × 100 mL), and then the combined filtrates were washed with 5 N aq

sodium thiosulfate (50 mL) and water until neutral. The solvents were then evaporated under reduced pressure to leave a residue consisting of practically pure **1** (5.3 g, 98%) as an oil. C<sub>11</sub>H<sub>12</sub>O<sub>2</sub>S<sub>2</sub> (240.3): calcd. C 54.97, H 5.03; found C 55.13, H 5.04. <sup>1</sup>H NMR (200 MHz): δ = 3.25 (s, 4 H, 4-H and 5-H), 4.35 (d, *J*<sub>6,2</sub> = 7.1 Hz, 2 H, 6-H), 4.79 (t, *J*<sub>2,6</sub> = 7.1 Hz, 1 H, 2-H), 7.38–7.52 (m, 3 H, *para*-H and *meta*-H), 8.03 (d, *J*<sub>ortho</sub> = 7.9 Hz, 2 H, *ortho*-H). <sup>13</sup>C NMR (200 MHz): δ = 37.8 (C-4, C-5), 50.4 (C-2), 68.0 (C-6), 120.5 (C-arom), 128.2–132.9 (C-arom), 165.8 (C=O).

**Compound 2. Typical Procedure:** Diethyl D-tartrate (1.4 mL, 9.6 mmol) dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL) was added under vigorous magnetic stirring at room temperature to a solution of Ti<sup>IV</sup> isopropoxide (0.7 mL, 2.4 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL). After 10 min, the resulting yellow homogeneous solution was cooled to –20 °C. *tert*-Butyl hydroperoxide (1.6 mL, 5.7 mmol) was then added dropwise, followed after a few minutes by 1,3-dithiolane **1** (2.9 g, 12.1 mmol) dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL). The reaction mixture was maintained at –20 °C whilst stirring for 14 h, then quenched with cold H<sub>2</sub>O (30 mL) and warmed up to room temperature. The resulting emulsion was cleared by filtration through Celite and the organic layer was then separated. The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 × 50 mL) and the combined organic phases were washed with 10% aq. sodium metabisulfite (100 mL), 5% aq. sodium hydroxide, and brine until neutral, then dried (Na<sub>2</sub>SO<sub>4</sub>) and the solvents evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (CHCl<sub>3</sub>) and, in addition to the unreacted starting material (1.5 g, 6.2 mmol), two diastereoisomeric sulfoxides were separated (overall yield 1.35 g, 90%). (*E*)-**2** (1.29 g, 5.03 mmol), m.p. 82–83 °C (from hexane). [α]<sub>D</sub><sup>25</sup> = –75.8 (*c* = 0.7, CHCl<sub>3</sub>). C<sub>11</sub>H<sub>12</sub>O<sub>3</sub>S<sub>2</sub> (256.3): calcd. C 51.54, H 4.72; found C 51.67, H 4.70. <sup>1</sup>H NMR (400 MHz): δ = 2.80–2.94 (m, 1 H, 5-H<sub>a</sub>), 3.48–3.62 (m, 2 H, 4-H), 3.75–3.86 (m, 1 H, 5-H<sub>b</sub>), 4.35 (dd, *J*<sub>6a,2</sub> = 9.3, *J*<sub>6a,6b</sub> = 12.4 Hz, 1 H, 6-H<sub>a</sub>), 4.53 (dd, *J*<sub>6b,2</sub> = 4.9, *J*<sub>6b,6a</sub> = 12.4 Hz, 1 H, 6-H<sub>b</sub>), 4.59 (dd, *J*<sub>2,6b</sub> = 4.9, *J*<sub>2,6a</sub> = 9.3 Hz, 1 H, 2-H), 7.43–7.51 (m, 2 H, *meta*-H), 7.60 (t, *J*<sub>ortho</sub> = 7.4 Hz, 1 H, *para*-H), 8.02 (d, *J*<sub>ortho</sub> = 7.4 Hz, 2 H, *ortho*-H). [The signal of the 2-H proton is split by addition of Eu(hfc)<sub>3</sub> from δ = 4.59 (dd) to δ = 5.06 (br. s, 96%) and δ = 5.17 (br. s, 4%).] <sup>13</sup>C NMR (400 MHz): δ = 32.8 (C-4), 55.1 (C-5), 63.6 (C-6), 72.3 (C-2), 128.9, 129.0, 130.1, 133.9 (C-arom), 166.1 (C=O). HPLC (DIACEL, 60% MeCN plus 0.01% TFA in H<sub>2</sub>O): *R*<sub>t</sub> = 15.48 min (4%), 19.99 min (96%).

(*E*)-**3** was prepared (yield 90%) under the same conditions from **1** using diethyl L-tartrate: m.p. 71–73 °C (from hexane). [α]<sub>D</sub><sup>25</sup> = +74.9 (*c* = 0.6, CHCl<sub>3</sub>). C<sub>11</sub>H<sub>12</sub>O<sub>3</sub>S<sub>2</sub> (256.3): calcd. C 51.54, H 4.72; found C 51.41, H 4.73. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra superimposable on those of (*E*)-**2**. HPLC (DIACEL, 60% MeCN plus 0.01% TFA in H<sub>2</sub>O): *R*<sub>t</sub> = 15.48 min (97%), 19.99 min (3%).

**Compounds 4a and 4b. Typical Procedure:** Trimethylsilyl triflate (TMSOTf) (0.9 mL, 4.8 mmol) and triethylamine (0.6 mL, 4.8 mmol) were added dropwise sequentially to a magnetically stirred suspension of *N*<sup>4</sup>-acetylcytosine (0.1 g, 1.2 mmol) in dry toluene (15 mL), at 0 °C and under nitrogen atmosphere. After 20 min a suspension of sulfoxide **2** [(*E*)-pair] (0.2 g, 0.8 mmol) in dry toluene (5 mL) was added and the resulting mixture was stirred overnight. After partial evaporation of the solvent under reduced pressure the residue was diluted with EtOAc (15 mL), washed with 5% aq NaHCO<sub>3</sub> (2 × 10 mL) and water until neutral, dried (Na<sub>2</sub>SO<sub>4</sub>), and then the solvents were evaporated under reduced pressure. After purification by column chromatography on silica gel (EtOAc/petroleum ether, 7:3) two diastereoisomeric cytosinyl dithiolanes (overall yield 0.22 g, 70%) were obtained.

Higher *R*<sub>f</sub> product **4a** (*trans*) (0.12 g), m.p. 194–196 °C (MeOH). [α]<sub>D</sub><sup>25</sup> = +78.0 (*c* = 0.2, CHCl<sub>3</sub>). C<sub>17</sub>H<sub>17</sub>N<sub>3</sub>O<sub>4</sub>S<sub>2</sub> (391.4): calcd. C 52.12, H 4.38; found C 52.26, H 4.37. <sup>1</sup>H NMR (500 MHz): δ = 2.23 (s, 3 H, COCH<sub>3</sub>), 3.46 (dd, *J*<sub>5a,4</sub> = 1.3, *J*<sub>5a,5b</sub> = 13.5 Hz, 1 H, 5-H<sub>a</sub>), 3.63 (dd, *J*<sub>5b,4</sub> = 4.6, *J*<sub>5b,5a</sub> = 13.5 Hz, 1 H, 5-H<sub>b</sub>), 4.37 (dd, *J*<sub>6a,2</sub> = 7.2, *J*<sub>6a,6b</sub> = 11.4 Hz, 1 H, 6-H<sub>a</sub>), 4.49 (dd, *J*<sub>6b,2</sub> = 7.2, *J*<sub>6b,6a</sub> = 11.4 Hz, 1 H, 6-H<sub>b</sub>), 5.02 (t, *J*<sub>2,6a</sub> = *J*<sub>2,6b</sub> = 7.2 Hz, 1 H, 2-H), 6.78 (dd, *J*<sub>4,5a</sub> = 1.3, *J*<sub>4,5b</sub> = 4.6 Hz, 1 H, 4-H), 7.41 (d, *J*<sub>5',6'</sub> = 7.5 Hz, 1 H, H-5'), 7.45–7.48 (m, 2 H, *meta*-H), 7.60 (t, *J*<sub>ortho</sub> = 7.4 Hz, 1 H, *para*-H), 8.04 (d, *J*<sub>ortho</sub> = 7.4 Hz, 2 H, *ortho*-H), 8.24 (d, *J*<sub>6',5'</sub> = 7.5 Hz, 1 H, 6'-H), 8.42 (br. s, 1 H, NH); [2-H, 4-H protons: no NOE effect]. <sup>13</sup>C NMR (200 MHz): δ = 24.7 (COCH<sub>3</sub>), 43.8 (C-5), 51.6 (C-2), 67.1 (C-4), 70.2 (C-6), 96.0 (C-5'), 128.4, 129.1, 129.6, 133.3 (C-arom), 146.3 (C-6'), 155.2 (C=O), 162.9 (C=N), 165.7 (C=O), 171.1 (C=O). HPLC (Cyclobond I 2000 RSP, 50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O): *R*<sub>t</sub> = 25.98 min (96%), 29.09 min (4%).

Lower *R*<sub>f</sub> product **4b** (*cis*) (0.1 g), m.p. 148–150 °C (MeOH). [α]<sub>D</sub><sup>25</sup> = –85.0 (*c* = 0.2, CHCl<sub>3</sub>). C<sub>17</sub>H<sub>17</sub>N<sub>3</sub>O<sub>4</sub>S<sub>2</sub> (391.4): calcd. C 52.16, H 4.38; found C 52.33, H 4.41. <sup>1</sup>H NMR (500 MHz): δ = 2.24 (s, 3 H, COCH<sub>3</sub>), 3.48 (dd, *J*<sub>5a,4</sub> = 2.6, *J*<sub>5a,5b</sub> = 13.5 Hz, 1 H, 5-H<sub>a</sub>), 3.62 (dd, *J*<sub>5b,4</sub> = 4.6, *J*<sub>5b,5a</sub> = 13.5 Hz, 1 H, 5-H<sub>b</sub>), 4.78 (dd, *J*<sub>6a,2</sub> = 5.6, *J*<sub>6a,6b</sub> = 11.8 Hz, 1 H, 6-H<sub>a</sub>), 4.81 (dd, *J*<sub>6b,2</sub> = 5.6, *J*<sub>6b,6a</sub> = 11.8 Hz, 1 H, 6-H<sub>b</sub>), 5.02 (t, *J*<sub>2,6a</sub> = *J*<sub>2,6b</sub> = 5.6 Hz, 1 H, 2-H), 6.71 (dd, *J*<sub>4,5a</sub> = 2.6, *J*<sub>4,5b</sub> = 4.6 Hz, 1 H, 4-H), 7.29 (d, *J*<sub>5',6'</sub> = 7.5 Hz, 1 H, 5'-H), 7.49–7.53 (m, 2 H, *meta*-H), 7.64 (t, *J*<sub>ortho</sub> = 7.6 Hz, 1 H, *para*-H), 8.07 (d, *J*<sub>ortho</sub> = 7.4 Hz, 2 H, *ortho*-H), 8.51 (d, *J*<sub>6',5'</sub> = 7.5 Hz, 1 H, 6'-H); [2-H, 4-H protons: NOE effect]. <sup>13</sup>C NMR (200 MHz): δ = 24.7 (CH<sub>3</sub>CO), 45.3 (C-5), 53.6 (C-2), 64.7 (C-4), 69.6 (C-6), 96.1 (C-5'), 128.5, 129.6, 130.8, 133.6 (C-arom), 146.3 (C-6'), 155.0 (C=O), 162.7 (C=N), 165.8 (C=O), 171.0 (C=O). HPLC (Cyclobond I 2000 RSP, 50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O): *R*<sub>t</sub> = 13.37 min (96%), 15.76 min (4%).

Under the same conditions the sulfoxides **3** [(*E*)-pair] afforded their enantiomeric nucleosides (**8a** and **8b**) in 72% yield, *trans/cis* = 1.1:1.0):

**8a** (*trans*), m.p. 193–195 °C (MeOH). [α]<sub>D</sub><sup>25</sup> = –80.0 (*c* = 0.2, CHCl<sub>3</sub>). C<sub>17</sub>H<sub>17</sub>N<sub>3</sub>O<sub>4</sub>S<sub>2</sub> (391.4): calcd. C 52.16, H 4.38; found C 52.21, H 4.38. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra superimposable on those of **4a**. HPLC (Cyclobond I 2000 RSP, 50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O): *R*<sub>t</sub> = 27.40 min (97%), 29.45 min (3%).

**8b** (*cis*), m.p. 149–151 °C (MeOH). [α]<sub>D</sub><sup>25</sup> = +84 (*c* = 0.2, CHCl<sub>3</sub>). C<sub>17</sub>H<sub>17</sub>N<sub>3</sub>O<sub>4</sub>S<sub>2</sub> (391.4): calcd. C 52.16, H 4.38; found C 52.25, H 4.39. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra superimposable on those of **4b**. HPLC (Cyclobond I 2000 RSP, 50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O): *R*<sub>t</sub> = 13.74 min (3%), 16.32 min (97%).

**Compounds 6a and 6b:** Trimethylsilyl triflate (TMSOTf) (0.44 mL, 2.4 mmol) and triethylamine (0.4 mL, 2.4 mmol) were added dropwise sequentially to a magnetically stirred suspension of 6-chloropurine (0.26 g, 1.6 mmol) in dry toluene (10 mL) at 0 °C under an N<sub>2</sub> atmosphere. After 4 h a suspension of sulfoxide **2** [(*E*)-pair] (0.2 g, 0.8 mmol) and ZnI<sub>2</sub> (0.08 g, 0.24 mmol) in the same solvent (5 mL) was added and the resulting mixture was stirred overnight. After partial evaporation of the solvent under reduced pressure, the residue was redissolved in EtOAc (15 mL), washed with 5% aq NaHCO<sub>3</sub> (2 × 10 mL) and water until neutral, dried (Na<sub>2</sub>SO<sub>4</sub>), and then the solvents were evaporated under reduced pressure. After purification by column chromatography on silica gel (EtOAc/petroleum ether, 7:3) two diastereoisomeric purinyl dithiolanes **6a** and **6b** (0.4 g, yield 65%) were obtained.

Higher  $R_f$  product **6a** (*trans*) (0.2 g), m.p. 156–157 °C (MeOH).  $[\alpha]_D^{25} = +43.3$  ( $c = 0.2$ , MeOH).  $C_{16}H_{13}ClN_4O_2S_2$  (392.9): calcd. C 48.91, H 3.34; found C 48.80, H 3.35.  $^1H$  NMR (500 MHz):  $\delta = 3.42$  (dd,  $J_{5a,4} = 1.5$ ,  $J_{5a,5b} = 13.4$  Hz, 1 H, 5-H<sub>a</sub>), 3.79 (dd,  $J_{5b,4} = 4.1$ ,  $J_{5b,5a} = 13.4$  Hz, 1 H, 5-H<sub>b</sub>), 4.45 (dd,  $J_{6a,2} = 7.3$ ,  $J_{6a,6b} = 11.4$  Hz, 1 H, 6-H<sub>a</sub>), 4.56 (dd,  $J_{6b,2} = 7.3$ ,  $J_{6b,6a} = 11.4$  Hz, 1 H, 6-H<sub>b</sub>), 5.15 (t,  $J_{2,6a} = J_{2,6b} = 7.3$  Hz, 1 H, 2-H), 7.12 (dd,  $J_{4,5a} = 1.5$ ,  $J_{4,5b} = 4.1$  Hz, 1 H, 4-H), 7.50 (t,  $J_{ortho} = 7.3$  Hz, 2 H, *meta*-H), 7.62 (t,  $J_{ortho} = 7.3$  Hz, 1 H, *para*-H), 8.07 (d,  $J = 7.3$  Hz, 2 H, *ortho*-H), 8.82 (s, 1 H, 2'-H), 8.91 (s, 1 H, 8'-H); [2-H, 4-H protons: no NOE effect].  $^{13}C$  NMR (200 MHz):  $\delta = 45.0$  (C-5), 51.8 (C-2), 66.8 (C-4), 69.4 (C-6), 121.7 (C-5'), 128.6, 129.2, 130.8, 133.6 (C-arom), 142.4 (C-4'), 148.5 (C-8'), 152.8 (C-2'), 162.8 (C-6'), 165.9 (C=O).

Lower  $R_f$  product **6b** (*cis*) (0.2 g), amorphous.  $[\alpha]_D^{25} = -34.4$  ( $c = 0.2$ , MeOH).  $C_{16}H_{13}ClN_4O_2S_2$  (392.9): calcd. C 48.91, H 3.34; found C 48.83, H 3.33.  $^1H$  NMR (500 MHz):  $\delta = 3.65$  (dd,  $J_{5a,4} = 2.5$ ,  $J_{5a,5b} = 13.4$  Hz, 1 H, 5-H<sub>a</sub>), 3.81 (dd,  $J_{5b,4} = 4.4$ ,  $J_{5b,5a} = 13.4$  Hz, 1 H, 5-H<sub>b</sub>), 4.73 (dd,  $J_{6a,2} = 6.4$ ,  $J_{6a,6b} = 11.7$  Hz, 1 H, 6-H<sub>a</sub>), 4.79 (dd,  $J_{6b,2} = 6.4$ ,  $J_{6b,6a} = 11.7$  Hz, 1 H, 6-H<sub>b</sub>), 5.17 (t,  $J_{2,6a} = J_{2,6b} = 6.4$  Hz, 1 H, 2-H), 7.06 (dd,  $J_{4,5a} = 2.5$ ,  $J_{4,5b} = 4.4$  Hz, 1 H, 4-H), 7.49 (t,  $J_{ortho} = 7.3$  Hz, 2 H, *meta*-H), 7.62 (t,  $J_{ortho} = 7.3$  Hz, 1 H, *para*-H), 8.04 (d,  $J = 7.3$  Hz, 2 H, *ortho*-H), 8.92 (s, 1 H, 2'-H), 9.02 (s, 1 H, 8'-H); [2-H, 4-H protons: NOE effect].  $^{13}C$  NMR (200 MHz):  $\delta = 46.3$  (C-5), 53.4 (C-2), 65.5 (C-4), 68.7 (C-6), 120.5, (C-5'), 128.0, 129.1, 133.6 (C-arom), 142.1 (C-4'), 148.2 (C-8'), 152.7 (C-2'), 162.6 (C-6'), 165.8 (C=O).

**Typical Deprotection Procedure. Compound 5a:** NaOMe (5.4 mg, 0.1 mmol) was added at room temperature to a stirring solution of **4a** (0.04 g, 0.1 mmol) in MeOH (5 mL) under an N<sub>2</sub> atmosphere. After 6 h the reaction mixture was quenched with glacial acetic acid (excess), the MeOH was removed under reduced pressure, and the crude residue chromatographed on silica gel (CHCl<sub>3</sub>/MeOH, 9:1) to give pure **5a** (0.02 g, yield 81%), m.p. 205 °C dec. (EtOAc).  $[\alpha]_D^{25} = +26.5$  ( $c = 0.7$ , MeOH).  $C_8H_{11}N_3O_2S_2$  (245.3): calcd. C 39.17, H 4.52; found C 39.06, H 4.55.  $^1H$  NMR (400 MHz, [D<sub>6</sub>]DMSO):  $\delta = 3.32$ –3.58 (m, 4 H, 5-H and 6-H), 4.72 (t,  $J_{2,6} = 6.9$  Hz, 1 H, 2-H), 5.35 (t,  $J_{OH,6} = 5.6$  Hz, 1 H, OH), 5.71 (d,  $J_{5',6'} = 7.6$  Hz, 1 H, 5'-H), 6.48 (dd,  $J_{4,5a} = 1.2$ ,  $J_{4,5b} = 4.6$  Hz, 1 H, H-4), 7.22 (br. d, 2 H, NH<sub>2</sub>), 7.88 (d,  $J_{6',5'} = 7.6$  Hz, 1 H, 6'-H).  $^{13}C$  NMR (500 MHz, [D<sub>6</sub>]DMSO):  $\delta = 42.6$  (C-5), 55.1 (C-2), 65.9 (C-4), 68.0 (C-6), 93.2 (C-5'), 142.5 (C-6'), 155.1 (C=O), 165.6 (C=N). HPLC (Cyclobond I 2000 RSP, 50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O):  $R_t = 9.68$  min (4%), 10.63 min (96%).

The following nucleosides were also obtained under the same deprotection conditions.

**Compound 5b from 4b:** 83% yield, m.p. 110–112 °C (EtOAc).  $[\alpha]_D^{25} = +18.5$  ( $c = 0.06$ , MeOH).  $C_8H_{11}N_3O_2S_2$  (245.3): calcd. C 39.17, H 4.52; found C 39.26, H 4.53.  $^1H$  NMR (400 MHz, [D<sub>6</sub>]DMSO):  $\delta = 3.36$  (dd,  $J_{5a,4} = 4.8$ ,  $J_{5a,5b} = 12.7$  Hz, 1 H, 5-H<sub>a</sub>), 3.46 (dd,  $J_{5b,4} = 4.6$ ,  $J_{5a,5b} = 12.7$  Hz, 1 H, 5-H<sub>b</sub>), 3.73 (t,  $J_{6,2} = 6.0$  Hz, 2 H, 6-H), 4.60 (t,  $J_{2,6} = 6.0$  Hz, 1 H, 2-H), 5.49 (t,  $J_{OH,6} = 5.9$  Hz, 1 H, OH), 5.74 (d,  $J_{5',6'} = 7.3$  Hz, 1 H, 5'-H), 6.47 (dd,  $J_{4,5a} = 4.8$ ,  $J_{4,5b} = 4.6$  Hz, 1 H, H-4), 7.15 (bd, 2 H, NH<sub>2</sub>), 8.04 (d,  $J_{6',5'} = 7.3$  Hz, 1 H, 6'-H).  $^{13}C$  NMR (500 MHz, [D<sub>6</sub>]DMSO):  $\delta = 42.3$  (C-5), 55.9 (C-2), 64.7 (C-4), 67.6 (C-6), 93.7 (C-5'), 142.1 (C-6'), 155.2 (C=O), 165.6 (C=N). HPLC (Cyclobond I 2000 RSP, 50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O):  $R_t = 9.76$  min (4%), 10.81 min (96%).

**Compound 9a from 8a:** 80% yield, m.p. 202 °C dec. (EtOAc).  $[\alpha]_D^{25} = -28.5$  ( $c = 0.7$ , MeOH).  $C_8H_{11}N_3O_2S_2$  (245.3): calcd. C 39.17, H 4.52; found C 39.29, H 4.51.  $^1H$  NMR and  $^{13}C$  NMR spectra superimposable on those of **5a**. HPLC (50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O):  $R_t = 9.70$  min (97%), 10.65 min (3%).

**Compound 9b from 8b:** 78% yield, m.p. 108–110 °C (EtOAc).  $[\alpha]_D^{25} = -20.5$  ( $c = 0.1$ , MeOH).  $C_8H_{11}N_3O_2S_2$  (245.3): calcd. C 39.17, H 4.52; found C 39.22, H 4.52.  $^1H$  NMR and  $^{13}C$  NMR spectra superimposable on those of **5b**. HPLC (50% MeOH plus 0.1 M NH<sub>4</sub>OAc in H<sub>2</sub>O):  $R_t = 9.75$  min (97%), 10.79 min (3%).

**Compound 7a from 6a:** 94% yield, m.p. 192–194 °C (MeOH).  $[\alpha]_D^{25} = +25.8$  ( $c = 0.34$ , MeOH).  $C_9H_9ClN_4OS_2$  (288.8): calcd. C 37.43, H 3.14; found C 37.36, H 3.12.  $^1H$  NMR (500 MHz):  $\delta = 3.39$  (dd,  $J_{5a,4} = 1.5$ ,  $J_{5a,5b} = 13.2$  Hz, 1 H, 5-H<sub>a</sub>), 3.64 (dd,  $J_{5b,4} = 4.2$ ,  $J_{5b,5a} = 13.2$  Hz, 1 H, 5-H<sub>b</sub>), 3.72–3.78 (m, 2 H, 6-H), 4.87 (t,  $J_{2,6} = 6.6$  Hz, 1 H, 2-H), 6.85 (dd,  $J_{4,5a} = 1.5$ ,  $J_{4,5b} = 4.2$  Hz, 1 H, 4-H), 8.61 (s, 1 H, 2'-H), 8.64 (s, 1 H, 8'-H).  $^{13}C$  NMR (500 MHz):  $\delta = 44.5$  (C-5), 54.3 (C-2), 66.2 (C-4), 69.5 (C-6), 112.0 (C-5'), 144.9 (C-8'), 152.3 (C-8'), 156.7 (C-4'), 161.9 (C-5').

**Compound 7b from 6b:** 95% yield, amorphous.  $[\alpha]_D^{25} = -32.7$  ( $c = 0.34$ , MeOH).  $C_9H_9ClN_4OS_2$  (288.8): calcd. C 37.43, H 3.14; found C 37.35, H 3.15.  $^1H$  NMR (500 MHz):  $\delta = 3.53$  (dd,  $J_{5a,4} = 2.9$ ,  $J_{5a,5b} = 13.1$  Hz, 1 H, 5-H<sub>a</sub>), 3.72 (dd,  $J_{5b,4} = 4.4$ ,  $J_{5b,5a} = 13.1$  Hz, 1 H, 5-H<sub>b</sub>), 3.74–3.81 (m, 2 H, 6-H), 4.91 (t,  $J_{2,6} = 5.6$  Hz, 1 H, 2-H), 6.78 (dd,  $J_{4,5a} = 2.9$ ,  $J_{4,5b} = 4.4$  Hz, 1 H, 4-H), 8.65 (s, 1 H, 2'-H), 8.84 (s, 1 H, 8'-H).  $^{13}C$  NMR (500 MHz):  $\delta = 46.2$  (C-5), 56.9 (C-2), 65.5 (C-4), 69.1 (C-6), 111.5 (C-5'), 144.5 (C-8'), 152.3 (C-2'), 156.0 (C-4'), 160.2 (C-6').

## Acknowledgments

Work supported by Regione Campania (L. 41/94) and Consorzio Interuniversitario Nazionale La Chimica per l'Ambiente (M.I.U.R., L. 488/92, Cluster 11-A).  $^1H$  and  $^{13}C$  NMR spectra were recorded at Centro Interdipartimentale di Metodologie Chimico-Fisiche, Università di Napoli Federico II. The authors are gratefully indebted to Prof. F. Giordano (Dip. di Chimica, Università di Napoli Federico II) for the X-ray crystallographic analysis of the chiral sulfoxide (*E*)-**3**.

- J. C. Graciet, P. Faury, M. Camplo, A. S. Charvet, N. Mourier, C. Trabaud, V. Niddam, V. Simon, J. L. Kraus, *Nucleosides* **1995**, *14*, 1379–1392.
- E. De Clercq, *Aids Research and Human Retroviruses* **1992**, *8*, 119–134.
- P. A. Furman, J. A. Fyle, M. H. St. Clair, K. Weinhold, J. L. Rideout, G. A. Freeman, S. Nusinoff-Lehrman, D. P. Bolognesi, S. Broder, H. Mitsuya, D. W. Barry, *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 8333–8337.
- R. Wittington, R. N. Brogden, *Drugs* **1992**, *44*, 656–683.
- D. Faulds, R. N. Brogden, *Drugs* **1992**, *44*, 94–116.
- T.-S. Lin, R. F. Schinazi, W. H. Prusoff, *Biochem. Pharmacol.* **1987**, *36*, 2713–2718.
- L. S. Jeong, R. F. Schinazi, J. W. Beach, H. O. Kim, K. Shanmuganathan, S. Nampalli, M. W. Chun, W.-K. Chung, B. G. Choi, C. K. Chu, *J. Med. Chem.* **1993**, *36*, 2627–2638.
- D. W. Kimberlin, D. M. Coen, K. K. Biron, J. I. Cohen, R. A. Lamb, M. McKinlay, E. A. Emini, R. J. Whitley, *Antiviral Res.* **1995**, *26*, 369–401.

- [9] J. A. V. Coates, N. Cammack, H. J. Jenkinson, *Antimicrob. Agents Chemother.* **1992**, *36*, 202–205.
- [10] C. Thibaudeau, J. Chattopadhyaya, *Nucleoside Nucleotides* **1997**, *16*, 523–529.
- [11] R. F. Schinazi, C. K. Chu, A. Peck, *Antimicrob. Agents Chemother.* **1992**, *36*, 672–676.
- [12] C. Simons, in: *Nucleoside Mimetics*, Gordon and Breach Science, **2001**, chapter 3, pp. 63–79.
- [13] N. Nguyen-Ba, W. Brown, N. Lee, B. Zacharie, *Synthesis* **1998**, 759–762.
- [14] N. Nguyen-Ba, W. L. Brown, L. Chan, N. Lee, L. Brasili, D. Lafleur, B. Zacharie, *Chem. Commun.* **1999**, 1245–1246.
- [15] R. Caputo, A. Guaragna, G. Palumbo, S. Pedatella, *Eur. J. Org. Chem.* **1999**, 1455–1458.
- [16] R. Caputo, E. Cassano, L. Longobardo, D. Mastroianni, G. Palumbo, *Synthesis* **1995**, 141–143 and papers cited therein.
- [17] E. V. Efimtseva, S. N. Mikhailov, S. Meshkov, T. Hankamäki, M. Oivanen, H. Lönnberg, *J. Chem. Soc., Perkin Trans. 1* **1995**, 1409–1415.
- [18] R. Caputo, C. Ferreri, G. Palumbo, *Synthesis* **1987**, 386–389.
- [19] F. Di Furia, G. Licini, G. Modena, *Gazz. Chim. Ital.* **1990**, *120*, 165–170.
- [20] B. Belleau, L. Brasili, L. Chan, M. P. DiMarco, B. Zacarie, N. Nguyen-Ba, H. J. Jenkinson, J. A. V. Coates, J. M. Cameron, *Bioorg. Med. Chem. Lett.* **1993**, *3*, 1723–1728.

Received July 15, 2002  
[O02391]