

Synthesis of *p*-trifluoroacetamidophenyl (4,6-dideoxy-4-formamido-3-*C*-methyl-2-*O*methyl- α -L-mannopyranosyl)-(1 \rightarrow 3)-(2-*O*methyl- α -D-rhamnopyranosyl)-(1 \rightarrow 3)-(2-*O*methyl- α -L-fucopyranosyl)-(1 \rightarrow 3)-(α -Lrhamnopyranosyl)-(1 \rightarrow 2)-6-deoxy- α -Ltalopyranoside: a spacer-armed pentasaccharide glycopeptidolipid antigen of *Mycobacterium avium* serovar 14

István Bajza^{a,*}, Katalin E. Kövér^b, András Lipták^{a, c,*}

^a Research Group for Carbohydrates of the Hungarian Academy of Sciences, Debrecen, P.O.B 55, H-4010 Hungary

^b Institute of Organic Chemistry, L. Kossuth University, Debrecen, P.O.B. 20, H-4010 Hungary ^c Institute of Biochemistry, L. Kossuth University, Debrecen, P.O.B 55, H-4010 Hungary

Received 19 September 1997; accepted 6 February 1998

Abstract

Syntheses of *p*-trifluoroacetamidophenyl glycosides of the haptenic pentasaccharide and the nonreducing disaccharide unit of the title pentasaccharide are reported. The synthesis of the terminal *N*formylkansosamine unit started from methyl 6-deoxy-2,3-*O*-isopropylidene- α -L-*lyxo*-hexopyran-4uloside which, after C-3 methylation, was transformed into a glycosyl donor [3-*O*-benzyl-4-*N*-benzylformamido-4,6-dideoxy-3-*C*-methyl-2-*O*-methyl- α , β -L-mannopyranosyl trichloroacetimidate (**20**), and used for the synthesis of *p*-trifluoroacetamidophenyl (4-formamido-4,6-dideoxy-3-*C*methyl-2-*O*-methyl- α -L-mannopyranosyl)-(1 \rightarrow 3)-6-deoxy-2-*O*-methyl- α -D-mannopyranoside (**29**). Ethyl (3-*O*-benzyl-4-*N*-benzylformamido-4,6-dideoxy-3-*C*-methyl-2-*O*-methyl- α -L-mannopyranosyl)-(1 \rightarrow 3)-4-*O*-benzyl-6-deoxy-2-*O*-methyl-1-thio- α -D-mannopyranoside (**31**), prepared by glycosylation of ethyl 4-*O*-benzyl-6-deoxy-2-*O*-methyl-1-thio- α -D-mannopyranoside with **20**, served as glycosyl donor in a 2+3 block synthesis of the title pentasaccharide. © 1998 Elsevier Science Ltd. All rights reserved

Keywords: Branched-chain sugar; Spacer, p-trifluoroacetamidophenyl; Oligosaccharide synthesis; Block synthesis

^{*} Corresponding authors.

1. Introduction

The pioneering work of Brennan [1] and Aspinall et al. [2] laid the foundation of the molecular basis of the exact serodiagnosis of infections caused by mycobacteria. Many pulmonary infections, especially in patients suffering from acquired immunodeficiency syndrome (AIDS), are connected with mycobacteria of the Mycobacterium avium-Mycobacterium scrofulaceum complex. Different serovars of this complex possess highly antigenic glycopeptidolipids (GPLs) on their cell surface. Thus the outer oligosaccharide haptens, after conjugation with suitable proteins, might aid the serodiagnosis of mycobacterial infections. Among these haptens, the pentasaccharide of M. avium serovar 14 has the most complex structure [3,4] (Scheme 1). Here rhamnose occurs in both enantiomeric forms and the terminal N-formyl- α -L-(4-N-formamido-4,6-dideoxy-3-Ckansosamine methyl-2-*O*-methyl- α -L-mannopyranose) is а highly functionalised branched-chain aminosugar. Although kansosamine has not been isolated in free form, its derivatives were found in the surface antigen of *M. kansasii* [5] [4-(2-*O*-methyl)propionamido] and in the antibiotic sibiromycin [6] (4-N-methylated). For the preparation of N-formyl- α -L-kansosamine, four different methods have been published [4,7–9], and the synthesis of some oligosaccharide fragments of the pentasaccharide have also been reported. The B-A disaccharide (with benzyl [10], methyl [11], and *p*-nitrophenyl [12,13] aglycons), and the E-D [4] and D-C [4] disaccharides (with the allyl aglycon) were also reported.



Scheme 1.

Two trisaccharide segments (C–B–A with the *o*-aminopropyl [14] and *p*-nitrophenyl aglycons [12], and E–D–C [4] with the allyl aglycon), and the tetrasaccharide D–C–B–A [12] with the *p*-nitrophenyl aglycon have also been prepared.

2. Results and discussion

For the synthesis of *N*-formyl-L-kansosamine on a preparative scale we applied two known synthetic strategies [15,16]. Firstly, the addition of methyl magnesium iodide to 2 (Scheme 2) obtained by oxidation of methyl 4-O-benzyl-2-O-methyl- α -Lrhamnopyranoside [17] (1) with pyridinium chlorochromate [18] gave diastereomers 3 and 4 in a ratio of 1:1.2. The stereochemical molar assignment of compounds 3 and 4 was based on homonuclear NOE measurements [19]. In compound 3, the irradiation of the 3-C-methyl protons resulted in the signal enhancement of H-2 (14%) and H-5 (8%), while no effect was observed at the resonance frequency of H-4. These spectral data showed the proximity of the 3-C-Me group to H-5, confirming its axial position. In the case of compound 4, signal intensity enhancements were observed at H-4 (9%) and at H-2 (11%) indicating the equatorial orientation of the 3-C-Me group. The low yields of 3(19%) and 4(13%) prompted us to proceed with our second synthetic strategy.

Methyl 6-deoxy-2,3-O-isopropylidene- α -L-lyxohexopyran-4-uloside (5) was C-methylated [20] with methyl iodide in the presence of strong inorganic or organic bases, to give methyl 6-deoxy-2,3-O-isopropylidene-3-C-methyl- α -L-lyxo-hexopyran-4-uloside (6). Reduction of compound 6 with $NaBH_4$ yielded exclusively 6-deoxy-talopyranoside derivative 7, which was converted into 4-O-triflate 8. Treatment of compound 8 with sodium azide afforded methyl 4,6-dideoxy-2,3-O-isopropylidene-3-*C*-methyl- α -L-*erythro*-hex-4-enopyranoside (9), instead of the desired 4-azido-L-rhamnopyranoside derivative. The reason of this elimination may have been the presence of the axial 3-C-Me group, which prevented the attack of the nucleophile, since successful azide displacements of the 4-O-trifluoromethanesulphonyl derivative in 6-deoxy- α -Ltalopyranoside [21] and D-talopyranoside [22] were reported to yield 4-azido-L-rhamno- and 4-azido-D-mannopyranoside derivatives. Catalytic reduction (Pd-C) of oxime 10, prepared from 6, gave exclusively the 6-deoxy-talo isomer (11), but the





LiA1H₄ reduction of **10** resulted in a 1:1 mixture of 6-deoxy-L-*talo*- (**11**) and 6-deoxy-L-*manno*-isomers (**12**) [23]. Compound **12** was isolated in 41% yield. After *N*-formylation of **12** with acetic-formic anhydride (\rightarrow **13**) and *N*-benzylation (\rightarrow **14**), the 2,3-*O*-isopropylidene group was removed (\rightarrow **15**) and the HO-2 function was selectively methylated to give compound **16**. The remaining free HO-3 group of **16** was benzylated to yield the fully protected *N*-formyl- α -L-kansosamine derivative (**17**). Acetolysis of compound **17** (\rightarrow **18**), followed by deacetylation (\rightarrow **19**) and imidation in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) [24] gave donor **20**.

For the preparation of *p*-nitrophenyl 4-*O*-benzyl-2-*O*-methyl- α -D-rhamnopyranoside (26), *p*-nitrophenyl 2,3-*O*-isopropylidene- α -D-mannopyranoside [25] was selectively tosylated at HO-6 (\rightarrow 21), and the tosyloxy group was transformed into a 6deoxy-6-iodo function (22). Compound 22 was reduced by LiA1H₄ to give 23, and the latter was benzylated and deisopropylidenated to give 25. Treatment of 25 with methyl iodide under phasetransfer condition [26] gave 26. The structure of all these compounds was verified by ¹H and ¹³C NMR data, and the physical parameters of compounds 23–25 were compared with those of their L-enantiomers [27]. The glycosylation of compound **26** with trichloroacetimidate **20** (Scheme 3), catalysed by trimethylsilyl triflate, yielded the fully protected disaccharide **27** in 35% yield, whose structure was confirmed by ¹H and ¹³C NMR spectroscopy (Table 1). The nitro group in **27** was catalytically reduced, and the amine formed was trifluoroacetylated to give **28**. Deprotection of **28** was achieved by catalytic hydrogenolysis yielding the spacer-armed disaccharide **29** (for NMR data, see Table 2).

Ethyl 4-*O*-benzyl-2-*O*-methyl-1-thio- α -D-rhamnopyranoside [12] (**30**) was glycosylated with donor **20**, yielding disaccharide **31** (75%) as the E–D synthon (Scheme 1; Table 1) for the block synthesis of the target pentasaccharide. The considerable difference in yields of the glycosylations of **26** and **30** with donor **20** may have been due to the different electronic effects of the aglycons since the substitution pattern of the two acceptors was the same.

Previously, we have reported [12] the synthesis of the *p*-nitrophenyl glycoside of the tetrasaccharide hapten isolated from *M. avium* serovar 20. This tetrasaccharide was the D–C–B–A part of our target pentasaccharide. An intermediate of the unprotected tetrasaccharide [12], *p*-nitrophenyl (3-*O*-acetyl-4-*O*-benzyl-2-*O*-methyl- α -D-rhamnopyranosyl)-(1–3)-(4-*O*-benzyl-2-*O*-methyl- α -L-fuco-



Scheme 3.

Table 1 NMR data for disaccharides **27** and **31** in CDCl₃ (δ in ppm)

Atom no.	27 a		27 b		31 a		31 b	
	δ ¹³ C	δ ¹ H						
1	95.88	5.64	96.22	5.60	80.61	5.38	80.61	5.34
2	76.39	3.84	77.2	3.81	77.96	3.75	78.70	3.71
3	73.98	4.20	74.16	4.27	74.38	3.97	74.78	4.04
4	79.54	3.54	80.09	3.55	80.02	3.48	80.33	3.47
5	69.51	3.76	69.30	3.71	68.38	4.10	68.20	4.0
6	18.33	1.30	18.13	1.23	18.12	1.33	18.03	1.26
1'	94.45	5.11	96.76	5.08	93.44	5.00	95.49	4.99
2'	81.03	3.46	82.53	3.77	80.90	3.45	82.25	3.66
3'	78.5		80.15		78.5		80.23	
4′	62.50	4.40	65.81	3.23	62.50	4.40	65.43	3.21
5'	65.27	4.13	63.77	5.04	65.14	4.04	63.89	5.00
6'	17.35	0.63	18.69	0.73	17.33	0.62	18.58	0.71
3'-CMe	17.50	1.45	18.30	1.53	17.33	1.40	18.30	1.54
2-OMe	59.20	3.58	59.68	3.55	58.83	3.48	59.37	3.52
2'-OMe	58.94	3.51	58.94	3.56	57.94	3.48	57.78	3.46
4-OCH ₂ Ph	74.38	4.69 ± 4.75	74.73	4.57 ± 4.98	74.43	4.71 + 4.73	74.94	4.54 + 4.98
$3'-OC\tilde{H_2}Ph$	63.36	4.45 + 4.48	63.72	4.45	63.26	4.43	63.52	4.40
NCH ₂ Ph	50.2	3.85 ± 5.40	55.97	4.58 ± 4.98	50.09	3.83 + 5.4	55.92	4.40
NCHŌ	161.61	7.49	163.32	8.36	161.71	7.49	163.27	8.33
SCH ₂ CH ₃					25.48	2.63	25.36	2.61
SCH_2CH_3					14.98	1.29	15.02	1.26
Ph	126.0 -		126.0 -		126.0 -	7.1 - 7.4	126.0 -	7.1-7.4
	140.0		140.0		140.0		140.0	
PNP-C1	161.14		161.45					
PNP-C2,C6	116.33	7.15	116.38	7.13				
PNP-C3,C5	125.83	8.19	125.76	8.17				
PNP-C4	142.72		142.53					

pyranosyl)- $(1\rightarrow 3)$ -(2,4-di-O-benzyl- α -L-rhamnopyranosyl)- $(1\rightarrow 2)$ -endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside, was saponified to afford the tetrasaccharide acceptor for the preparation of our target pentasaccharide. Unfortunately neither imidate **20** nor the previously prepared phenyl 4-(Nbenzylformamido)-4,6-dideoxy-3-C-methyl-2-Omethyl-1-thio- α -L-mannopyranoside reacted with this acceptor.

The trisaccharide acceptor *p*-nitrophenyl (4-*O*-benzyl-2-*O*-methyl- α -L-fucopyranosyl)- $(1\rightarrow 3)$ -(2,4-di-*O*-benzyl- α -L-rhamnopyranosyl)- $(1\rightarrow 2)$ -*endo*-

3,4-*O*-benzylidene-6-deoxy- α -L-talopyranoside [12] (32) reacted readily with the disaccharide donor 31 in the presence of NIS/TfOH as promotor [28], yielding the fully protected pentasaccharide 33 in a yield of 55% (Scheme 4). The complete ¹H and ¹³C NMR assignment of compound 33 was not possible since signals of the CH₂ protons of the benzyl groups appeared in the informative region of the ¹H NMR spectrum. The *E* and *Z* isomerism of the N-CHO group also made the interpretation of the spectrum more difficult. The assignable chemical shift values for compound 33 are presented in Table 3.

Table 2 NMR data for disaccharide **29** in CD₃OD (δ in ppm)

Atom no.	δ^1	Н	$\delta^{13}C$		
	29 a	29 b	29 a	29 b	
1	5.	5.60		96.3	
2	3.	3.88		76.8	
3	4.	4.06		75.1	
4	3.	3.49		71.0	
5	3.	3.68		69.5	
6	1.	1.24		17.2	
OMe	3.30-	3.30-3.32		\sim 58.9	
Ph	7.56-	7.56-7.11		122.8-117	
1'	5.04	5.15	94.6	94.8	
2'	3.12	3.17	84	.3	
4′	3.	3.26		60.3	
5'	4.00	4.05	67.5		
6'	1.17	1.20	17	.7	
OMe	3.49-	3.49-3.52		\sim 58.9	
NCHO	8.16	8.55	136.9	131	
CH ₃ (3')	1.32	1.39	1	9	

The fully protected pentasaccharide **33** was converted into *p*-trifluoroacetamidophenyl glycoside **34**, and then deprotected as described for disaccharide **29**. Full assignment of the ¹H and ¹³C spectra of the pentasaccharide **35** was achieved by ¹H-¹H COSY and TOCSY experiments as well as ¹H/¹³C HSQC and HSQC-TOCSY experiments [29,30] (Table 4). The spectral parameters were in full accord with the target structure.

3. Experimental

General.-Melting points (uncorrected) were determined on a Kofler hot-stage apparatus. NMR spectra were recorded at 25 °C with a Bruker WP 200 SY (¹H, 200 MHz; ¹³C, 50 MHz) or a Bruker Avance DRX 500 (¹H, 500.13 MHz; $^{13}C.$ 125.76 MHz) spectrometer for solutions in CDCl₃ (internal Me₄Si) or in CD₃OD. Proton chemical shifts (δ) are given in ppm relative to the signal for internal Me₄Si (CDCl₃). Carbon chemical shifts were referenced to the solvent signal. Column chromatography was performed on Kieselgel 60 (Merck, 230 mesh) and fractions were monitored by TLC on Kieselgel 60 F_{254} (Merck) by detection with UV light and then charring with H₂SO₄. Unless noted otherwise, optical rotations were measured for solutions in CHCl₃ at 20 °C with a Perkin Elmer 241 polarimeter, using a 10 cm (1 mL) cell. Concentration of solutions was performed at 40 °C.

Methyl 4-O-benzyl-2-O-methyl- α -L-arabino-hexopyranosid-3-ulose (2).—Methyl 4-O-benzyl-6-deoxy-2-O-methyl- α -L-mannopyranoside (1) (1.96 g, 6.94 mmol) was dissolved in CH₂Cl₂ (200 mL), pyridinium chlorochromate (6.5 g, 34.7 mmol) was added, and the mixture was stirred for 12 h at room temperature. The mixture was chromatographed (7:3 hexane–EtOAc) to yield 2 (1.62 g, 83%) as a



α-Kanp4NFo-(1 \rightarrow 3)-2-OMe-α-D-Rhap-(1 \rightarrow 3)-α-L-Rhap-(1 \rightarrow 2)-α-L-Rhap-(1 \rightarrow 2)-6d-α-L-Talp-(1 \rightarrow OPTFAAP)

Table 4

Residue	Atom no.	$\delta^{13}C$	$\delta^1 H$	
6d-α-L-Talp	1	98.7	5.64	
1	2	71.44	4.26	
	3	74.84	4.60	
	6	17-19	1.2-1.4	
	benzylidene	104.74	5.85	
	$J_{\rm C1,H1}$	175.1	Hz	
α-L-Rhap	1	94.40	5.11	
1	2	77.5	3.68	
	3	76.82	4.27	
	6	17-19	1.2-1.4	
	$J_{\rm C1,H1}$	168.2 Hz		
α-l-Fucp	1	98.97	5.08	
1	2	79.85	3.85	
	3	79.50	3.55	
	4	78.33	3.83	
	5	67.58	3.90	
	6	18	0.95	
	2-OMe	58.89	3.12	
	$J_{\rm C1,H1}$	167.3	Hz	
α-D-Rhap	1	98.45	5.26	
	2	80.30	3.47	
	3	74.09	4.09	
	4	76.32	3.68	
	5	68.78	3.90	
	6	17-19	1.2-1.4	
	2-OMe	58.89 or 58.38 3.45 or 3.		
	$J_{\rm C1,H1}$ 17		2.1 Hz	
α-Kan <i>p</i> -4NFo	1	93.50	5.07	
	6	17-19	1.2-1.4	
	2-OMe	58.89 or 58.38	3.45 or 3.41	
	CHO	162.31	8.32	
	N-Bzl	50.03	3.84	
	$J_{\rm C1,H1}$	166.3 Hz		

Table 3 NMR data for pentasaccharide **33** in CDCl₃ (δ in ppm)

Residue	Atom no.	$\delta^1 H$	δ^{13} C
6d-α-l-Talp	1	5.62	98.1
	2	4.03	77.8
	3	4.12	71
	4	3.63	70.6
	5	3.80	69.8
	6	1.25	17.2
α-L-Rhap	1	5.03	103.44
-	2	4.10	75.1
	3,4	3.82,3.79	78.3
	5	3.63	72.1
	6	1.12	16.7
α-L-Fucp	1	5.17	99.3
1	2	3.73	77.5
	3,4,5	3.93,3.98,4.03	75,67.1,70
	6	1.21	17.4
	2-OMe	3.49,3.52,3.54	58.8-59
α-D-Rhap	1	5.39	98.7
1	2	3.67	78.2
	3	4.05	76.3
	4	3.75	72.3
	5	4.02	68.3
	6	1.20	17.5
	2-OMe	3.49,3.52,3.54	58.8-59
α-Kan <i>p</i> -4NFo	1	5.08	94.9
-	2	3.11 3.15	84.6
	4	3.24 3.28	53.5
	5	4.08	70.6
	6	1.22	15.8
	2-OMe	3.49,3.52,3.54	58.8-59
	NCHO	8.2 and 8.0	131 and 137
	$CH_{3}(3)$	1.37 and 1.31	19.1

colourless syrup; $[\alpha]_{D}$ –183.5° (*c* 0.45); IR: 1730 cm⁻¹ (C=O); ¹H NMR: δ 7.20–7.30 (m, 5 H, Ph), 4.79 (d, 1 H, $J_{1,2}$ 2 Hz, H-1), 4.01 (d, 1 H, $J_{4,5}$ 9.2 Hz, H-4), 3.88 (dd, 1 H, $J_{5,Me(6)}$ 6 Hz, H-5), 3.55 (d, 1 H, H-2), 3.28 and 3.26 (2 s, each 3 H, 2 OCH₃), 1.32 (d, 3 H, CH₃). Anal. Calcd for C₁₅H₂₀O₅: C, 64.27; H, 7.19. Found: C, 64.11; H, 7.05.

Methyl 4-O-benzyl-6-deoxy-3-C-methyl-2-Omethyl- α -L-manno-(3) and altro-pyranoside (4).— A solution of compound 2 (196 mg, 0.7 mmol) in Et₂O (2 mL) was added dropwise to a solution of Grignard reagent prepared from CH₃I (875 mg, 14.1 mmol) and Mg (343 mg, 14.1 mmol) in Et₂O (5 mL), and the mixture was stirred for 1 h at room temperature. The reaction was quenched by addition of a chilled NH₃-solution (10 mL), then the mixture was diluted with Et₂O (20 mL), extracted with water (2×10 mL), dried, and concentrated. Chromatography of the residue (8:2 hexane– EtOAc) gave pure **3** (40 mg, 19%) and **4** (28 mg, 13%), isolated as colourless syrups. Compound **3**: $[\alpha]_{\rm D} - 67.1^{\circ} (c \ 0.43)$; ¹H NMR: δ 7.40 (m, 5 H, Ph), 4.72 (d, 1 H, $J_{1,2}$ 1 Hz, H-1), 3.61 (dd, 1 H, $J_{4,5}$ 10, $J_{5,\text{Me}(6)}$ 6 Hz, H-5), 3.48 and 3.36 (2 s, each 3 H, 2 OCH₃), 3.20 (d, 1 H, H-4), 3.08 (d, 1 H, H-2), 1.38 (s, 3 H, CH₃), 1.25 (d, 3 H, CH₃); Compound **4**: $[\alpha]_{\rm D} - 56.1^{\circ} (c \ 0.3)$; ¹H NMR: δ 7.4 (m, 5 H, Ph), 4.72 (d, 1 H, $J_{1,2}$ 1.5Hz, H-1), 4.66 (s, 2 H, CH₂Ph), 3.92 (dd, 1 H, $J_{4,5}$ 9.5, $J_{5,6}$ 6Hz, H-5), 3.46 and 3.41 (2 s, each 3 H, 2 OCH₃), 3.19 (d, 1 H, H-4), 3.09 (d, 1 H, H-2), 1.15 (s, 3 H CH₃), 1.14 (d, 3 H, CH₃). Anal. Calcd for C₁₆H₂₄O₅: C, 64.84; H, 8.16. Found: C, 64.90; H, 8.05.

Methyl 6-deoxy-2,3-O-isopropylidene- α -L-lyxohexopyran-4-uloside (5).—To a solution of methyl 6-deoxy-2,3-O-isopropylidene- α -L-mannopyranoside (1.0 g, 4.6 mmol) in dry CH₂Cl₂ (50 mL) was added pyridinium chlorochromate (5.0 g, 23 mmol), and the mixture was stirred for 3.5 h at room temperature. Conventional work-up (see **2**) yielded pure **5** (0.92 g, 91%), isolated as a colourless syrup; $[\alpha]_{\rm D}$ -35.8° (*c* 0.77); IR: 1740 cm⁻¹ (C=O); ¹H NMR: δ 4.85 (s, 1 H, H-1), 4.44 (s, 2 H, H-2,3), 4.26 (d, 1 H, $J_{5,Me(6)}$ 7 Hz, H-5), 3.47 (s, 3 H, OCH₃), 1.49 and 1.37 (2 s, each 3 H, Ip CH₃), 1.41 (d, 3 H, CH₃). Anal. Calcd for C₁₀H₁₆O₆: C, 55.54; H, 7.46. Found: C, 55.03; H, 7.18.

Methyl 6-deoxy-2,3-O-isopropylidene-3-C-methyl- α -L-lyxo-hexopyran-4-uloside (6).—A solution of BuLi (15% in hexane; 2.1 mL, 4.8 mmol) was added dropwise at -40 °C under Ar to a solution of diisopropylamine (672 µL, 4.8 mmol) in dry THF (50 mL). The light-yellow mixture was stirred for 20 min at -40 °C, then cooled to -78 °C, and a solution of 5 (0.91 g, 4.2 mmol) in dry THF (5 mL) was added dropwise. After stirring for 1h at -78 °C, hexamethylphosphoric triamide (1.1 mL) and CH₃I (2.2 mL, 34 mmol) was added, and the mixture was allowed to warm up to room temperature. Then, an aqueous solution of NH₄Cl (10%, 12mL) was added, the mixture was diluted with CH_2Cl_2 (200 mL), the organic layer was washed with water, dried, and concentrated. The residue was chromatographed (8:2 hexane-EtOAc) to yield pure 6 (800 mg, 83%), isolated as a colourless syrup; $[\alpha]_{\rm D} - 65.8^{\circ}$ (c 0.38); IR: 1740 cm⁻¹ (C=O); ¹H NMR: δ 4.92 (d, 1 H, $J_{1,2}$ 1.3 Hz, H-1), 4.34 (d, 1 H, J_{5,Me(6)} 6.8 Hz, H-5), 4.07 (d, 1 H, H-2), 350 (s, 3 H, OCH₃), 1.49 and 1.37 (2 s, each 3 H, Ip CH₃), 1.43 (s, 3 H, CH₃), 1.41 (d, 3 H, CH₃). Anal. Calcd for C₁₁H₁₈O₅: C, 57.38; H, 7.88. Found: C, 56.96; H, 7.70.

Methyl 6-deoxy-2,3-O-isopropylidene-3-C-methyl- α -L-talopyranoside (7).—To the chilled solution of 6 (500 mg, 2.18 mmol) in MeOH (10 mL) was added NaBH₄ (150 mg, 3.9 mmol), and the mixture was stirred for 1h, then boiled under reflux for 20 min, and concentrated. CH₂Cl₂ (50 mL) was added, the organic layer was washed with water until neutral, dried, and concentrated to yield 7 (480 mg, 95%), isolated as a colourless syrup; $[\alpha]_{\rm D}$ -58.0° (c 0.35); ¹H NMR: δ 4.93 (s, 1 H, H-1), 3.88 (dd, 1 H, $J_{4,5}$ 1, $J_{5,Me(6)}$ 6.5 Hz, H-5), 3.75 (s, 1 H, H-2), 3.40 (s, 3 H, OCH₃), 3.17 (dd, 1 H, J_{4,OH} 5 Hz, H-4), 2.48 (d, 1 H, OH), 1.57 and 1.38 (2 s, each 3 H, Ip CH₃), 1.42 (s, 3 H, CH₃), 1.34 (d, 3 H, CH₃). Anal. Calcd for $C_{11}H_{20}O_5$: C, 56.88; H, 8.68. Found: C, 56.16; H, 8.42.

Methyl 6-deoxy-2,3-O-isopropylidene-3-C-methyl-4-O-trifluoromethanesulfonyl- α -L-talopyranoside (8).—To the solution of 7 (220 mg, 0.95 mmol) in dry CH₂Cl₂ (8 mL) containing pyridine (400 µL) was added dropwise triflic anhydride (210 µL, 1.24 mmol) at -70 °C, and the mixture was stirred for 90 min. After work-up via extractions and purification on a short column of silica (9:1 hexane–EtOAc) pure **8** was obtained as a colourless syrup (304 mg, 88%); [α]_D -17.8° (*c* 0.95); ¹H NMR: δ 4.94 (s, 1 H, H-1), 4.35 (s, 1 H, H-4), 4.03 (d, 1 H, $J_{5,Me(6)}$ 6 Hz, H-5), 3.78 (s, 1 H, H-2), 3.40 (s, 3 H, OCH₃), 1.55 and 1.40 (2 s, each 3 H, Ip CH₃), 1.50 (s, 3 H, CH₃), 1.38 (d, 3 H, CH₃). Anal. Calcd for C₁₂H₁₉F₃O₇S: C, 39.56; H, 5.26. Found: C, 39.16; H, 5.02.

Methyl 4,6-dideoxy-2,3-O-isopropylidene-3-C*methyl*-α-L-erythro-*hex-4-enopyranoside* (**9**).—To the solution of 8 (300 mg, 0.82 mmol) in dry DMF (5 mL) was added NaN₃ (150 mg, 2.46 mmol) and dicyclohexyl-18-crown-6 (10 mg, 0.03 mmol), and the mixture was stirred for 6h at room temperature. The mixture was then poured into 0.1 M HCl (10 mL), extracted with Et_2O (2×20 mL), and the combined organic layer was washed with water until neutral, dried, and concentrated. Chromatography (95:5 hexane–EtOAc) of the residue yielded **9** (150 mg, 87%), isolated as a colourless oil; $[\alpha]_{\rm D}$ -18.3° (c 0.65); ¹H NMR: δ 5.02 (d, 1 H, $J_{1,2}$ 2 Hz, H-1), 4.60 (s, 1 H, H-4), 3.81 (d, 1 H, H-2), 3.45 (s, 3 H, OCH₃), 1.80, 1.48, 1.42, and 1.43 (4 s, each 3 H, 4 CH₃). Anal. Calcd for C₁₁H₁₉O₄: C, 61.37; H, 8.90. Found: C, 60.56; H, 8.92.

Methyl 6-deoxy-2,3-O-isopropylidene-3-C-methyl- α -L-lyxo-hexopyran-4-uloside-oxime (10).—To a solution of 6 (5.8 g, 25.2 mmol) in EtOH (150 mL) was added hydroxylamine hydrochloride (8.85 g, 127.4 mmol) and Na₂CO₃ (15 g, 141.5 mmol), and the mixture was stirred for 15 h at 80 °C. Inorganic salts were filtered off, the filtrate was concentrated, and CH_2Cl_2 (300 mL) was added. The organic layer was washed with water until neutral, dried, and concentrated to yield 10 (6.2 g, 99%) as a colourless oil which was sufficiently pure for the next transformation; $[\alpha]_{\rm D} = -78.0^{\circ}$ (c 0.55); ¹H NMR: δ 4.72 (dd, 1 H, J_{5.6} 5 Hz, H-5), 4.57 (s, 1 H, H-1), 3.92 (s, 1 H, H-2), 3.87 (s, 3 H, OCH₃), 3.38 (s, 3 H, OCH₃), 1.55 (s, 3 H, CH₃), 1.53 and 1.45 (2 s, each 3 H, Ip CH₃), 1.44 (d, 3 H, CH₃). Anal. Calcd for C₁₁H₁₉NO₅: C, 53.86; H, 7.81. Found: C, 54.16; H, 7.92.

Methyl 4-amino-4,6-dideoxy-2,3-O-isopropylidene-3-C-methyl- α -L-mannopyranoside (12).—A solution of 10 (6.0 g, 24.5 mmol) and LiAlH₄ (8.83 g, 233 mmol) in 1,4-dioxane (400 mL) was stirred for 4 h at 100 °C. The mixture was chilled, and the excess of hydride was decomposed by addition of EtOAc (200 mL) and water. Then the organic layer was washed with water until neutral, dried, and concentrated. Chromatography (9:1 hexane–EtOAc) of the residue yielded **12** (2.27 g, 41%), isolated as a syrup. The corresponding L-*talo* isomer was isolated in 38% yield; $[\alpha]_{\rm D}$ -40.1° (*C* 0.99); ¹H NMR: δ 4.87 (d, 1 H, $J_{1,2}$ 1 Hz, H-1), 3.79 (d, 1 H, H-2), 3.51 (dd, 1 H, $J_{4,5}$ 10, $J_{5,\text{Me(6)}}$ 6.5 Hz, H-5), 3.38 (s, 3 H, OCH₃), 2.84 (d, 1 H, H-4), 1.51 and 1.37 (2 s, each 3 H, Ip CH₃), 1.31 (s, 3 H, CH₃). Anal. Calcd for C₁₁H₂₁NO₄: C, 57.12; H, 9.15; N, 6.05. Found: C 57.10; H, 9.00; N, 5.95.

Methyl 4,6-dideoxy-4-formamido-2,3-O-isopropy*lidene-3-C-methyl-α-L-mannopyranoside* (13).—To a solution of 12 (880 mg, 3.8 mmol) in MeOH (20 mL) was added acetic-formic anhydride (3 mL, 35.6 mmol) and the mixture was stirred for 30 min, then concentrated. Chromatography (8:2 hexaneacetone) of the residue gave 13 (980 mg, 99%), isolated as a syrup; $[\alpha]_{\rm D}$ -76.0° (c 0.72); ¹H NMR: δ 8.18 and 7.98 (2 d, 1 H, CHO), 7.22 and 6.70 (2 m, 1 H, NH), 4.64 and 4.54 (2 d, 1 H, $J_{1,2}$ 1 Hz, H-1), 3.05 and 3.04 (2 d, 1 H, J_{4,5} 10 Hz, H-4), 3.75 and 3.65 (2 dd, 1 H, J_{5,Me(6)} 6 Hz, H-5), 3.43 (d, 1 H, H-2), 3.30 and 3.28 (2 s, 3 H, OCH₃), 1.49 and 1.40 (2 s, each 3 H, Ip CH₃), 1.23 and 1.22 (2 s, 3 H, CH₃), 1.19 and 1.17 (2 d, 3 H, CH₃). Anal. Calcd for C₁₂H₂₁NO₅: C, 55.58; H, 8.16. Found: C, 55.04; H, 8.01.

Methyl 4-N-benzylformamido-4,6-dideoxy-2,3-O-isopropylidene-3-C-methyl- α -L-mannopyranoside (14).—A solution of 13 (980 mg, 1.77 mmol), benzyl bromide (1 mL, 3.9 mmol) and powdered KOH (2g) in dry DMF (20mL) was stirred for 1.5h at room temperature. Then the mixture was diluted with EtOAc (200 mL), washed with water until neutral, dried, and concentrated. Chromatography (8:2 hexane–EtOAc) of the residue yielded 14 (1.0 g, 75%), isolated as a syrup; $[\alpha]_{\rm D} - 2.9^{\circ} (c \ 0.6)$; ¹H NMR: δ 8.2 and 8.05 (2 d, 1 H, CHO), 7.3–7.2 (m, 5 H, Ph), 4.64 and 4.59 (2 s, 1 H, H-1), 4.08 (dd, J_{4,5} 10, J_{5,6} 6 Hz, H-5), 3.58 and 3.51 (2 s, 1 H, H-2), 332 (s, 3 H, OCH₃), 2.91 (d, 1 H, H-4), 1.41 and 1.40 (2 s, 3 H, CH₃), 0.85 (d, 3 H, CH₃). Anal. Calcd for C₁₉H₂₇NO₅: C, 65.31; H, 7.79. Found: C, 64.90; H, 7.80.

Methyl 4-N-benzylformamido-4,6-dideoxy-3-Cmethyl- α -L-mannopyranoside (15).—Compound 14 (280 mg, 0.8 mmol) was dissolved in aq 70% HOAc (20 mL) and the solution was kept for 4 h at 80 °C, then concentrated. The residue was chromatographed (9:1 CH₂Cl₂–MeOH) to yield syrupy **15** (208 mg, 84%); $[\alpha]_{\rm D}$ –11.41° (*c* 0.60); ¹H NMR: δ 8.35 and 8.05 (2 s, 1 H, CHO), 7.2–7.3 (m, 5 H, Ph), 5.40 (bs, 1 H, OH), 4.64 and 4.62 (2 s, 1 H, H-1), 4.0 (dd, 1 H, $J_{4,5}$ 10, $J_{5,Me(6)}$ 6Hz, H-5), 3.57 and 3.52 (2 s, 1 H, H-2), 3.30 (s, 3 H, OCH₃), 2.90 (d, 1 H, H-4), 1.40 and 1.39 (2 s, 3 H, CH₃), 1.25 (s, 1 H, OH), 0.80 (d, 3 H, CH₃). Anal. Calcd for C₁₆H₂₃NO₅: C, 62.12; H, 7.49. Found: C, 62.16; H, 7.52.

Methyl 4-N-benzylformamido-4,6-dideoxy-3-Cmethyl-2-O-methyl- α -L-mannopyranoside (16).—To a solution of 15 (200 mg, 0.65 mmol) in dry DMF (10 mL) was added NaH (29 mg, 0.98 mmol), and the mixture was stirred at room temperature for 40 min, then chilled, and CH_3I (52 µL, 0.85 mmol) was injected. After 15 min, MeOH (0.5 mL) was added, the mixture was concentrated, and the residue was chromatographed (7:3 hexane-acetone) to yield 16 (169 mg, 80%), isolated as a syrup; $[\alpha]_{\rm D}$ -32.01° (c 0.87); ¹H NMR: δ 8.40 and 8.15 (2 s, 1 H, CHO), 7.20–7.40 (m, 5 H, Ph), 4.50 (d, 1 H, J_{1 2} 1 Hz, H-1), 4.00 (dd, 1 H, J_{4,5} 10, J_{5,Me(6)} 6 Hz, H-5), 3.50 and 3.48 (2 s, 3 H, OCH₃), 3.37 and 3.34 (2 s, 3 H, OCH₃), 3.10 and 3.03 (2 d, 1 H, H-2), 2.58 (bs, 1 H, OH), 1.41 and 1.32 (2 s, 3 H, CH₃), 0.77 (d, 3 H, CH₃). Anal. Calcd for $C_{17}H_{27}NO_5$: C, 63.14; H, 7.79. Found: C, 63.06; H, 7.90.

Methyl 3-O-benzyl-4-N-benzylformamido-4,6dideoxy-3-C-methyl-2-O-methyl-a-L-mannopyranoside (17).—To a solution of 16 (1.56 g, 4.8 mmol) in dry DMF (70 mL) was added NaH (300 mg, 9.6 mmol), and the mixture was stirred for 2h at room temperature. Benzyl bromide (1.5 mL, 6.2 mmol) was added and the mixture was stirred for 3h. After processing (see 16) pure 17 (1.0g, 50%) was isolated as a colourless glass; $[\alpha]_{\rm D} - 48.0^{\circ}$ (*c* 0.35); ¹H NMR: δ 8.32 and 8.15 (2 s, 1 H, CHO), 7.20 and 7.43 (2 m, 10 H, 2 Ph), 4.91 and 4.05 (2 dd, 1 H, J_{4,5} 11, J_{5,Me(6)} 6 Hz, H-5), 4.72 and 4.70 (2 d, 1 H, J_{1,2} 1 Hz, H-1), 4.50 and 4.42 (2 d, each 1 H, PhCH₂), 4.49 (s, 2 H, PhCH₂), 3.47 and 3.45 (2 s, 3 H, OCH₃), 3.41 (d, 1 H, H-2), 3.37 and 3.50 (2 s, 3 H, OCH₃), 3.21 (d, 1 H, H-4), 1.60 and 1.45 (2 s, 3 H, CH₃), 0.78 (d, 3 H, CH₃). Anal. Calcd for C₂₄H₃₁NO₅: C, 69.71; H, 7.56. Found: C, 70.06; H, 7.82.

1-O-Acetyl-3-O-benzyl-4-N-benzylformamido-4,6-dideoxy-3-C-methyl-2-O-methyl-α-L-mannopyranose (18).—To a solution of 17 (300 mg, 0.72 mmol) in Ac₂O (2 mL) was added H₂SO₄ in Ac₂O (4%, 2 mL), and the mixture was stirred for 30 min at room temperature. The solution was poured into ice-cold aq NaHCO₃, extracted with CH₂Cl₂ (20 mL), and the organic layer was separated, dried, and concentrated. Chromatography (97:3 CH₂Cl₂-EtOAc) of the residue yielded syrupy **18** (221 mg, 70%); $[\alpha]_D - 28.0^\circ$ (*c* 0.55); ¹H NMR: δ 8.32 and 8.13 (2 s, 1 H, CHO), 7.20–7.50 (m, 10 H, Ph), 6.12 and 6.20 (2 d, 1 H, $J_{1,2}$ 1 Hz, H-1), 5.08 and 4.20 (2 dd, 1 H, $J_{4,5}$ 10, $J_{5,Me(6)}$ 6 Hz, H-5), 4.50 (d, 1 H, H-2), 4.50-4.43 (m, 4 H, PhCH₂), 3.52 and 3.51 (2 s, 3 H, OCH₃), 3.32 (d, 1 H, H-4), 2.10 and 2.12 (2 s, 3 H, Ac), 1.52 and 1.60 (2 s, 3 H, CH₃), 0.75 (d, 3 H, CH₃). Anal. Calcd for C₂₅H₃₁NO₆: C 68.01; H, 7.08. Found: C, 67.76; H, 7.02.

3-O-Benzyl-4-N-benzylformamido-4,6-dideoxy-3-C-methyl-2-O-methyl-a-L-mannopyranosyl trichloroacetimidate (20).—To a solution of 18 (100 mg, 0.23 mmol) in dry DMF (5 mL) was added hydrazine acetate (30 mg, 0.3 mmol), and the solution was kept for 30 min at 50 °C. Then, EtOAc (20 mL) was added, and the organic layer was washed with water, dried, and concentrated to give **19** (80 mg, 87%) which was used without purification ($[\alpha]_{\rm D}$ $+33.2^{\circ}$ (c 0.29)). A solution of the foregoing material (80 mg, 0.2 mmol), trichloroacetonitrile (190 µL, 1.9 mmol) and DBU (50 mL, 0.34 mmol) in dry CH_2Cl_2 (5 mL) was stirred for 10 min at room temperature, then concentrated. The residue was chromatographed (7:3 hexane-acetone +1% Et₃N) to yield **20** (100 mg, 91%; 6:4, E:Z) as a syrup; $[\alpha]_{\rm D} = -21.3^{\circ} (c \ 0.42)$; ¹H NMR: $\delta 8.68$ and 8.56 (2 s, 1 H, NH), 8.34 and 8.13 (2 s, 1 H, CHO), 7.20-7.50 (m, 10 H, Ph), 6.35 and 6.29 (2 d, 1 H, J_{1,2} 2 Hz, H-1), 5.17 and 4.29 (2 dd, 1 H, $J_{4,5}$ 10, $J_{5,Me(6)}$ 6 Hz, H-5), 4.50 (bs, 4 H, PhC H_2), 3.72 and 3.65 (2 bs, 1 H, H-2), 3.56 and 3.54 (2 s, 3 H, OCH₃), 3.30 (d, 1 H, H-4), 1.60 (s, 3 H, CH₃), 0.79 (d, 3 H, CH₃). Anal. Calcd for C₂₅H₂₉C₁₃N₂O₅: C, 55.21; H, 5.38. Found: C, 55.16; H, 5.12.

p-*Nitrophenyl* 2,3-O-*isopropylidene*-6-O-p-*tol-uenesulfonyl*- α -D-*mannopyranoside* (21).—*p*-Toluenesulfonyl chloride (290 mg, 1.5 mmol) was added to a solution of *p*-nitrophenyl 2,3-O-isopropylidene- α -D-mannopyranoside (100 mg, 0.3 mmol) in dry pyridine (8 mL), and the solution was kept for 1.5 h at 60 °C. After concentration, chromato-graphy (7:3 hexane–acetone) of the residue yielded **21** (96 mg, 70%), isolated as a syrup; $[\alpha]_{\rm D}$ + 52.6° (*c* 0.63); ¹H NMR: δ 8.15 and 7.10 (2 m, each 2 H, *p*-subst. Ph), 7.70 and 7.32 (2 m, each 2 H, *p*-Ts),

5.78 (s, 1 H, H-1), 4.38 (d, 1 H, $J_{2,3}$ 5 Hz, H-2), 4.3 (m, 3 H, H-3,4,5), 3.50 (m, 2 H, H-6a,6b), 2.90 (s, 1 H, OH), 2.45 (s, 3 H, *p*-Ts CH₃), 1.55 and 1.42 (2 s, each 3 H, Ip CH₃); ¹³C NMR: δ 115.66 (acetalic C), 94.71 (C-1), 67.63 (C-6), 25.1 and 26.82 (2 Ip CH₃), 20.32 (*p*-Ts CH₃). Anal. Calcd for C₂₂H₂₅NO₁₀S: C, 57.01; H, 5.44. Found: C, 56.96; H, 5.12.

p-Nitrophenyl 6-deoxy-6-iodo-2,3-O-isopropy*lidene-* α -D-*mannopyranoside* (22).—A solution of 21 (580 mg, 1.2 mmol) and NaI (4 g, 24 mmol) in dry DMF (10 mL) was kept for 5 h at 100 °C, then cooled, diluted with CH_2Cl_2 (200 mL), extracted with water, and concentrated. Chromatography (7:3 hexane-EtOAc) of the residue afforded 22 (380 mg, 73%), isolated as a syrup; $[\alpha]_{\rm D}$ + 85.3° (c 0.36); ¹H NMR: δ 8.22 and 7.20 (2 m, each 2 H, *p*subst. Ph), 5.90 (d, 1 H, J_{1 2} 1 Hz, H- 1), 4.42 (dd, 1 H, J_{2,3} 6 Hz, H-2), 4.32 (dd, 1 H, J_{3,4} 7 Hz, H-3), 3.50 (ni, 2 H, H-5,6a), 3.31 (dd, 1 H, J_{5,6a} 7, J_{6a,6b} 11 Hz, H-6b), 3.12 (ddd, 1 H, J_{4,5} 7, J_{4,OH} 2 Hz, H-4), 2.05 (s, 1 H, OH), 1.60 and 1.42 (2 s, each 3 H, Ip CH₃); ¹³C NMR: δ 116.82 (acetalic C), 96.17 (C-1), 26.2 and 27.95 (2 Ip CH₃), 5.11 (C-6). Anal. Calcd for C₁₅H₁₈INO₇: C, 39.93; H, 4.02. Found: C, 40.16; H, 3.92.

p-Nitrophenyl 6-deoxy-2,3-O-isopropylidene- α -D-mannopyranoside (23).—To a solution of 22 (380 mg, 0.84 mmol) in dry THF (15 mL) was added LiAlH₄ (96 mg, 2.5 mmol), and the mixture was stirred for 20h at room temperature. The excess of hydride was decomposed by addition of EtOAc (100 mL) and water, and the organic layer was washed with water until neutral, dried, concentrated. Chromatography (7:3 CH₂Cl₂-acetone) of the residue yielded 23 (144 mg, 53%), isolated as a syrup; $[\alpha]_{\rm D}$ + 115.4° (*c* 0.24); ¹H NMR: δ 8.20 and 7.28 (2 m, each 2 H, p-subst. Ph), 5.82 (d, 1 H, J_{1,2} 1 Hz, H-1), 4.45 (dd, 1 H, J_{2,3} 6, J_{3,4} 6.5 Hz, H-3), 4.39 (dd, 1 H, H-2), 3.75 (dd, 1 H, J_{4.5} 9.5, J_{5.Me(6)} 6 Hz, H-5), 3.40 (dd, 1 H, H-4), 1.55 and 1.40 (2 s, each 3 H, Ip CH₃), 1.21 (d, 1 H, CH₃). Anal. Calcd for C₁₅H₁₉NO₇: C, 55.38; H, 5.89. Found: C, 55.16; H, 5.92.

p-Nitrophenyl 4-O-benzyl-6-deoxy-2,3-O-isopropylidene- α -D-mannopyranoside (24).—To a chilled solution of 23 (140 mg, 0.43 mmol) and benzyl bromide (67 μ L, 0.56 mmol) in DMF (10 mL) was added KOH (2.5 g), and the mixture was stirred for 40 min at 0 °C. EtOAc (100 mL) was added, and the organic layer was washed with water until neutral, dried, and concentrated. Chromatography (95:5 hexane–EtOAc) of the residue yielded **24** (133 mg, 75%), isolated as a syrup; $[\alpha]_{\rm D}$ + 121° (*c* 0.17); ¹H NMR: δ 8.19 and 7.13 (2 m, each 2 H, *p*-subst. Ph), 7.32 (m, 5 H, Ph), 5.79 (d, 1 H, $J_{1,2}$ 1 Hz, H-1), 4.92 and 4.66 (2 d, each 1 H, PhCH₂), 4.45 (t, 1 H, $J_{2,3}$ 6, $J_{3,4}$ 6 Hz, H-3), 4.38 (dd, 1 H, H-2), 3.72 (dd, 1 H, $J_{4,5}$ 9.5, $J_{5,Me(6)}$ 6 Hz, H-5), 3.31 (dd, 1 H, H-4), 1.56 and 1.42 (2 s, each 3 H, Ip CH₃), 1.22 (d, 1 H, CH₃). Anal. Calcd for C₂₂H₂₅NO₇: C, 63.60; H, 6.07. Found: C, 63.46; H, 6.22.

p-Nitrophenyl 4-O-benzyl-6-deoxy- α -D-mannopyranoside (25).—To a solution of 24 (500 mg, 1.2 mmol) in CH₂Cl₂ (10 mL) were added CF₃CO₂H (1 mL) and water (0.5 mL), and the mixture was stirred vigorously for 10 min at room temperature. After concentration, flash chromatography (7:3 hexane–EtOAc) of the residue gave 25 (405 mg, 90%), isolated as a syrup; [α]_D + 105.2° (*c* 0.58); ¹H NMR: δ 8.20 and 7.28 (2 m, each 2 H, *p*subst. Ph), 7.32 (m, 5 H, Ph), 5.90 (d, 1 H, *J*_{1,2} 1 Hz, H-1), 4.91 and 4.60 (2 d, each 1 H, CH₂Ph), 5.2 and 3.2 (2 s, each 1 H, OH). Anal. Calcd for C₁₉H₂₁NO₇: C, 60.79; H, 5.64. Found: C, 61.0; H, 5.88.

p-Nitrophenyl 4-O-benzyl-6-deoxy-2-O-methyl- α -D-mannopyranoside (26).—A mixture of 25 (200 mg, 0.53 mmol), Bu₄NBr (300 mg), and CH₃I (3 mL, 9.3 mmol) in CH₂Cl₂ (10 mL) and aq 20% NaOH (6mL) was stirred vigorously overnight. The organic layer was separated, washed with water, concentrated, and the residue was chromatographed (7:3 hexane-EtOAc) to yield 26 (186 mg, 90%) as a syrup; $[\alpha]_{\rm D}$ +118.0° (*c* 0.15); ¹H NMR: δ 8.20 and 7.30 (2 m, each 2 H, p-subst. Ph), 7.32 (m, 5 H, Ph), 5.85 (d, 1 H, J_{1.2} 2 Hz, H-1), 5.21 (d, 1 H, J_{OH,3} 7 Hz, OH), 4.88 and 4.58 (2 d, each 1 H, PhC*H*₂), 3.95 (m, 1 H, *J*_{2,3} 3.5, *J*_{3,4} 9 Hz, H-3), 3.58 (dd, 1 H, H-2), 3.47 (dd, 1 H, J_{5.6} 6, J_{4.5} 9.5 Hz, H-5), 3.46 (s, 3 H, OCH₃), 3.30 (t, 1 H, H-4), 1.1 (d, 3 H, CH₃). Anal. Calcd for C₂₀H₂₃NO₇: C, 61.69; H, 5.95. Found: C, 61.50; H, 5.92.

p-Nitrophenyl (3-O-benzyl-4-N-benzylformamido-4,6-dideoxy-3-C-methyl-2-O-methyl- α -L-mannopyranosyl)-(1 \rightarrow 3)-4-O-benzyl-6-deoxy-2-O-methyl- α -D-mannopyranoside (27).—A mixture of 26 (29 mg, 0.074 mmol) and 20 (50 mg, 0.09 mmol) in dry CH₂Cl₂ (5 mL), containing 4 Å molecular sieves (100 mg) was stirred for 30 min at 0 °C under Ar, then cooled to -30 °C. A solution of TMSOTf (17 µL, 0.3 equiv.), in dry CH₂Cl₂ (1 mL) was injected and after 15 min the reaction was quenched with pyridine (0.5 mL). Then the mixture was diluted with CH_2Cl_2 (20 mL), filtered, and concentrated. Chromatography (7:3 hexane–EtOAc) of the residue yielded **27** (20 mg, 35%), isolated as a syrup; $[\alpha]_D$ + 52.9° (*c* 1.03). ¹H and ¹³C NMR data (500 and 125 MHz, respectively) are presented in Table 1. "a" and "b" denote the two isomers present due to the formamido group. Anal. Calcd for $C_{43}H_{50}N_2O_{11}$: C, 66.99; H, 6.54; N, 3.64. Found: C, 66.85, H, 6.48, N, 3.65.

p-Trifluoroacetamidophenyl (4-formamido-4,6dideoxy-3-C-methyl-2-O-methyl-a-L-mannopyranosyl)- $(1 \rightarrow 3)$ -6-deoxy-2-O-methyl- α -D-mannopyranoside (29).—Compound 27 (15 mg, 0.03 mmol) in EtOAc (10 mL) was treated with H_2 in the presence of Adam's catalyst (PtO_2 , 10 mg) for 2 h at room temperature. Pyridine (200 µL) and trifluoroacetic anhydride $(150 \,\mu\text{L})$ were added, and the mixture was stirred for an additional 1 h. After filtration, the filtrate was concentrated and the residue was chromatographed (7:3 hexane-EtOAc) to yield 28 (11 mg). A mixture of 28 (11 mg) and $Pd(OH)_2$ (40 mg) in MeOH (5 mL) was stirred vigorously under H₂ overnight. After filtration and concentration, the residue was purified on a short column of silica gel (98:2 CH_2Cl_2 –MeOH) to yield **29** (4 mg, 24%) as a glass; $[\alpha]_{D} - 4.52^{\circ}$ (*c* 0.24; MeOH). ¹H and ¹³C NMR data (recorded in CD₃OD at 500 and 125 MHz, respectively) are presented in Table 2.

Ethyl (3-O-benzyl-4-N-benzylformamido-4,6dideoxy-3-C-methyl-2-O-methyl-a-L-mannopyranosyl)- $(1 \rightarrow 3)$ -4-O-benzyl-6-deoxy-2-O-methyl-1-thio- α -D-mannopyranoside (31).—A mixture of ethyl 4-O-benzyl-6-deoxy-2-O-methyl-1-thio-α-D-mannopyranoside (**30**) (23 mg, 0.074 mmol), **20** (50 mg, 0.09 mmol) and 4 A molecular sieves (100 mg) in dry CH_2Cl_2 (3 mL) was stirred for 30 min at 0 °C under Ar. The mixture was cooled to -30 °C, and TMSOTf (17μ L, 0.3 equiv.) in CH₂Cl₂ (1 mL) was injected. After 15 min, pyridine (500 mL) was added, and the mixture was diluted with CH₂Cl₂, filtered, and the filtrate was concentrated. Chromatography (7:3 hexane-EtOAc) of the residue gave **31** (38 mg, 75%), isolated as a syrup; $[\alpha]_{\rm D}$ $+56.8^{\circ}$ (c 0.37). ¹H and ¹³C NMR data (500 and 125 MHz, respectively) are presented in Table 1. "a" and "b" denote the two isomers present due to the formamido group. Anal. Calcd for $C_{39}H_{51}$ NO₈S: C, 67.50; H, 7.41. Found: C, 67.42; H, 7.23.

p-Nitrophenyl (3-O-benzyl-4-N-benzylformamido-4,6-dideoxy-3-C-methyl-2-O-methyl- α -L-mannopyranosyl)-(1 \rightarrow 3)-(4-O-benzyl-6-deoxy-2-O-methyl- α - D-mannopyranosyl)- $(1 \rightarrow 3)$ -(4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)- $(1 \rightarrow 3)$ -(2,4-di-O-benzyl-6-deoxy- α -L-mannopyranosyl)- $(1 \rightarrow 2)$ -3,4-O-endo-benzylideneα-L-talopyranoside (33).—A mixture of 31 (30 mg, 0.046 mmol), **32** [12] (48 mg, 0.074 mmol) and 4 A molecular sieves (100 mg) in dry CH_2Cl_2 (5 mL) was stirred for 30 min at 0 °C, then cooled to -40 °C. A solution of NIS (14 mg, 0.055 mmol) and TfOH (0.6 mL, 0.005 mmol) in CH₂Cl₂ (1 mL) was added dropwise and the mixture was allowed to warm up to 0 °C. After 40 min the mixture was diluted with CH_2Cl_2 (20 mL), extracted with a Na₂S₂O₃ (10 mL) and aq NaHCO₃ (10 mL), and the organic layer was dried and concentrated. The residue was chromatographed (8:2 hexane–EtOAc) to yield 33 (40 mg, 55%), isolated as a syrup; $[\alpha]_{\rm D} -23.2^{\circ}$ (c 0.19); ¹H and ¹³C NMR data (500 and 125 MHz, respectively) are given in Table 3. Anal. Calcd for $C_{90}H_{104}$ N₂O₂₃: C, 68.34; H, 6.63. Found: C, 68.12; H, 6.58. p-Trifluoroacetamidophenyl (4-formamido-4,6dideoxy-3-C-methyl-2-O-methyl-a-L-mannopyranosyl)- $(1 \rightarrow 3)$ -(6-deoxy-2-O-methyl- α -D-mannopyranosyl)- $(1 \rightarrow 3)$ -(2-O-methyl- α -L-fucopyranosyl)- $(1 \rightarrow 3)$ - $(6\text{-}deoxy-\alpha-L\text{-}mannopyranosyl)-(1\rightarrow 2)-\alpha-L\text{-}talopyr$ anoside (35).—A solution of 33 (35 mg, 0.02 mmol) in EtOAc (10 mL) was stirred vigorously under H_2 in the presence of Adam's catalyst (PtO₂, 10 mg) for 2 h at room temperature. Pyridine (200 µL) and trifluoroacetic anhydride $(150 \,\mu\text{L})$ were added, and the mixture was stirred for an additional 1 h. After filtration and concentration of the filtrate, resulting 34 (16 mg) was dissolved in MeOH (10 mL), and $Pd(OH)_2$ (50 mg) was added. The mixture was stirred vigorously under H₂ overnight, then filtrated followed by concentration of the filtrate. The resi-

due was purified on a column of silica (98:2 CH₂Cl₂–MeOH) to yield pure **35** (5 mg, 25%), isolated as a syrup; $[\alpha]_{\rm D} - 17.9^{\circ}$ (*c* 0.21; MeOH). ¹H and ¹³C NMR data (500 and 125 MHz, respectively) are given in Table 4.

Acknowledgements

This work was supported in part by an International Research Scholar's award from the Howard Hughes Medical Institute. Support from the Hungarian Science Fund (OTKA F 019482) to I. Bajza, (OTKA T 014982 and OTKA D 23749) to K.E. Kövér, from the Ministry of Education (FKFP 0500/1997) to K.E. Kövér, and from the Hungarian Academy of Sciences (AKP96-121) to A. Lipták is also acknowledged. The purchase of the DRX 500 NMR spectrometer used in this study was supported by OMFB Mec-93-0098, Phare-Accord H-9112-0198 and OTKA A084.

References

- [1] P.J. Brennan, Rev. Infect. Dis., 11 (1989) S420-430.
- [2] G.O. Aspinall, D. Chatterjee, and P.J. Brennan, Adv. Carbohydr. Chem. Biochem., 51 (1995) 169–242.
- [3] M. McNeil, H. Gaylord, and P.J. Brennan, Carbohydr. Res., 177 (1988) 185–198.
- [4] G.O. Aspinall, N.K. Khare, R.K. Sood, D. Chatterjee, B. Rivoire, and P.J. Brennan, *Carbohydr. Res.*, 216 (1991) 357–373.
- [5] S.W. Hunter, T. Fujiwara, R.C. Murphy, and P.J. Brennan, J. Biol. Chem., 259 (1984) 9729–9734.
- [6] J. Yoshimura, K.I. Sato, and R.B. Singh, Chem. Lett., (1985) 69–70.
- [7] J. Yoshimura, A. Aqeel, K.I. Sato, R.B. Singh, and H. Hashimoto, *Carbohydr. Res.*, 166 (1987) 252– 256.
- [8] R.M. Giuliano and S. Kasperowicz, *Carbohydr. Res.*, 183 (1988) 277–285.
- [9] I. Bajza and A. Lipták, *Carbohydr. Res.*, 205 (1990) 435–439.
- [10] G.O. Aspinall and K. Takeo, *Carbohydr. Res.*, 121 (1983) 61–77.
- [11] M.K. Gurjar and G. Viswanadham, *Tetrahedron Lett.*, 32 (1991) 6191–6194.
- [12] J. Kerékgyártó, Z. Szurmai, and A. Lipták, Carbohydr. Res., 245 (1993) 65–80.
- [13] I. Bajza, J. Kerékgyártó, J. Hajkó, L. Szilágyi, and A. Lipták, *Carbohydr. Res.*, 253 (1994) 111–120.
- [14] H.M. Zuurmond, G.H. Veeneman, G.A. van der Marel, and J.H. van Boom, *Carbohydr. Res.*, 241 (1993) 153–164.
- [15] B. Flaherty, W.G. Overend, and N.R. Williams, J. Chem. Soc. C, (1966) 398–403.
- [16] D.R. Hicks and B. Fraser-Reid, Can. J. Chem., 53 (1975) 2017–2023.
- [17] V. Pozsgay, Carbohydr. Res., 69 (1979) 284-286.
- [18] R.F. Butterworth and S. Hanessian, Synthesis, (1971) 70–88.
- [19] D. Neuhaus and M. Williamson, *The Nuclear Overhauser Effect in Structural and Conformational Analysis*, VCH Publishers, Cambridge (1988).
- [20] A. Klemer and H. Beerman, J. Carbohydr. Chem., 2 (1983) 457–459.
- [21] DR. Bundle, M. Gerken, and T. Peters, *Carbohydr. Res.*, 174 (1988) 239–251.
- [22] G.W.J. Fleet, M.J. Gough, and P.W. Smith, *Tet-rahedron Lett.*, 25 (1984) 1853–1856.
- [23] K. Miyai and R.W. Jeanloz, Carbohydr. Res., 21 (1972) 45–55.

- [24] R.R. Schmidt, Angew. Chem. Int. Ed. Engl., 25 (1986) 212–235.
- [25] S.S. Rana, J.J. Barlow, and K.L. Matta, Carbohydr. Res., 96 (1981) 79–85.
- [26] P.J. Garegg, T. Iversen, and S. Oscarson, *Carbohydr. Res.*, 50 (1976) C12–C14.
- [27] A. Borbás and A. Lipták, Carbohydr. Res., 241 (1993) 99–116.
- [28] G.H. Veeneman, S.H. van Leeuwen, and J.H. van Boom, *Tetrahedron Lett.*, 31 (1990) 275–278.
- [29] R.R. Ernst, G. Bodenhausen, and A. Wokaum, Principles of Nuclear Magnetic Resonance in One and Two Dimensions, Oxford University Press, Oxford (1986).
- [30] R. Freeman, *Handbook of Nuclear Magnetic Spectroscopy*, Longmans, Harlow, UK (1988).